The androgen-induced protein AIbZIP facilitates proliferation of prostate cancer cells through downregulation of p21 expression

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Supplementary Information

Supplementary Figure S1.



Supplementary Figure S1. WB analysis for AIbZIP in LNCaP cells treated with tunicamycin LNCaP cells were treated with 3 μ g/ml tunicamycin (Tm). Asterisk: nonspecific bands. β -actin was used as a loading control.

Supplementary Figure S2.

Supplementary Figure S2. The GGA(A/T) core sequences within the -1 kb upstream of *AIbZIP* Sequences colored by green indicate the GGA(A/T) core sequences.

Supplementary Figure S3.



Supplementary Figure S3. The analysis for the expressions of *AIbZIP* in LNCaP cells transfected with siRNAs targeting *AIbZIP*

RT-PCR analysis for *AIbZIP* in LNCaP cells transfected with siRNAs targeting *AIbZIP* or scramble. Each siRNA (#1, #2, #3) for *AIbZIP* decreased the expression levels of *AIbZIP* to 27%, 57%, 27% respectively. A mixture of these siRNAs reduced the mRNA level of *AIbZIP* up to 78%. The numbers represent fold changes of mRNA levels. *GAPDH* was used as an internal control.

Supplementary Figure S4.



Supplementary Figure S4. Proliferation assay for LNCaP tet-off cells stably expressing FLAGtagged AIbZIP

(a) WB analysis for FLAG-tagged AIbZIP in the LNCaP tet-off cells treated with doxycycline or not. (b) LNCaP tet-off cells were cultured with or without doxycycline, and the numbers of cells were counted at day 1, 3 and 5 (means \pm s.d., n = 3; ***P < 0.001, vs. Doxycycline+).

Supplementary Figure S5.



Supplementary Figure S5. RT-PCR analysis for OASIS in indicated cancer cell lines.

Supplementary Figure S6.



Supplementary Figure S6. Schematic representation for the mechanism of OASIS activation

In response to various stimuli, OASIS is translocated from the ER to the Golgi apparatus, and sequentially cleaved by site-1 protease (S1P) and site-2 protease (S2P). Cleaved N-terminal OASIS containing transcription activation and basic leucine zipper (bZIP) domains translocate to the nucleus to promote the transcription of target genes including p21.

Supplementary Figure S7.



Supplementary Figure S7. Quantification for protein levels of full-length, S1P-, and S2P-cleaved OASIS in Figure 7(e). (means \pm s.d., n = 3).

Supplementary Figure S8.



Figure 1(g) β-actin

Supplementary Figure S8. Full-length gels/blots of Figure 1(b), (c), (d), (f) and (g)

Supplementary Figure S9.



Supplementary Figure S9. Full-length gels of Figure 2(a), (c), (e), (g) and (i)

Supplementary Figure S10.



Figure 3(b) WB: anti-FLAG



Figure 3(d) P1

Figure 3(d) P2



Figure 3(d) P3

Figure 3(d) P4

Supplementary Figure S10. Full-length gels/blots of Figure 3(b) and (d)

Supplementary Figure S11.



Figure 4(e) β-actin

Supplementary Figure S11. Full-length gels/blots of Figure 4(d) and (e)

Supplementary Figure S12.



Figure 5(a) AlbZIP







Figure 5(c) AlbZIP



Figure 5(f) AlbZIP

Figure 5(f) *p21*



Figure 5(f) GAPDH



Supplementary Figure S12. Full-length gels/blots of Figure 5(a), (c), (f) and (h)

Supplementary Figure S13.









Figure 6(a) *p21*



Figure 6(c) p21



Figure 6(c) β-actin



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Figure 6(e) AlbZIP

Figure 6(e) OASIS



Figure 6(e) Myc-AlbZIP

Figure 6(e) FLAG-OASIS

Figure 6(e) β-actin

Supplementary Figure S14.





Figure 7(e) β-actin

Supplementary Figure S14. Full-length blots of Figure 7(a) and (e)

Supplementary Table 1.

List of primers used for plasmids construction				
Gene	e Primer forward primer			
FLAG-AlbZIP	forward	5'-GGTACCATGGACTACAAGGATGACGATGACAAGGATCTCGGAATCCCTGAC-3'		
	reverse	5'-TCACATCTCATCTGCATGCAG-3'		
Myc-AlbZIP	forward	5'-ATGGAACAGAAACTGATCTCTGAAGAAGACATGACAAGGATCTCGGAATCCCTGAC-3'		
	reverse	5'-TCACATCTCATCTGCATGCAG-3'		
Myc-AlbZIP∆bZIP	forward	5'-TCCTCCAGACGCTAATTGCTCAAAC-3'		
	reverse	5'-GTCTGGAGGACCCTCTCCTCTGCC-3'		
FLAG-SPDEF	forward	5'-CCACCATGGACTACAAGGATGACGATGACAAGATGGGCAGCGCCAG-3'		
	reverse	5'-TCAGATGGGGTGCACGAACTGGTAG -3'		
	forward	5'-GCGGCCGCTCGAGTCTAGAGGGCCC-3'		
FLAG-SFDEFDETS	reverse	5'-TCACTGCCCGGAGCATGATGAGTCCACC-3'		
	forward	5'-GATGAAAGAGCGGACTTCACC-3'		
FLAG-SFDEFASAM	reverse	5'-GCCCACCACCATGGACTGCAC-3'		
	forward	5'-CAAGTGACTTATTCAGGGAGACTGC-3'		
AIDZIF-5K	reverse	5'-CGGAAAGAGTCAAGAGACCTG-3'		
AlbZID Ek to 1 7k	forward	5'-CAAGTGACTTATTCAGGGAGACTGC-3'		
AIDZIP-5K to -1.7K	reverse	5'-CCTTCCGACGAAACAGAGAAGTGATG-3'		
	forward	5'-CTGGGCAACAACAGCGAAAC-3'		
AIDZIP-3k to -1.7k	reverse	5'-CCTTCCGACGAAACAGAGAAGTGATG-3'		
AlbZIP-1.7k	forward	5'-GGAAGGCTGCTTTGTCTCTCACTAC-3'		
	reverse	5'-CGGAAAGAGTCAAGAGACCTG-3'		
	forward	5'-CCTGGACCATCAGACATTAGAC-3'		
AIDZIP-1.0K	reverse	5'-CGGAAAGAGTCAAGAGACCTG-3'		

Supplementary Table 2.

List of antibodies					
antibody name	species	type	Manufacturer		
anti-β-actin	mouse	monoclonal	A2228, Sigma-Aldrich		
anti-AlbZIP	mouse	monoclonal	H00148327-AP51, Abnova		
anti-FLAG M2	mouse	monoclonal	F3165, Sigma-Aldrich		
anti-FLAG	rabbit	polyclonal	600-401-383, ROCKLAND		
anti-p21	mouse	monoclonal	SC-6246, Santa Cruz Biotechnology		
anti-p53	mouse	monoclonal	#9282, Cell Signaling Technology		
anti-phospho-p53 (Ser15)	rabbit	polyclonal	#9284, Cell Signaling Technology		
anti-Myc	mouse	monoclonal	M192-3, MBL		
anti-Myc	rabbit	polyclonal	562, MBL		
anti-calnexin	mouse	monoclonal	ADI-SPA-860, Enzo		
anti-GM130	rabbit	polyclonal	D6B1, Cell Signaling Technology		
anti-Histone H3	mouse	monoclonal	SC-10809, Santa Cruz Biotechnology		
anti-mouse IgG	mouse	monoclonal	G3A1, Cell Signaling Technology		
anti-rabbit IgG	rabbit	polyclonal	Cell Signaling Technology		

Supplementary Table 3.

List of primers used for RT-PCR				
Gene	forward primer	revers primer		
AlbZIP	5'-GGTTCCGGTAACTAGGCT-3'	5'-AGACGCTTCTCCTCATCG-3'		
PSA	5'-CCAGACACTCACAGCAAGGA-3'	5'-CTGAGGGTTGTCTGGAGGAC-3'		
BiP	5'-GTTTGCTGAGGAAGACAAAAAGCTC-3'	5'-CACTTCCATAGAGTTTGCTGATAAT-3'		
XBP1	5'-CAGCAGGTGCAGGCCCAGTTGTC-3'	5'-GACACTAATCAGCTGGGGAAAGAC-3'		
AR	5'-TCCAAATCACCCCCCAGGAA-3'	5'-GACATCTGAAAGGGGGCATG-3'		
SPDEF	5'-GTCAGCGGCCTGGATGAAAGA-3'	5'-AAGATGCCCTTCTCCTTGTTG-3'		
p21	5'-GGAAGACCATGTGGACCTGT-3'	5'-GGCGTTTGGAGTGGTAGAAA-3'		
p53	5'-AGTCACAGCACATGACGGAGG-3'	5'-TGGAGTCTTCCAGTGTGATGATG-3'		
p27	5'-CAGCTTGCCCGAGTTCTACTACAG-3'	5'-AGGTCGCTTCCTCATCCCTG-3'		
CCNE1	5'-CCAGGAAGAGGAAGGCAAACG-3'	5'-GTGTTGCTCAAGAAAGTGCTG-3'		
CCNA2	5'-GCATGTCACCGTTCCTCCTTG-3'	5'-GTGATGTCTGGCTGTTTCTTC-3'		
CDK2	5'-GGGCCTAGCTTTCTGCCATTC-3'	5'-GGAAACTTGGCTTGTAATCAGG-3'		
CDK4	5'-CTTCCCATCAGCACAGTTCG-3'	5'-GGTGTAAGTGCCATCTGGTAG-3'		
OASIS	5'-GAACATGGAGGACTTCTCCAATG-3'	5'-CGGGCTCTGCTCCTGCTTCAC-3'		
GAPDH	5'-AGGTGAAGGTCGGAGTCAAC-3'	5'-GACGGTGCCATGGAATTTGC -3'		

Supplementary Table 4.

List of siRNA sequences					
Gene	sense	anti-sense			
scramble	5'-UUCUCCGAACGUGUCACGU-3'	5'-ACGUGACACGUUCGGAGAA-3'			
AlbZIP#1	5'-GAACCAAGAAUUACAGAAA-3'	5'-UUUCUGUAAUUCUUGGUUC-3'			
AIbZIP#2	5'-CAGAAAAUCUGGAGACCCA-3'	5'-UGGGUCUCCAGAUUUUCUG-3'			
AIbZIP#3	5'-GGAAAUAAGUUUUGAGUGA-3'	5'-UCACUCAAAACUUAUUCC-3'			
AR	5'-AAGAAGGCCAGUUGUAUGGAC-3'	5'-GUCCAUACAACUGGCCUUCUU-3'			
SPDEF	5'-CCUUCUACCUCUCCUACUU-3'	5'-AAGUAGGAGAGGUAGAAGG-3'			

Supplementary Table 5.

List of primers used for ChIP assay				
primer number	forward primer	revers primer		
1	5'-CTGAGTTTTGGTATTACTACTTTC-3'	5'-CATACAGTTACCGGGAACAT-3'		
2	5'-TGCTGTTCTCCATTAACGCCACAC-3'	5'-AGTCAACAGCCCGCTGACCC-3'		
3	5'-TGGGGTCAGCGGGCTGTTGAC-3'	5'-GTCAGGCGGGGATTGGGTTC-3'		
4	5'-ACCCAATCCCCGCCTGACAAATAAG-3'	5'-TGGCTGTTCAGGAGAGTCAGGCCAA-3'		