

Supplementary Fig.1 Measurement of iGluSnFR signal fluorescence in plaque
 microenvironment and manipulation of glutamate signaling pathway

6 Intracortical injection of AAV.hSynapsin.iGluSnFR in somatosensory cortex area (APPPS1 n=7-7 9, 3-7 imaged brain areas/mouse). A) Fluorescence of iGluSnFR to calculate relative glutamate 8 levels was measured in ROI1 and ROI4 by averaging baseline frames 1-25 over all APPPS1 9 animals. ROI1 shows a significantly higher level of fluorescence in comparison to ROI4 pointing 10 towards higher glutamate levels within this area. B-E) Open brain experiments after intracortical injection of AAV.hSynapsin.iGluSnFR in somatosensory cortex area (APPPS1 n=2, 2 imaged 11 brain areas/mouse) were used to manipulate the glutamate pathway. B) Cadmium chloride ( $Cd^{2+}$ , 12 13 1mM) was applied topically to the brain (dura mater was left intact) to block all presynaptic 14 transmitter release and thus release of glutamate. The detected levels of glutamate fluorescence changed upon treatment. Levels in ROI1 were significantly reduced however stayed elevated in 15 comparison to ROI4 levels. C) Treatment with TBOA was used to block glutamate uptake. In 16 this experiment fluorescent levels of ROI1 and ROI4 were significantly increased, with ROI1 17 displaying permanently higher levels throughout the baseline, treatment and wash-out phase. 18 19 Both treatments could be reversed after a wash-out phase of 10 min. D-E) Confirmation of the 20 change in glutamate signaling can be seen in the average traces of all recorded animals. Upon application of  $Cd^{2+}$  the sensory evoked response is abolished and returns after a 10 min wash-out 21 phase (panel D). Upon treatment with TBOA the hindlimb response is longer and wider resulting 22 in a longer rate of decay which also reversed to its baseline level after a 10 min wash- out phase. 23 \*\*\*P<0.001, \*\*P<0.01, \*P<0.05. 24





Supplementary Fig.2 Detection of altered glutamate dynamics in Aβ plaque
 microenvironment using tplsm linescan imaging
 30

A) Intracortical injection of AAV.hSvnapsin.iGluSnFR in somatosensory cortex area (APPPS1 31 32 mice n=9, 3-4 imaged brain areas/mouse). Glutamate fluctuations were imaged using linescan 33 tplsm (16x128 Pixels, 57Hz). Relative overall fluctuation of glutamate is shown in false colour RMS map. Methoxy X04 stained amyloid deposit is coloured in purple. APPPS1 animals show 34 35 altered glutamatergic activity near plaque. Scalebar 20um B) Traces representing glutamate 36 dynamics differ significantly in relation to AB plaque distance. High fluctuations were measured in ROI1 in which no stimulus locked response was detected. ROI3 displayed a stimulus-evoked 37 response with significantly altered characteristics in comparison to ROI4. C) The highest average 38 39 RMS was measured in the direct vicinity of A $\beta$  plaques with its average RMS of  $\Delta$ F/F being 40 significantly different in comparison to all other ROIs. D) The maximal response to the stimulation was detected in the ROI imaged farthest away from the AB plaque (ROI4). E) A 41 42 significant difference in the area under the response between ROI3 and ROI4 was detected. F) 43 The decay rates of glutamate calculated for ROIs3 and 4 are significantly different. Error bars 44 indicate SEM. \*\*\*P<0.001, \*\*P<0.01, \*P<0.05.

- 45
- 46
- 47
- 48



## 50 Supplementary Fig.3 Reduction of possible background noise in ROI1

A) To exclude the possibility that a stimulus evoked response of glutamate is hidden in the high fluctuation in ROI1, the hindlimb stimulation was repeated using 100 trials. RMS of average RMS of  $\Delta F/F$  was still significantly higher in ROI1 in comparison to all other ROIS and the mean of all measured glutamate traces did still not result in a stimulus locked response in ROI1 and 2 B). \*\*\*P<0.001, \*\*P<0.05.



# Supplementary Fig.4 Non-uniform response to hindlimb stimulation in transgene negative animals

72

A) Three individual examples of transgene negative animals responding to a hindlimb stimulus.
The responding areas are shown in false colour displaying the relative maximum amplitudes at

75 the timepoint of stimulation. B) The randomly placed ROIs show an average maximum

amplitude that is not significantly different when comparing the ROIs. C) The taus obtained from

77 the randomly placed ROIs show no significant difference when comparing the different traces, as

- can also be seen in the individually plotted average traces in D. Error bars indicate SEM.
- 79
- 80
- 81
- 82
- 83
- 84
- 85



## Supplementary Fig. 5 Ca<sup>2+</sup> imaging in relation to plaque distance

88

89 Intracortical injection of AAV.hSynapsin.iGluSnFR and AAV1.Syn.Flex.NESjRGECO1a.WPRE.SV40 in somatosensory cortex area (APPPS1 mice n=3, 3-4 imaged brain 90 91 areas/mouse). Glutamate and Calcium fluctuations were imaged using tplsm. A) Merge of brain region with jRGECO expressing neurons (white) and methoxy X04 stained amyloid deposit 92 (green). Scalebar 20µm. B) Traces of single calcium recordings (circled in vellow) in different 93 regions of interest in relation to the amyloid deposit shown in green. C) Mean of recorded  $Ca^{2+}$ 94 traces from all responding cells in the different regions of interest. D) Mean of recorded 95 Glutamate traces from the different regions of interest. No stimulus evoked response was 96 detected in ROI1 and 2. E) Even though Ca<sup>2+</sup> recordings showed stimulus evoked responses in 97 all ROIs, the overall percentage of responding cells was lower in the immediate plaque 98 99 microenvironment.

100

101

102

103

104

105

106



### 

Alexa 594 was puffed into the brain adjacent to an amyloid plaque to test whether the rate of diffusion and or rate of decay is changed in ROI1 in comparison to ROI4. A) Alexa 594 is shown in red and amyloid plaques are stained with methoxy\_X04 in green. Time point 1 (TP1) shows the start of the diffusion. B) The Alexa 594 is reaching the amyloid plaque in TP2. C) The decay of the dye around the plaque is shown in TP3. The following taus were calculated for the decay rate of Alexa 594 in ROI1 tau=11.290+/-0.2 sec and ROI4 tau=11.30+/-0.2 sec. Thus, no difference in the rate of decay was detected in this experiment.

Supplementary Fig.6 Diffusion of Alexa594 around amyloid plaques



141 Supplementary Fig.7 GFAP staining around amyloid deposits after Ceftriaxone and vehicle 142 treatment

- 144 Immunohistological stainings of astrocytes using GFAP in red and amyloid deposits in green are
- shown in 3 different examples after treatment with either Ceftriaxone or vehicle.
- 146 We observed a similar expression pattern when comparing GFAP after Ceftriaxone and vehicle
- 147 treatment.