

## Supplementary Information

### Importance of the C-terminal histidine residues of *Helicobacter pylori* GroES for Toll-like receptor 4 binding and interleukin-8 cytokine production

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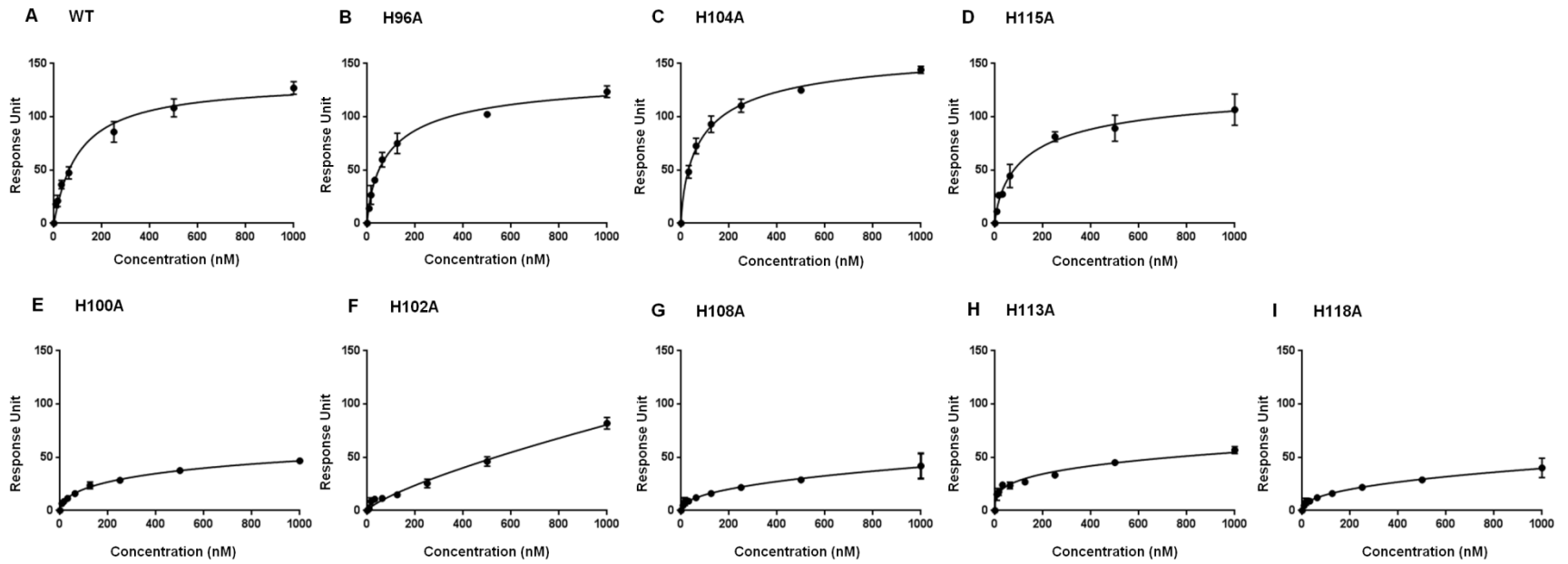
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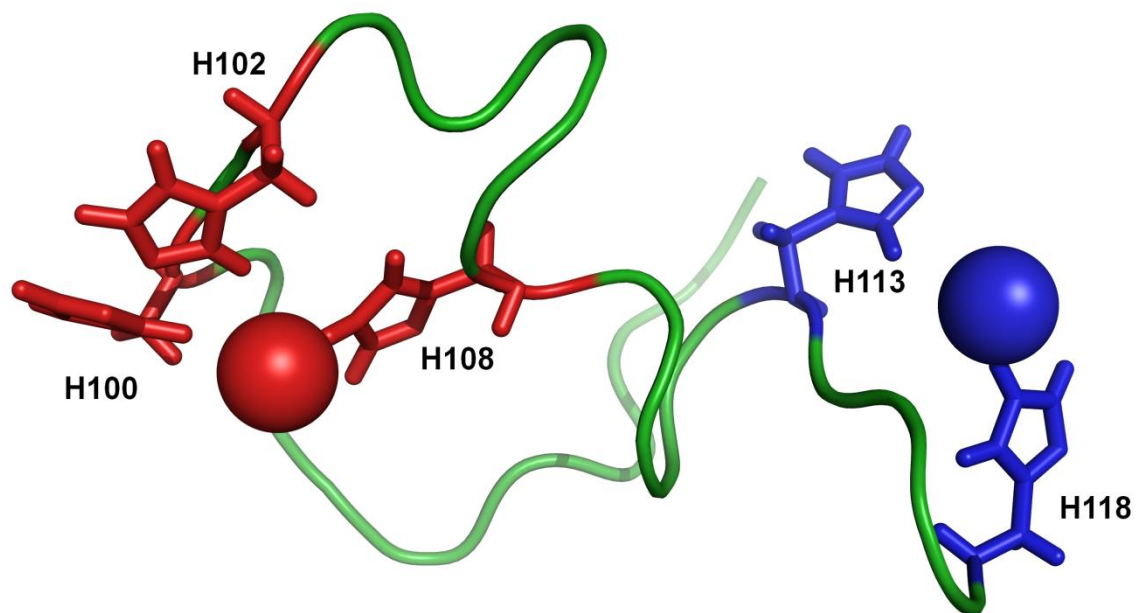
## **Supplementary Method**

### ***Homology modeling of domain B***

We used the domain B sequence (SGSCCHTGNHDKHAKHEACCHDHKKH) of *HpGroES* as our target. The predicting model of domain B from I-TASSER<sup>1</sup> and subsequently applied molecular docking simulation method for the complex structure of the two nickel ions/domain B. Next, autodock via docking software<sup>2</sup> was used to dock the two nickel ions into the domain B structure. The best model was then selected for subsequent molecular dynamics (MD) simulations. Initially, the best model was inserted into the tip3p water box. The MD simulations were performed in the canonical ensemble with a simulation temperature of 310 K, unless stated otherwise, by using the Verlet integrator with an integration time step of 0.002 ps and SHAKE constraints<sup>3</sup> of all covalent bonds involving hydrogen atoms. In the electrostatic interactions, atom-based truncation was performed using the PME method<sup>4</sup>, and the switch van der Waals function was used with a 2.00 nm cut-off for atom-pair lists. The complex structure was minimized for 10000 conjugate gradient steps, and was then subjected to a 100 ns isothermal, constant volume MD simulation. The MD calculations were carried out with the Amber14 program with Amber FF99SB and additional nickel-bound force fields<sup>5</sup>. Moreover, the final structures from these simulations were used in the solvated interaction energies (SIE) and alanine scanning calculations.

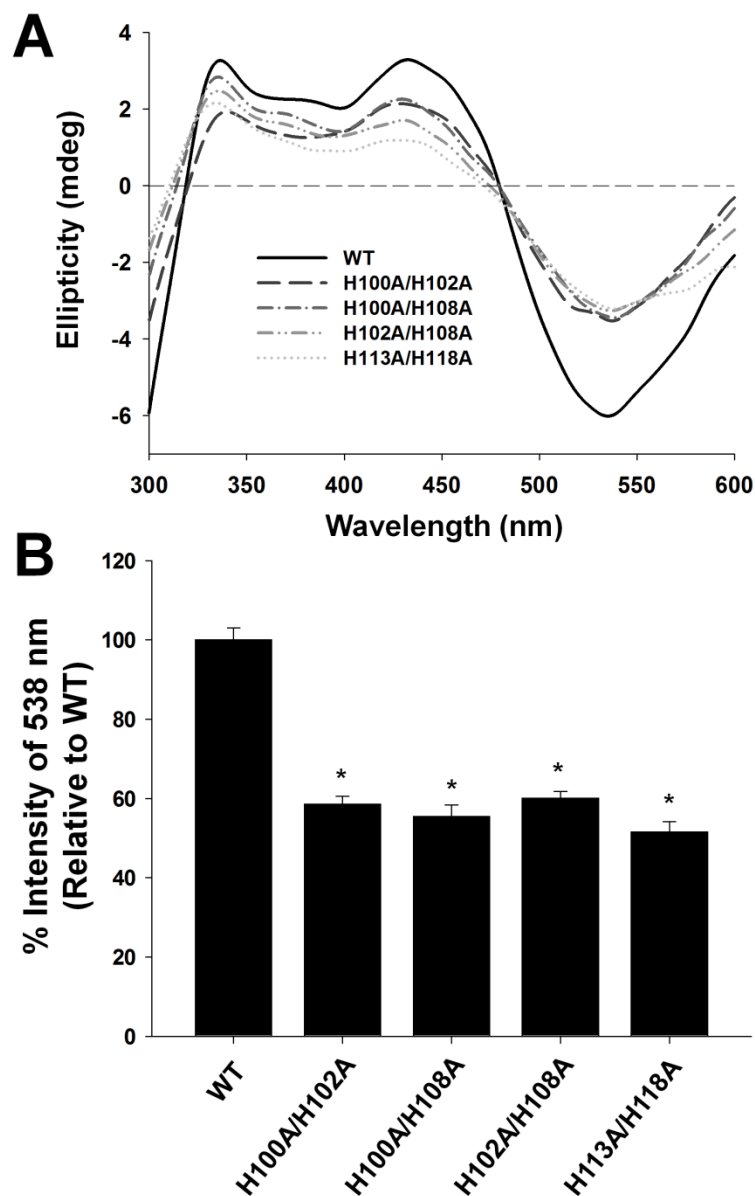


**Supplementary Figure S1.** The steady state binding affinity analysis of WT and histidine mutants to immobilized TLR4 by surface plasmon resonance (SPR) assay



**Supplementary Figure S2.** Model of the domain B with putative nickel binding residues

The configuration calculated for nickel binding to various histidine residues. The two nickel ions are colored in red and blue.



**Supplementary Figure S3.** Visible Circular dichroism (CD) spectra of Ni<sup>2+</sup> bound to WT and various histidine double mutants.

(A) Visible CD spectra of Ni<sup>2+</sup> bound to 20 μM WT and histidine double mutants. The differences in ellipticity values were approximated at 440 and 538 nm, indicating different amounts of Ni<sup>2+</sup> binding to the proteins. Data are representative of three independent experiments. (B) Intensities of the negative band at 538 nm of WT and histidine double mutants. Data were normalized and compared to WT (whereby intensity of WT was defined as 100%).

**Supplementary Table SI.** Primer sequences used for *HpGroES* constructs

<b>Construct</b>	<b>Primer Sequence</b>
<i>HpGroES</i> WT	F: 5-GGATCCATGAAGTTTCAGCCATTAGGAGA-3 R: 5-GGTACCTTAGTGTTTTTTGTGATCATGACA-3
<i>HpGroES</i> H96A	F: 5-GGCTCTTGTTGT <u>GCT</u> ACGGATAGTCAT-3 R: 5-TGAGCCCACAATACCTAGAATGTCTTCT-3
<i>HpGroES</i> H100A	F: 5-TCATACGGATAGT <u>GCT</u> GACCATAAACATG-3 R: 5-CAACAAGAGCCTGAGCCCACAATAC-3
<i>HpGroES</i> H102A	F: 5-GATAGTCATGAC <u>GCT</u> AAACATGCTAAAG-3 R: 5-CGTATGACAACAAGAGCCTGAGC-3
<i>HpGroES</i> H104A	F: 5-CATGACCATAAAG <u>GCT</u> GCTAAAGAGCAT-3 R: 5-ACTATCCGTATGACAACAAGAGCCTG-3
<i>HpGroES</i> H108A	F: 5-ATGCTAAAGAG <u>GCT</u> GAAAGCTTGCT-3 R: 5-GTTTATGGTCATGACTATCCGTATGACAAC-3
<i>HpGroES</i> H113A	F: 5-AGCTTGCTGT <u>GCT</u> GATCACAAAAA-3 R: 5-TCATGCTCTTTAGCATGTTTATGGTCAT-3
<i>HpGroES</i> H115A	F: 5-GCTGTCATGAT <u>GCC</u> AAAAAACACTA-3 R: 5-AAGCTTCATGCTCTTTAGCATGTTTA-3
<i>HpGroES</i> H118A	F: 5-GATCACAAAAA <u>AGCCT</u> AAGGTACCCC-3 R: 5-ATGACAGCAAGCTTCATGCTCTTTAG-3
<i>HpGroES</i> H100A/H102A	F: 5-GATAGTGCTGAC <u>GCT</u> AAACATGCTAAAG-3 R: 5-ATGTTT <u>AGCGT</u> CAGCACTATCCGTATG-3
<i>HpGroES</i> H100A/H108A	F: 5-ATGCTAAAGAG <u>GCT</u> GAAAGCTTGCT-3 R: 5-AGCTTCAGCCTCTTT <u>AGCAT</u> GTTTATGG-3
<i>HpGroES</i> H102A/H108A	F: 5-ATGCTAAAGAG <u>GCT</u> GAAAGCTTGCT-3 R: 5-AGCTTCAGCCTCTTT <u>AGCAT</u> GTTTATAGC-3
<i>HpGroES</i> H113A/H118A	F: 5-GATCACAAAAA <u>AGCCT</u> AAGGTACCCC-3 R: 5-ACCTTAG <u>GCT</u> TTTTTTGTGATCAGCAC-3

Mutation sites are underlined

F, forward; R, reverse.

**Supplementary Table SII.** The percentage of secondary structure content of WT and histidine mutants calculated by the CDSSTR program

<b>Sample</b>	<b>Secondary structure %</b>					
	<b><math>\alpha</math> Helix</b>	<b>3/10 Helix</b>	<b><math>\beta</math> Sheet</b>	<b><math>\beta</math> Turns</b>	<b>poly(Pro)II</b>	<b>Unordered</b>
<b>WT</b>	36.3	14.6	1.4	12.6	18.3	16.6
<b>H100A</b>	34.4	14.7	1.2	13.1	18.8	16.8
<b>H102A</b>	31.7	12.4	4.2	11.4	16.9	22.6
<b>H108A</b>	34.7	14.0	2.5	9.3	18.0	22.2
<b>H113A</b>	31.3	14.8	2.0	12.2	19.3	21.1
<b>H118A</b>	23.2	14.2	7.5	10.5	19.2	25.9

## Reference

- 1 Zhang, Y. I-TASSER server for protein 3D structure prediction. *BMC Bioinformatics* **9**, 40 (2008).
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- 3 Ryckaert, J.-P., Ciccotti, G. & Berendsen, H. J. C. Numerical integration of the cartesian equations of motion of a system with constraints: molecular dynamics of n-alkanes. *Journal of Computational Physics* **23**, 327-341, doi:[http://dx.doi.org/10.1016/0021-9991\(77\)90098-5](http://dx.doi.org/10.1016/0021-9991(77)90098-5) (1977).
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