

Supplementary Information

Discovery of MRSA active antibiotics using primary sequence from the human microbiome

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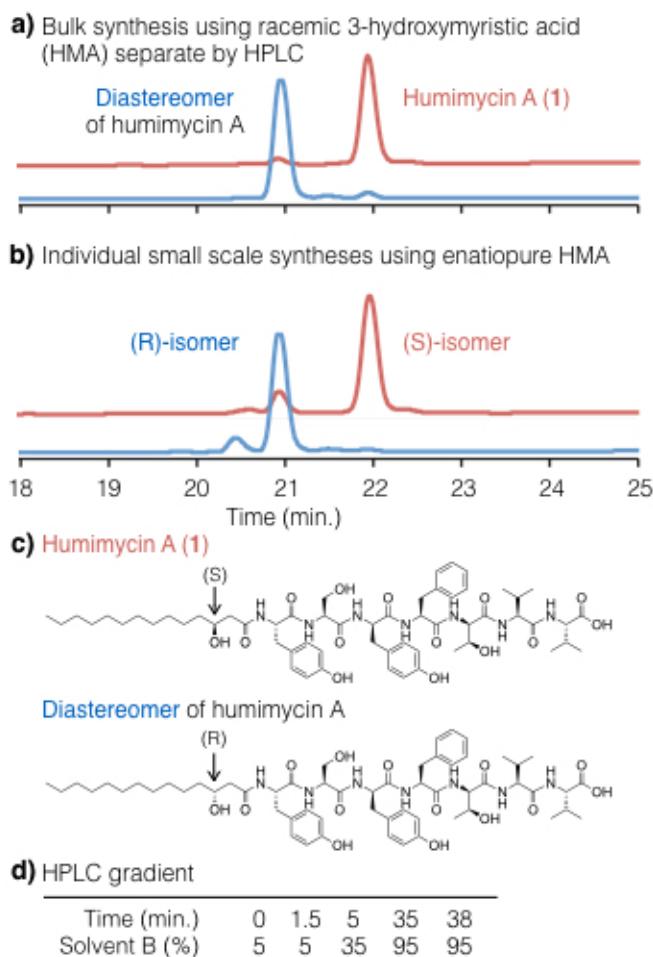
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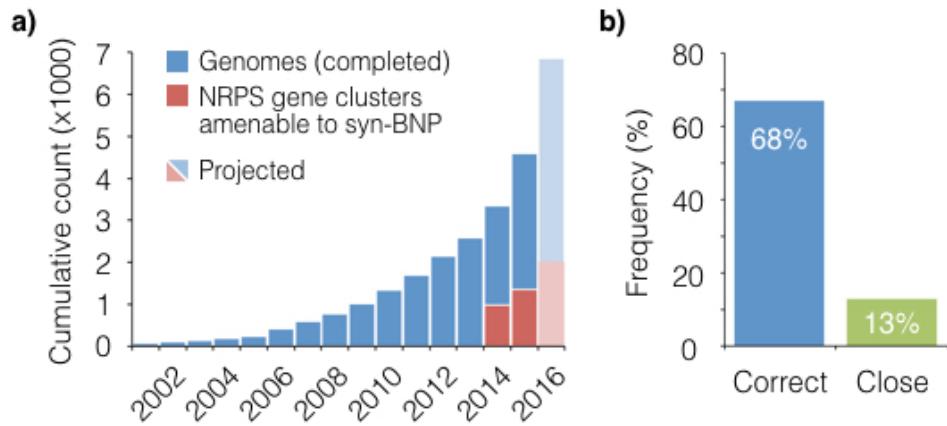
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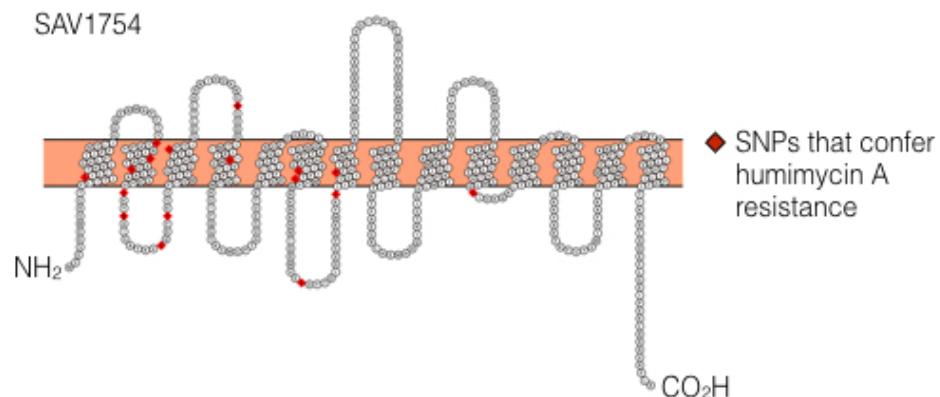
Supplementary Figure 1. Stereochemistry assignment of humimycin A. **a)** Humimycin A used in this study was synthesized in bulk using a racemic mixture of 3-hydroxymyristic acid (HMA), and the resulting pair of diastereomers were separated by HPLC. **b)** The absolute stereochemistry of humimycin A was assigned by comparison to small scale re-syntheses of humimycins using enantiopure (R)- and (S)-HMA. Humimycin A (**1**) refers to the peptide *N*-acylated with (S)-HMA and shows more potent antibiotic activity (**Supplementary Table 3**). **c)** Structures of humimycin A and its diastereomer. **d)** Solvent gradient for HPLC runs. Characterization and purification were carried out on an Agilent instrument at 4 mL/min using an XBridge BEH C18 column (130 Å, 5 um, 4.6 x 150 mm, Waters Corporation). Solvent A and B are water and acetonitrile with 0.1% TFA, respectively.



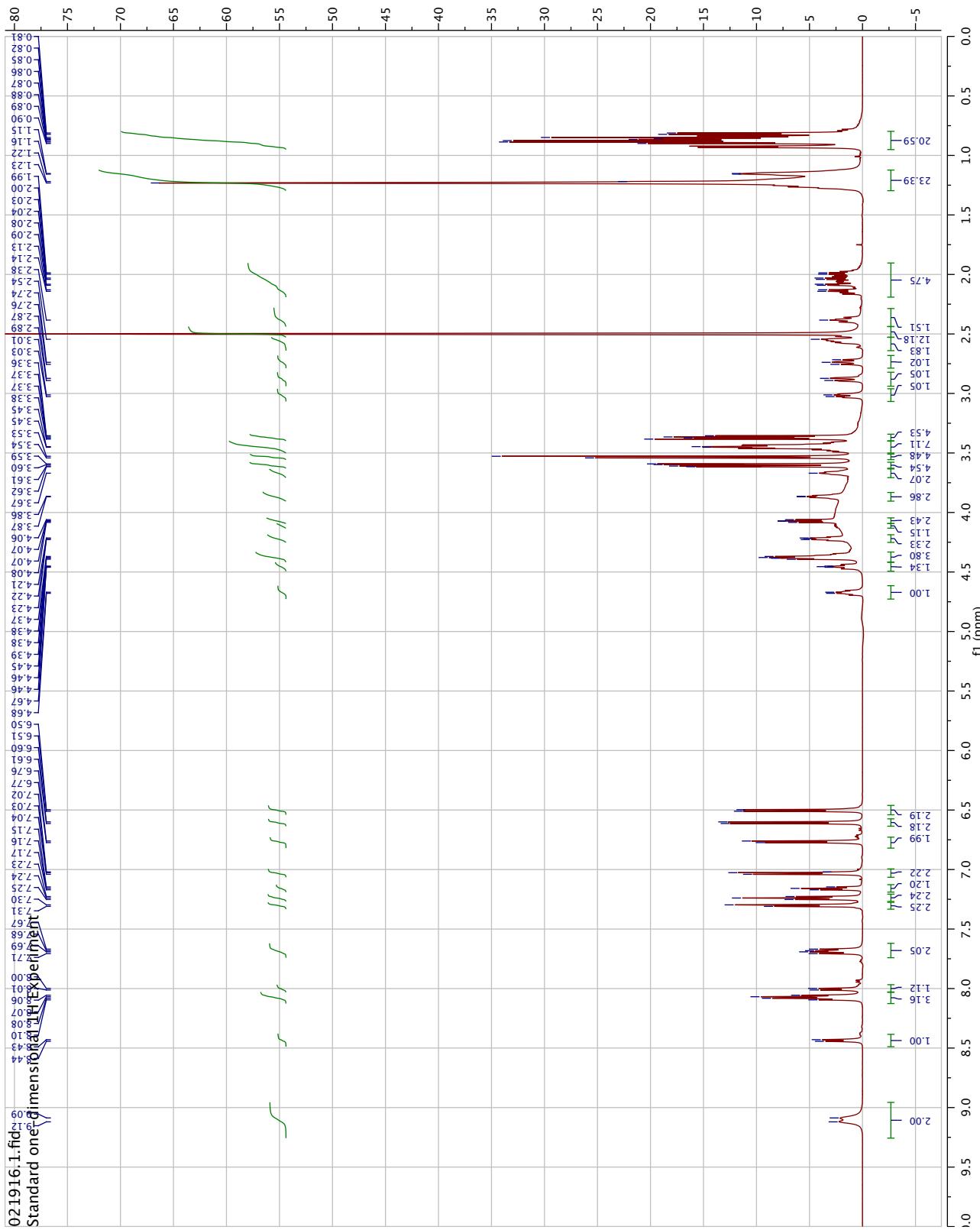
Supplementary Figure 2. Scope of the syn-BNP approach. **a)** Cumulative counts of completed bacterial genomes in GenBank (blue bars) and NRPS gene clusters amenable to the syn-BNP approach (red bars). **b)** Our analysis of Minimum Information about a Biosynthetic Gene Cluster (MIBiG, <http://mibig.secondarymetabolites.org>) indicates that the existing bioinformatic tools correctly predicted the core peptide encoded by the vast majority of well studied NRPS gene clusters. Overall 303 adenylation domains from 42 NRPS gene clusters (containing ≥ 3 modules, full information available) were analyzed. “Correct” predictions include those identical to the natural product as well as the following nearly exact matches: Val/Leu/Ile, Dab/Orn/Lys, Asp/Glu, Asn/Gln. Predictions were deemed “close” in case of “correct” predictions that have additional modifications, e.g., β -hydroxylation, β -methylation, etc.



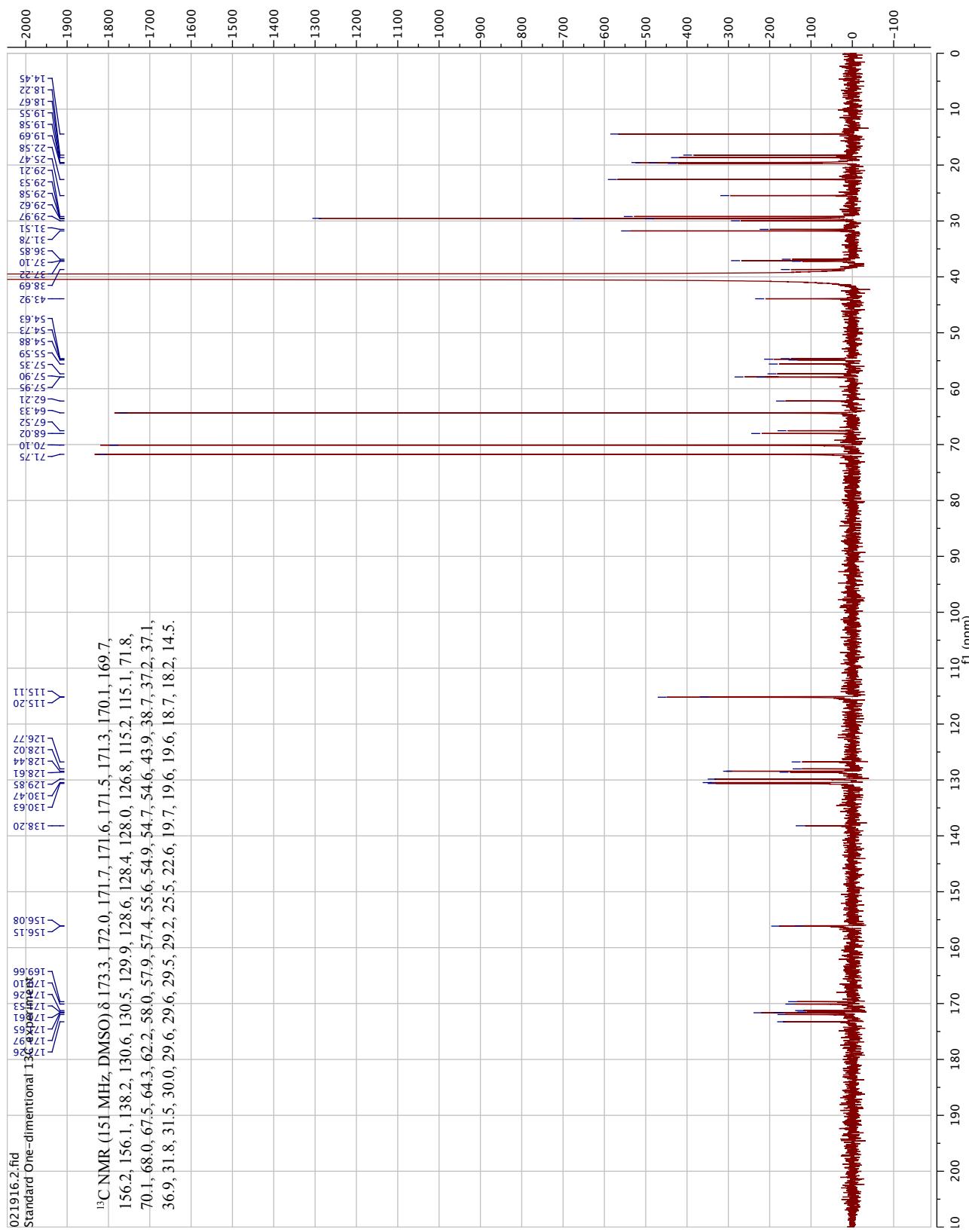
Supplementary Figure 3. SNPs in SAV1754 that confer humimycin A resistance. We selected *S. aureus* USA300 mutants that could survive on 2.5 times the MIC (20 µg/mL) and sequenced the genome of 23 resistant mutants. Upon comparison to the parent strain, we found that all 23 mutants contained one non-synonymous mutation in SAV1754. Sixteen of the 23 are unique mutations and displayed below.



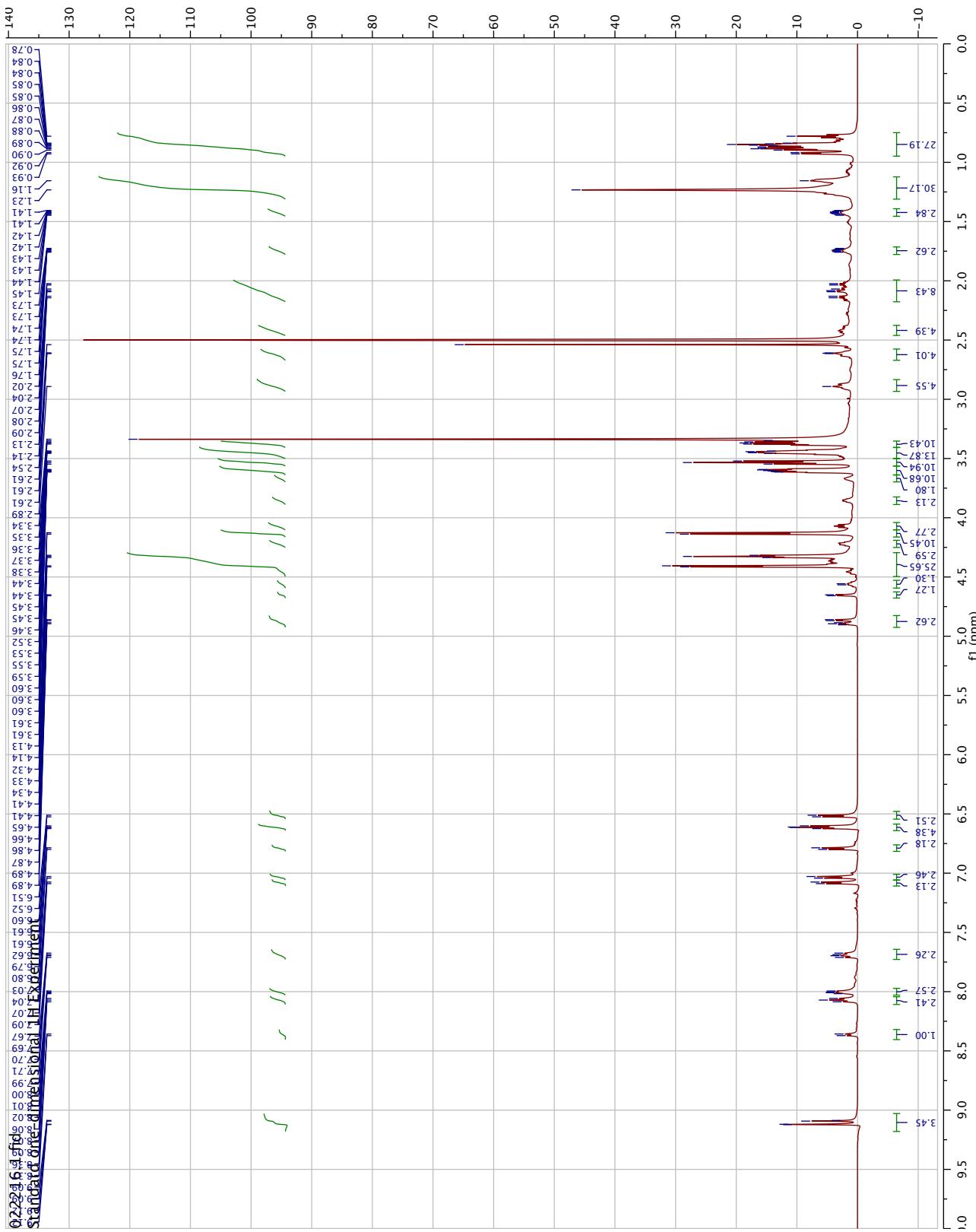
Supplementary Figure 4. ^1H NMR spectrum of humimycin A



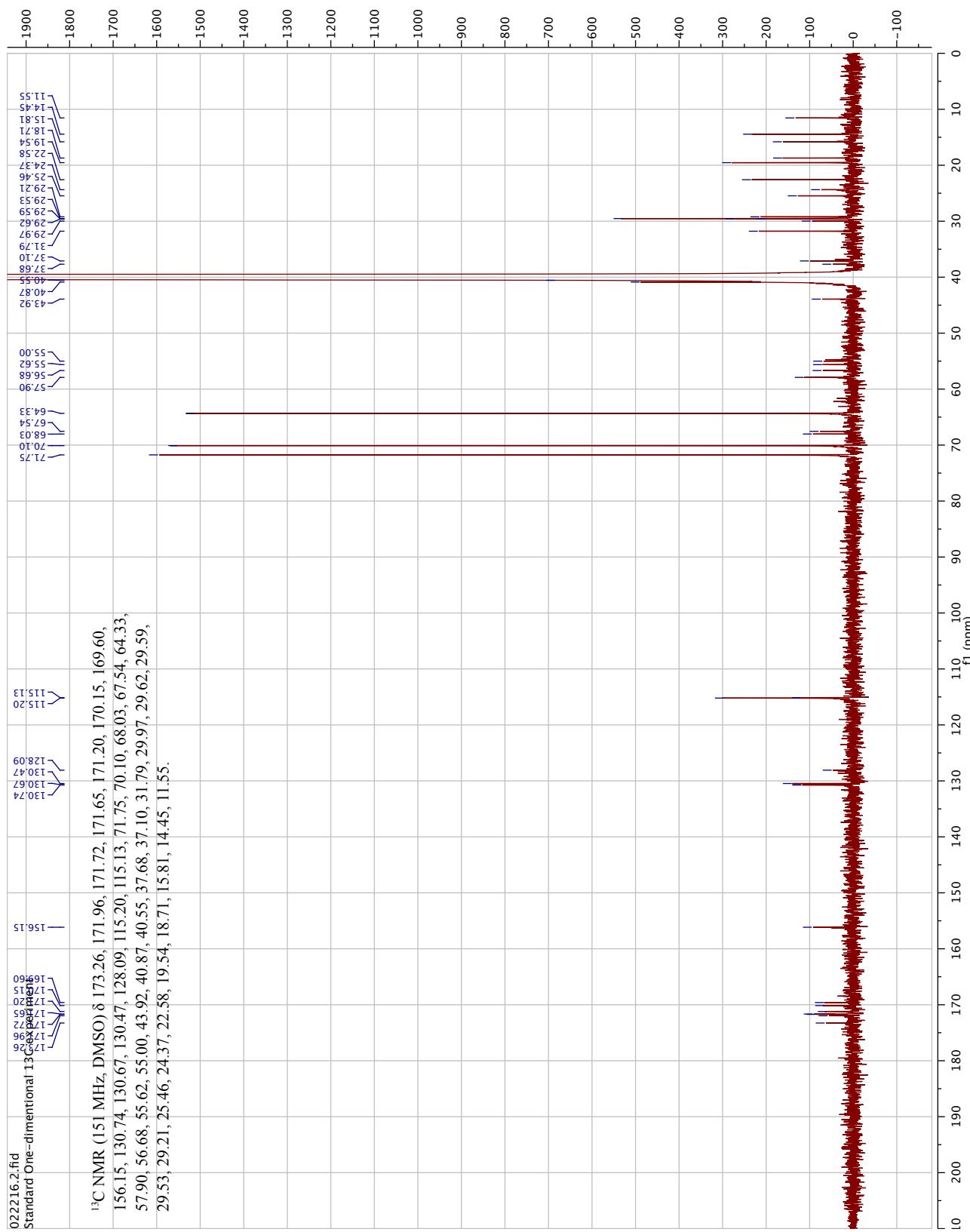
Supplementary Figure 5. ^{13}C NMR spectrum of humimycin A



Supplementary Figure 6. ^1H NMR spectrum of humimycin B



Supplementary Figure 7. ^{13}C NMR spectrum of humimycin B



Supplementary Table 1. List of all syn-BNPs for this study

No.	Name	N-Cap	1	2	3	4	5	6	7	8	9	10	11	12	13	MW	IsoMS	Obsd MS	Ion	Organism of origin	Genome cl.
1	Human.1	β -Hydroxymyristic acid	Tyr	Ser	D-Tyr	Tyr	D-Thr	Ile	Val	◀ Humanycin B	1134.4	1133.6	1155.6 [M+Na] ⁺	Rhodococcus erythropolis SK121	gj_225919436	2					
2	Human.2	none (NH ₂)	D-Tyr	Ser	Ser	Ser	Thr	Gly	Ser	687.7	687.2	688.3							gj_2259191953	1	
3	Human.3	β -Hydroxymyristic acid	Tyr	Ser	D-Tyr	Phe	D-Thr	Val	Val	◀ Humanycin A	1104.4	1103.6	1104.2	Rhodococcus erythropolis ATCC 33107	gj_316528562	2					
4	Human.4	none (NH ₂)	Gly	Orn	D-Ser	Ser	Gly	Phe	Val	893.1	892.5	893.55	Pseudomonas sp. HG47	gj_334134556	1						
5	Human.5v1	β -Hydroxymyristic acid	D-Thr	Ser	Ala	Ser	Leu	D-Gln	Orn	946.2	945.6	946.6	Bacillus sp. 7-6-55CFAA, CT2	gj_305164252	2						
6	Human.5	none (NH ₂)	Leu	D-Ser	D-Arg	Lys	Orn	Thr	717.9	717.5	718.45	Pseudomonas sp. 2-1-26	gj_35564830	2							
7	Human.7	none (NH ₂)	Ala	Phe	D-Ala	Ala	D-Ala	Phe	596.7	596.3	597.35	Mycobacterium parsonsophilaceum ATCC BAA-614	gj_2965172066	2							
8	Human.8v1	β -Hydroxymyristic acid	Tyr	Glu	D-Asp	D-Gln	D-Gln	D-Lys	D-Glu	909.0	908.4	909.5	Rhodococcus equu ATCC 33707	gj_326528562	3						
9	Human.8v2	β -Hydroxymyristic acid	Tyr	Lys	D-Lys	D-Glu	D-Glu	921.5	922.1	922.55	Rhodococcus equu ATCC 33707	gj_326528562	3								
10	Oral.1	β -Hydroxymyristic acid	Glu	D-Orn	Tyr	D-Thr	Glu	D-Val	Pro	1482.7	1481.8	1481.55 [M-H] ⁺	Bacillus subtilis BS56	gj_320017650	1						
11	Oral.2	none (NH ₂)	D-Ser	Lys	Gly	Orn	D-Ser	Ser	Gly	965.1	964.5	965.55	Pseudomonas fluorescens SS101	gj_367957393	2						
12	Oral.3	none (NH ₂)	Asp	D-Orn	Dab	Thr	Ala	D-Ala	D-Orn	804.9	804.5	805.5	Pseudomonas protegens Pf-5	gj_68342549	3						
13	Oral.4v1	none (NH ₂)	D-Asp	Orn	D-Phe	Gly	D-Glu	Gly	Glu	895.9	895.4	897.4	Bradyrhizobium elkanii WSM1741	gj_54896015	1						
14	Oral.4v2	none (NH ₂)	D-Asp	Orn	D-Phe	Gly	D-Glu	Gly	Orn	865.9	865.4	866.45	Bradyrhizobium elkanii WSM1741	gj_54896015	1						
15	Oral.5	β -Hydroxymyristic acid	Phe	Glu	Asp	Ile	Thr	Leu	Ala	1131.4	1129.5	[M-H] ⁺	Uncultured bacterium ACD 77C00477	gj_406883084	1						
16	Oral.6	none (NH ₂)	D-Leu	D-Thr	D-Ile	D-Leu	D-Ser	Leu	D-Ser	859.1	858.5	859.55	Pseudomonas fluorescens SS101	gj_387957393	1						
17	Oral.7	β -Hydroxymyristic acid	Asn	D-Tyr	D-Asn	Gln	Pro	D-Ser	Asn	1062.2	1061.5	1084.5 [M+Na] ⁺	Bacillus subtilis subsp. spizizenii N23	gj_305410941	4						
18	Oral.8	none (NH ₂)	D-Ser	Arg	D-Ser	Orn	Lys	Orn	Thr	907.0	906.5	907.5	Pseudomonas denitrificans ICBP-P-PA14	gj_11583796	4						
19	Oral.9v1	none (NH ₂)	D-Ser	D-Orn	Orn	Gly	D-Thr	Ser	Orn	693.8	692.4	693.4	Pseudomonas denitrificans PA7	gj_150595624	3						
20	Oral.9v2	none (NH ₂)	D-Ser	D-Orn	Gly	D-Thr	Ser	Glu	722.7	723.35	Pseudomonas denitrificans PA7	gj_150595624	3								
21	Oral.10v1	none (NH ₂)	Glu	Orn	Asp	Dab	Asp	D-Orn	Gly	762.8	762.4	763.3	Robusta picketti/DTP0602	gj_546540292	1						
22	Oral.10v2	none (NH ₂)	Asp	Dab	Glu	Asp	D-Orn	Orn	Gly	762.8	762.4	763.35	Rakotonia picketti/DTP0602	gj_546540292	1						
23	Oral.11	β -Hydroxymyristic acid	Leu	D-Ser	Pro	Val	Cys	Gly	801.1	800.5	801.4	Streptococcus mutans TCI-145	gj_34996068	1							
24	Oral.12	none (NH ₂)	Ser	D-Asp	Ser	Orn	Ser	508.5	508.2	509.25	Achromobacter xylosoxidans R8	gj_310757913	1								
25	Oral.13	none (NH ₂)	Ala	Leu	MeOrn	Ser	Thr	532.6	532.3	533.35	Mycobacterium neauraum ATCC 25795	gj_635284330	1								
No.	Name	N-Cap	1	2	3	4	5	6	7	8	9	10	11	12	13	MW	IsoMS	Obsd MS	Ion	Organism of origin	Genome cl.

Failed peptides synthesis

Human.5v2	β -Hydroxymyristic acid	D-Thr	Gin	Ala	Ser	Leu	D-Glu	Glu	1003.2	1002.6		Bacillus sp. 7-6-55CFAA, CT2	gj_365164252	2						
Human.9	β -Hydroxymyristic acid	Ser	Glu	Phe	Asp	Ser	Ser	Phe	1652.8	1651.8		Rhodococcus equu ATCC 33107	gj_326528562	7						
Human.10	β -Hydroxymyristic acid	D-Ala	Thr	D-Ala	Leu	Phe			962.2	961.6		Rhodococcus equu ATCC 33107	gj_326528562	5						
Human.11	β -Hydroxymyristic acid	Phe	Leu	Thr	Tyr				1029.4	1028.6		Rhodococcus equu ATCC 33107	gj_326528562	6						
Oral.14	β -Hydroxymyristic acid	D-Leu	D-Asp	D-Thr	D-Ile	D-Ser	Ile	Leu	D-Ser	1313.7	1312.9		Pseudomonas protegens Pf-5	gj_68342549	1					

Supplementary Table 2. Bacteria strain information

a)	Name of Bacteria	Strain	Media	Resistance
ESKAPE pathogens	<i>Enterococcus faecium</i>	Com15	LB	
	<i>Staphylococcus aureus</i>	USA300	LB	Methicillin
	<i>Klebsiella pneumonia</i>	ATCC 10031	LB	
	<i>Acinetobacter baumannii</i>	ATCC 17978	LB	
	<i>Pseudomonas aeruginosa</i>	PAO1	LB	
	<i>Enterobacter cloacae</i>	ATCC 14037	LB	
Various MRSA strains	<i>Staphylococcus aureus</i>	ATCC BAA-42	LB	Methicillin
	<i>Staphylococcus aureus</i>	NRS100	LB	Methicillin, tetracycline
	<i>Staphylococcus aureus</i>	NRS108	LB	Gentamicin
	<i>Staphylococcus aureus</i>	ATCC BAA-1721	LB	Hyper-virulent
	<i>Staphylococcus aureus</i>	NRS281	LB	Erythromycin
	<i>Staphylococcus aureus</i>	NRS22	LB	Vancomycin
	<i>Staphylococcus aureus</i>	USA800	LB	Methicillin
	<i>Staphylococcus aureus</i>	USA500	LB	Methicillin
	<i>Staphylococcus aureus</i>	USA300	LB	Methicillin
	<i>Staphylococcus aureus</i>	USA200	LB	Methicillin
	<i>Staphylococcus aureus</i>	USA100	LB	Methicillin
Staphylococci	<i>Staphylococcus intermedius</i>	NCTC 11048	LB	
	<i>Staphylococcus delphini</i>	8086	LB	
	<i>Staphylococcus pseudintermedius</i>	ED99	LB	
b)	Name of Bacteria	Strain	Media	Resistance
Actinobacteria	<i>Bifidobacterium adolescentis</i> *	L2-32	LYBHI	
	<i>Bifidobacterium longum</i> *	CCUG 52486	LYBHI	
	<i>Corynebacterium amycolactum</i> [§]	SK46	LYBHI	
	<i>Rhodococcus erythropolis</i>	ATCC 11048	BHI	
	<i>Rothia dentocariosa</i>	M567	LYBHI	
	<i>Rothia mucilaginosa</i> [§]	CC87LB	LYBHI	
Bacteroidetes	<i>Bacteroides dorei</i> *	5_1_36/D4	LYBHI	
	<i>Bacteroides finegoldii</i> *	CL09T03C10	LYBHI	
	<i>Parabacteroides merdae</i> *	CL03T12C32	LYBHI	
	<i>Prevotella melaninogenica</i> *	D10	LYBHI	
Firmicutes	<i>Bacillus subtilis</i>	168 1A1	LYBHI	
	<i>Eubacterium</i> sp. *	3_1_31	LYBHI	
	<i>Staphylococcus aureus</i>	RN4220	LB/BHI	
	<i>Staphylococcus aureus</i>	USA300	LB	Methicillin
	<i>Staphylococcus epidermidis</i>	RP62A	LB	Methicillin
	<i>Streptococcus mitis</i>	B6	LYBHI	
	<i>Streptococcus mutans</i>	UA159	LYBHI	
	<i>Streptococcus pneumoniae</i>	TCH8431/19A	BHI	
Proteobacteria	<i>Streptococcus sanguinis</i>	SK36	BHI	
	<i>Escherichia coli</i>	LF82	LB	
	<i>Neisseria mucosa</i>	ATCC 25996	LYBHI	
	<i>Salmonella enterica</i>	subsp. <i>enterica</i>	LYBHI	

1. Superscripts denote special growth conditions, i.e., anaerobic atmosphere (*) and 30°C incubation([§]).

2. The LYBHI medium contains (per liter) brain heart infusion (BHI, 37 g), yeast extract (5 g), maltose (1 g), cellobiose (1 g), cysteine (0.5 g), and hemin (5 mg).

Supplementary Table 3. MICs of humimycin A and its alanine scan analogs

Name ^c	Sequence ^{de}	MIC ($\mu\text{g/mL}$) ^{ab}	
		(R) diastereomer ^f	(S) diastereomer ^f
Original syn-BNP	HMA-YSyFtVV	32	8 (humimycin A, 1)
Ala1	HMA- A SyFtVV	>	>
Ala2	HMA-Y A yFtVV	>	>
Ala3	HMA-YSaFtVV	>	>
Ala4	HMA-YSy A tVV	>	>
Ala5	HMA-YSyF a VV	>	>
Ala6	HMA-YSyFt A V	>	32
Ala7	HMA-YSyFt V A	>	>

^a Minimal inhibitory concentration (MIC) values are expressed in units of $\mu\text{g/mL}$.

^b ">" denotes MIC values greater than 128 $\mu\text{g/mL}$ (highest concentration tested).

^c Alanine substitution analogs were synthesized on 2-chlorotriptyl resins. The first amino acid (1.2 equiv.) was loaded in the presence of DIEA (3 equiv.) and capped by DCM/MeOH/DIEA (80:15:5) treatment for 30 min. The rest of the SPPS steps were identical to those described in *Peptide Synthesis* section of the *Online Methods*.

^d HMA denotes *N*-terminal modification with 3-hydroxymyristic acid (MHMA).

^e Amino acids are abbreviated using standard 1-letter codes; upper and lower case letters denote amino acids in L- and D- forms, respectively.

^e Humimycin A (**1**) refers to the syn-BNP *N*-acylated with the (S)-HMA (**Supplementary Figure S1**).

Supplementary Table 4. MICs of humimycin A against various clinical isolates of MRSA

Strain	MIC ($\mu\text{g/mL}$)
USA300	8
BAA-42	8
NRS108	8
NRS22	8
USA100	8
USA200	16
USA800	16
BAA-1721	32
NRS100	128

Supplementary Table 5. List of SNPs in SAV1754 that confer humimycin A resistance

Mutations within SAV1754											Summary		
MUTANT CHROM	POS	TYPE	REF	ALT	EVIDENCE	STRAND	NT POS	AA POS	EFFECT	LOCUS TAG	GENE PRODUCT		
1	NC_007793	1875948	SNP	C	T:65 C:0	-	169/1662	57/553	missense_variant c.169G>A p.Ala57Thr	RS09285	polysaccharide biosynthesis protein		
2	NC_007793	1874892	SNP	C	T:50 C:0	-	1225/1662	409/553	missense_variant c.1225G>A p.Gly409Ser	RS09285	polysaccharide biosynthesis protein		
3	NC_007793	1875468	SNP	G	A:144 G:0	-	649/1662	217/553	missense_variant c.649C>T p.Leu217Phe	RS09285	polysaccharide biosynthesis protein		
4	NC_007793	1876068	SNP	T	C:43 T:0	-	49/1662	17/553	missense_variant c.49A>G p.Ser17Gly	RS09285	polysaccharide biosynthesis protein		
5	NC_007793	1874892	SNP	C	T:34 C:0	-	1225/1662	409/553	missense_variant c.1225G>A p.Gly409Ser	RS09285	polysaccharide biosynthesis protein		
6	NC_007793	1875894	SNP	T	A:69 T:0	-	233/1662	75/553	missense_variant c.233A>T p.Asn75Tyr	RS09285	polysaccharide biosynthesis protein		
7	NC_007793	1875791	SNP	G	A:69 G:0	-	326/1662	109/553	missense_variant c.326C>T p.Ala109Val	RS09285	polysaccharide biosynthesis protein		
8	NC_007793	1875948	SNP	C	T:57 C:0	-	169/1662	57/553	missense_variant c.169G>A p.Ala57Thr	RS09285	polysaccharide biosynthesis protein		
9	NC_007793	1875903	SNP	C	T:66 C:0	-	214/1662	72/553	missense_variant c.66G>A p.Ala72Thr	RS09285	polysaccharide biosynthesis protein		
10	NC_007793	1875416	SNP	G	A:120 G:0	-	701/1662	234/553	missense_variant c.701C>T p.Ser234Phe	RS09285	polysaccharide biosynthesis protein		
11	NC_007793	1875350	SNP	G	T:41 G:1	-	767/1662	256/553	missense_variant c.767C>A p.Pro256Gln	RS09285	polysaccharide biosynthesis protein		
12	NC_007793	1875737	SNP	C	A:32 C:0	-	380/1662	127/553	missense_variant c.380G>T p.Trp127Leu	RS09285	polysaccharide biosynthesis protein		
13	NC_007793	1875974	SNP	G	A:43 G:0	-	143/1662	48/553	missense_variant c.143C>T p.Ala48Val	RS09285	polysaccharide biosynthesis protein		
14	NC_007793	1875736	SNP	C	G:28 C:0	-	383/1662	127/553	missense_variant c.383G>C p.Trp127Cys	RS09285	polysaccharide biosynthesis protein		
15	NC_007793	1875869	SNP	C	T:15 C:0	-	248/1662	83/553	missense_variant c.248G>A p.Ser83Asn	RS09285	polysaccharide biosynthesis protein		
16	NC_007793	1875372	SNP	C	G:99 C:0	-	745/1662	249/553	missense_variant c.745C>G p.Glu124Gln	RS09285	polysaccharide biosynthesis protein		
17	NC_007793	1875792	SNP	C	T:44 C:0	-	325/1662	109/553	missense_variant c.325G>A p.Ala109Thr	RS09285	polysaccharide biosynthesis protein		
18	NC_007793	1875930	SNP	C	A:85 C:2	-	215/1662	72/553	missense_variant c.215C>T p.Ala72Val	RS09285	polysaccharide biosynthesis protein		
19	NC_007793	1875698	SNP	C	G:67 C:0	-	187/1662	63/553	missense_variant c.187G>T p.Val63Leu	RS09285	polysaccharide biosynthesis protein		
20	NC_007793	1875480	SNP	C	T:206 C:0	-	419/1662	140/553	missense_variant c.419G>C p.Ser140Thr	RS09285	polysaccharide biosynthesis protein		
21	NC_007793	1875857	SNP	T	A:102 T:0	-	637/1662	213/553	missense_variant c.637G>A p.Tyr87Phe	RS09285	polysaccharide biosynthesis protein		
22	NC_007793	1874892	SNP	C	T:56 C:1	-	260/1662	87/553	missense_variant c.1225G>A p.Gly409Ser	RS09285	polysaccharide biosynthesis protein		
23	NC_007793	1875870	SNP	T	A:30 T:0	-	1225/1662	409/553	missense_variant c.1225G>A p.Ser83Cys	RS09285	polysaccharide biosynthesis protein		
2	NC_007793	813007	SNP	A	G:93 A:0	-	239/642	80/213	missense_variant c.239T>C p.Val80Ala	RS03940	hypothetical protein		
5	NC_007791	2378	SNP	G	A:6692 G:0	+	778/1242	260/413	missense_variant c.778G>A p.Glu260Lys	RS14730	Plasmid recombination enzyme type 3		
8	NC_007791	2378	SNP	G	A:6862 G:8	+	778/1242	260/413	missense_variant c.778G>A p.Glu260Lys	RS14730	Plasmid recombination enzyme type 3		
10	NC_007793	2363468	SNP	G	T:234 G:1	-	221/360	74/119	missense_variant c.221C>A p.Thr74Asn	RS12070	50S ribosomal protein L18		
12	NC_007793	626985	SNP	C	T:50 C:0	-	419/666	140/221	missense_variant c.626C>T p.Ile202Met	RS02945	bacillithiol biosynthesis deacylase BshB2		
12	NC_007793	123206	SNP	G	C:100 G:0	-	606/873	202/290	missense_variant c.606C>G p.Ile202Met	RS00565	membrane protein		
14	NC_007793	2433481	SNP	A	C:30 A:0	-	778/1242	83/97	missense_variant c.243C>T p.Thr83Ile	RS11970	helicase		
17	NC_007793	2350655	SNP	G	A:47 G:0	-	248/294	778/1242	missense_variant c.248C>T p.Thr83Ile	RS14730	Plasmid recombination enzyme type 3		
18	NC_007791	2378	SNP	G	A:9202 G:6	+	778/1242	260/413	missense_variant c.778G>A p.Glu260Lys	RS14730	Plasmid recombination enzyme type 3		
22	NC_007791	2378	SNP	A	A:8436 G:102	+	778/1242	260/413	missense_variant c.778G>A p.Glu260Lys	RS14730	Plasmid recombination enzyme type 3		

Supplementary Table 6. Overexpression of SAV1754 confers humimycin A resistance.

a) Primers for the amplification of SAV1754^a

Primer F	gcgcgagctcagaggaggactct <u>ATG</u> ACTGAAAGTAAAGAAATGGTGC
Primer R	gcgcggta <u>TCATCGTAAAACCTA</u> ACTCTACGTC

b) PCR protocol

-
- Step 1 98°C for 8 min
 - Step 2 98°C for 30 sec
 - Step 3 62°C for 30 sec
 - Step 4 72°C for 2 min
 - Step 5 Go to Step 2 for 32 times
 - Step 6 72°C for 8 min
-

c) Susceptibility to humimycin A of *S. aureus* hosting various plasmids^{bc}

Insert	Vector	MIC ($\mu\text{g/mL}$) ^d
Empty vector	pRMC2	8
SAV1754 (WT)	pRMC2	>128
SAV1754 (mutant no.8)	pRMC2	>128

^a Start and stop codons of the SAV1754 gene (upper case) are in bold; restriction sites for cloning are underlined (*SacI/KpnI*); ribosome binding sequence was built into the forward primer (lower case).

^b The SAV1754 gene was placed under the control of the tetracycline-inducible promoter P_{xyl/tetO}.

^c Empty and recombinant plasmids were transformed into *S. aureus* RN4220.

^d Minimal inhibitor concentrations (MIC) were tested in the presence of 5 ng/mL of anhydrotetracycline and recorded after 48 h of growth; ">128" denotes MIC values greater than 128 $\mu\text{g/mL}$ (highest concentration tested).