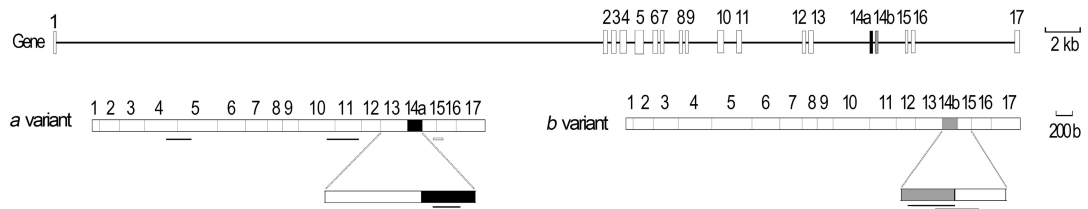


Table S1. Primers used in RT-PCR, RACE, ORF verification, dsRNA synthesis, and qPCR.

Fragment name	Forward primer	Reverse primer
RT-PCR		
<i>LdChSA-1</i>	GTCAAAGGTGCGATGTTA	AGAAACGGAAACTGGAAT
<i>LdChSA-2</i>	CGCTACTCGGTTTCCTAC	ACACTGTTCCGATGGTTT
<i>LdChSA-3</i>	CTTGCCCTCTACCTAACC	TTCGATACTGCCTTCGTC
<i>LdChSB-1</i>	TGACTTAGCGGACGACTC	AAGAAACAGCGAAGAGGG
<i>LdChSB-2</i>	GCAGCCATAATGAGTGTT	CTGAGTATGCCTGGGAAG
RACE		
<i>LdChSA</i> 5'-GSP		TTCCCACCAGCCGACAGA
<i>LdChSA</i> 5'-NGSP		AAGTCTGCTCGCCCCTCC
<i>LdChSB</i> 5'-GSP		CGCTGCGTAATTGACGGATT
<i>LdChSB</i> 5'-NGSP		CATGCGTAGATGCGAGTA
<i>LdChSA</i> 3'-GSP	ACCAATGACGATGACGAAGG	
<i>LdChSA</i> 3'-NGSP	CCAACGAAACCATCGGAACA	
<i>LdChSB</i> 3'-GSP	GGACAAAATGGTGGTGAC	
<i>LdChSB</i> 3'-NGSP	AAACTGAGAAACCCAAGGCACC	
ORF verification		
<i>LdChSA</i>	GGCGAGTGGATTGCGACC	CGATTATTTGTTCCCATC
<i>LdChSB</i>	ATGCAGAGGCGATATCAGT	TCATAATCTGAGAGATGCAG
dsRNA synthesis		
<i>dsChSA-1</i>	TTTCGACCAGACGAGATA	GTAGGTTAGGTAGGAGGC
<i>dsChSA-2</i>	ATTCCTTATGTTGGTGGG	CCCAGGATACATTGTTTAGA
<i>dsChSAa</i>	ATCAGTCTTTGCTTTCTT	ACTTTATGTGGAGGTTGT
<i>dsChSAb</i>	GAACTGCGGAACAAGTCG	CTCGGAAGTCTCCTCAATAT
<i>dsChSB-1</i>	AGGGTACTCTATTATTCATG	GTCAACATAGCTCCTTTT
<i>dsChSB-2</i>	GTAGTGGCTGCCGTCTTG	GCATTTGGACCTTTGAGT
<i>dse_{gfp}</i>	AAGTTCAGCGTGTCCG	CACCTTGATGCCGTTT
qPCR		
<i>qLdChSA</i>	TTGGAACCATAGCTCACATCTT	AATCTCCACTGCCTGCTTATC
<i>qLdChSAa</i>	TAAGGAAGAGAAGGCCCGTA	AAGGGCCACTTTATGTGGAG
<i>qLdChSAb</i>	ACGTTAAGTGCCGTTAGGA	GACCAAGATGAGTGCGAAGA
<i>qLdChSB</i>	AGACTTCTGGTGTGCTCTTC	GTAGGCGCATTCGTCCTTAT
<i>qLdRP4</i>	AAAGAAACGAGCATTGCCCTTCCG	TTGTCGCTGACACTGTAGGGTTGA
<i>qLdRP18</i>	TAGAATCCTCAAAGCAGGTGGCGA	AGCTGGACCAAAGTGTTCACCTGC
<i>qLdARF1</i>	CGGTGCTGGTAAAACGACAA	TGACCTCCCAAATCCCAAAC
<i>qLdARF4</i>	GTGCTCGTGAACCATGTGAA	AACCTCCAATCCCTCGTGAA

A *LdChSA*



B *LdChSB*

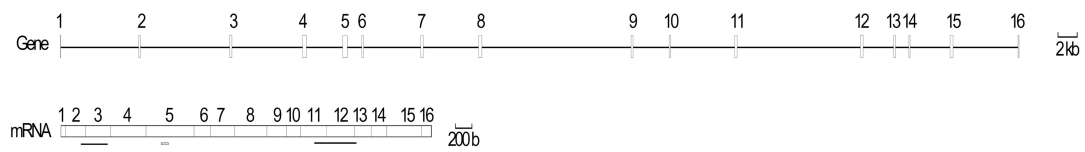


Figure S1. Exon/intron and mRNA structures of putative *ChSs* from *Leptinotarsa decemlineata*. Boxes mark exons. Lines mark introns. *LdChSA* and *LdChSB* genes respectively contains 17 and 16 exons, and 17 and 15 introns. For *LdChSA*, alternative splicing of exon 14a or 14b forms two splicing variants, *LdChSAa* and *LdChSBb*. The dsRNA and qRT-PCR sequences were marked with black and gray lines.

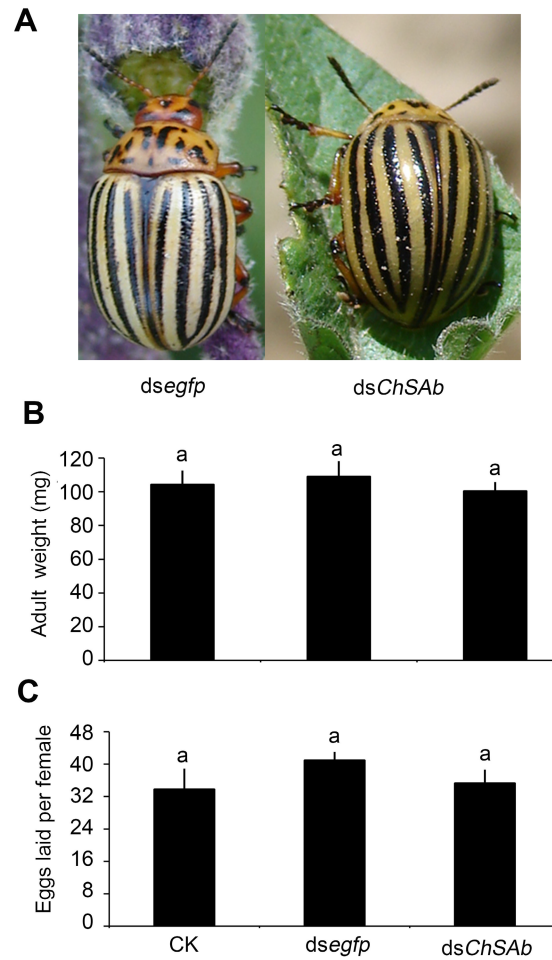


Figure S2. Effects of RNAi of *LdChSAb* in *L. decemlineata* second-instar larvae on adult performance. The resulting adults did not have obvious defective phenotypes and had similar size (A), weight (B) and fecundity (C) to control adults. The bars represent values (\pm SE). Different letters indicate significant difference at P value < 0.05 .

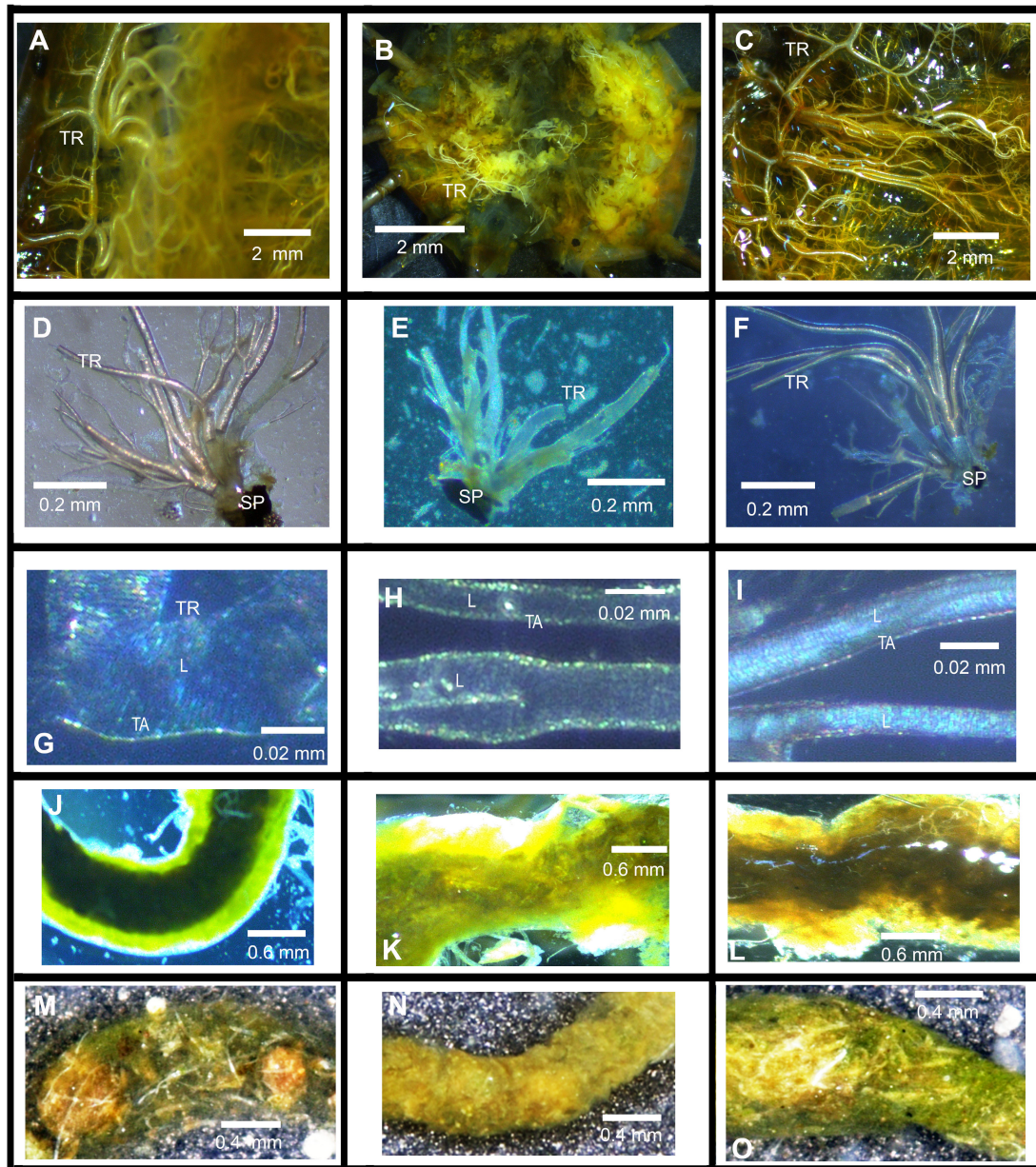


Figure S3. Knockdown of *LdChSaa* and *LdChSab* on chitin-containing structures in *L. decemlineata*. The PBS (the left column)-, *dsChSaa* (the middle column)-, and *dsChSab* (the right column)-ingested larvae are dissected and observed under a light microscope. The tracheae were directly seen (A-F), or are incubated with 10 M NaOH at 95 °C for 2 hrs (G-I). SP, spiracle; TR, tracheae; L, tracheal lumen; TA, taenidia. Larvae previously fed PBS and *dsChSab* have well developed tracheae (A, C, D, F), there are distinct taenidia in the tracheae (G, I). The taenidia run around the tracheal tube and form parallel transverse folds lining the lumen of the tracheae (G, I). In contrast, the *dsChSaa*-fed larvae possess underdeveloped tracheae (B), the taenidia are thinned (E, H). Moreover, the larvae previously fed PBS, *dsChSaa* and *dsChSab* have clear gut lumen, which was full of food (J, K, L). After removal of the midgut epithelia cells, integrate peritrophic matrix envelops food in these larvae (M, N, O).