
SUPPLEMENTARY INFORMATION

A CRISPR-Cas9 Assisted Non-Homologous End-Joining Strategy for One-step Engineering of Bacterial Genome

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HindIII-linearized pUC19 SmaI-linearized pUC19 CRISPR-cleaved pUC-lacZ

Supplementary Figure 1. Blue-white screening of the transformants to determine the efficiency and fidelity of the heterogenous NHEJ pathway in *E. coli*. White colonies represent the *lacZ* genotype and blue colonies represent the *lacZ⁺* genotype.

HindIII-linearized pUC19

WT	TTACGCCAAGCTGATGCCCTGCAGGTGACTCTAGAGGATCCC GG TACCGAGCTGAATTCACTGGCCCTCGTTTACAACGTCGTG
8 random	TTACGCCAAGCTGATGCCCTGCAGGTGACTCTAGAGGATCCC GG GAGCTGAATTCACTGGCCCTCGTTTACAACGTCGTG
selected	TTACGCCAAGCTGATGCCCTGCAGGTGACTCTAGAGGATCCC GG GAGCTGAATTCACTGGCCCTCGTTTACAACGTCGTG
white	TTACGCCAAGCTGATGCCCTGCAGGTGACTCTAGAGGATCCC GG GAGCTGAATTCACTGGCCCTCGTTTACAACGTCGTG
colonies	TTACGCCAAGCTGATGCCCTGCAGGTGACTCTAGAGGATCCC GG GAGCTGAATTCACTGGCCCTCGTTTACAACGTCGTG TTACGCCAAGCTGATGCCCTGCAGGT TC -N ₁₆₃

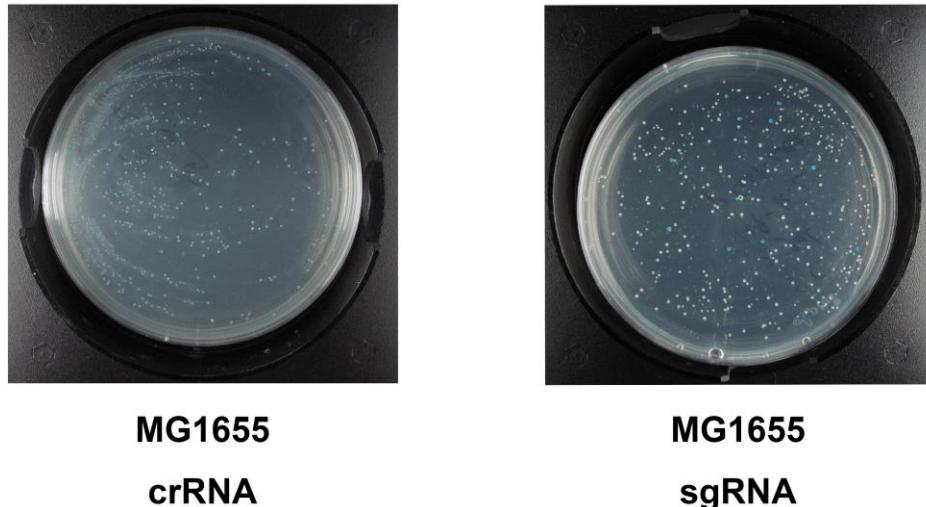
SmaI-linearized pUC19

WT	TTGTGTGGAATTGTGAGCGGATAACAATTCACACAGGAACAGCTATGACCATGATTACGCC AGCTT GCATGCCCTGCAGGTGACTCTA
8 random	TTGTGTGGAATTGTGAGCGG GG -CTGCAGGTGACTCTA
selected	TTGTGTGGAATTGTGAGCGGATAACAATTCACACAGGAACAGCTATGACCATGATTACGCC AGCT -TGCACTGAGGTGACTCTA
white	TTGTGTGGAATTGTGAGCGGATAACAATTCACACAGGAACAGCT ATG -CTGCAGGTGACTCTA
colonies	TTGTGTGGAATTGTGAGCGGATAACAATTCACACAGGAACAGCTATGACCATGATTACGCC AGCT -TGCACTGAGGTGACTCTA N ₃₄ -GGTCAGTCTA

CRISPR-cleaved pUC-lacZ

WT	GGGTACCGAGCTGAATTCACTGG CCGTCGT TTACAACGTCGTGACTGGAAAACCCTGGCGTTACCAACTTAATGCCCTGCAGCAC
8 random	GGGTACCGAGCTGAATTCACTGG CCGTCGT -CGTACTGGAAAACCCTGGCGTTACCAACTTAATGCCCTGCAGCAC
selected	GGGTACCGAGCTGAATTCACTGG CCGTCGT -GACTGGAAAACCCTGGCGTTACCAACTTAATGCCCTGCAGCAC
white	GGGTACCGAGCTGAATTCACTGG CCGTCGT -GACTGGAAAACCCTGGCGTTACCAACTTAATGCCCTGCAGCAC
colonies	GGGTACCGAGCTGAATTCACTGG CCGTCGT -GGGAAAACCCTGGCGTTACCAACTTAATGCCCTGCAGCAC GGGTACCGAGCTGAATTCACTGG CCGTCGT -CGTACTGGAAAACCCTGGCGTTACCAACTTAATGCCCTGCAGCAC GGGTACCGAGCTGAATTCACTGG CCGTCGT -N ₈₁

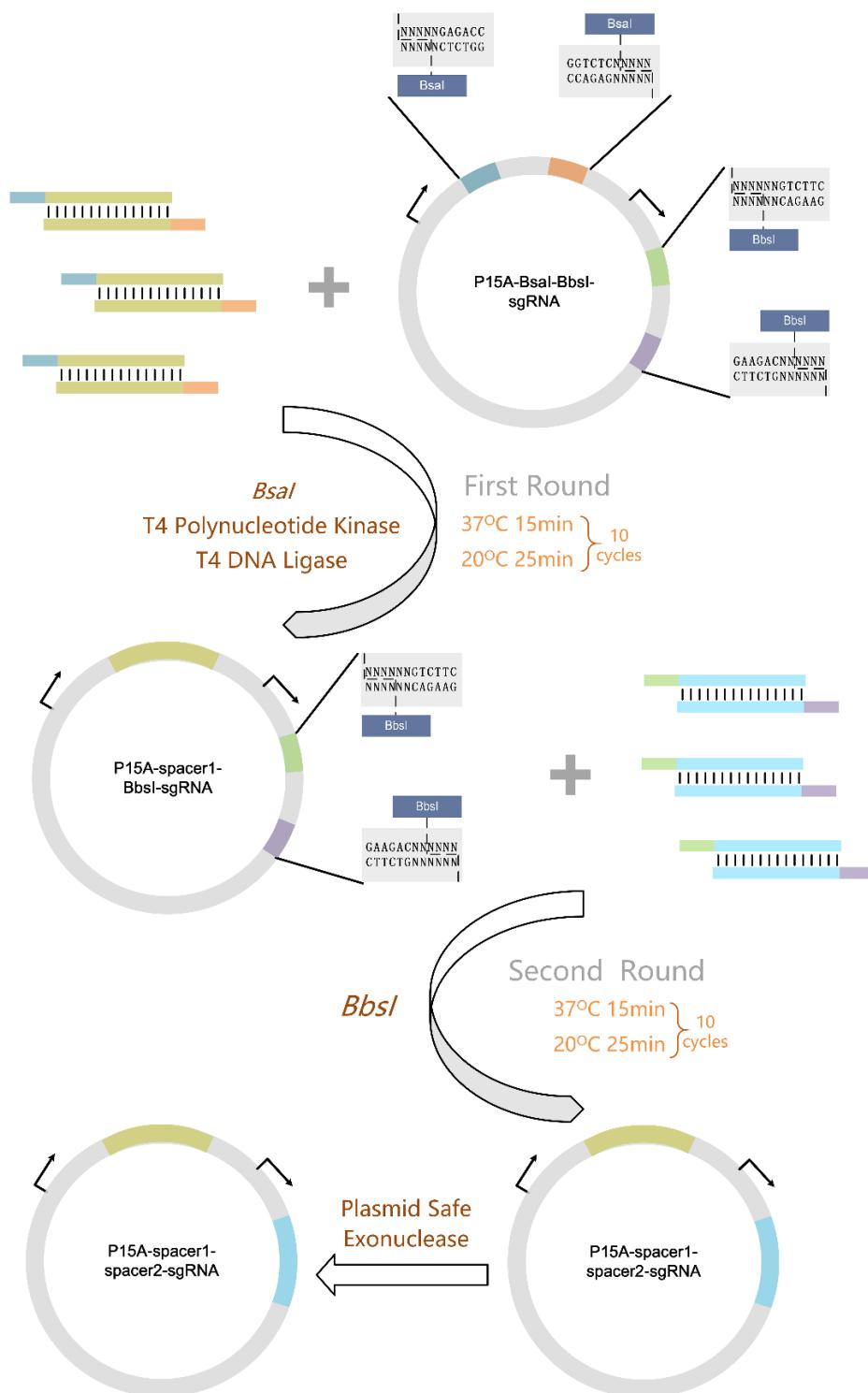
Supplementary Figure 2. Sanger sequencing analysis of 8 random selected white colonies (*lacZ*⁻ genotype) generated by repair of *HindIII/SmaI*-linearized pUC19 or Cas9-cleaved pUC-lacZ, respectively. The recognition sequences of *HindIII/SmaI* and the LR4-CRISPR target are shown in orange. The PAM sequences are shown in red. The light blue color indicates micro-homology sequence for ending-joining and the dashed line highlighted in yellow represents the deleted nucleotides by NHEJ.



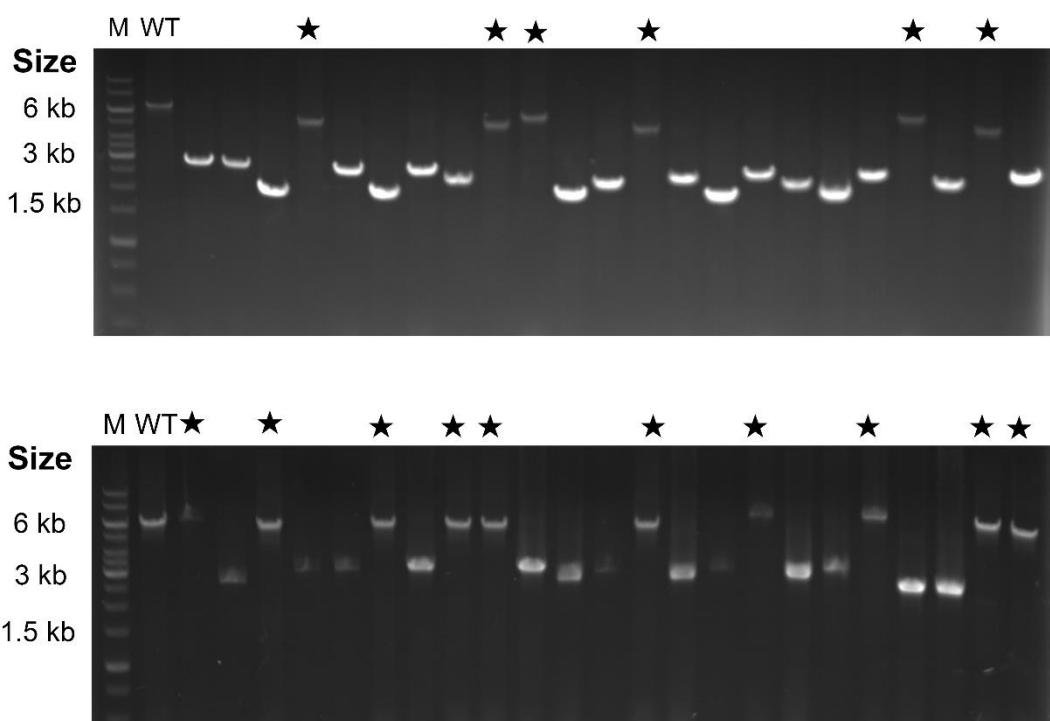
Supplementary Figure 3. Blue-white screening of the transformants to evaluate the efficiency and positivity rate of the improved CA-NHEJ system using L4 target site in *E. coli* MG1655. White colonies represent the *lacZ* genotype and blue colonies represent the *lacZ⁺* genotype.



Supplementary Figure 4. Sanger sequencing analysis of the LR4 CRISPR array in 8 random selected blue colonies grown on the X-gal plate. The repeat sequences and the LR4 spacer of CRISPR array are shown in blue and orange, respectively. The dashed line highlighted in yellow indicates the deleted sequences.



Supplementary Figure 5. One-step digestion-ligation method based on golden gate cloning to easy clone of specific sgRNA pair cassettes in plasmid p15A-BsaI-BbsI-sgRNA. The one-step digestion-ligation cloning strategy for ligation of the sgRNA pairs into the p15A-BsaI-BbsI-sgRNA plasmid was accomplished through two rounds iterative golden gate cloning in a 30- μ l reaction system. Tan: spacer 1; Light blue: spacer 2.



Supplementary Figure 6. Gel electrophoresis of the PCR products to distinguish the mutation types of *lacZ* by sgRNA pair L4&LR8. Asterisk indicates the frameshift mutation of *lacZ*.

Supplementary Table 1. Primers used in this study (The homologous sequences used for assembling were in bold, restriction sites were underlined).

Primers	Nucleotide sequence(5'-3')
Primers for plasmids construction	
lacZ-F	CCCAA <u>AGCTT</u> ACAGCTATGACCATGATTACGGATT
lacZ-R	TCCAG <u>ATCTA</u> GACCTTACCGCAAATACGGGCAGACAT
Cm-Ori-F	TA <u>GTGTGAGAT</u> CTCATGGGCTTACTCGATGCATGC <u>GCTAACGTTT</u> TATCAGGCTCTGGG
Cm-Ori-R	ATCCGTACGCCTGCAGGTCTAGATTA <u>ATTAACGCCGGCGGCCGCTATGGACAGTTTCC</u> CCTTGATAT
cas9-F	GC <u>GGCCGCCGCCGGCGTTA</u> ATTAATCTAGACCTGCAGGCGTACGGATTACGAAATCATCCTGTGGAGC
cas9-R	CG <u>GGTTAGCGCATGCATCGAGTAAGCCC</u> GATGAGATCTCACACTACTCTTCTTTGCCTATTATAAC
23119-lgd-F	ACCGTA <u>AGAT</u> TTGACAGCTAGCTAGTC <u>CTAGGTATA</u> ATGCTAG <u>CTAGAGAAAGAGGA</u> AATA <u>CTAGATGG</u> TT CGCGTCGGAGCA
23119-lgd-R	TTAATT <u>ATTCGCGACCACCTCA</u> CTGG
mku-F	GC <u>GGCCGGACAAGAAACCC</u> AGTGAGGTGGTGC <u>CGCAATGA</u> ATTAA <u>AGAGGA</u> AA <u>ACTAGATGC</u> GAGCCATTGGA CGGG
mku-R	ACA <u>ATGATGC</u> ATTAGCGCAAGAAGACAAAA <u>ATCAC</u> CTTGCG <u>CTAATG</u> CTGTTACAGTCACGGAGGCGTGGGACG
CRISPR-F	TCCAG <u>ATCTA</u> GATGC <u>CTCTAG</u> CACCGTACCAT
CRISPR-R	TCCAG <u>ATCTA</u> GATGACCG <u>AAATTCA</u> ACTCAACAA <u>AGT</u>
NHEJ-F	ACCG <u>ATACTAG</u> TTGACAGCT <u>CACTAG</u> GTATA <u>ATG</u> CTAGC
NHEJ-R	ACCG <u>ATACTAG</u> TTAGCG <u>CAAGAAGAC</u> AAAA <u>ATCAC</u> CTTGCGC

Spc-F	TCTGTTGTTGTCGGTGA ACTGGATCCCTAGTAAAGCCCTCGCTAGATT
Spc-R	ACCATCATACACTAAATCAGTAAGTGGCAGCATCACCGACGGCTCGGCCAGGCAGA
P15A-F	CGTCGGGTGATGCTGCCAACTTA
P15A-R	ACTTATATCGTATGGGGCTGACTTC
sgRNA-F	AGCACCTGAAGTCAGCCCCATACGATA AGTCTATAAAAAAGGCGTATCACGAGGC
sgRNA-R	TTACTAAGGGATCCAGTTCACCGACAAACA CAGA

Primers for gene knockout

lac-F	CGATACCGAAGACAGCTCATGT
lac-R	TATCATGCCGGCTTGCCCCGT

Primers for fragment deletion analysis

LacZ-JF	ATGTCCGCCGTAGCCCCTCCGATGAT
LacZ-JR	CCAGCAGGAACGGTACTTCAAAC
Lac-JF	AGCGCCTCGTCATCAATA
Lac-JR	TTCCCACAAGACAACAA
LC-JF	CAGAACAAATGAGCAGACGGAATA
LC-JR	GGAGGCTAACAGTGTCGAATAAC
MLC-JF	AGCCCTTCCATCAGAGCGACCA
MLC-JR	CACACAACCGGCACAAACCACC

Supplementary Table 2. Plasmids used in this study.

Plasmids	Relevant genotype	Reference
pUC19	Cloning vector, Amp ^R	Lab stock
pCRISPR	pUC19 containing CRISPR array, kan ^R	1
pCas9	pACYC184 containing tracr RNA, <i>cas9</i> and CRISPR array, Cm ^R	1
pwtCas9	pUC19 containing aTc-inducible promoter P _L tetO-1 expressing <i>cas9</i> , Amp ^R	2
pTKRED	Temperature-conditional replicon containing γ , β , <i>exo</i> (red recombinase), Spc ^R	3
pCP20	Helper plasmid, Cm ^R	4
pUC-lacZ	pUC19 containing <i>lacZ</i> gene, Amp ^R	This study
pCas9 (Ts)	Temperature-conditional replicon containing <i>cas9</i> , Cm ^R	This study
pCas9 (Ts)-NHEJ	pCas9 (Ts) containing p23119- <i>mku-ligd</i> , Cm ^R	This study
pCas9 (Ts)-LR4	pCas9 (Ts) containing LR4-CRISPR array, Cm ^R	This study
pCas9 (Ts)-NHEJ-LR4	pCas9 (Ts)-NHEJ containing LR4-CRISPR array, Cm ^R	This study
pcurCas9 (Ts)-NHEJ	pCas9 (Ts)-NHEJ containing <i>lacI</i> and trc-p15A-sgRNA, Cm ^R	This study
pwtCas9-NHEJ	pwtCas9 containing p23119- <i>mku-ligd</i> , Amp ^R	This study
p15A-gRNA	p15A replicon containing sgRNA, spc ^R	This study
p15A-BsaI-BbsI-gRNA	p15A replicon containing two sgRNA modules for one-step constructing sgRNA pairs, spc ^R	This study
pCRISPR-L4	pCRISPR containing L4 CRISPR array, kan ^R	This study
pCRISPR-LR4	pCRISPR containing LR4 CRISPR array, kan ^R	This study
p15A-L4	p15A-gRNA containing sgRNA L4, spc ^R	This study
p15A-L5	p15A-gRNA containing sgRNA L5, spc ^R	This study
p15A-LR4	p15A-gRNA containing sgRNA LR4, spc ^R	This study
p15A-LR6	p15A-gRNA containing sgRNA LR6, spc ^R	This study

p15A-LR7	p15A-gRNA containing sgRNA LR7, spc ^R	This study
p15A-LR8	p15A-gRNA containing sgRNA LR8, spc ^R	This study
p15A-L4&LR8	p15A-BsaI-BbsI-gRNA containing sgRNA pair L4&LR8, spc ^R	This study
p15A-LI10&LA0	p15A-BsaI-BbsI-gRNA containing sgRNA pair LI10&LA0, spc ^R	This study
p15A-LI10&CR0	p15A-BsaI-BbsI-gRNA containing sgRNA pair LI10&CR0, spc ^R	This study
p15A-ME17&CR0	p15A-BsaI-BbsI-gRNA containing sgRNA pair ME17&CR0, spc ^R	This study

Supplementary Table 3. CRISPR target sequences used in this study.

Spacer name	Nucleotide sequence (5'-3')
L4-CRISPR	GGTTTCCCAGTCACGACGTGTAACACGA
LR4-CRISPR	TCCGCCGTTGTTCCCACGGAGAACCGAC
L4	GTCACGACGTTGTAAACACGA
L5	GTGAGCGAGTAACAACCCGT
LR4	GTTCCCACGGAGAACCGAC
LR6	CAACGTGACCTATCCCATT
LR7	CCATGCCATCTGCTGCACG
LR8	CTCCTGGAGCCCGTCAGTAT
LI10	TACGATGTCGCAGAGTATGC
LA0	ATTGGCAATAACGTCTGGAT
CR0	CCCGCCACTACTGGAGAGAA
ME17	GTTCGCGATCGACTCGTAC

Supplementary Note 1

The sequences of core elements in this study

PJ23119-*ligd-mku*

ttgacagetagctcgtccatggataatgtacttagatctactagAGAAAGAGGAGAAATACTAGATG
GGTTCGGCGTCGGAGCAACGGGTGACGCTGACCAACGCCGACAAGG
TGCTCTATCCGCCACCGGGACCACAAAGTCCGATATCTCGACTACT
ACGCCGGTGTGCCGAAGTCATGCTCGGCCACATCGCGGGACGGCCG
GCGACGCGCAAGCGCTGGCTAACGGCGTCGACCAACCCGCGTTCTT
CGAAAAGCAGTTGGCGTTGTCGGCGCCCTGGCTGTCACGTGCAA
CGGTGGCGCACCGGTCCGGGACGACGACCTATCCGATCATCGATAGC
GCAACCGGGCTGGCCTGGATGCCAACAGGCGCGCTGGAGGTGC
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GGACCAACCGAGGTTGCTGGACGAGCCGAAGACGTCTCCGACCTGC
TCGCCAAGCTGGAGGCCAGCGTGAAGGCGCGCTCGAAGGCCAACTCA
AACGTCCCACGCCTCCGTGA

tracRNA-Cas9-CRISPR array

AAAAAAAGCACCGACTCGTGCCACTTTCAAGTTGATAACGGACTA
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GA
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GATTACGAAATT
TTAGA
CA
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AC

P_{J23119}-BsaI-sgRNA

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TTTAGAGCTAGAAATAGCAAGTTAAAAT
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- 1 Jiang, W. Y., Bikard, D., Cox, D., Zhang, F. & Marraffini, L. A. RNA-guided editing of bacterial genomes using CRISPR-Cas systems. *Nat Biotechnol* **31**, 233-239, doi:10.1038/nbt.2508 (2013).
- 2 Qi, L. S. *et al.* Repurposing CRISPR as an RNA-guided platform for sequence-specific control of gene expression. *Cell* **152**, 1173-1183, doi:10.1016/j.cell.2013.02.022 (2013).
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