## Transition steps in peroxide reduction and a molecular switch for peroxide robustness of prokaryotic peroxiredoxins

Neelagandan Kamariah<sup>a,#</sup>, Mun Foong Sek<sup>b,#</sup>, Birgit Eisenhaber<sup>a</sup>, Frank Eisenhaber<sup>a,c,d</sup> and Gerhard Grüber<sup>a,b,\*</sup>

<sup>a</sup> Bioinformatics Institute, Agency for Science, Technology and Research (A\*STAR), 30 Biopolis Street, #07-01 Matrix, Singapore 138671, Republic of Singapore

<sup>b</sup> School of Biological Sciences, Nanyang Technological University, 60 Nanyang Drive, Singapore 637551, Republic of Singapore

<sup>c</sup> School of Computer Engineering, Nanyang Technological University (NTU), 50 Nanyang Drive, Singapore 637553, Republic of Singapore

<sup>d</sup> Department of Biological Sciences, National University of Singapore, 8 Medical Drive, Singapore 117597, Republic of Singapore

<sup>#</sup> Authors contributed equally to this work

\* To whom correspondence may be addressed:

Prof. Dr. Gerhard Grüber, Tel.: + 65 - 6316 2989; Fax: + 65 - 6791 3856

E-mail: ggrueber@ntu.edu.sg

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**Supplementary Figure S1.** (*A*) Comparison of the ten subunits (chains A-J) of decameric oxidized  $EcAhpC_{1-186}$ -YFSKHN. Each subunit is shown with different color and the C<sub>P</sub>47 adopts LU conformation in all subunits. The C-terminal tail is not shown for clarity. (*B*) The dimeric unit (chain A and B: shown in dark and light colors) shows that the C-terminal tail of chain A is stabilized in the oxidized  $EcAhpC_{1-186}$ -YFSKHN structure by the packing interactions with the symmetry related molecule (grey color). (*C*) The C-terminus of chain H is stabilized by the packing interactions with the symmetry related molecule. Similarly, the C-terminal tail in chain G and J are also stabilized by packing interactions.

**Supplementary Figure S2.** (*A*) Comparison of the active site conformations between the ten subunits (chains A-J) of decameric reduced  $EcAhpC_{1-186}$ -YFSKHN. Each subunit are shown with different color and the C-terminal tail is not shown for clarity. The C<sub>P</sub>47 adopts two different conformation; in one subunit (chain A; red) the C<sub>P</sub>47 adopts a loop conformation and chains B-J adopt helical conformation. The influence of crystal packing interactions on the active site conformation for (*B*) chain A and (*C*) G is shown as a representative for the rest of the subunits in the decameric reduced  $EcAhpC_{1-186}$ -YFSKHN. No symmetry related molecule is in contact with the C<sub>P</sub>47 of the reduced structure.

**Supplementary Figure S3.** Non-reducing SDS-PAGE analysis of oxidation of (*A*) WT *Ec*AhpC, (*B*)  $EcAhpC_{1.172}$  and (*C*)  $EcAhpC_{1187G}$  by  $H_2O_2$ . The reduced AhpC proteins were treated with varying  $H_2O_2$  concentrations (0 to 30 mM) for 5 min. The SDS-PAGE analysis shows that the reduced AhpC proteins without  $H_2O_2$  run as a major band corresponding to a monomer (*Lane 1*). At 0.1 mM of  $H_2O_2$  *Ec*AhpC becomes oxidized and undergoes the transition from a monomer to a dimer (*Lane 2*). The monomer band reappears with high concentrations of  $H_2O_2$  (1 to 30 mM) likely due to over-oxidized *EcAhpC*. The analysis supports that *EcAhpC*<sub>1-172</sub> and *EcAhpC*<sub>1187G</sub> were sensitive to over-oxidation compared to WT *EcAhpC*. *D* indicates the dimeric and *M* the monomeric state.

**Supplementary Figure S4.** Comparison of interaction patterns of the reactive C<sub>P</sub>47 with the conserved active site residues between the typical FF (left panel; PDB ID: 4MA9) and FF<sub>like</sub> (right panel) conformations. A shift in C<sub>P</sub>47, located at helix  $\alpha 2$  of the FF<sub>like</sub> active site of reduced *Ec*AhpC<sub>1-186</sub>-YFSKHN, alters its interaction pattern with the conserved active site residues.

**Supplementary Figure S5.** Gel filtration elution profile of (*A*)  $EcAhpC_{1-186}$ -YFSKHN, (*B*)  $EcAhpC_{S86A,T88A}$ , (*C*) EcTrx and (D) EcTrxR.  $EcAhpC_{1-186}$ -YFSKHN,  $EcAhpC_{S86A,T88A}$ , and EcTrx were purified by Ni-NTA affinity- and size exclusion chromatography using a Superdex 75 column. In case of EcTrxR, a Superdex 200 gel filtration column was used. The volume of the shaded peak area was pooled and 3 µl of pure eluted protein were applied on a 17% SDS gel to estimate its purity (*inset*).



Supplementary Figure S1A-C



Supplementary Figure S2A-C





Supplementary Figure S3A-C



Supplementary Figure S4



Supplementary Figure S5A-D

**Supplementary Table S1:** The backbone dihedral angles for active site residues at different conformations

Conformation	Residue	Phi (°)	Psi (°)
	P48	-60.6	-36.5
FF	C47	-49.2	-53.6
	V46	-71.8	-35.4
	F45	115.8	-61.7
	P48	-88.6	48.5
LU	C47	-55.5	132.7
	V46	-58.1	132.6
	F45	-79.2	-21.3
	P48	-61.0	-34.5
FFlike	C47	-64.3	-38.9
ince	V46	-85.9	-30.8
	F45	-86.7	-35.6
	P48	-65.5	52.0
LU <sub>like</sub>	C47	-53.1	150.4
	V46	-67.0	90.1
	F45	-99.9	-15.3