1 Supplementary Information

- A peptidoglycan recognition protein acts in whitefly
 (*Bemisia tabaci*) immunity and involves in Begomovirus
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- ⁶ Zhi-Zhi Wang ^a, Min Shi ^a, Yi-Cun Huang ^a, Xiao-Wei Wang ^a, David Stanley^b and
 ⁷ Xue-Xin Chen^{a*}
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- 9 *^a Ministry of Agriculture Key Lab of Agricultural Entomology, Institute of Insect*
- 10 Sciences, Zhejiang University, 866 Yuhangtang Road, Hangzhou 310058, China
- ^b Biological Control of Insects Research Laboratory, Agricultural Research Service, U.S.
- 12 Department of Agriculture, 1503 S. Providence Road, Columbia MO 65203, USA

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14 ***Corresponding author:** <u>*xxchen@zju.edu.cn*</u>, phone number: +86 571 88982868

Prime r na me	Primer sequences (5'-3')	Use
PGRP-F 1	CACCACTCCCGACTTCCACCTTGC	3'RACE
PGRP-F 2	CTCGTGGGCTACTCGGAGCAGGAC	3'RACE
PGRP-R 1	AAGGTGGAAGTCGGGAGTGGTGGAT	5'RACE
PGRP-R 2	CGCCGAGAAATGCTATGTTGATGC	5'RACE
PGRP-BamH	CGGGATCCATTGAGGGTCGCCGAGATTCGTGGTACG	Vector construction
PGRP-Hind III	CCCAAGCTTCTATTCGACGAGGAGGGGGGGGG	Vector construction
V61	ATACTTGGACACCTAAT GG	IC-PCR
C473	AGTCACGGGCCCTTACAA	IC-PCR
actin-F	TGGAGATGGTGTTTCCCACAC	qRT-PCR
actin-R	CCAGCCAAGTCCAAACGAAG	qRT-PCR
q <i>PGRP</i> -R	TTTCGTGGATTTCTTTGC	qRT-PCR
q <i>PGRP-</i> F	CAAGGTGGAAGTCGGGAG	qRT-PCR
dsPGRP-F	GAGGCGATGGATCTGTTTAT	RNAi
dsPGRP-R	GGTGGCTGAACAGGGAGGAC	RNAi
PGRP-T7-F	ggatccTAATACGACTCACTATAGGGAGGCGATGGATCTGTT	RNAi
PGRP-T7-R	ggatccTAATACGACTCACTATAGGGGTGGCTGAACAGGGAGG	RNAi
dsGFP-F	AAGGGCGAGGAGCTGTTCACCG	RNAi
dsGFP-R	CAGCAGGACCATGTGATCGCGC	RNAi
GFP-T7-F	ggatceTAATACGACTCACTATAGGAAGGGCGAGGAGCTGTTC	RNAi
GFP-T7-R	ggatccTAATACGACTCACTATAGGCAGCAGGACCATGTGATC	RNAi

Table S1. Primer sequences used in this paper

Buffer No.	Buffer recipe	
1	25mM Tris buffer (pH 8.0) containing 0.2M NaCl, 5mM DTT, 1mM EDTA and 6M urea	
2	25mM Tris buffer containing 50mM NaCl, 1mM EDTA, 4M urea, 0.1mM l-arginine (Arg), 0.2mM oxidized glutathione (GSSG) and 2mM reduced glutathione (GSH) at pH 8.0	
3	25mM Tris buffer containing 20mM NaCl, 1mM EDTA, 2M urea, 0.1mM Arg, 0.2mM GSSG and 2mM GSH at pH8.0	
4	25mM Tris buffer containing 10mM NaCl, 1mM EDTA, 1M urea, 0.1mM Arg, 1mM DTT, 0.2mM GSSG and 2mM GSH at pH 7.4	
5	25mM Tris buffer containing 10mM NaCl, 1mM EDTA, 0.5M urea, 0.1mM Arg, 1mM DTT, 1% Gly, 0.2mM GSSG and 2mM GSH at pH 7.4	

Table S2. Dialysis buffers used in succession to refold rBtPGRP.



Figure S1. Expression of the BtPGRP gene in E. coli and purification of the BtPGRP

22 peptide. (A) Expression of the BtPGRP gene in *E. coli* B121 (DE3) and enrichment of the

rBtPGRP protein, analyzed on 12% SDS-PAGE gels. Lane M: protein molecular weight

24 markers; Lane 1: the total proteins in uninduced *E. coli* Bl21; Lane 2: the expressed

25 product of pET 28a-BtPGRP induced with 1 mM IPTG; Lane 3: Highly enriched

recombinant protein. (B) Western blot analysis of purified rBtPGRP.



- **Figure S2.** Immunofluorescence staining shows the binding of rBtPGRP to *C. albicans*.
- *C. albicans* was treated with rBtPGRP for 10 min. rBtPGRP (green) is not seen bound to
- 32 the fungal cell walls of *C. albicans*.



Figure S3. Enzymatic degradation of peptidoglycan. rBtPGRP or egg white lysozyme

37 was incubated with insoluble peptidoglycan from *E. coli* and *S. aureus* (InvivoGen,

USA), and the optical density (OD) at 540nm was recorded every fifth minute during a
120 min period.

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Figure S4. Microbial binding and bactericidal activity of His -Thrombin tag peptide. (a) 43 Western blot analysis of the microbial binding activity of tag protein. Live S. aureus and 44 E. coli were incubated with tag protein for 10 min. Bound tag protein (P) was separated 45 from free tag protein (S) in the supernatant by centrifugation. Tag protein without added 46 microorganisms was used as a control (left lane). The samples were analyzed by western 47 blotting using an anti-His antibody. (c) Bactericidal activity of tag protein against E. coli 48 (left) and S. aureus (right). Diluted bacteria samples were incubated with tag protein for 1 49 h or 2 h and then spread on LB agar plates. Viability was recorded as CFUs/ml after 50 incubation for 18 h (n = 3). 51



55 Figure S5. Analysis of *BtPGRP* mRNA expression. (a) *BtPGRP* mRNA expression in the

the indicated whitefly tissues and life stages. M, midgut; O, ovary; F, fat body; E, egg; 1-

4, 1st to 4th instar larvae; A, adults. (b) *BtPGRP* mRNA expression in the *B. tabaci* in

response to host plants. Whiteflies feeding on different host plants (cotton, cabbage and

tobacco) at least for three generations were used for RNA isolation. The data represent

60 the mean \pm SD of 3 biological samples.