

**CRISPR/Cas9 mediated somatic correction of a novel coagulator factor IX gene
mutation ameliorates hemophilia in mouse**

Supplementary Information List

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A

	<u>PAM</u>	<u>sgRNA</u>	
WT	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
F0 No. #1	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC .	-1bp
#2	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#3	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#4	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#5	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#6	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#7	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#8	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#9	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#10	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#11	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#12	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#13	GGCTTCCAT	TCTTCAGTCCCTTAGAGTTCC	A>C mutation
#14	GGCTTCCAT	TCTTCAGTCCCTTAGAGTTCC	A>C mutation
#15	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#16	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#17	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#18	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#19	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#20	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#21	GGCTTCCAT	TCTTCAGTCCCTTAGAGTTCC	A>C mutation

B

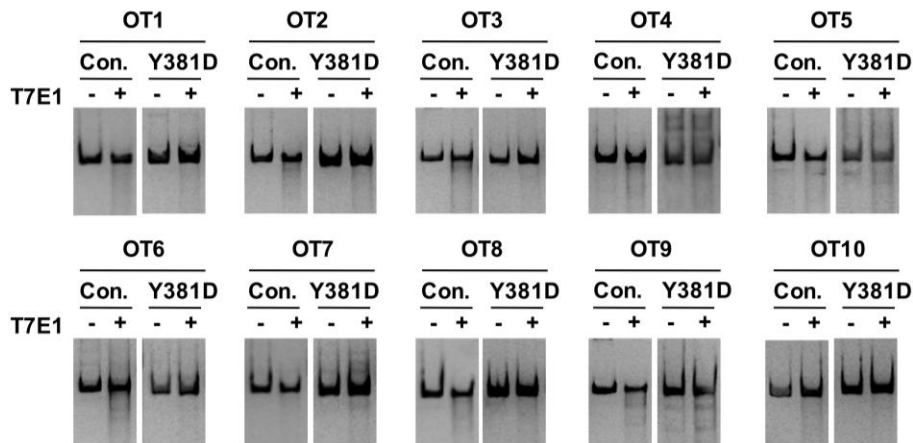
	<u>PAM</u>	<u>sgRNA</u>	
WT	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
F0 No. #1	GGCTTCCAT	TCT . CAGTACCTTAGAGTTCC	-1bp
#2	GGCTTCCAT	TCTTCAGTACCTT T . AG TTCC	-1bp Stop codon
#3	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#4	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#5	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#6	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#7	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#8	GGCTTCCAT	T CG . CAGTACCTTAGAGTTCC	-1bp
	GGCTTCCAT	T C . . . AGTACCTTAGAGTTCC	-3bp
#9	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#10	GGCTTCCAT	TCTTT T CAGTACCTTAGAGTTCC	+1bp
#11	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#12	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#13	GGCTTCCAT	T . . . CAGTACCTTAGAGTTCC	-3bp
#14	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#15	GGCTTCCAT	TCTTCAGTACCTTAGAGTTCC	
#16	GGCTTCCAT	TCTTCAG G ACCTTAGAGTTCC	T>G mutation

Appendix Figure S1. Generation of variant mouse strains with *F9* mutation via Cas9-mediated genome editing.

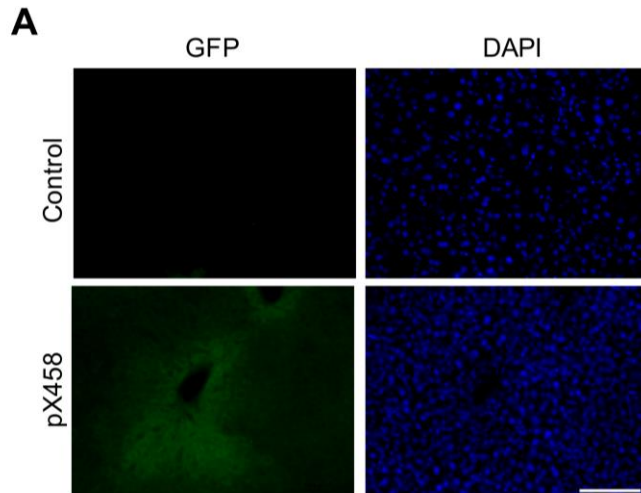
A The founder sequences of *F9*^{Y381S} mouse strain are shown. Nucleotide substitutions were indicated by a red letter. **B** The founder sequences of *F9*^{Y381D} and *F9*^{382Stop} strains are shown. Nucleotide substitutions were indicated by a red letter and deletions by dots; premature stop codon is boxed in red.

A

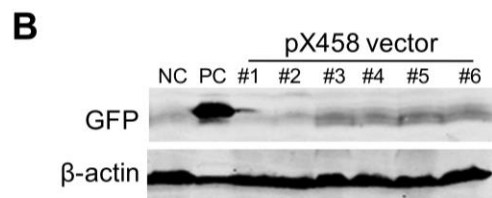
	sequence	score	UCSC gene	locus
OT1	CCTTTCTTCAGAACCATAGAGAT	1.4	NM_001164818	chr2:-156393557
OT2	CTTTTCTTCAGTACCTGAGTTTA	0.9	NM_007882	chr18:-20146875
OT3	CTCTTCTTCAGGACCTGAAAGTT	0.9	NM_001033460	chr5:+30657485
OT4	CTTCTCTTCAGTACCATATGGTT	0.7	NM_177184	chr9:-67795908
OT5	CTCTTCTTCAGTAAGTAAGGGTT	0.5	NM_175089	chr8:+63578702
OT6	CTCTTCTTCAGAACCCTGATATTT	0.5	NM_001080924	chr11:-5238542
OT7	CCGTTCTTCAGAACCCTTGGGCT	0.4	NR_102726	chr1:-173531247
OT8	CTGTCCTTCAGCACCTTAAAGGT	0.3	NM_025281	chr5:+38625321
OT9	CCATTCTTCACTACATCAGAATT	0.2	NM_027823	chr9:+8995857
OT10	CTATTCATCAGAACCCTTGGATT	0.2	NM_194055	chr4:+11294015

B**Appendix Figure S2. Off-target analysis of the sgRNA used for generation of *F9* mutant mice.**

A The 10 most likely potential off-target sites predicted by Optimized CRISPR Design website (<http://crispr.mit.edu/>) are listed with their sequence, score, gene ID and genomic locus. **B** T7E1 assays were performed with PCR products from the founder mouse of the *F9*^{Y381D} strain. No off-target mutation was detected by sequencing of 50 TA-clones of each PCR product.



ROIs	Positive	Total	Ratio
1	228	929	24.5%
2	341	1136	30.0%
3	210	1022	20.5%
4	360	1607	22.4%
5	39	942	4.1%
6	105	987	10.6%
7	270	1185	22.8%
8	111	1052	10.6%
Average	208	1107.5	18.2%

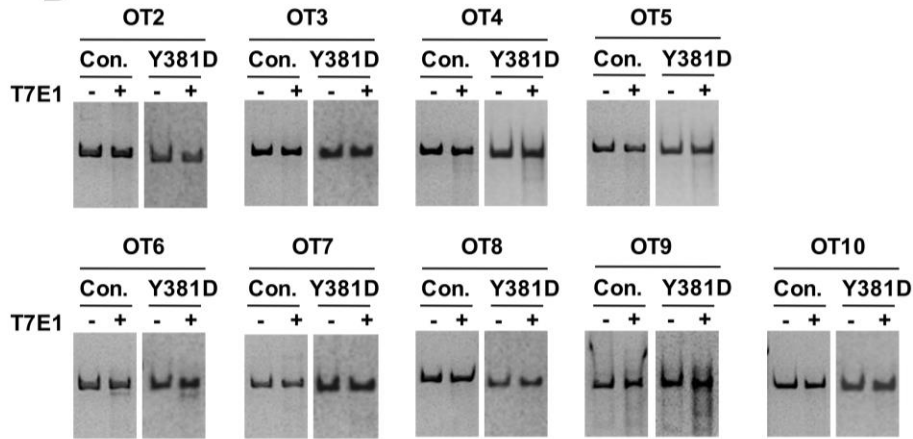


Appendix Figure S3. Expression of Cas9-2A-GFP protein in liver tissue from mice 20h after hydrodynamic injection of 120 μ g pX458 construct.

A Frozen sections were prepared from mouse liver tissues with or without plasmid DNA treatment. Upper panel: the sections were stained with 2-(4-Amidinophenyl)-6-indolecarbamide dihydrochloride (DAPI) and fluorescence images were taken with a microscope. Scale bar: 100 μ m. Lower panel: list of individual region of interests (ROIs). In each ROI, the GFP positive and total cell numbers were counted and the average percentage of GFP signals was calculated. **B** GFP expression was detected through western blotting with antibodies against GFP. β -actin was used as an internal loading control. NC: negative control without plasmid injection; PC: positive control of 293T cells transfected with pX458. The other lanes are individual mice that received pX458 through hydrodynamic tail vein injection.

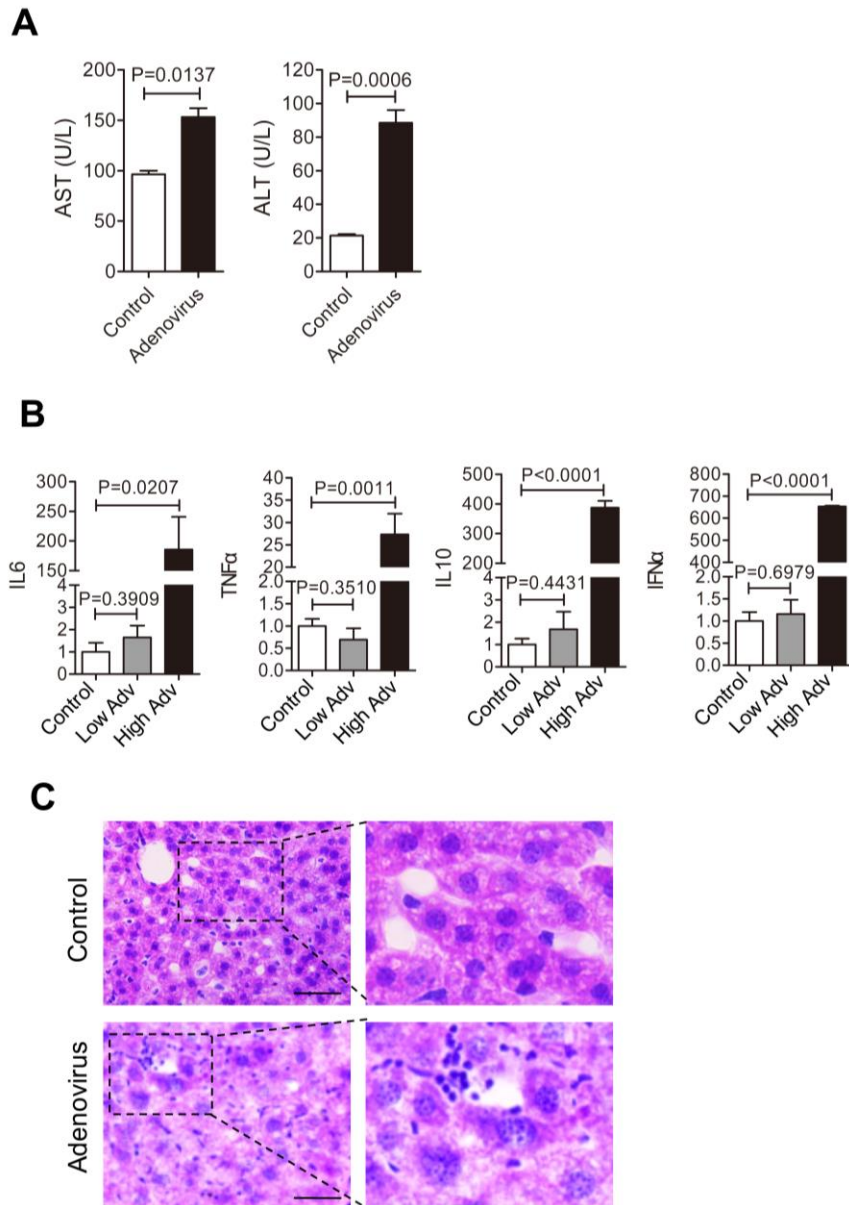
A

	sequence	score	UCSC gene	locus
OT1	CCATTCTTCAGTACCTTAGAGTT	49.2	NM_007979	chrX:-57282327
OT2	CTCTTCTTCAGGACCTGAAAGTT	3.7	NM_001033460	chr5:+30657485
OT3	CCTTTCTTCAGAACCATAGAGAT	1.4	NM_001164818	chr2:-156393557
OT4	CTCTTCTTCAGGACCTCAAAGAC	0.9	NM_011086	chr1:-65301691
OT5	CTTTTCTTCAGGGCCTTAAAATA	0.8	NM_013930	chr6:-23024686
OT6	CCATTCTTCATGAGCTCAGAGTT	0.6	NM_001165944	chr10:-129403721
OT7	CCCTTCTTCATGAGCTTAGGGTG	0.5	NM_011439	chr1:+135279172
OT8	CCATTCTTCAGGCCCATGGAGTG	0.5	NM_182745	chr5:+107974825
OT9	CTCTTCTTCAGAACCCTGATATTT	0.5	NM_001080924	chr11:-5238542
OT10	CCGTTCTTCAGAACCCTTGGGCT	0.4	NR_102726	chr1:-173531247

B

Appendix Figure S4. Off-target analysis of the sgRNA used for correction of the F9^{Y381D} mutation in mice.

A The 10 most likely potential off-target sites predicted by Optimized CRISPR Design website (<http://crispr.mit.edu/>) software are listed with their sequence, score, gene ID and genomic locus. **B** T7E1 assays were performed with PCR products amplified from DNA extracted from liver tissues of the mice treated with 1×10^{10} and 7×10^{10} vector genomes of AdvCas9 and AdvG/T, respectively. No off-target mutation was detected by sequencing of 50 TA-clones of each of 9 PCR products.



Appendix Figure S5. Liver damage was evaluated through molecular and histological studies.

A Aspartate aminotransferase (AST) and alanine aminotransferase (ALT) were tested in peripheral blood from F9^{Y381D} mice treated with or without 1×10^{10} and 7×10^{10} vector genomes of AdvCas9 and AdvG/T for 8 weeks (n=5). **B** mRNA levels of inflammatory cytokines were determined by qRT-PCR. mRNA samples were harvested in control or Adv treated mice described in (A). **C** Typical H&E staining images of liver tissues from mice treated with or without Adv. Scale bar=100µm.

Appendix Table S1 sgRNA sequences.

ID	Sequence (5'-3')
F9 ^{WT}	CTCTAAGGTACTGAAGAA
F9 ^{Y381D}	AACTCTAAGGTCCTGAAGAA

Appendix Table S2 ssODN sequences.

ID	Sequence (5'-3')
F9 T>G	TGGCTGGGGAAAAGTCTTCAACAAAGGGAGACAGGCTT CCATTCTTCAGGACCTTAGAGTTCCACTGGTGGATAGAG CCACATGCCTTAGGTCCACAAC
F9 A>C	GGCTGGGGAAAAGTCTTCAACAAAGGGAGACAGGCTTC CATTCTTCAGTCCCTTAGAGTTCCACTGGTGGATAGAGCC ACATGCCTTAGGTCCACAACA
F9 ^{Y381D} G>T	TGTCAGTGGCTGGGGAAAAGTCTTCAACAAAGGGAGAC AGGCTTCCATTCTTCAGTACCTTAGAGTTCCACTGGTGGAA TAGAGCCACATGCCTTAGGTCCACAACATTCATCTAT AA

Appendix Table S3 primer sequences for off target analysis of sgRNA for generation of F9^{Y381D} mutant mice.

ID	Sequence (5'-3')
F9-OT1-F	GGTGACAAGGAAGATGATGGG
F9-OT1-R	CAGAAGGACTCCAGAGTACAGC
F9-OT2-F	CCATTAGCTCTTCAGTCGTGG
F9-OT2-R	CTCTAGTGTCTGGTACACTGATGC
F9-OT3-F	CAGAGCTGGGCTATGCATGTATAG
F9-OT3-R	CTGCCGTATCTGCTTGGTGTAT
F9-OT4-F	GTCTGTGTACCATATGCATGCC
F9-OT4-R	AGCAAGTTGTCCTACACCTCCC
F9-OT5-F	GTTGCAACAATCAGAGTTGGGG
F9-OT5-R	GGGCCTTTTGGGATAGCATT
F9-OT6-F	GGCCCCAAAGATGTAAGGTACAG
F9-OT6-R	TCTTAGATGCACCCACTAGGCCT
F9-OT7-F	GTCTGCTCATCCGCTGGTTCTA
F9-OT7-R	CTCTGACTTCAAACCTCAGGAGCC
F9-OT8-F	CTGAGGCAGCTGCTTATCTTC
F9-OT8-R	AACAACACCCCAACCTCATC

F9-OT9-F	CCTTGACCAAGGAGCTTAGCA
F9-OT9-R	GCCATCGTCATTAACAGGAGTG
F9-OT10-F	AGACAGACAGGAAGGCTATAGGA
F9-OT10-R	GTCATGCTGGGATTTAATCCC

Appendix Table S4 primer sequences for off target analysis of *in vivo* genome editing sgRNA of F9^{Y381D} mutant mice.

ID	Sequence (5'-3')
F9 ^{Y381D} -OT2-F	GTGGATACATGCATGTGAGTGG
F9 ^{Y381D} -OT2-R	ACCGGTTTCAGTTTCCTCTTCTG
F9 ^{Y381D} -OT3-F	CAGGTGACAAGGAAGATGATGG
F9 ^{Y381D} -OT3-R	CTCCAAGGAAACAAGAGGAACC
F9 ^{Y381D} -OT4-F	CACTGGCAAAGATACTTCTGGG
F9 ^{Y381D} -OT4-R	AAACCAGGGGAAAGGCAGTT
F9 ^{Y381D} -OT5-F	GAGCAAATGTCTGTCTGCCAGA
F9 ^{Y381D} -OT5-R	CCATTCTGAGGTCTTGTGACTGAG
F9 ^{Y381D} -OT6-F	CTTATGGTCTTGAGCTGTACCAGC
F9 ^{Y381D} -OT6-R	GAATGGGTTGTTCTTCTCACAGG
F9 ^{Y381D} -OT7-F	GCTGTATCCTCTGGGTGTTTGT
F9 ^{Y381D} -OT7-R	GGAGAGAAATAGGGTCAACAGC
F9 ^{Y381D} -OT8-F	CACACCTGGCTTGAAAGGTT
F9 ^{Y381D} -OT8-R	CAACTCTGGTCACCCTGACTTAA
F9 ^{Y381D} -OT9-F	TTAGATGCACCCACTAGGCCTA
F9 ^{Y381D} -OT9-R	GATTATTGAGTATGGCCCCAGG
F9 ^{Y381D} -OT10-F	TTCAAACCTCAGGAGCCTCCAGA
F9 ^{Y381D} -OT10-R	GTCATCCGCTGGTTCTAATGT

Appendix Table S5 primer sequences for sequencing and T7E1 assay.

ID	Sequence (5'-3')
F9-F1	TCGAACTATCCCTCATCACCAG
F9-R1	GTCTGTAAAGGGCATCACCCAT
F9-F2	GAACCCATAAAGCACTCTCAGC
F9-R2	GGCTGGACTTACAGAAATGGTG
F9-F3	CTTCCTCAAGTTTGGTTCTGGC
F9-R3	CTTCACACGAATCTTTGCCTCC

Appendix Table S6 primer sequences for QPCR.

ID	Sequence (5'-3')
IL6-F	GGGACTGATGCTGGTGACAA
IL6-R	ACAGGTCTGTTGGGAGTGGT
TNF α -F	AGAGCTACAAGAGGATCACCAGCAG
TNF α -R	TCAGATTTACGGGTCAACTTCACAT
IL10-F	GCTCTTACTGACTGGCATGAG
IL10-R	CGCAGCTCTAGGAGCATGTG
actin-F	CGTCATACTCCTGCTTGCTG
actin-R	GTACGCCAACACAGTGCTG