

**ENERGY & MATERIALS** 

# Supporting Information

# Tandem Catalytic Depolymerization of Lignin by Water-Tolerant Lewis Acids and Rhodium Complexes

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# **Contents**



#### **S1 Experimental Procedures**

#### **Materials**

1,4-dioxane (99+%, a.c.s. grade, Alfa Aesar), chloro(1,5-cyclooctadiene)rhodium(I) dimer (98%, ABCR), 1,3-bis(diphenylphosphino)propane (98%, ABCR), scandium(III) triflate (95%, Acros), indium(III) triflate (99%, ABCR), gallium(III) triflate (98%, ABCR) and ytterbium(III) triflate (98%, ABCR) were all used as received. Lignin model compounds 2-phenoxy-1-phenylethan-1-ol  $(1a)$ ,  $[1]$  1- $(3,4$ dimethoxyphenyl)-2-(2-methoxyphenoxy)ethan-1-ol<sup>[1]</sup> (1b) and 1-(3,4dimethoxyphenyl)-2-(2-methoxyphenoxy)propane-1,3-diol (**1d,** obtained as a 5:1 mixture of *erythro*:*threo*) were synthesized according to previously published procedure. Dioxasolv lignin was isolated from poplar sawdust (sieved through a 1mm grid), brewers' spent grains, and commercial pine wood shavings according to a previously published procedure, affording 9, 4 and 3.5 wt% of lignin respectively.[2] The brewers' spent grains were sourced from Daftmill Distillery, Cupar, UK and were air dried to a constant weight before use. Poplar sawdust used in reactions was Soxhlet extracted overnight, dried under vacuum, and ball-milled for 25 days at 60 rpm to afford a very fine powder.<sup>[3]</sup>

#### **Physical Measurements**

GC measurements were performed on a Varian GC equipped with a VF-5 ms capillary column and an FID detector. Mesitylene was used an internal standard; response factors were determined for all products for which reference materials were available, for other compounds these were extrapolated by number of carbon atoms from structurally similar compounds. GC-MS measurements were performed on a Shimadzu GC-2010 using a VF5-ms column, coupled to a Shimadzu GC-MS-QP2010

mass spectrometer. 2D GC×GC-MS (1st column: VF-5ms 30 m, 0.25 mm ID, df 0.25 μm; 2nd column: VF-17ms, 1 m, 0.15 mm ID, df 0.15 μm) was performed in a GC-MS2010 Ultra (Shimadzu) equipped with a ZX1 thermal modulation system (Zoex). The temperature program started with an isothermal step at 40 °C for 5 min. Next, the temperature was increased to 280  $\degree$ C by 2.5  $\degree$ C min<sup>-1</sup>. The program finished with an isothermal step at 280 °C for 5 min. The modulation applied for the comprehensive GC×GC analysis was a hot jet pulse (375 ms) every 7000 ms. The 2D chromatograms were processed with GC Image software (Zoex). NMR spectra were recorded on a Varian spectrometer operating at 400 MHz ( ${}^{1}$ H) or 100 MHz ( ${}^{13}$ C) in DMSO-d6 (unless mentioned otherwise) and referenced against the signal of the residual protio impurity of the solvent  $(^1H 2.50$  ppm,  $^{13}C 39.52$  ppm). HSQC spectra were recorded on a Bruker Avance II 600 MHz spectrometer equipped with a 5 mm CPTCI <sup>1</sup>H- $13C/15N/2H$  cryogenic probe with z-gradients at 25 °C using the Q-CAHSQC pulse program.[4] Matrices of 2048 data points for the <sup>1</sup>H-dimension and 128 data points for the  $^{13}$ C-dimension were collected with a relaxation delay of 6 s and spectral widths from 13 to -1 ppm and from 160 to 0 ppm for the  $\rm{^1H}$  and  $\rm{^{13}C}$  dimensions, respectively. The lignins and lignin products were dissolved in DMSO-d6 after overnight stirring (200 mg/750 μL) and chemical shifts were referenced to the solvent signal (2.50/39.5 ppm). The spectra were processed and analysed using MestReNova software. Prior to Fourier transformation, FIDs were apodized with a  $\pi/2$  sine square bell function in both dimensions and zero-filled up to 512 points in the  $^{13}$ C-dimension and 4096 points in the  ${}^{1}$ H-dimension. A semi-quantitative analysis of the HSQC spectra was performed by integration of correlations peaks in the different regions of the spectra with MestReNova. The relative quantity of side chains involved in the interunit and terminal substructures was expressed as a number per 100 aromatic units (S+G). In the aliphatic oxygenated region, interunit linkages were estimated from Cα−Hα correlations. GPC measurements were performed on a Polymer Labs GPC 50 system, equipped with a series of three PLGel Mixed-E columns and guard a guard column and using THF spiked with 0.1 vol% acetic acid as mobile phase. Detection was done with an external Knauer UV detector at 280 nm and molecular weight determinations were based on calibration with polystyrene standards.

#### **Catalytic experiments**

In a typical experiment, a 40 mL stainless steel Parr autoclave was charged with model compound (2.40 mmol), lignin (300 mg) or poplar sawdust (2.0 g), chloro(1,5 cyclooctadiene)rhodium(I) dimer (29.0 mg, 0.060 mmol), 1,3 bis(diphenylphosphino)propane (99.0 mg, 0.240 mmol), scandium(III) triflate (59.1 mg, 0.120 mmol) and 22 mL 9:1 1,4-dioxane:water mixture. After sealing, the autoclave was flushed three times with 50 bars argon and after release of pressure heated to the reaction temperature. After reacting for 2 hours, the heating mantle was removed and the reactor rapidly cooled with an ice-water bath. From the resulting solution was taken a 200 µL sample, which was concentrated *in vacuo* and redissolved in the GPC mobile phase for GPC analysis. To the remainder was added mesitylene and a 0.5 mL sample was passed over 0.5 g of silica gel padded with 0.2 g of magnesium sulfate to remove water and the inorganic compounds; organics were subsequently eluted with 3 mL ethyl acetate and analyzed by GC and GC-MS. In so far possible, quantification was performed against authentic standards; if not response factors were determined by interpolating from known compounds (Table S2).

#### **2-bromo-3',4',5'-trimethoxyacetophenone**

To 120 mL of ethyl acetate is added 40.3 g (180 mmol) of finely ground copper(II) bromide and brought to reflux. 3',4',5'-trimethoxyacetophenone (18.9 g, 90 mmol) was dissolved in 120 mL chloroform and added to the refluxing dark green copper(II) bromide suspension. After refluxing for 4 hours the suspension had turned brownish and HBr evolution ceased; the suspension was allowed to cool to room temperature and the solids removed filtration. The filtrate was concentrated in vacuo to afford 22.8 g of brown oil that solidifies upon standing. The crude material was used directly in the subsequent step without further purification. <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  7.24 (s, 2H), 4.41 (s, 2H), 3.92 (s, 9H).

### **2-(2-methoxyphenoxy)-1-(3,4,5-trimethoxyphenyl)ethan-1-one**

The crude material from the previous step was dissolved in 250 mL acetone, to which was added 9.31 g (75 mmol) guaiacol and 11.3 g (81.8 mmol) potassium carbonate. The mixture was refluxed for 3 hours and after cooling to room temperature the solids were removed by filtration. Concentration of the filtrate afforded a yellow-brown solid, which was subjected to recrystallization from 5:1 ethanol:toluene, affording the title compound as 12.5 g (37.6 mmol, 42%) of yellow crystals. Analytical data was in agreement with literature. <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  7.33 (s, 2H), 6.97 (dt, J = 8.6, 4.2 Hz, 1H), 6.91 (dt, J = 8.1, 1.0 Hz, 1H), 6.86 (dd, J = 4.6 Hz, 1.1 Hz, 2H), 5.26 (s, 2H), 3.92 (s, 3H), 3.90 (s, 6H), 3.87 (s, 3H). <sup>13</sup>C NMR (CDCl3) δ 193.91, 153.27, 149.82, 147.50, 143.27, 129.89, 122.62, 120.96, 114.82, 112.24, 106.01, 72.54, 61.07, 56.41, 55.95.

#### **2-(2-methoxyphenoxy)-1-(3,4,5-trimethoxyphenyl)ethan-1-ol**

2-(2-methoxyphenoxy)-1-(3,4,5-trimethoxyphenyl)ethan-1-one (11.4 g, 34.3 mmol) was added to a mixture of 250 mL tetrahydrofuran, 75 mL methanol and 75 mL methanol and gently heated to ca. 50 °C until all material had dissolved. Sodium borohydride (2.64 g, 96.8 mmol) was carefully added in small portions with vigorous gas evolution. After addition of all sodium borohydride, the mixture was stirred for another 2 hours and then quenched with 250 mL of saturated ammonium chloride solution. The organic layer was separated and the aqueous layer extracted three times with 100 mL ethyl acetate. The combined organic layers were washed with brine, dried with magnesium sulfate and concentrated in vacuo to afford 13.3 g of pale yellow oil. The compound was purified by silica gel column chromatography with a 1:1 ethyl acetate:hexanes mixture(Rf  $\sim$  0.45) as eluent, to afford 9.00 g (27.0 mmol, 79%) of a colorless viscous oil, which eventually crystallized as white needles.  ${}^{1}H$ NMR (CDCl<sub>3</sub>) δ 7.04 – 6.87 (m, 4H), 6.66 (s, 2H), 5.03 (dd, J = 9.2, 3.0 Hz, 1H), 4.18 (dd, J = 10.0, 3.0 Hz, 1H), 3.97 (dd, J = 10.0, 9.2 Hz, 1H), 3.89 (s, 3H), 3.87 (s, 6H), 3.84 (s, 3H). <sup>13</sup>C NMR (CDCl3): δ 153.47, 150.27, 148.08, 137.72, 135.38, 122.78, 121.24, 116.28, 112.13, 103.34, 76.55, 72.59, 60.96, 56.26, 55.97; Elemental analysis calculated for  $C_{18}H_{22}O_6$ : C 64.66; H 6.63; found C 64.54 H 6.63

### **S2 Lignin Characterization**





*Figure S1: HSQC NMR spectra of poplar dioxasolv lignin before (a) and the freezedried reaction mixture after reaction (b, for labelling scheme see Figure S7). After reaction both the signals for the*  $\beta$ *-O-4 and*  $\beta$ *-5 linkages have disappeared, demonstrating complete conversion of those ether fragments. Reaction conditions: 300 mg lignin, 0.060 mmol [Rh(cod)Cl]2, 0.240 mmol dppp, 0.120 mmol (ScOTf)3; 175 °C, 2 h, 22 mL 1,4-dioxane:water*



*Figure S2: Molecular weight distribution of poplar sawdust dioxasolv lignin before and after reaction, based on GPC data (reaction conditions: 300 mg lignin, 0.060 mmol [Rh(cod)Cl]2, 0.240 mmol dppp, 0.120 mmol (ScOTf)3; 175 °C, 2 h, 22 mL 1,4 dioxane:water; GPC: 0.1% AcOH in THF eluent, polystyrene standards).*

*Table S1: Quantification of side chains and aromatic units present in poplar lignin the freeze-dried reaction mixture after reaction. Reaction conditions: 300 mg lignin, 0.060 mmol [Rh(cod)Cl]2, 0.240 mmol dppp, 0.120 mmol (ScOTf)<sup>3</sup> ; 175 °C, 2 h,* 

				22 mL 1,4-dioxane:water		
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<sup>a</sup> Linkage content per 100 aromatic units

 $<sup>b</sup>$  Molar percentage (S+G+H=100)</sup>

<sup>c</sup> The presence of signals of  $p$ -hydroxy benzoate and  $p$ -hydroxy units cannot be unequivocally established due to overlap with the dppp signals.

Elemental Analysis (wt %): C (60.75%), H (6.23%), N (0.02%), S (0.00%)



*Figure S3: Molecular weight distribution of dioxasolv brewer's grain determined by GPC (THF eluent; polystyrene standards).*



*Figure S4: HSQC NMR of brewer's grain lignin. For labelling scheme see Figure S7.*

*Table S2: Quantification of side chains and aromatic units present in brewer's grain*

	Brewer's grain lignin
$\beta$ -O-4 $(A)^a$	42.0
$\beta$ -5 $(B)$ <sup>a</sup>	9.8
$\beta$ - $\beta$ (C) <sup>a</sup>	2.5
$p$ -coumarate (Pca) <sup>a</sup>	3.7
Tricin $(T)^a$	0.8
$S(%)^b$	30.6
$G (%)^b$	65.0
$H (%)^b$	4.5

## *lignin*

<sup>a</sup> Linkage content per 100 aromatic units

 $b$  Molar percentage (S+G+H=100)

Elemental Analysis (wt %): C (57.45%), H (5.76%), N (0.66%), S (0.04%)

*Pine Lignin*



*Figure S5: Molecular weight distribution of dioxasolv pine lignin determined by GPC (THF eluent; polystyrene standards).*



*Figure S6: HSQC NMR of pine lignin. For the labelling scheme see Figure S7.*

	Pine lignin	
$\beta$ -O-4 $(A)^a$	29.0	
$\beta$ -5 $(B)^a$	11.9	
$\beta$ - $\beta$ (C) <sup>a</sup>	4.2	
Stilbene $(St)^a$	0.6	
$S(%)^b$		
$G \left( % \right)^{b}$	96	
$H (%)^b$		

*Table S3: Quantification of side chains and aromatic units present in pine lignin*

<sup>a</sup> Linkage content per 100 aromatic units

 $<sup>b</sup>$  Molar percentage (S+G+H=100)</sup>

Elemental Analysis (wt %): C (64.03%), H (6.86%), N (0.00%), S (0.00%)



*Figure S7: Labelling scheme used in assignments in HSQC NMR spectra.*



*Figure S8: Molecular weight distribution of reaction mixture after reaction of 1b and 1c determined by GPC (THF eluent; polystyrene standards). Conditions: 2.4 mmol substrate, 0.060 mmol [Rh(cod)Cl]2, 0.240 mmol dppp, 0.120 mmol Sc(OTf)3; 175 °C, 2 h, 22 mL 9:1 1,4-dioxane:water.*



*Figure S9: Comparison of molecular weight distribution of reaction mixtures after reaction of poplar lignin and sawdust determined by GPC (THF eluent; polystyrene standards). Conditions: 300 mg lignin or 2.0 g sawdust, 0.060 mmol [Rh(cod)Cl]2, 0.240 mmol dppp, 0.120 mmol Sc(OTf)3; 175 °C, 2 h, 22 mL 9:1 1,4-dioxane:water.*



*Figure S10: Molecular weight distributions from GPC data of reaction mixtures after depolymerization with different amounts of catalyst. Conditions: 300 mg lignin, 0.5 eq [Rh(cod)Cl]2, 2 eq dppp, Sc(OTf)<sup>3</sup> as listed; 175 °C, 2 h, 22 mL 9:1 1,4 dioxane:water.*



*Figure S11: Molecular weight distributions from GPC data of reaction mixtures after depolymerization with different acids. Conditions: 300 mg lignin, 0.060 mmol [Rh(cod)Cl]2, 0.240 mmol dppp, 0.120 mmol acid; 175 °C, 2 h, 22 mL 9:1 1,4 dioxane:water.*

### **S4 GC Calibration**



# Table S4: Experimentally-determined response factors

<sup>a</sup> Effective Carbon Number



*Figure S12: Linear regression between ECN and experimentally determined response factors, which was used to estimate response factors for other compounds.*

### **S5 GC/GC-MS Data for Model Compound Reactions**



*Figure S13: GC-MS Chromatogram corresponding to Table 1, entry 1; showing the* 

*formation of 2-phenylethanol.*



*Figure S14: GC-MS Chromatogram corresponding to Table 1, entry 2; showing the* 

*formation of 2-phenylnaphthalene.*



*Figure S15: GC-MS Chromatogram corresponding to Table 1, entry 3; showing the formation of homoveratryl aldehyde. The compound is not present in the NIST database; the peak at m/z = 180 corresponds to the molecular ion, whereas the fragment at m/z = 151 is attributed to loss of the carbonyl.*



*Figure S16: GC-MS Chromatogram corresponding to Table 1, entry 3; showing the formation of* **7b***. The compound is not present in the NIST database;*  $m/z = 324$ *corresponds the molecular ion.*



*Figure S17: Overlaid GC chromatograms corresponding to Table 1, entry 6 and an authentic sample of* cis*-5b, demonstrating the predominant formation of the* cis *isomer. The trans isomer is present as a minor impurity in the authentic sample and is also observed in the reaction mixture.*



*Figure S18: GC-MS Chromatogram corresponding to Table 1, entry 11; showing the formation of 1-veratrylethanone. The compound is not present in the NIST database; the peak at m/z = 194 corresponds to the molecular ion, whereas the fragment at m/z = 151 is attributed to loss of the acetyl functionality.*

# **S6 Complete Product Listing for Reactions with Lignin**

# *Table S5: Table 2, Entry #1*



# *Table S6: Table 2, Entry #2*



# *Table S7: Table 2, Entry #3*



### *Table S8: Table 2, Entry #4*



# *Table S9: Table 2, Entry #5*



### *Table S10: Table 2, Entry #6*



# *Table S11: Table 2, Entry #7*



## *Table S12: Table 2, Entry #8*



# *Table S13: Table 2, Entry #9*



# *Table S14: Table 2, Entry #10*



### *Table S15: Table 2, Entry #11*



# *Table S16: Table 3, Entry #1*



### *Table S17: Table 3, Entry #2*



# *Table S18: Table 3, Entry #3*



# *Table S19: Table 3, Entry #4*



### *Table S20: Table 3, Entry #5*



### **References**

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