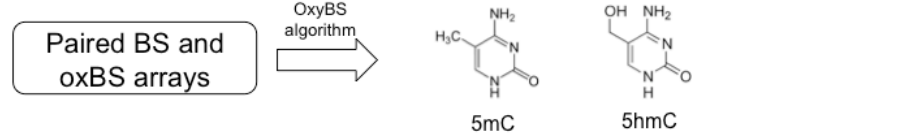


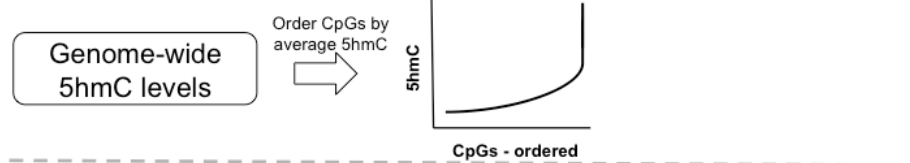
SUPPLEMENTARY FIGURES:

Supplementary Figure 1

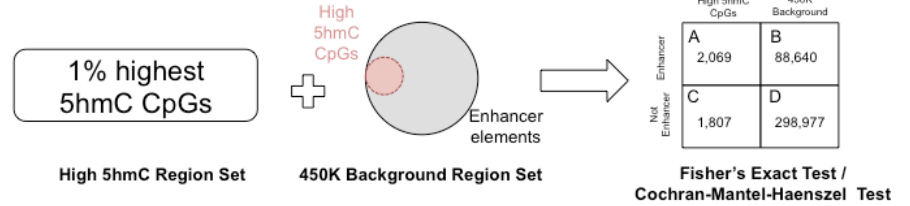
1. Characterize 5hmC distribution and abundance



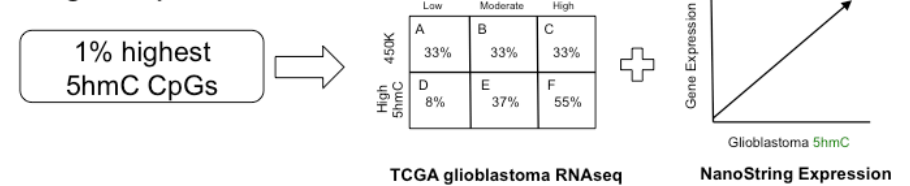
2. Identify high 5hmC CpGs



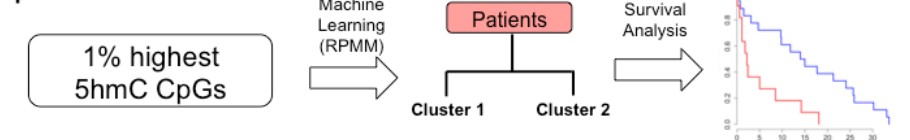
3. Functional annotation



4. Test relation between 5hmC and gene expression



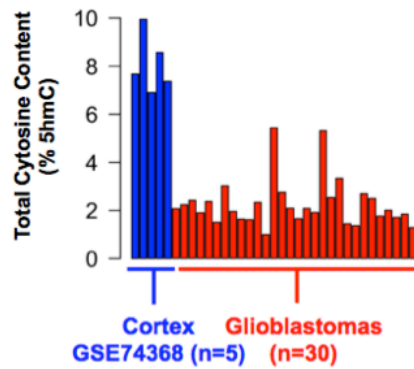
5. Cluster patients by 5hmC profiles



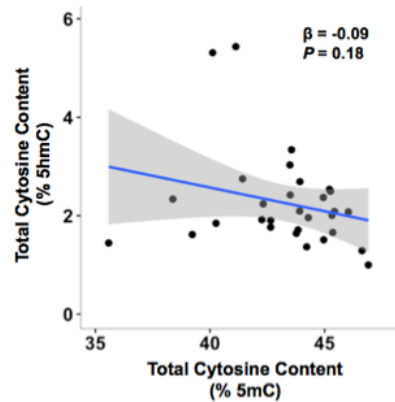
Supplementary Figure 1. Glioblastoma 5hmC quantified by paired BS and oxBS treated DNA hybridized to Infinium DNA methylation arrays. Workflow depicts analytic approach using the BS-oxBS protocol and OxyBS software. 1. Nucleotide level 5mC and 5hmC are resolved from paired arrays. 2. The CpGs in the glioblastoma genome that exhibit frequent levels of high 5hmC are identified. 3. The enrichment of genomic regions with high 5hmC is tested against the background of the 450K array genomic regions. 4. Correlations between 5hmC and glioblastoma are assessed in glioblastoma TCGA RNAseq data and confirmed using NanoString nCounter candidate genes. 5. The association between patient 5hmC profiles and survival are analyzed.

Supplementary Figure 2

A

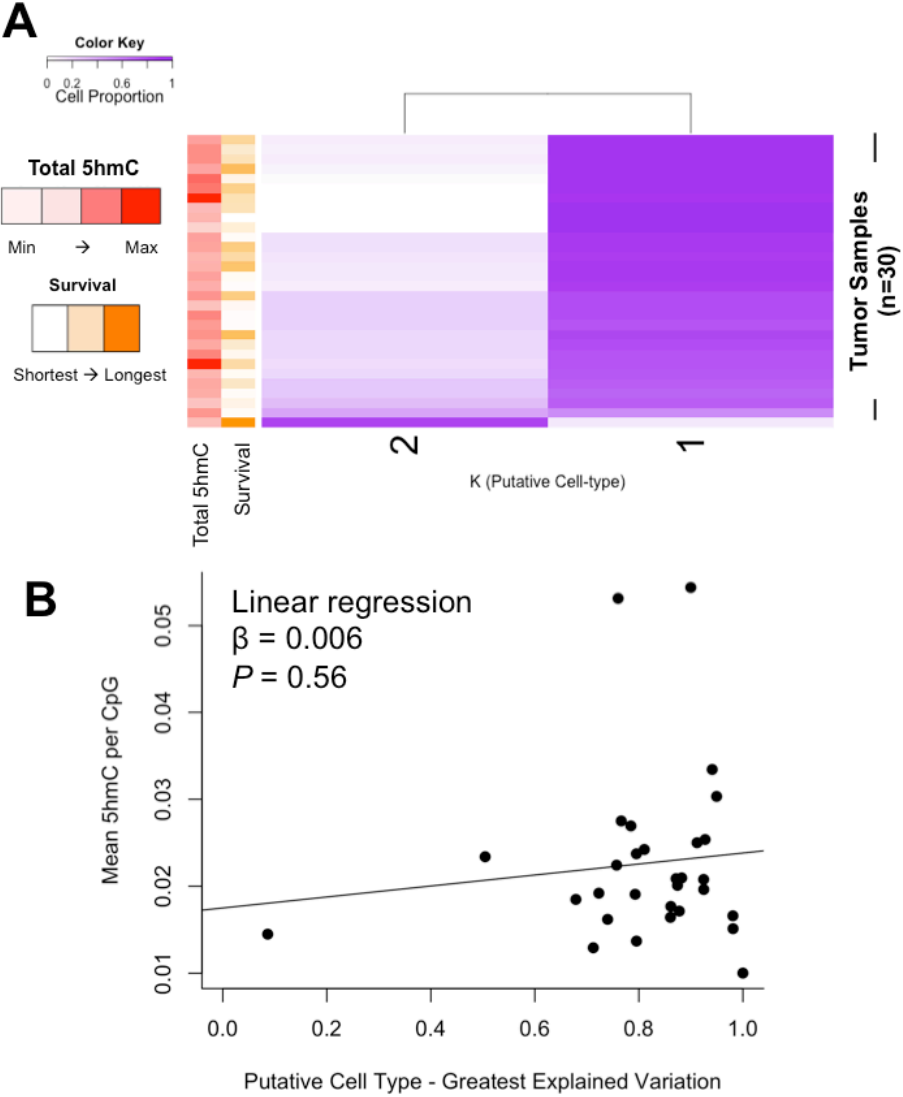


B



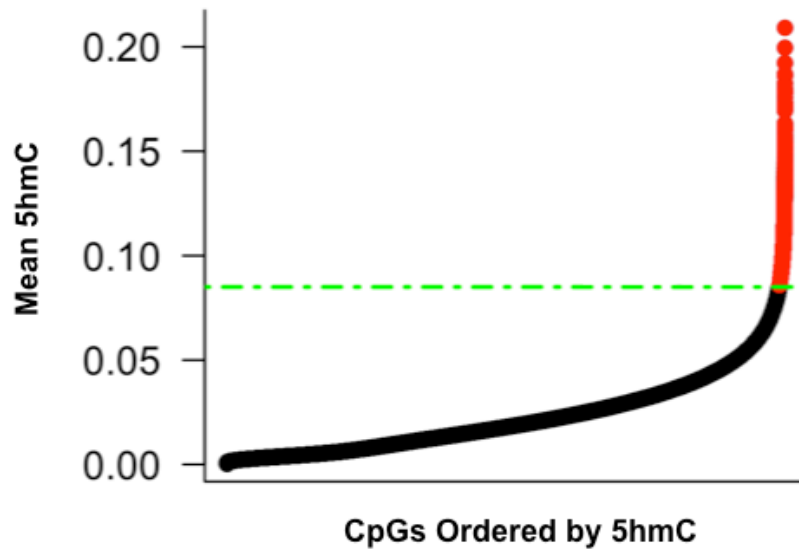
Supplementary Figure 2. (A) Proportion of 5hmC in total cytosine content (a measure of total 5hmC) across prefrontal cortex samples from GSE74368 (n=5) is elevated compared with glioblastomas (n = 30). **(B)** Proportion of 5hmC in total cytosine content is not significantly associated with the proportion of 5mC in total cytosine content in glioblastoma (n=30).

Supplementary Figure 3



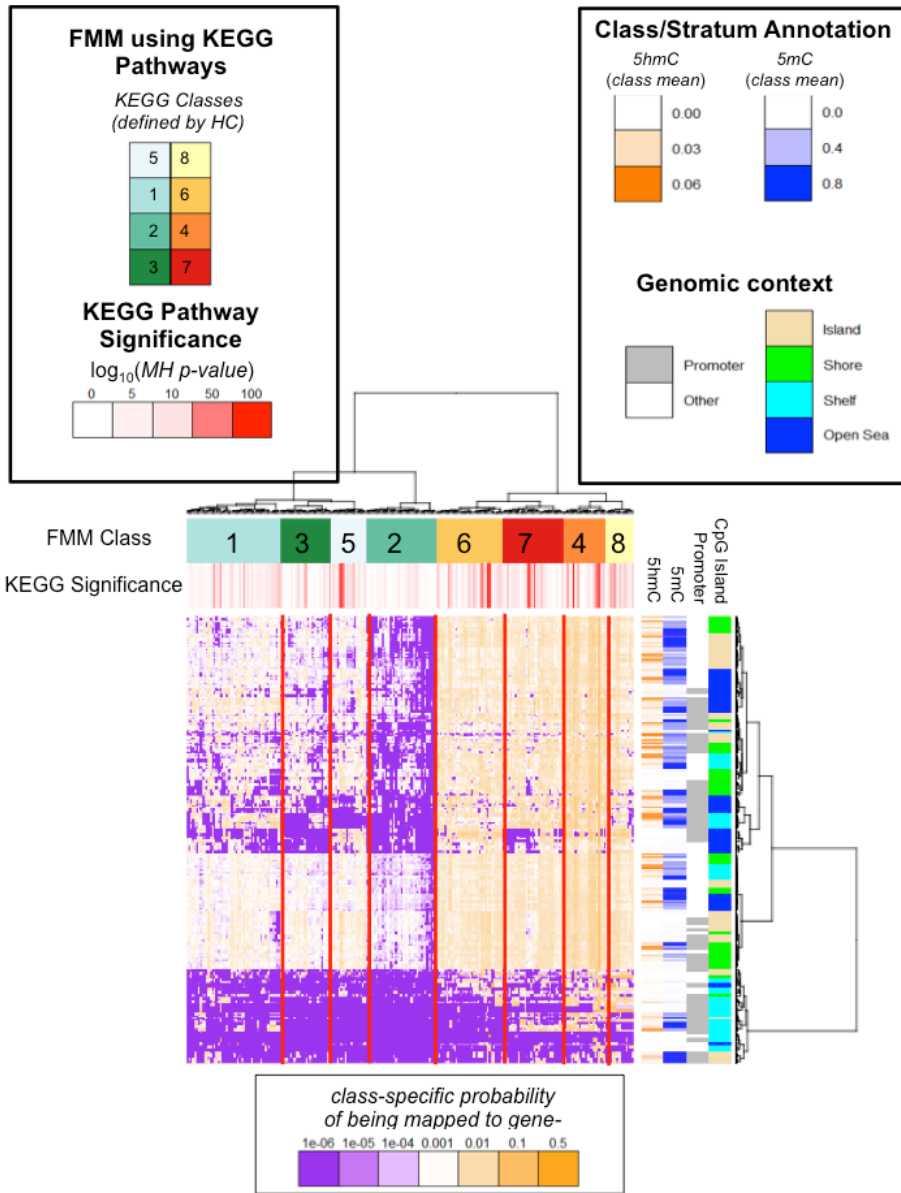
Supplementary Figure 3. (A) Clustering heatmaps of cell proportion matrix for glioblastoma data set (purple intensity indicates cell proportion). **(B)** Putative cell-type 1 (source of greatest explained variation) was not significantly associated with proportion of 5hmC in total cytosine content ($P > 0.05$, linear regression).

Supplementary Figure 4

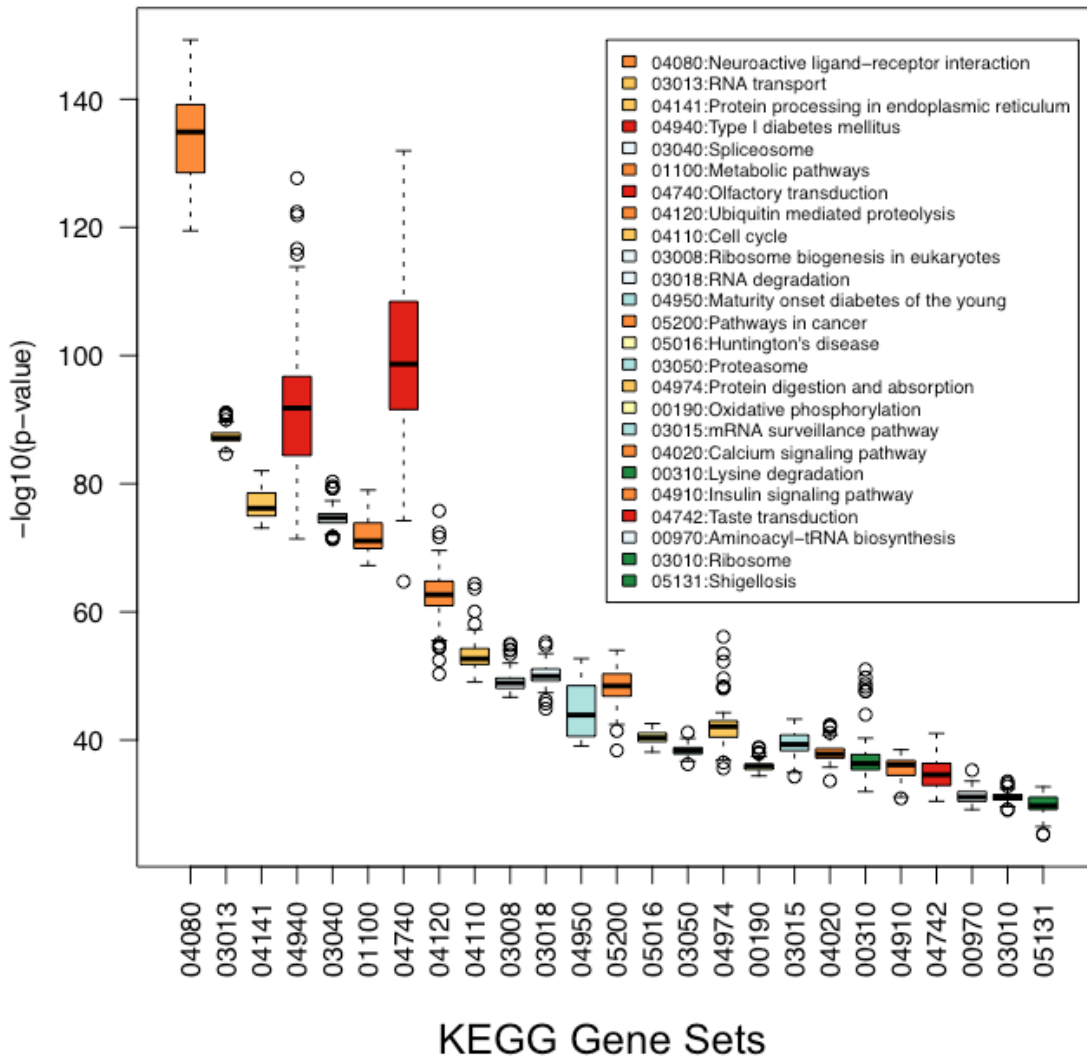


Supplementary Figure 4. CpG-specific mean 5hmC values were calculated across the 450K array and ordered by increasing magnitude. The CpGs with the highest one-percent 5hmC values were termed high 5hmC CpGs (red points).

Supplementary Figure 5B

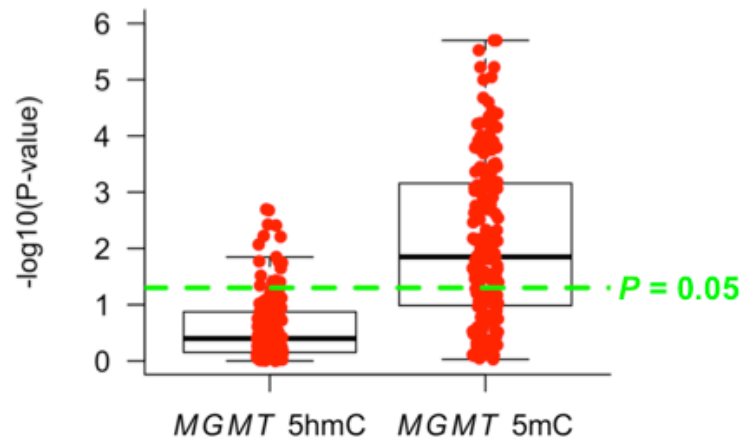


Supplementary Figure 5C



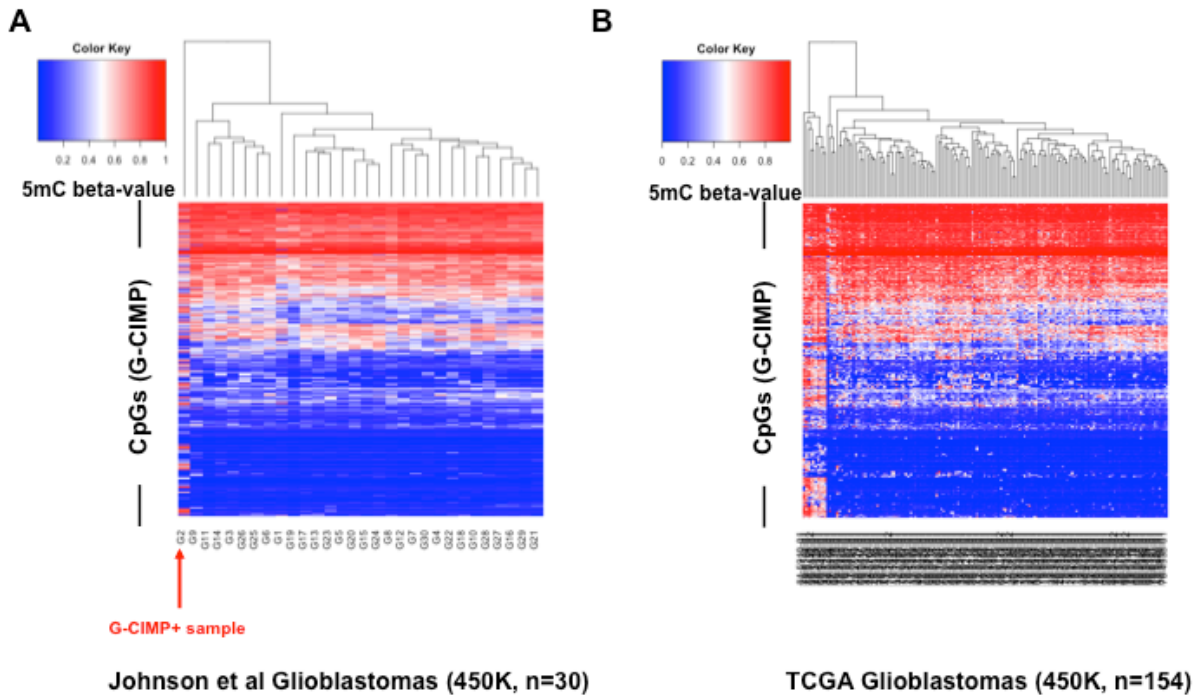
Supplementary Figure 5. (A) Analysis strategy for agnostic consensus clustering approach. **(B)** Consensus clustering using KEGG gene-sets to determine functional relevance of 5hmC and 5mC patterns in glioblastoma. **(C)** Boxplots of top-associated gene-sets.

Supplementary Figure 6



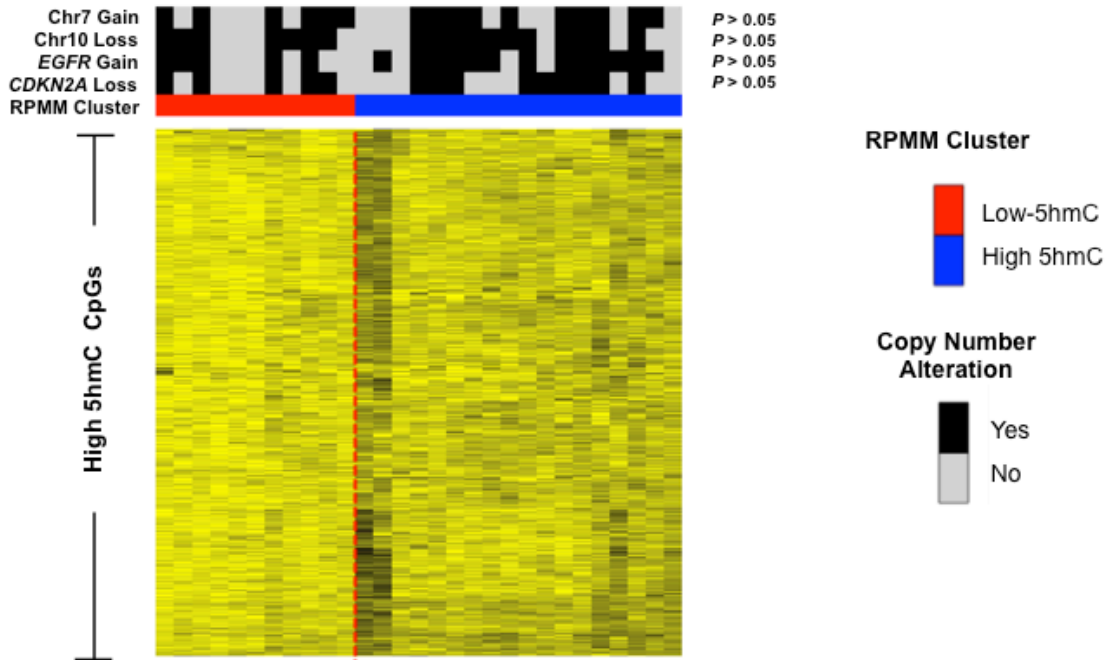
Supplementary Figure 6. Comparison of significance tests (Spearman's rho test) for CpG-specific 5hmC and 5mC values and *MGMT* gene expression (n=176 CpGs).

Supplementary Figure 7



Supplementary Figure 7. Identification of non-IDH-mutant Glioma-CpG Island Methylator Phenotype (G-CIMP) tumor sample in glioblastoma cohort (n=30). **(A)** Heat map of 5-methylcytosine values and unsupervised hierarchal clustering of CpGs from G-CIMP genes identified in Noushmehr et al⁸. High levels of methylation, that is outlier DNA methylation, at these genes is suggestive of a G-CIMP phenotype for the sample in the furthest left portion of the dendrogram **(B)** Unsupervised hierarchal clustering of CpGs from G-CIMP genes in the TCGA glioblastoma data set (n=154). High levels of DNA methylation at G-CIMP genes are present in the far left branch of the dendrogram.

Supplementary Figure 8



Supplementary Figure 8. Recursively Partitioned Mixture Model (RPMM) of tumor samples based on highest 5hmC CpGs (n=3,876 CpGs). In the heat map, each row represents a single CpG and each column represents a single tumor sample. Present or absence of copy number alterations for chromosome 7 gain, chromosome 10 loss, *EGFR* gain, and *CDKN2A* loss were not associated with cluster membership ($P > 0.05$, Fisher’s exact test).

SUPPLEMENTARY TABLES:

Supplementary Table 1. Correlations between 5hmC levels and gene expression of epigenetic enzymes

Gene	Spearman Correlation Coefficient	Spearman P-value
<i>DNMT1</i>	-0.106	0.630
<i>DNMT3A</i>	0.272	0.209
<i>DNMT3B</i>	-0.190	0.385
<i>IDH1</i>	-0.302	0.161
<i>IDH2</i>	0.019	0.933
<i>TET1</i>	0.158	0.469
<i>TET2</i>	-0.069	0.754
<i>TET3</i>	0.151	0.489

**Supplementary Table 2.
Genes actively transcribed
and with alternative mRNA
splicing are characterized
by 5-
hydroxymethylcytosine in
glioblastoma.**

Gene transcription levels (RNAseq, n=172 TCGA glioblastoma)

	Gene transcription levels (TCGA, n=172)			P-value
	Low	Moderate	High	
All Genes on 450K array (n=20,621 genes)	6,397 (32.6 %)	6,582 (33.6 %)	6,621 (33.8 %)	
Genes with 5hmC sites (n=2,121 genes)	174 (8.1 %)	785 (36.7 %)	1,183 (55.2 %)	5.20E-139

*Pearson's Chi_Squared test

Supplementary Table 3

TCGA glioblastoma-specific splicing events (RNAseq, n=160 TCGA glioblastoma)

Alternative Transcription Events	All Genes on 450K* array (n=20,621 genes)	Genes with 5hmC sites (n=2,121 genes)	Enrichment (95 % CI)	P-value
Exon Skip (ES)	5,008 (24.3 %)	837 (39.5 %)	2.03 (1.85 - 2.23)	2.23E-48
Alternate Donor Site (AD)	1,569 (7.6 %)	179 (8.4 %)	1.12 (0.95 - 1.31)	1.70E-01
Alternate Acceptor Site (AA)	1,821 (8.8 %)	269 (12.7 %)	1.50 (1.30 - 1.72)	2.36E-08
Retained Intron (RI)	1,258 (6.1 %)	123 (5.8 %)	0.95 (0.78 - 1.15)	6.30E-01
Mutually Exclusive Exons (ME)	96 (0.4 %)	22 (1.0 %)	2.24 (1.33 - 3.60)	2.00E-03
Alternate Terminator (AT)	2,327 (11.1%)	353 (16.6 %)	1.57 (1.38 - 1.78)	3.44E-12
Alternate Promoter (AP)	2,580 (12.5 %)	483 (22.8 %)	2.06 (1.84 - 2.30)	9.75E-35

* Genes on the 450K array with CpGs that passed QC (n=387,617). The level of enrichment was determined by Fisher's exact test by testing binary comparisons (e.g., gene region of interest vs. others)