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Supplementary Figure 1: H2AK119u1 and H3K27me3 domains at pericentric
 heterochromatin in *Aebp2<sup>tr/tr</sup>*, *Jarid2* KO and complemented *Jarid2* KO mESCs.

H2AK119u1 in  $Aebp2^{WT}$  and  $Aebp2^{tr/tr}$  (**A**) and *Jarid2* WT and KO (**B**) is observed at the pericentric heterochromatin (PCH) upon transient expression of MBD-RPCD. Arrowhead indicates a PCH focus. Scale bar is 5 µm. (**C**) Transient expression of MBD-Ezh2 in *Jarid2* WT and KO mESCs and immunofluorescence staining for H3K27me3. In both mESCs lines H3K27me3 is visualised at PCH (arrowhead). Scale bar is 5 µm. (**D**) Quantification of cells in figure (**C**). Error bars display standard deviation. A minimum of 300 cells were counted in 3 biological repeats.



Supplementary Figure 2: Characterisation of Jarid2 KO mESCS in Dnmt1<sup>F/F</sup> 32 33 background. (A) Immunoblot of whole cell extracts from mESCs showing depletion of 34 JARID2 in the Jarid2 KO line. Asterisks indicate species cross-reacting with the antibody. 35 (B) Southern blot showing loss of DNA methylation at major satellite repeats upon tamoxifen induction of Dnmt1<sup>F/F</sup> mESCs. Genomic DNA from control mESCs :WT and TKO (Dnmt1<sup>-/-</sup>, 36 Dnmt3a<sup>-/-</sup> Dnmt3b<sup>-/-</sup>) or test mESCs : Dnmt1<sup>F/F</sup> Jarid2 WT or KO, plus or minus tamoxifen (to 37 induce deletion of Dnmt1) was digested with the methylation-sensitive HpyCH41V and the 38 39 blot was probed for major satellite repeats. (C) Immunoblot of histone extracts of Dnmt1<sup>F/F</sup> 40 Jarid2 WT or KO probed for H3 and H3K27me3. (D) Immunofluorescence staining of Dnmt1<sup>F/F</sup> Jarid2 WT and Dnmt<sup>F/F</sup> Jarid2 KO mESCs for H2AK119u1 and H3K27me3 either 41 42 without tamoxifen induction (left) or with 12 days tamoxifen induction (right). Arrowhead 43 indicates a pericentric heterochromatin focus. (E, F) Transient expression of MBD-Ezh2 in Dnmt1<sup>F/F</sup> Jarid2 WT and KO mESCs and immunofluorescence staining for H3K27me3 (E). 44 45 quantified in graph (F). In both mESC lines H3K27me3 is visualised at PCH (arrowhead). (G) Stable EZH2-GFP mESC lines were created in both Dnmt1<sup>F/F</sup> Jarid2 WT and Dnmt<sup>F/F</sup> 46 47 Jarid2 KO backgrounds and localisation to PCH was assessed by GFP fluorescence (left) 48 and quantified (right). P value Student's t-test, unpaired. Scale bars represent 5 µm. 49 Arrowheads indicate a PCH domain. In all experiments a minimum of 300 cells were 50 counted in 3 biological repeats. Error bars indicate standard deviation.





54 Supplementary Figure 3: Deletion of Jarid2 diminishes Xist RNA dependent 55 recruitment of PRC2. (A) The mESC cell line BglXist1 has a doxycyclin-inducible Xist 56 transgene which contains BgIG stem loops on an autosome. The binding of BgI-mCherry 57 identifies the Xist coated chromosome. (B) Immunoblot showing levels of the indicated

58	proteins in the Jarid2 KO line. (C) Immunoblot showing levels of the indicated proteins in
59	Suz12 and Atrx KO cell lines. (D) Immunofluorescence for H3K27me3 and PRC2
60	components EZH2 and SUZ12 in WT, Jarid2, Suz12 and Atrx KO mESCs after induction of
61	Xist. Arrowhead indicates an inactive chromosome focus. (E) Quantification of foci observed.
62	A minimum of 300 cells were counted in 3 biological repeats. Error bars indicate standard
63	deviation.



$$\bigcirc$$
 Jarid2<sup>flox/flox</sup> Zp3::Cre<sup>+</sup> **X**  $\bigcirc$  Jarid2<sup>flox/ko</sup> — F1 50%  $\Delta$ m/flox  
= full ko

b





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Supplementary Figure 4: JARID2 absence disrupts EED recruitment to the inactive X
during preimplantation development. (A) Schematic of mating scheme to generate
complete Jarid2 knockout embryos. (B) Immunofluorescence and Xist RNA FISH of
heterozygote maternal Jarid2 KO/paternal Jarid2<sup>F/F</sup> (mko/flox) embryos at late
morula/blastocyst stage. JARID2 (grey) and EED (green) recruitment on Xist-coated
chromosomes (red; white arrow). (C) As (B), but homozygous Jarid2 KO embryos

- 73 (maternal Jarid2 KO/paternal Jarid2<sup>-/-</sup> (mko/ko). In (B) and (C), lower panel indicates zoom
- from the selected area in upper panel. DAPI is counterstained in blue. Scale bar is 10  $\mu$ m.



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78 Supplementary Figure 5: Absence of JARID2 disrupts PRC2 recruitment to the 79 inactive X during preimplantation development. (A) Immunofluorescence and Xist RNA FISH of heterozygote maternal Jarid2 KO/paternal Jarid2<sup>F/F</sup> (mko/flox) embryos at blastocyst 80 81 EED (green) recruitment and H3K27me3 enrichment (grey) on Xist-coated stage.

82 chromosomes (red; white arrow) are shown. Lower panel indicates zoom from the selected 83 area in upper panel. DAPI is counterstained in blue. Scale bar is 10 µm. (**B**) As (**A**), but with 84 homozygous *Jarid2* KO embryos (maternal *Jarid2* KO/paternal *Jarid2<sup>-/-</sup>* (mko/ko)). (**C**) % of 85 *Xist*-coated chromosomes counted from embryos as in (**A**) or (**B**). n represents numbers of 86 blastomere cells counted. 87

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90 Supplementary Figure 6: The N-terminus and UIM of Jarid2 complement PRC2 91 recruitment to H2AK119u1 chromatin. (A) Immunoblot of whole cell extracts of 92 complemented Jarid2 KO cell lines normalised for H3. (B) Immunoblot of reciprocal co-93 immunoprecipitation of HA-tagged JARID2 fragments and EZH2. (**C**) Co-94 immunoprecipitation of EZH2 with HA-tagged Jarid2 44-512 and Jarid2 K116R (D) 95 H2AK119u1 is observed at the PCH upon expression of MBD-RPCD in complemented

96 Jarid2 KO mESCs. (E) H3K27me3 is observed at the PCH after complementation with 97 deletion constructs containing the N-terminal region, but not with a construct containing the 98 C-terminus or with the construct JARID2-44-541. Quantification is shown in Fig. 3B. Scale 99 bar is 5 μm. (F) Examples illustrating analysis of Xist dependent EZH2 recruitment in WT, 100 Jarid2 null and two independent cell lines complemented with JARID2 1-541 or JARID2 44-101 541. Quantification is shown in Fig. 3E. Examples for Jarid2 null mESCs illustrate absent 102 (left panel) or weak (right panel) EZH2 staining. Arrowheads indicate a position of Xist 103 domain in all panels. Scale bars represent 5 µm.

а



1: Jarid2 KO 2: Jarid2 KO + WT 3: Jarid2 KO + 121-1234 4: Jarid2 KO + 542-1234 5: Jarid2 KO + 1-583 6: Jarid2 KO + 1-542

**b** anti-HA 30 s exposure







#### anti-Ezh2 10 s exposure



1: Jarid2 KO 2: Jarid2 KO + 1-583 3: Jarid2 KO + 542-1234 4: Jarid2 KO + 1-512 5: Jarid2 KO + 44-542 6: Jarid2 KO + 44-512 7: Jarid2 KO + FL K116R

- 106 Supplementary Figure 7: Full Immunoblots. (A) Complete immunoblots for the
- 107 experiment shown in Supplementary Figure 6b. (B) Complete immunoblots for the
- 108 experiment shown in Supplementary Figure 6c. The red boxes indicate the regions of the
- 109 blots that have been cropped.
- 110



staining of polyacrylamide gel of histone octamers reconstituted with WT H2A, H2AK119u1

and H2AK119u1I44A. Molecular weight markers are as indicated on the left. (C) Western

117 blot of JARID2-1-530 purified after expression in insect cells. Molecular weight markers are

- 118 as indicated on the left.
- 119

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112

113

WT Cells

JARID2 KO Cells

anti-JARID2







#### anti-RYBP











anti-H4

- 1. Input
- 2. Beads only
- 3. Unmodified Nuc
- 4. H2AK119u1 Nuc
- 5. H2AK119u1 Ub I44A Nuc
- 6. Input
- 7. Beads only
- 8. Unmodified Nuc
- 9. H2AK119u1 Nuc

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### 122 Supplementary Figure 9: Full Immunoblots for the experiment shown in Figure 4a.

# **Supplementary Table 1**. Oligonucleotides used in this study

Cloning primer	Sequence 5'- 3'
J2F1	GTCGACATGAGCAAGGAAAGACCCAAGAGG
J2F121	GTCGACATGCAGCCGAATAGTCCCAGCAC
J2F542	GTCGACATGAAGGGGAGCGGCAAGTCTGGG
J2R1234	GCGGCCGCTCAGGCATAGTCAGGCACGTCATAAGGATAAAC
	CTTTCTCTTCTTTTTGGTGAGGATGGGAGCCGAGATGG
J2R583	GCGGCCGCTCAGGCATAGTCAGGCACGTCATAAGGATAAAC
	CTTTCTCTTCTTTTTGGCCCGTACTTCTCCACCTG
J2R541	GCGGCCGCTCAGGCATAGTCAGGCACGTCATAAGGATAAAC
	CTTTCTCTTCTTTTTGGCTCTGGCTTGCCCTGTGGG
sgRNA target sites	Sequence (PAM in bold)
Jarid2 (exon 3)	ATTTTGAAGAAGGGCCGTCG <b>AGG</b>
Atrx (exon 9)	ATGTCTTCTGGAACCGAGGAAGG
Suz12 (exon 7)	CCAATAAGACAAGTCCCTAC <b>TGG</b>
KO validation primer	Sequence
JARID2_HRMA_F2	CCCTTTTCTCACCTGTAGGG
JARID2_HRMA_R2	GATTCACACTTGCTCCCATGT
ATRX_HRM_F3	AGCTCTCCAGTTCTGGCTCAGTC
ATRX_HRM_R3	TAGTGGGGAACATAAGGGTTCAGG
SUZ12_HRM_F2	TCAATCTTTTCTTTGCCAGGAT
SUZ12_HRM_R2	CCAATATTTCATTGGTTTCTCC
EZH2_HRM_R2	GCAAAAGATGGAGAATGTCTAAGGA
Genotyping primer	Sequence
JD2-5ARM-F	CCTTTATCTAGAACAGTAGTTCTCAGCC
JD2-3ARM-R	CACTCACTGGGTAGATTGATTACATAC
JD2-CASS-R	AGGAATGCCCAGCCAAAATC
FLP-F	CCCATTCCATGCGGGGTATCG
FLP-R	GCATCTGGGAGATCACTGAG
CRE-F	GCCTGCATTACCGGTCGATGCAACGA
CRE-R	GTGGCAGATGGCGCGGCAACACCATT