

# Supporting Information

## Supplementary Figures

Incomplete and delayed Sox2 deletion defines residual ear neurosensory development and maintenance

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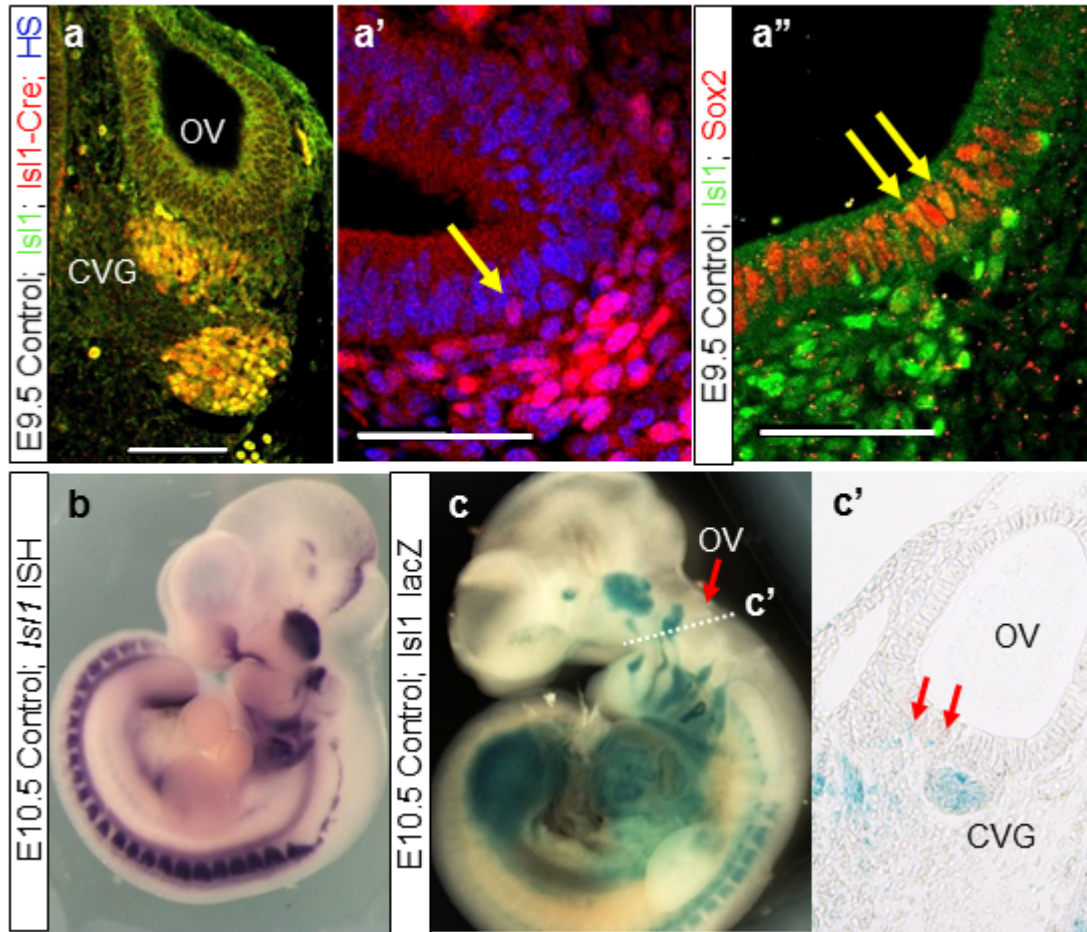
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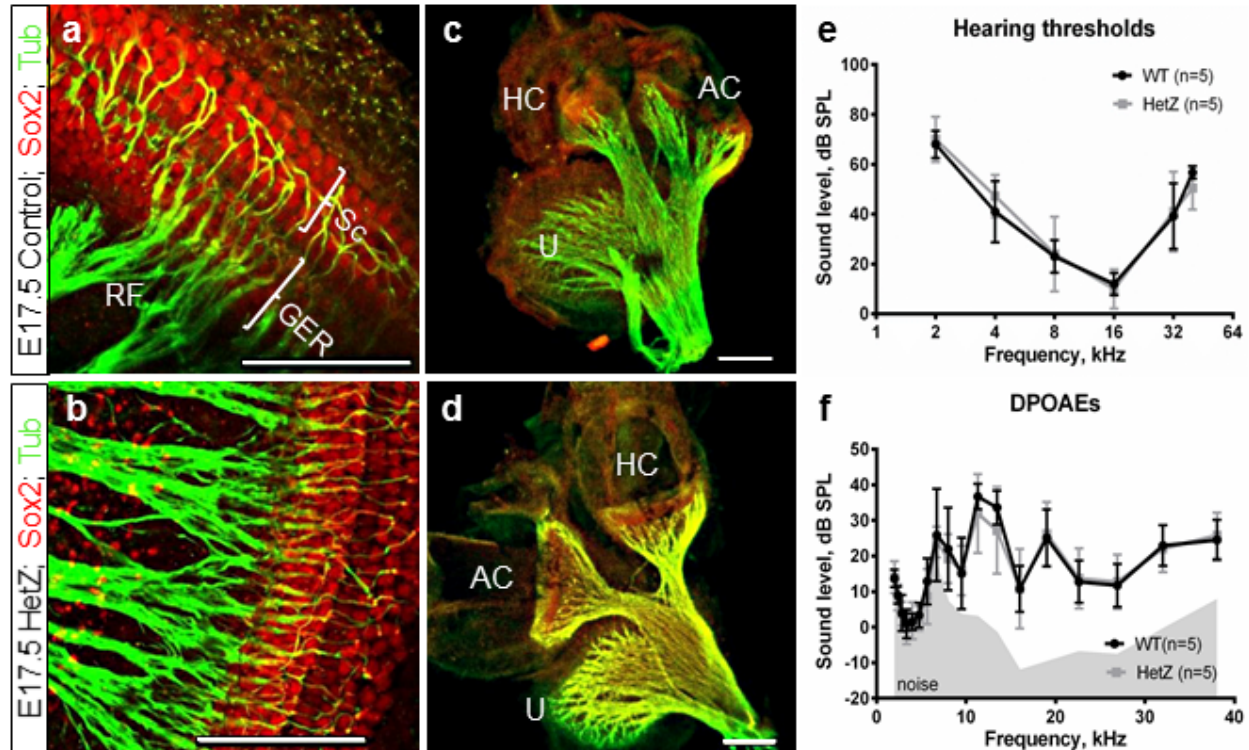
#Indicates co-senior author

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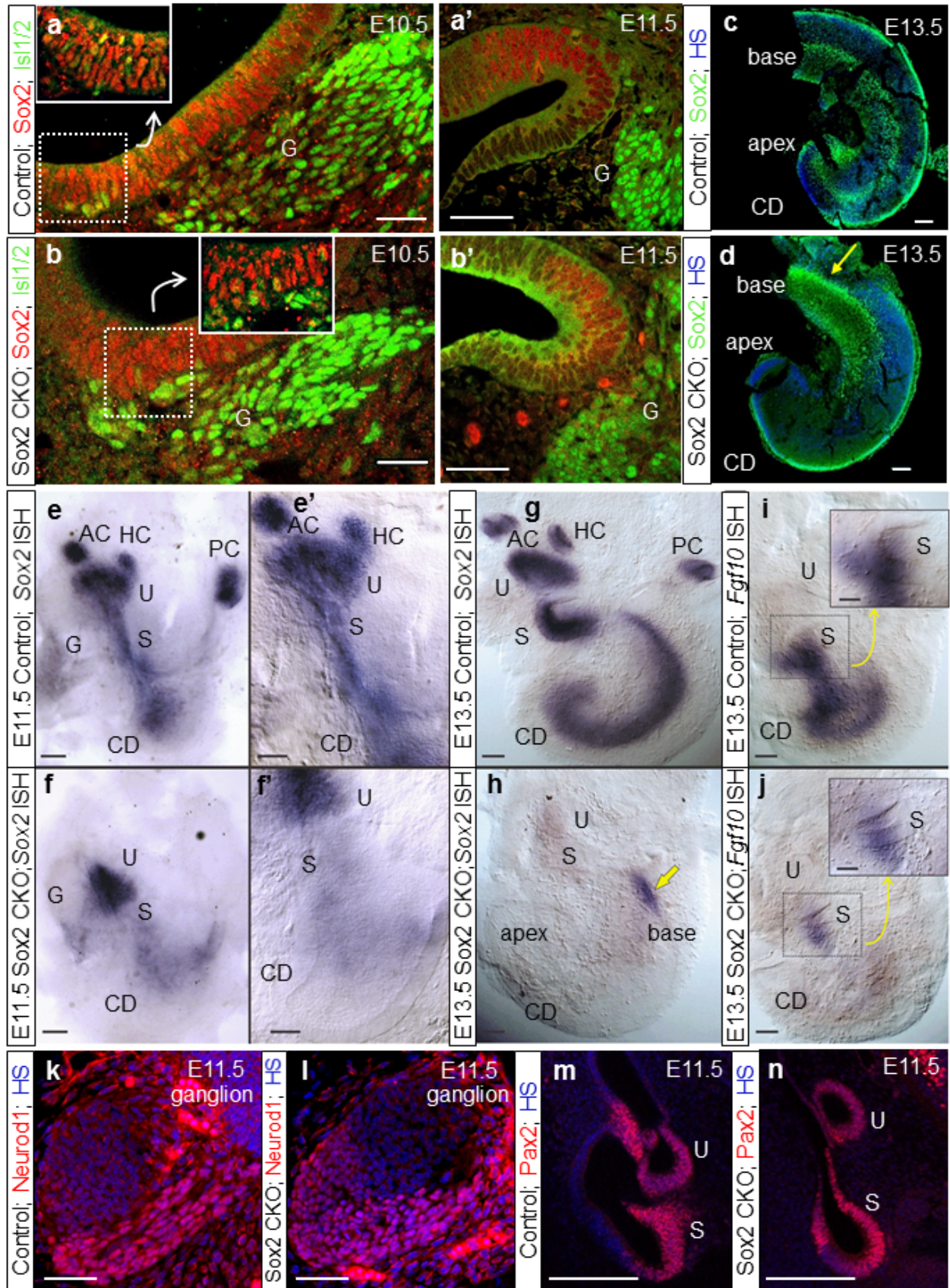


**Supplementary Figure 1. *Isl1*-cre recombination corresponds to the expression of *Isl1* in the otocyst.** (a) At E9.5, the expression of *Isl1*-cre correlates with the expression of *Isl1* protein (green). (a') *Isl1*-cre (red) is detected in the delaminated cochleovestibular (CVG) neurons and also within the otic epithelium (arrow). (a'') *Isl1* (green) is co-expressed with *Sox2* (red) in the otic neuroepithelium at E9.5 (arrow). (b) *Isl1* mRNA is expressed in the progenitor cells of sensory epithelium and in otic ganglion at E10.5. Cre-mediated  $\beta$ -gal reporter expression shows Cre recombination in the tissues corresponding to the *Isl1*<sup>+</sup> domain in E10.5 embryos (c, c'). OV, otic vesicle; CVG, cochleovestibular ganglion. Scale bar: 100  $\mu$ m.



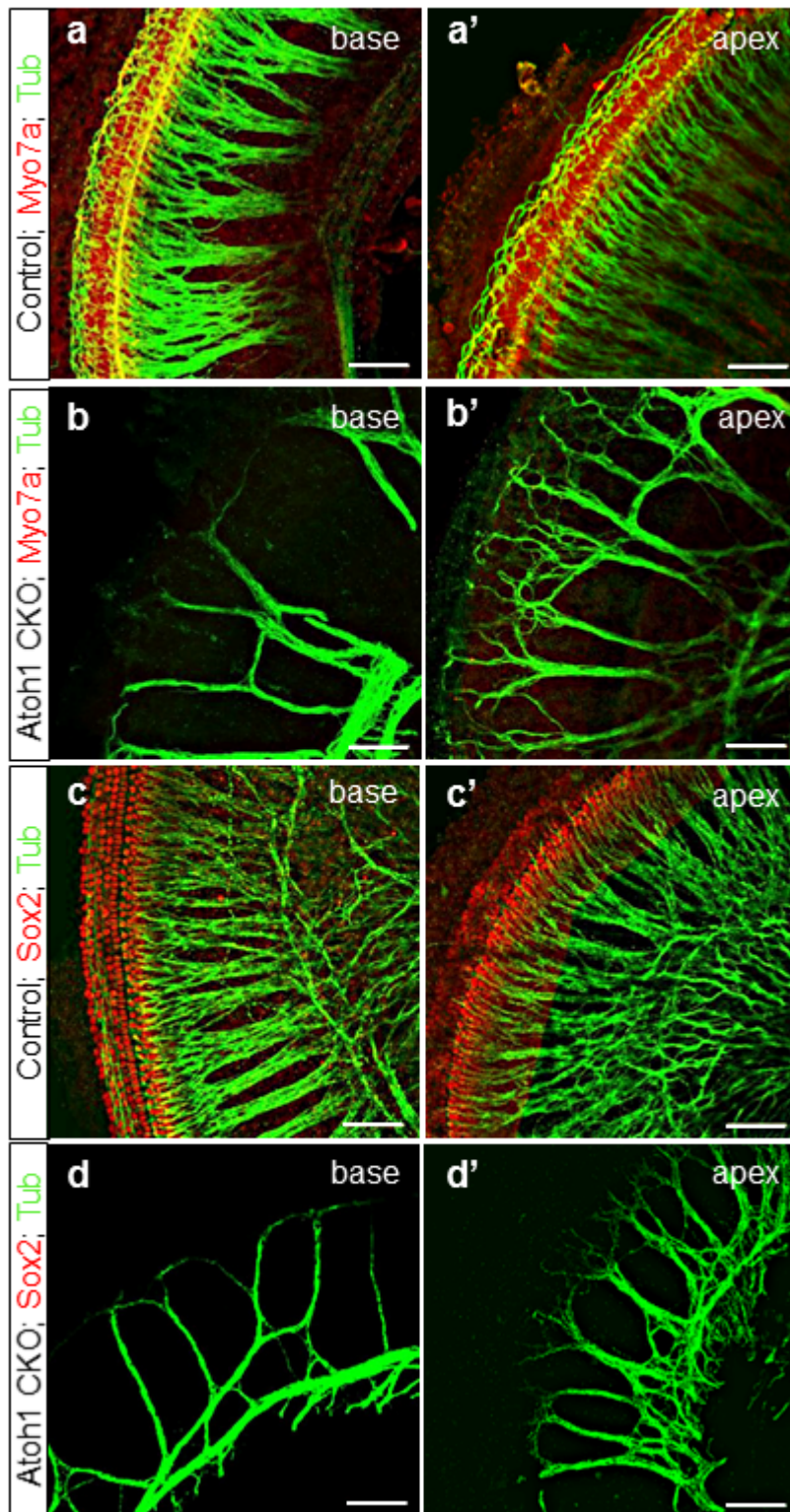
**Supplementary Figure 2. The development of the inner ear sensory organs of heterozygous (*Hetz*) *Isl1-cre/+;Sox2<sup>f/+</sup>* mutant is not altered. (a-d) *Sox2* expression (red) and the pattern of innervation (tubulin, green) in the sensory organs of the inner ear is comparable between heterozygous *Isl1-cre/+;Sox2<sup>f/+</sup>* (*Hetz*) and wild type littermate controls. (e, f) Hearing functions of *Hetz* are similar compared to wild type littermates as shown by hearing thresholds (ABRs) and deteriorated distortion product otoacoustic emissions (DPOAEs). Results of hearing tests of 1 month old mice are presented as means  $\pm$  SD. Scale bars: 50  $\mu$ m (a, b), 100  $\mu$ m (c, d). AC, anterior crista; HC, horizontal crista; GER, greater epithelial ridge; RF, radial fibers; Sc, supporting cells; U, utricle.**





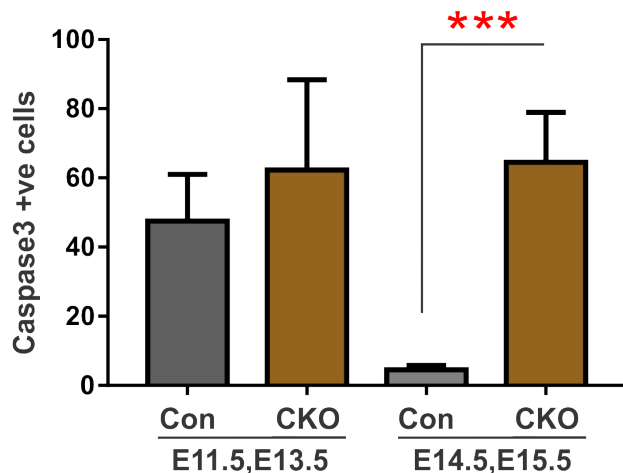
**Supplementary Figure 3. Gene expression pattern is effectively altered in Sox2 CKO mutants.** Reduced overlap of Isl1 and Sox2 immunostaining is apparent in Sox2 CKO ears at E10.5 (**a, b**). Some Sox2 protein can still be detected in the inner ear up to E11.5 (**a', b'**). At E13.5, Sox2 is expressed only in the base of the Sox2 CKO cochlea, whereas in the controls, the expression of Sox2 (green) occurs throughout the entire length of the cochlea (**c, d**). The Sox2 mRNA reduction is apparent in the utricle, saccule and cochlear base at E11.5 with no expression in the semicircular canal cristae and in the apical half of the growing cochlear duct (**e-f'**). At E13.5, there is only a weak expression of Sox2 mRNA in the base of the cochlea compared to controls (**g, h**). No *Fgf10* mRNA could be detected in canal cristae and only limited *Fgf10* labeling is in saccular neurons of the Sox2 CKO (**i, j**). The expression of Neurod1 in the otocyst-derived ganglion neurons is comparable between Sox2 CKO and controls at E11.5 (**k, l**). The expression of Pax2 appears unaffected in the Sox2 CKO inner ear compared to controls at E11.5 (**m, n**). Scale bars: 100  $\mu\text{m}$ , except a and b that indicates 50  $\mu\text{m}$ . AC, anterior crista; CVG, cochleovestibular ganglion; CD, cochlear duct; G, ganglion; HC, horizontal crista; PC, posterior crista; U, utricle; S, saccule.



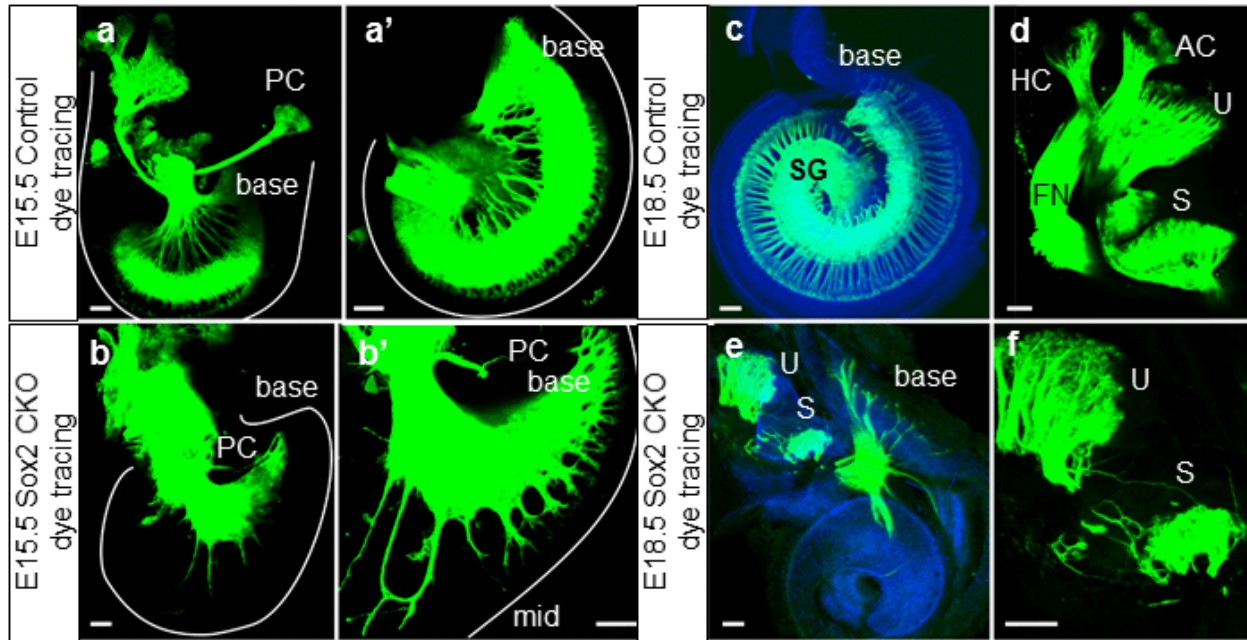


**Supplementary Figure 4.** *Isl1-cre;Atoh1<sup>fl/fl</sup>* (Atoh1 CKO) mutants show a complete loss of all HCs and Sox2 expression. *Isl1-cre* mediated recombination of floxed *Atoh1*

results in a complete loss of HCs in the cochlear base (**a, b**) and apex (**a', b'**), as indicated by the expression of HC marker, *Myo7a*. These data show a uniform action of *Isl1-cre* along the entire cochlea. Our analysis of *Isl1-cre*-mediated  $\beta$ -gal reporter expression showed *cre* recombination in the organ of Corti (OC) as well as in the greater epithelial ridge (GER) in the cochlea at P0. Accordingly, there is a complete loss of *Sox2* expression in OC and GER of *Atoh1* CKO mice (**c, c', d, d'**), suggesting that the absence of *Atoh1* or/and the organ of Corti causes the loss of *Sox2* expression in GER and supporting cells in the cochlea. Thus, neither the base nor the apex of these mutants show any *Myo7a* positive HCs (**b, b'**), suggesting that incomplete recombination of *Sox2* by *Isl1-cre* is not the explanation of the patchy retention of some *Myo7a* positive cells in *Isl1-cre;Sox2<sup>ff</sup>* mice. Scale bar, 50  $\mu$ m.

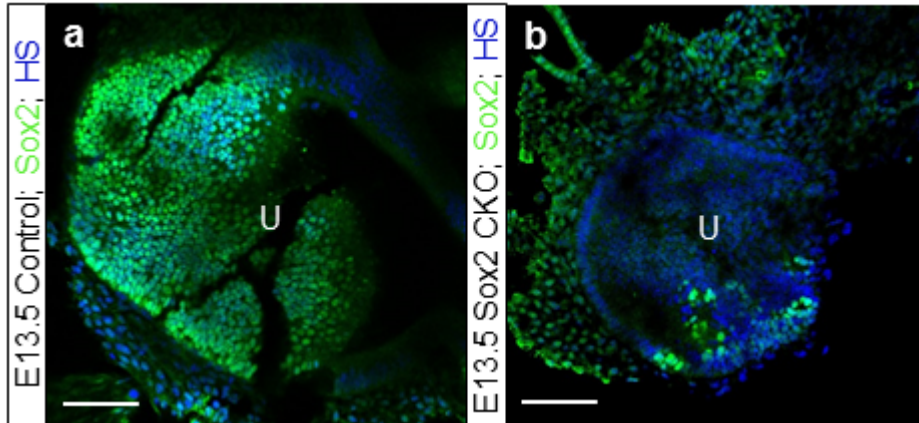


**Supplementary Figure 5. Caspase-3 mediated neuronal death massively progresses in the Sox2 CKO inner ear after E13.5 compared to controls.** The total number of Caspase3<sup>+</sup> cells was determined in the vestibular ganglia (E11.5-E13.5) and superior vestibular ganglia (E14.5 and E15.5) of the control (Con) and Sox2 CKO ear at indicated embryonic days. Caspase3 positive cells were quantified after whole mount immunostaining with anti-Caspase3 antibody using LAS AF Lite draw counter. The values represent means  $\pm$  SD (N = 4 individuals/group). \*\*\*P<0.0001, *t*-test.

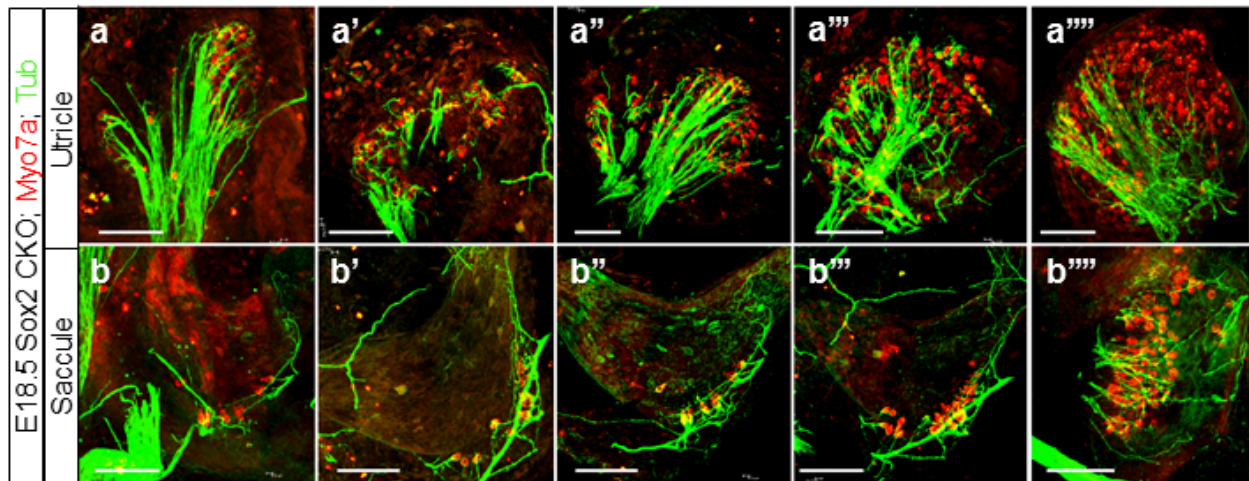


**Supplementary Figure 6. The pattern of innervation in the inner ear is profoundly changed in Sox2 CKO mutants.** The pattern of innervation differs dramatically between control and mutant littermates at E15.5 (**a, b**) and E18.5 (**c-f**). Spiral ganglion neurons (SG) are present only in the basal turn in mutants (**a, a', b, b'**) with only few fibers extending toward the middle turn (**a', b'**). The posterior vertical canal crista (PC) receives a prominent fiber track in control mice (**a**) but only few fibers extend some distance toward this epithelium in mutants (**b, b'**). Due to further reduction of SG by apoptosis only a small population remains near the basal tip of the cochlea in the mutant (**c, e**) and only limited innervation remains to the saccule (**d-f**). AC, anterior canal crista; HC, horizontal canal crista; PC, posterior canal crista; S, saccule; SG spiral ganglion neurons; U, utricle. Scale bars, 100  $\mu$ m.





**Supplementary Figure 7. Delayed deletion of Sox2 by Isl1-Cre leaves some Sox2<sup>+</sup> cells in the utricule.** Compared to controls (a), the size of Sox2<sup>+</sup> domain is substantially decreased in the Sox2 CKO utricule (b). U, utricule. Scale bars, 100  $\mu$ m.



**Supplementary Figure 8. Variation of HC numbers is detected in the Sox2 CKO.** Numbers of HCs and their distribution in the utricle (a-a''') and saccule (b-b''') vary between individual animals but are overall dramatically reduced compared to littermate controls. Scale bars, 100  $\mu$ m.

**Table S1. Quantification of HCs in the utricle, saccule and cochlea of E18.5 control and Sox2 CKO mice.**

|                | Control Utricle | Sox2 CKO Utricle | Control Saccule | Sox2 CKO Saccule | Control Cochlea* | Sox2 CKO Cochlea |
|----------------|-----------------|------------------|-----------------|------------------|------------------|------------------|
|                | 1,031           | 132              | 880             | 15               | 908              | 21               |
|                | 1,068           | 247              | 910             | 59               | 774              | 42               |
|                | 978             | 112              | 1,190           | 28               | 719              | 31               |
|                | 1,005           | 197              | 817             | 12               | 871              | 22               |
|                |                 | 88               |                 | 94               | 774              | 36               |
|                |                 | 334              |                 | 95               | 717              | 28               |
|                |                 |                  |                 |                  | 809              |                  |
| <i>Average</i> | <i>1,021</i>    | <i>185</i>       | <i>949</i>      | <i>51</i>        | <i>796</i>       | <i>30</i>        |
| <i>SD</i>      | <i>38</i>       | <i>94</i>        | <i>165</i>      | <i>38</i>        | <i>72</i>        | <i>8</i>         |

Myo7a positive HCs are quantified after whole mount immunostaining using LAS AF Lite draw counter to avoid counting error. The total number of HCs was determined in the entire utricle and saccule, and in the entire Sox2 CKO cochlea. \*The number of HCs in the control cochlea represents the total number of HCs in 1.5 mm of the base.