Supplementary Data for:

Structural basis of damage recognition by Thymine DNA Glycosylase

Christopher T. Coey^{1,§}, Shuja S. Malik^{1,§}, Lakshimi S. Pidugu¹, Kristen Varney^{1,2,3}, Edwin Pozharski^{1,2,3,*}, and Alexander C. Drohat^{1,2,*}

¹Department of Biochemistry and Molecular Biology, University of Maryland School of Medicine, Baltimore, MD 21201, USA, ²University of Maryland Marlene and Stewart Greenebaum Cancer Center, Baltimore, MD 21201, USA, ³Center for Biomolecular Therapeutics, Institute for Bioscience and Biotechnology Research, Rockville, MD 20850, USA

* To whom correspondence should be addressed. Tel. 410-706-8118; Email: adrohat@som.umaryland.edu. Tel: 240-314-6255; Email: <u>EPozharskiy@som.umaryland.edu</u>

	F·S TDG ⁸²⁻³⁰⁸ G·U ^F	F·P TDG ⁸²⁻³⁰⁸	F·S TDG ¹¹¹⁻³⁰⁸ G·U ^F
	(PDBID: 5HF7)	(PDBID: 5FF8)	(PDBID: 5JXY)
Data collection			
Space Group	C2	C2	C2
Cell Dimensions			
a, b, c (Å)	97.0, 52.8, 81.2	96.9, 52.5, 81.3	88.5, 53.3, 82.6
β (°)	95.2	95.2	95.6
Resolution (Å)	32.52-1.54 (1.57-1.54)	31.08-1.70 (1.73-1.70)	39.03-1.71 (1.74-1.71)
R _{pim}	0.035	0.057	0.052
Mean I/oI	9.3 (0.6)	7.9 (0.7)	12.9 (0.5)
CC _{1/2}	0.998 (0.35)	0.998 (0.50)	0.999 (0.224)
Completeness (%)	100.0 (100.0)	100.0 (99.9)	95.7 (37.0)
Redundancy	5.0 (4.7)	7.5 (7.4)	19.8 (7.3)
Wilson B-factor (Å ²)	28.2	29.0	24.6
Refinement			
Program	BUSTER-TNT	BUSTER-TNT	BUSTER-TNT
Resolution (Å)	32.51-1.54	32.48-1.70	24.97-1.71
No. of reflections	59281	44159	39815
$R_{ m work}/R_{ m free}$	0.186/0.216	0.194/0.217	0.197/0.231
Number of atoms			
Protein	1603	1596	1544
DNA	1123	1123	1164
Water	330	235	228
B-factors (Å ²)			
Protein	36.8	37.4	27.6
DNA	53.2	55.3	43.8
Water	50.2	43.3	40.6
Ramachandran Plot			
Favoured (%)	98.1	97.5	96.4
Allowed (%)	100	100	100
Outliers (%)	0	0	0
RMSD from ideal			
Bond lengths (Å)	0.010	0.010	0.008
Bond angles (°)	1.0	0.98	0.98

Supplementary Table S1. Data collection and refinement statistics

Values shown in parenthesis are for highest resolution shell. The Ramachandran analysis was performed using Molprobity [1]. Wilson B-factor estimated by phenix.xtriage. Number of atoms includes all atom records explicitly included in the model, including alternate positions.

1. Chen, V.B., W.B. Arendall, III, J.J. Headd, D.A. Keedy, R.M. Immormino, G.J. Kapral, L.W. Murray, J.S. Richardson, and D.C. Richardson, *MolProbity: all-atom structure validation for macromolecular crystallography*. Acta Crystallographica Section D, 2010. **66**(1): p. 12-21.



Figure S1. Alignments show that the previous $TDG^{111-308}$ -G·U^F structure (2.97 Å, yellow) differs significantly from the new $TDG^{111-308}$ -G·U^F structure (green), with a percentile-based spread (p.b.s.) of 0.51 Å (backbone; see main text). The previous $TDG^{111-308}$ -G·U^F structure (PDBID: 3UFJ) was obtained from crystals in which $TDG^{111-308}$ binds DNA with 2:1 stoichiometry, one subunit (yellow) at the G·U^F site and the other subunit (cyan) at a nonspecific site. The new $TDG^{111-308}$ -G·U^F structure reported here features 1:1 stoichiometry. The N-terminal helix observed for structures with 1:1 binding (green helix with no yellow counterpart) is likely destabilized by the TDG:TDG dimer interface for complexes featuring 2:1 (TDG:DNA) binding.



Figure S2. Biochemical studies of TDG⁸²⁻³⁰⁸ and its R110A variant. (**A**) Equilibrium binding of TDG⁸²⁻³⁰⁸ to DNA (10 nM) containing a G·T^F mismatch, monitored by EMSA. For reference, the far left lane shows binding of native TDG⁸²⁻³⁰⁸ (20 nM) to G·T^F DNA. (**B**) Single-turnover kinetics for excision of T from a G·T DNA substrate by wild-type TDG⁸²⁻³⁰⁸ and the R110A mutant at 22 °C (R110A-TDG⁸²⁻³⁰⁸ is unstable at 37 °C). The rate constants are $k_{max} = 0.108 \pm 0.006 \text{ min}^{-1}$ for TDG⁸²⁻³⁰⁸ and $k_{max} = 0.030 \pm 0.001 \text{ min}^{-1}$ for R110A-TDG⁸²⁻³⁰⁸, giving a 3.7-fold mutational affect.



Figure S3. Crystal structure of an E·S complex for TDG^{82-308} bound to G·U^F DNA (PDBID: 5HF7), solved at 1.54 Å resolution, focusing on N-terminal residues. TDG is in cartoon format (blue) with some residues in stick format (C, white; N, blue; O, red). The $2F_0$ - F_c electron density map, contoured at 1.0 σ , is shown in yellow for residues 107 to 110 of TDG. DNA is shown in space and stick formats, with the dUrd-containing strand yellow and the complementary strand green. Water molecules are shown as red spheres and dashed lines represent hydrogen bonds.



Figure S4. ¹⁵N-HSQC NMR spectra for TDG⁸²⁻³⁰⁸ (black peaks) and TDG¹¹¹⁻³⁰⁸ (red). Four resonances that are nearly identical in both spectra (marked with *) likely represent disordered C-terminal residues (see main text). Resonances in the upper right (box with dotted lines) are from side chain amino groups. The NMR samples contained TDG⁸²⁻³⁰⁸ or TDG¹¹¹⁻³⁰⁸ in 0.02 M sodium phosphate pH 6.5, 0.15 M NaCl, 0.2 mM EDTA, 0.2 mM DTT, 7% D₂O. Experiments were collected at 18 °C on an 800 MHz NMR spectrometer.



Figure S5. The 2'-fluoroarabino-dUrd analog flipped into the TDG⁸²⁻³⁰⁸ active site exhibits a C1'-exo-O4'-endo sugar pucker. The DNA is shown in stick format colored yellow (cyan for 2'-F), with a $2F_0$ - F_c map contoured at 1.0 σ . Note that the sugar is oriented such that C2' and C3' are aligned, with the bond vector pointing toward the viewer and C3' in the foreground.



Figure S6. Structure of an E·S complex for TDG⁸²⁻³⁰⁸ bound to G·U^F features a solvent-filled channel from the active site to the enzyme surface, along the target DNA strand. TDG⁸²⁻³⁰⁸ is shown in space-filling and cartoon modes, the DNA is in stick format, with the target strand in yellow, the dUrd nucleotide cyan, and the complementary DNA strand in green. The $2F_{o}$ - F_{c} map, contoured at 1.0 σ , is shown in blue for the target DNA strand. Water molecules are shown as red spheres.



Figure S7. TDG contacts the Gua ("opposing G") of the G·U mispair. TDG⁸²⁻³⁰⁸ is shown in surface representation with residues that contact the opposing G shown as sticks. The Uracontaining DNA strand is yellow and the complementary strand green, and water molecules are red spheres. Dashed lines represent hydrogen bonds, with interatomic distances (Å). The backbone oxygen of Ala²⁷⁴ contacts the opposing Gua at both N1H (2.7 Å) and N2H₂ (2.8 Å), and the Gua is also contacted at N2H₂ by the backbone oxygen of Pro²⁸⁰ (3.0 Å).