

Supporting Information Appendix

Tethered IL-15 augments antitumor activity and promotes a stem-cell memory subset in tumor-specific T cells

Lenka V. Hurton^{1,2}, Harjeet Singh¹, Amer Najjar³, Kirsten Switzer¹, Tiejuan Mi¹, Sourindra Maiti¹, Simon Olivares¹, Brian Rabinovich¹, Helen Huls^{1,4}, Marie-Andrée Forget^{1,5}, Vrushali Datar³, Partow Kebriaei⁶, Dean A. Lee¹, Richard E. Champlin⁶, Laurence J.N. Cooper^{1,7}

¹Division of Pediatrics, The University of Texas MD Anderson Cancer Center, 1515 Holcombe Blvd., Houston, Texas, 77030, USA

²The University of Texas Graduate School of Biomedical Sciences, 6767 Bertner Ave., Houston, Texas, 77030, USA

³Cancer Systems Imaging, The University of Texas MD Anderson Cancer Center, 1515 Holcombe Blvd., Houston, Texas, 77030, USA

⁴Intrexon, 20358 Seneca Meadows Pkwy, Germantown, Maryland, 20876, USA

⁵Melanoma Medical Oncology, The University of Texas MD Anderson Cancer Center, 1515 Holcombe Blvd., Houston, Texas, 77030, USA

⁶Stem Cell Transplantation and Cellular Therapy, The University of Texas MD Anderson Cancer Center, 1515 Holcombe Blvd., Houston, Texas, 77030, USA

⁷ZIOPHARM Oncology, Inc., One First Ave., Parris Building 34, Navy Yard Plaza, Boston, Massachusetts, 02129, USA

SI Appendix Materials and Methods

Plasmid Design. mbIL15: Alternate codon usage and signal peptides can increase expression of IL-15. Therefore, signal peptides for IL-15 and IL-15Ra were omitted and the IgE signal peptide (gb|AAB59424.1) was used for the mbIL15 fusion protein. The mbIL15 construct fuses the human codon-optimized (hCoOp) GeneArt (Regensburg, Germany) IL-15 cDNA sequence (NM_000585.4) to the full-length IL-15Ra cDNA sequence (NM_002189.3) via a 26 amino acid glycine-serine linker [SG₃(SG₄)₃SG₃SLQ] and has a C-terminal (FLAG)₃ epitope tag (DYKDDDDK, repeated three times). The DNA plasmid expressing IL15-IL15Ra-FLAG_pMK-RQ was synthesized by GeneArt. IL15-IL15Ra-FLAG_pMK-RQ and GlySer-EGFP-mIgG1/pSBSO were digested with *NheI* and *XhoI* to liberate the mbIL15 insert and pSBSO Sleeping Beauty (SB) vector backbone, respectively, to generate IL15-IL15Ra-Flag/pSBSO. Alternatively, a clinically relevant design of mbIL15 (mIL15-IL15Ra(hCoOp)/pSBSO-FRA) was used in the established leukemia mouse model along with the *CAR. The clinical mbIL15 insert consisted of all of the same components as the original mbIL15 but for the removal of the FLAG tag. The clinical mbIL15 insert was codon-optimized and synthesized by GeneArt. Vector 13ABPHSP mIL15-IL15Ra/pMK-T and clinical vector CD19R-CD28Tm-41BBCyt-Z(hCoOp)/pSBSO-FRA were digested with *NheI* and *XhoI* to liberate the mbIL15 insert and pSBSO SB vector backbone, respectively to generate the clinical mIL15-IL15Ra(hCoOp)/pSBSO-FRA plasmid.

SB11: The hyperactive transposase (SB11) is under the control of a cytomegalovirus (CMV) promoter in the pKan-CMV-SB11 SB plasmid (1, 2).

CARs: Generation of the SB transposon encoding the second generation CD19-specific CAR (CD19RCD28/pSBSO) under control of the human elongation factor-1 α promoter has been described (1, 2) and is herein referred to as ‘CAR’.

A CD19-specific CAR co-expressing *ffLuc*, designated CAR-*ffLuc*, (CD19RCD28-T2A-Flag-*ffLuc*/pT2SBSO) was used for *in vivo* monitoring of T cells by bioluminescent imaging (BLI). The hCoOp CAR and firefly (*Photinus pyralis*) luciferase reporter genes, linked via a self-cleaving 2A peptide from the *Thosea asigna* virus, were derived from the previously made plasmid CD19RCD28mz-T2A-h Δ TK(CoOp)/p Δ SBSO. The SB inverted-direct repeats (IRs/DRs) T1 were replaced with the hyperactive IRs-DRs T2, to improve transposition, and the firefly luciferase reporter gene replaced the h Δ TK transgene. Plasmids CD19RCD28mz-T2A-h Δ TK(CoOp)/p Δ SBSO and CD19RCD28mZ(CoOp)/pT2SBSO were digested with *Stu I*-*Kpn I* and *Clal*-*Kpn I* restriction enzymes, respectively, with an extra blunting reaction after the *Clal* digestion, and ligated using T4 DNA ligase to create intermediate plasmid-I, CD19RCD28mz-T2A-h Δ TK(CoOp)/pT2SBSO. The T2A-h Δ TK transgene was replaced with a T2A-flag-tag fusion and a multiple cloning site (MCS). T2A-flag-MCS was created by PCR primer annealing and TOPO-TA cloning. Plasmids CD19RCD28mz-T2A-h Δ TK(CoOp)/pT2SBSO and T2A-Flag-MCS/TOPO were digested with *BspE I* and *Spe I* and ligated to create intermediate plasmid-II, CD19RCD28mz-T2A-Flag(CoOp)/pT2SBSO. The firefly luciferase gene was excised from the previously made plasmid Flag-FFLuc-HyTK(CoOp)/pT2SBSO using *SnaBI* and *EcoRV* restriction enzymes. The same restriction enzymes were used to digest the accepting pT2SBSO vector. Purified fragments were ligated to create the CD19RCD28mz-T2A-Flag-FFLuc(CoOp)/pT2SBSO.

The *SB* transposon encoding *CAR (CD19R-Fcm-CD28m-Z(hCoOp)/pSBSO-SIM), a second generation CD19-specific IgG4-Fc stalk modified CAR, was generated to abrogate unintended binding to innate Fc γ receptors. Source DNA was human codon-optimized and synthesized by GeneArt. The same CD19-specific scFv as used in our previous vectors (2) was connected to modified human IgG4 hinge and Fc regions (the hinge was previously mutated from amino acids CPSC and CPPC to enhance stability of the dimerized IgG4 heavy chain, and amino acid changes L235E, and N297Q were newly made to decrease unintended binding to innate Fc γ receptors). The Fc region was connected to the human CD28 transmembrane and cytoplasmic domains with the cytoplasmic endodomain modification RLLH to RGGH to improve CAR expression; and the CD28 endodomain was connected to the CD3-Z cytoplasmic domain. The GeneArt vector and vector CD19RCD28mZ(hCoOp)/pSBSO-SIM used in clinical trials were digested with *AvrII* and *XmnI* to obtain the insert and vector backbone, which included a portion of the CD19R-CAR. Ligation generated the final *CAR vector (CD19R-Fcm-CD28m-Z(hCoOp)/pSBSO-SIM).

rLuc-mKate reporter: *rLuc*8.6-535-T2A-mKateS158A/pLV430G is a lentiviral vector encoding both *Renilla luciferase* (*rLuc*) and mKate. They were created from sequences for *rLuc*8.6-535 and mKateS158A as reported (3, 4), codon-optimized as previously reported (5), and were synthesized by GeneArt to yield a B1 *rLuc*8.6-535-T2A-mKateS158A B2 construct. The B1 and B2 sites are lambda phage recombination sites used in Gateway Recombination Cloning Technology (Invitrogen) synthesizing the construct via BP reaction into pDONR221. Subsequently, *rLuc*8.6-535-T2A-mKateS158A/pDONR221 was recombined into pLV430G (6), a self-inactivating lentivirus encoding an internal U3 promoter from the murine stem cell virus.

Cell lines and propagation. Cell lines were maintained in complete media [CM; RPMI 1640, 10% heat-inactivated fetal bovine serum (Hyclone), and 1% Glutamax-100 (Gibco)] in humidified conditions at 5% CO₂ and 37°C. NALM-6 (pre-B cell) and THP-1 (human monocyte) were obtained from American Type Culture Collection (Manassas, VA). K562-derived AaPC (designated “clone 9”) (7), CD19⁺EL4 (2), and GFP⁺*effLuc*⁺NALM-6 (8) have been described. The *rLuc*⁺mKate⁺NALM-6 cells were generated by lentiviral transduction of NALM-6 with *rLuc*8.6-535-T2A-mKateS158A/pLV430G using standard techniques. Cell lines were negative for mycoplasma. Identity of cell lines was confirmed by STR DNA fingerprinting by the Characterized Cell Line Core, a Cancer Center Support Grant supported facility, at MD Anderson.

Generation and *ex vivo* expansion of genetically modified T cells. The CAR and mbIL15-CAR T cells were generated from mononuclear cell buffy coats from anonymized healthy donors (Gulf Coast Regional Blood Center, Houston, TX) but with modifications to the previously described method of generating clinical-grade CD19-specific CAR T cells (**SI Appendix, Fig. S1**) (2). Briefly, PBMC were co-electroporated with supercoiled DNA plasmids coding for mbIL15 and second-generation CAR separately expressed as two *Sleeping Beauty* (SB) transposons along with supercoiled DNA plasmid coding for SB11 transposase. Cells were then cocultured on γ-irradiated K562 (clone #9)-derived activating and propagating cells (AaPC), modified to co-express truncated CD19, CD64, CD86, and CD137L, and supplemented with recombinant soluble IL-21 (30 ng/mL). The genetic modification and expansion of CAR T cells (not expressing mbIL15) was similarly performed on AaPC clone #9, but electroporated with only DNA plasmid coding for CAR as SB transposon and DNA plasmid coding for the SB transposase with subsequent cultures supplemented with recombinant soluble IL-21 (30 ng/mL) and IL-2 (50 U/mL) or soluble IL-

15/IL-15R α -Fc complex (5 ng/mL, IL-15 complex). The AaPC were added once every 8 – 10 days at ratio of 1:1 (T cells to AaPC) and cytokine(s) were supplemented three times per week for all cultures. IL-15/IL15-R α -Fc (referred to as ‘IL-15 complex’) was supplemented in CAR T-cell culture medium or used as a control in certain assays. IL-15/IL15-R α -Fc complex is a 2:1 molar ratio combination of human IL-15 (PeproTech) and human IL-15R α -Fc chimera (R&D Systems) containing 5 ng/mL of IL-15 (dosing of complex is based on IL-15 content). If CD3^{neg}CD56⁺ NK cell outgrowth exceeded 15%, these were depleted using CD56 microbeads (Miltenyi Biotec) and LS columns (Miltenyi Biotec) per the manufacturer’s instructions. Viable T cells were enumerated after each stimulation cycle (8–10 days) based on Trypan blue exclusion. Inferred CAR T cell numbers were calculated by multiplying the CD3⁺CAR cell fold expansion from the first stimulation with the starting cell number that would then be multiplied by the CAR-specific T cell expansion from subsequent stimulation cycles.

Western Blot of mbIL15. Whole cell lysates (2.4 μ g) of propagated mbIL15-CAR T cells were separated by SDS-PAGE under reducing conditions using a “Simple Western” size-based assay (WesTM System, ProteinSimple, Santa Clara, CA) per the company’s protocol. The target protein (mbIL15) was identified using anti-human IL-15 antibody (20ug/mL, R&D Systems) followed by HRP-conjugated rabbit anti-goat secondary antibody (1:250).

Antibodies and Flow Cytometry. Up to 10⁶ cells were stained with fluorochrome-conjugated antibodies: fluorescein isothiocyanate (FITC), phycoerythrin (PE), peridinin chlorophyll protein conjugated to cyanine dye (PerCP-Cy5.5), allophycocyanin (APC), AlexaFluor-488, and AlexaFluor-647, which unless otherwise stated, were obtained from BD Biosciences. Unless

otherwise stated, antibodies were used at 1:20, 1:40, 1:25, 1:33, 1:20, and 1:20 dilutions, respectively, in a 100 μ l volume of buffer. Staining for cell surface markers and corresponding isotype controls was conducted in flow cytometry buffer (phosphate-buffered saline (PBS), 2% fetal bovine serum (FBS), 0.1% sodium azide) for 30 minutes at 4°C. Intracellular staining was performed following fixation and permeabilization for 20 minutes at 4°C with BD Cytofix/Cytoperm with antibodies diluted in 1 \times Perm/Wash buffer for 30 minutes at 4 °C (BD Biosciences). Annexin V staining was conducted in 1 \times annexin binding buffer (BD biosciences). Antibodies used for immunostaining included (against human): Bcl-2 (1:5 dilution, clone Bcl-2/100), Fc (to detect CAR; clone H10104, Invitrogen), IL-15 (1:6.7 dilution for APC; clone 34559, R&D Systems), Blimp-1 (clone 646702, R&D Systems), CD19 CAR idiotype (clone 136.20.1) (9), CD3 (clone SK7), CD4 (cloneRPA-T4), CD8 (clone SK1), CD25 (clone M-A251), CD27 (clone L128), CD28 (clone L293), CD19 (clone HIB19), CD45RA (clone HI100), CD45RO (clone UCHL1), CD56 (clone B159), CD62L (clone DREG-56), CD122 (clone Mik- β 3), CD127 (1:8.3 dilution; clone 40131, R&D Systems), CCR7 (1:45 dilution for PerCP-Cy5.5; clone G043H7, BioLegend), IL-2 (clone MQ1-17H12), IFN γ (clone B27), pSTAT5 (pY694) [1:5 dilution; clone 47/Stat5(pY694)], T-bet (1:20 for PerCP-Cy5.5; clone 4B10, BioLegend), and unconjugated rabbit anti-TCF1 (1:20 dilution; clone C725.7, Thermo Scientific). Polyclonal goat anti-rabbit IgG-DyLight650 (1:100 dilution; Thermo Scientific) was used as a secondary antibody to detect TCF1. Annexin V (1:20 dilution; BD Bioscience) and anti-FLAG (1:200 dilution; Prozyme) were also used. Data were acquired using a FACSCalibur with Cell Quest version 3.3 software (BD Biosciences) and analyzed with FlowJo software (version 7.6.5, TreeStar).

Phosflow of pSTAT5. Cells were washed in PBS to remove residual cytokines from culture media. The cells were then transferred to fresh serum-free culture media (RPMI supplemented with 1% Glutamax) at a density of 2×10^6 /mL and starved for 12 hours without serum and exogenous cytokine. Serum-starved negative control CAR T cells provided the baseline level of pSTAT5, and positive control CAR T cells were treated with 100 ng/mL IL-15 for one hour after “starving” prior to fixation. Cells were pelleted to remove the supernatant and processed according to the manufacturer’s protocol using PhosFlow Lyse/Fix Buffer (BD Biosciences), BD PhosFlow Perm Buffer III (BD Biosciences), and pSTAT5 (pY694, BD Biosciences).

Intracellular cytokine production. Production of cytokines was assessed by intracellular cytokine staining of T cells as described above. T cells were co-cultured with CD19⁺ or CD19^{neg} targets at an effector to target ratio of 1:5 in 200 µL of CM. T cells were incubated alone (negative control) and with the addition of Leukocyte Activation Cocktail (LAC, containing PMA and ionomycin, BD Biosciences; positive control). Brefeldin-A (GolgiPlug, BD Biosciences) was added, per manufacturer instructions, to all cultures to block exocytosis of cytokines. Cultures were incubated for 6 hours at 37°C, then stained for surface markers, followed by fixation, permeabilization and staining for expression of intracellular proteins.

Chromium release assay. The antigen-specific cytolytic activity of T cells was assessed in a standard 4-hour chromium release assay (CRA), as previously described (10). Genetically modified T cells were incubated with 2×10^5 of ⁵¹Cr-labeled target cells (CD19^{neg} parental EL4, CD19⁺ EL4, or CD19⁺ NALM-6). The percentage of specific lysis was calculated from the release of ⁵¹Cr using a TopCount NXT (Perkin-Elmer) (10).

In vitro antigen and cytokine withdrawal assays. After the fourth stimulation with AaPC, mbIL15[±]-CAR T cells were maintained with exogenous cytokine(s) for 10 days, then enumerated and transferred to fresh CM in exogenous cytokine-free (unless otherwise noted) and antigen-free culture conditions. CAR T cells supplemented with IL-15 complex (5 ng/mL, 2×/week) served as a positive control for an assay with cells enumerated on days 15, 30, 50, and 65. T cells persisting in culture for >65 days, referred to as “AWD-T cells”, were assessed for pSTAT5 and Bcl-2. Two days prior to staining for Bcl-2, baseline control CAR T cells were washed and resuspended in CM without cytokine supplementation.

The withdrawal assay assessing CCR7 subset T cell viability was performed as described above, but tested the following treatments: (i) mbIL15-CAR T cells, (ii) CAR T cells supplemented with IL-2 (50 U/ml, 2×/week), and (iii) CAR T cells supplemented with IL-15 (5 ng/ml, 2×/week). Live cell staining of AWD-T cell CCR7 subsets was performed on cells after 65 days. AWD-T cells were first stained with CCR7 antibody and gently washed once with FACS buffer, followed by staining with Annexin V (1:20 dilution; BD Bioscience) per the manufacturer’s protocol.

nCounter system digital gene expression analysis. Day 0 (i.e., 8 days after the fourth AaPC stimulation) CAR and mbIL15-CAR T cells and day 65 AWD-mbIL15-CAR T cells were frozen at -80°C and later analyzed in batch for gene expression analysis using the nCounter System (NanoString Technologies) according to the manufacturer’s protocol. At the time of analysis, RLT buffer (Qiagen) was added to the samples at a ratio of 166 µL of buffer per 10⁶ cells prior to thawing. RNA lysates were immediately processed and analyzed using the nCounter Analysis System (NanoString Technologies) to detect the expression of genes of interest. Multiplexed

target-specific, color-coded reporter and biotinylated capture probes for mRNA species of interest were hybridized for >12 hours at 65°C in a thermocycler (Peltier Thermal Cycler, Bio-Rad), processed in the nCounter PrepStation (NanoString Technologies), and then analyzed using the nCounter Digital Analyzer (NanoString Technologies). Reference genes (*ACTB*, *G6PD*, *OAZ1*, *POLR1B*, *POLR2A*, *RPL27*, *RPS13*, and *TBP*), which together span a dynamic range of RNA expression, were included in the CodeSet and were used to normalize transcript levels to account for variation of transcript levels between samples and of the amount of total RNA present in samples. The count data was positive-, negative-, and housekeeping gene-normalized per nCounter guidelines using the nCounter RCC Collector Worksheet (v1.6.0, NanoString Technologies), as previously described (11), and filtered to remove low-expression genes: those with counts of less than 10 in greater than 60% of samples. A lymphocyte-specific CodeSet (**SI Appendix, Table S1**) was generated from RefSeq accession numbers for chosen mRNA transcripts that were used to generate the reporter and capture probe pairs constituting the lymphocyte gene panel. Statistical analyses were performed using edgeR software (12) via the RobiNA Java interface (13). Statistical evaluation of differential gene expression implemented statistical methods based on a negative binomial dispersion. Genes considered to be significantly differentially expressed demonstrated an absolute fold change ≥ 2 , $P < 0.01$, and a FDR significance value of $q < 0.05$ with the Benjamini-Hochberg method used for multiple testing correction. Significant differentially expressed genes were represented in a heatmap generated using Qlucore Omics Explorer (v.2.3, Qlucore). Digital gene expression analysis for assessing V α /V β repertoire was performed as previously described (14).

G-banded Karyotyping. AWD-mIL15-CAR T cells that were in long-term continuous culture were restimulated with AaPC. Actively dividing T cell samples were submitted for G-banded karyotyping to the Texas Children's Hospital Clinical and Research Cytogenetic Laboratory.

PKH Dilution Assay. AWD-mIL15-CAR T cells that were in long-term continuous culture (1–2 years, depending on the donor) were labeled with PKH (Sigma) per manufacturer's instructions. Labeled cells were assessed for PKH dilution by flow cytometry 10 and 30 days after PKH labeling. The positive PKH dilution control was AWD-mIL15-CAR T cells stimulated with AaPC.

Mouse experiments. *In vivo* experiments used humanized NSG mice (NOD.Cg-Prkdc^{scid}IL2ry^{tm1Wjl}/SzJ) that were purchased from The Jackson Laboratory (Bar Harbor, ME) or bred at The University of Texas MD Anderson Cancer Center.

Bioluminescence imaging. Serial BLI was performed to measure temporal biodistribution and abundance of mbIL15-CAR and CAR T cells co-expressing *ffLuc* as previously described (8, 15). Serial BLI was used to measure tumor burden of *ffLuc*⁺ or *rLuc*⁺NALM-6 cells as previously described (15). Imaging was performed on anesthetized mice using a Xenogen IVIS 100 (Caliper Life Sciences). Imaging for T cells and tumor were performed on alternate days with an initial imaging of background BLI to check that no residual luminescence remained. T cell and tumor flux (photons/s/cm²/steradian) were quantified by measuring photon signal within a delineated region of interest (ROI) encompassing: (i) the body of the mouse, except the tail and (ii) over the femur. Living Image software (v2.50, Xenogen; Caliper Life Sciences) was used to process the BLI data.

Tissue preparation. Mice were euthanized, and peripheral blood, spleen, and BM from femur and tibia were harvested. Spleens were manually homogenized in 2% FBS/PBS and passed through a 40 μ m cell strainer (BD BioSciences). To obtain marrow, the femurs and tibia were flushed with 2% FBS/PBS using 30G needles (BD Biosciences). ACK lysing buffer (Gibco-Invitrogen) was used to lyse red blood cells in all samples.

In vivo persistence and characterization of genetically modified T cells in the absence of CAR activation. CAR T cells (2×10^7 per mouse) were injected i.v. into NSG mice in two cohorts ($n = 5$ /group): (i) mbIL15-CAR-*ffLuc* T cells and (ii) CAR-*ffLuc* T cells. Serial bioluminescent imaging (BLI) measured T-cell persistence and biodistribution until sacrifice at days 48-50. Tissues were harvested for analysis.

In vivo characterization of mbIL15-CAR T cells persisting in the absence of CAR activation. NSG mice were engrafted with 2×10^7 mbIL15-CAR T cells ($n = 9$). After 48-50 days, peripheral blood, spleen, and BM were harvested for flow cytometry analysis by for CD45RO and CCR7 expression. T cells from spleen, liver, and BM samples were *ex vivo* expanded on AaPC at an initial ratio of one isolated cell to five AaPC with subsequent weekly stimulations at a 1:1 ratio. Expanded T cells were tested for antigen-specific function by assessing intracellular staining of IFN γ in response to CD19 $^+$ versus CD19 neg cell targets.

In vivo persistence and antitumor efficacy of CAR and mbIL15-CAR T Cells in a preventative leukemia model. T cells were engrafted prior to tumor to mimic MRD. On day 0, treatment cohorts of NSG mice ($n = 5$ /group) were injected i.v. with 2×10^7 genetically modified T cells: (i) mbIL15-CAR-*ffLuc* T cells, or (ii) CAR-*ffLuc* T cells (cultured with IL-15 and IL-21). On day 6, 1×10^4 *rLuc* $^+$ NALM-6 were injected i.v. T cell and tumor BLI were monitored as described (6) until death

or sacrifice, when tissues were harvested for analysis of CD3 and CD19 by flow cytometry. For BLI, background fluorescence was defined by measuring animals given luciferin but without ffLuc-bearing cells (Fig. 5, T cell imaging). To assess therapeutic efficacy, a similar experiment was undertaken with survival at 98 days as the experimental endpoint in cohorts: (i) mbIL15-CAR-*ffLuc* T cells plus NALM-6 ($n = 7$), (ii) CAR-*ffLuc* T cells plus NALM-6 ($n = 8$), and (iii) NALM-6 only ($n = 8$). T-cell persistence and biodistribution was measured by BLI until death or euthanasia. Mice were considered to have reached the experimental endpoint if they developed hind-limb paralysis or were otherwise moribund.

In vivo persistence and antitumor efficacy of CAR and mbIL15-CAR T Cells in an established leukemia model. On day 0, NSG mice were injected i.v. with 1.5×10^4 *ffLuc*⁺eGFP⁺NALM-6. After 5 days of tumor engraftment, mice were imaged and randomized into treatment groups ($n = 7$, tumor only group; $n = 6$, all other T cell treatments). On day 6, mice were given 1×10^7 genetically modified T cells via intra-cardiac injection. Infused T cells were CD19-specific CAR T cells (expressing CAR or *CAR) with or without coexpression of mbIL15. The infused cell number was based on the CAR T cell number. Tumor BLI was measured until mortality or sacrifice, when tissues were harvested for analysis as above.

Statistical analysis. Statistical analyses were performed using Prism (GraphPad Software, v.6.02). Cell count and flux values were log-transformed for statistical analysis. Paired (*in vitro* data) and unpaired (*in vivo* data) t-tests were used to compare two groups. One-way ANOVA was used to compare three groups. Two-way ANOVA was used for multiple comparisons of two factors. Two-

way RM ANOVA (Tukey's post-test) compared multiple groups over time. Survival was analyzed with the log-rank test.

SI Appendix References

1. Davies JK, *et al.* (2010) Combining CD19 redirection and alloantigenization to generate tumor-specific human T cells for allogeneic cell therapy of B-cell malignancies. *Cancer Res* 70(10):3915-3924.
2. Singh H, *et al.* (2013) Manufacture of clinical-grade CD19-specific T cells stably expressing chimeric antigen receptor using Sleeping Beauty system and artificial antigen presenting cells. *PLoS One* 8(5):e64138.
3. Loening AM, Wu AM, & Gambhir SS (2007) Red-shifted *Renilla reniformis* luciferase variants for imaging in living subjects. *Nat Methods* 4(8):641-643.
4. Pletnev S, *et al.* (2008) A crystallographic study of bright far-red fluorescent protein mKate reveals pH-induced cis-trans isomerization of the chromophore. *J Biol Chem* 283(43):28980-28987.
5. Rabinovich BA, *et al.* (2008) Visualizing fewer than 10 mouse T cells with an enhanced firefly luciferase in immunocompetent mouse models of cancer. *Proc Natl Acad Sci U S A* 105(38):14342-14346.
6. Bai X, *et al.* (2011) Tracking long-term survival of intramyocardially delivered human adipose tissue-derived stem cells using bioluminescence imaging. *Mol Imaging Biol* 13(4):633-645.
7. Denman CJ, *et al.* (2012) Membrane-bound IL-21 promotes sustained ex vivo proliferation of human natural killer cells. *PLoS One* 7(1):e30264.
8. Singh H, *et al.* (2011) Reprogramming CD19-specific T cells with IL-21 signaling can improve adoptive immunotherapy of B-lineage malignancies. *Cancer Res* 71(10):3516-3527.
9. Jena B, *et al.* (2013) Chimeric antigen receptor (CAR)-specific monoclonal antibody to detect CD19-specific T cells in clinical trials. *PLoS One* 8(3):e57838.
10. Cooper LJ, *et al.* (2003) T-cell clones can be rendered specific for CD19: toward the selective augmentation of the graft-versus-B-lineage leukemia effect. *Blood* 101(4):1637-1644.
11. Maiti SN, *et al.* (2013) Sleeping beauty system to redirect T-cell specificity for human applications. *J Immunother* 36(2):112-123.
12. Robinson MD, McCarthy DJ, & Smyth GK (2010) edgeR: a Bioconductor package for differential expression analysis of digital gene expression data. *Bioinformatics* 26(1):139-140.
13. Lohse M, *et al.* (2012) RobiNA: a user-friendly, integrated software solution for RNA-Seq-based transcriptomics. *Nucleic Acids Res* 40(Web Server issue):W622-627.
14. Deniger DC, *et al.* (2013) Bispecific T-cells Expressing Polyclonal Repertoire of Endogenous gamma delta T-cell Receptors and Introduced CD19-specific Chimeric Antigen Receptor. *Molecular Therapy* 21(3):638-647.
15. Singh H, *et al.* (2007) Combining adoptive cellular and immunocytokine therapies to improve treatment of B-lineage malignancy. *Cancer Res* 67(6):2872-2880.

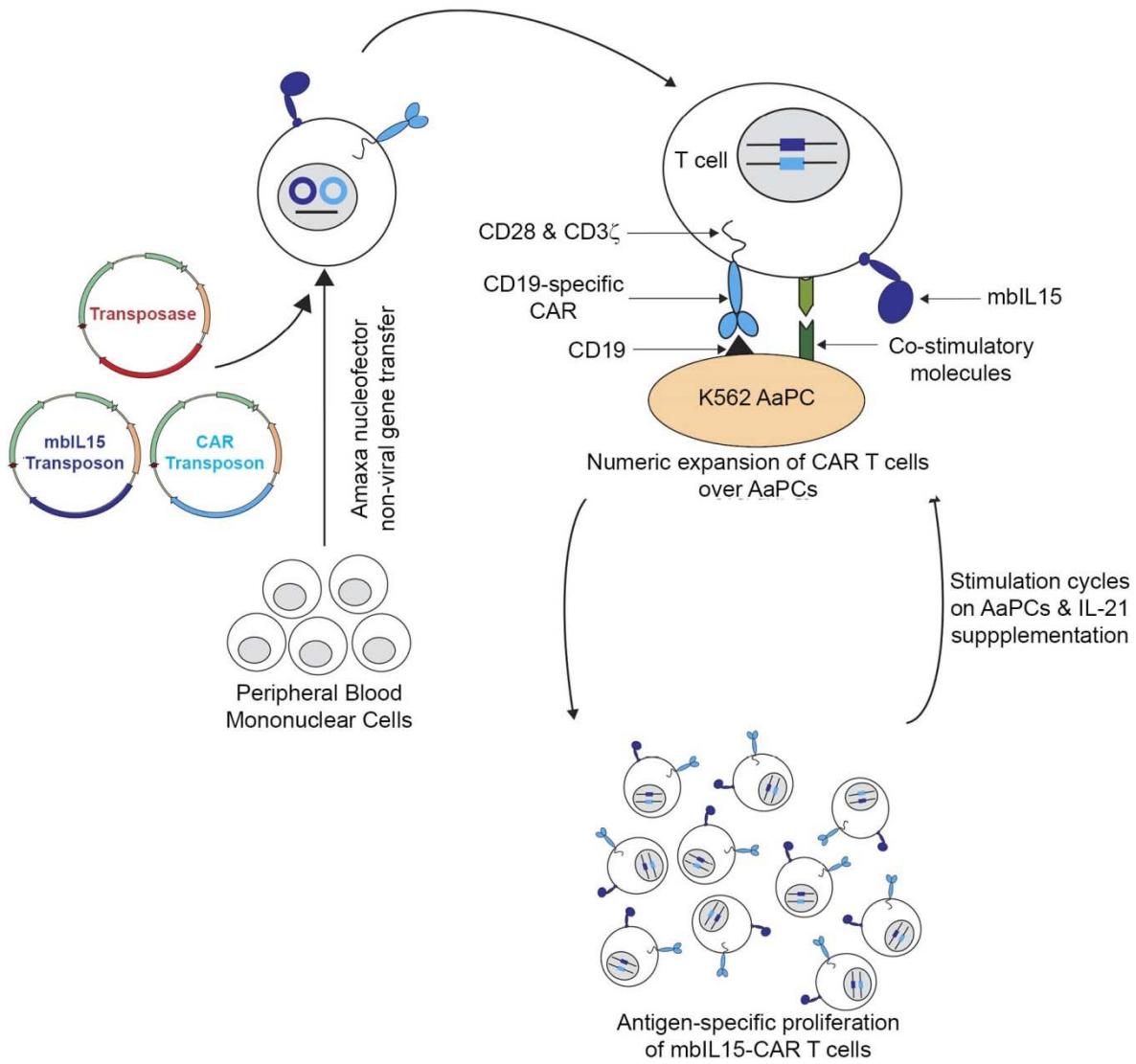


Fig. S1. Generation of CAR⁺mbIL15[±] CD19-specific T cells using a clinical platform. Schematic of the clinically compliant methods (2) adopted and modified for generating and numerically expanding mbIL15-CAR T cells. PBMC were co-electroporated with mbIL15 and second-generation CAR separately expressed as two *Sleeping Beauty* (SB) transposons along with the SB11 transposase. Cells were then cocultured on γ -irradiated K562 (clone #9)-derived activating and propagating cells (AaPC), modified to co-express truncated CD19, CD64, CD86, and CD137L, and supplemented with IL-21 (30 ng/mL). The genetic modification and expansion of CAR T cells was similarly performed, but used only CAR SB transposon and SB transposase during electroporation with subsequent cultures supplemented with IL-21 (30 ng/mL) and IL-2 (50 U/mL) or soluble IL-15/IL-15R α -Fc complex (5 ng/mL, IL-15 complex).

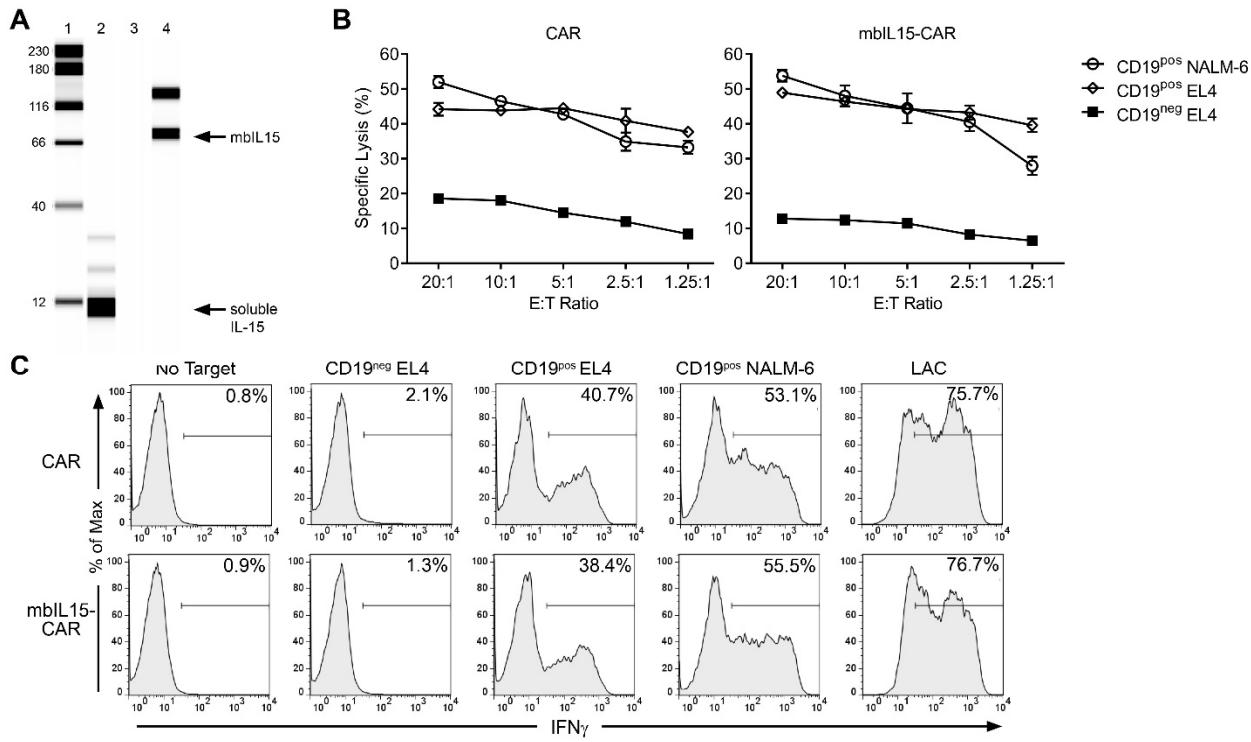


Fig.S2. In vitro phenotype and effector function is not altered by mbIL15 expression. (A) Western blot analysis of mbIL15 expressed by propagated human T cells. A representative blot from one of three donors is shown. Molecular weight marker (Lane 1), 0.1ng recombinant human IL-15 positive control (Lane 2), CAR T-cell negative control (Lane 3), and mbIL15-CAR T cells (Lane 4). The predicted molecular weight of monomeric recombinant human IL-15 is 14 kDa and that of mIL15 is approximately 72 kDa (incorporating possible post-translational modifications). (B) CAR T cell (left panel) and mbIL15-CAR T cell (right panel) specific lysis rates of CD19⁺ or CD19^{neg} targets from a four-hour CRA. Data are represented as mean \pm SD for triplicate wells of one of three normal donors. (C) IFN γ production by CAR and mbIL15-CAR T cells that were stimulated for 6 hours with CD19⁺ (tCD19⁺ EL4 and NALM-6) or CD19^{neg} (EL4) targets, Leukocyte Activating Cocktail (LAC; positive control), or media alone (negative control). Histograms represent CD3⁺CAR⁺ cells. Representative histograms are shown for one of four normal donors.

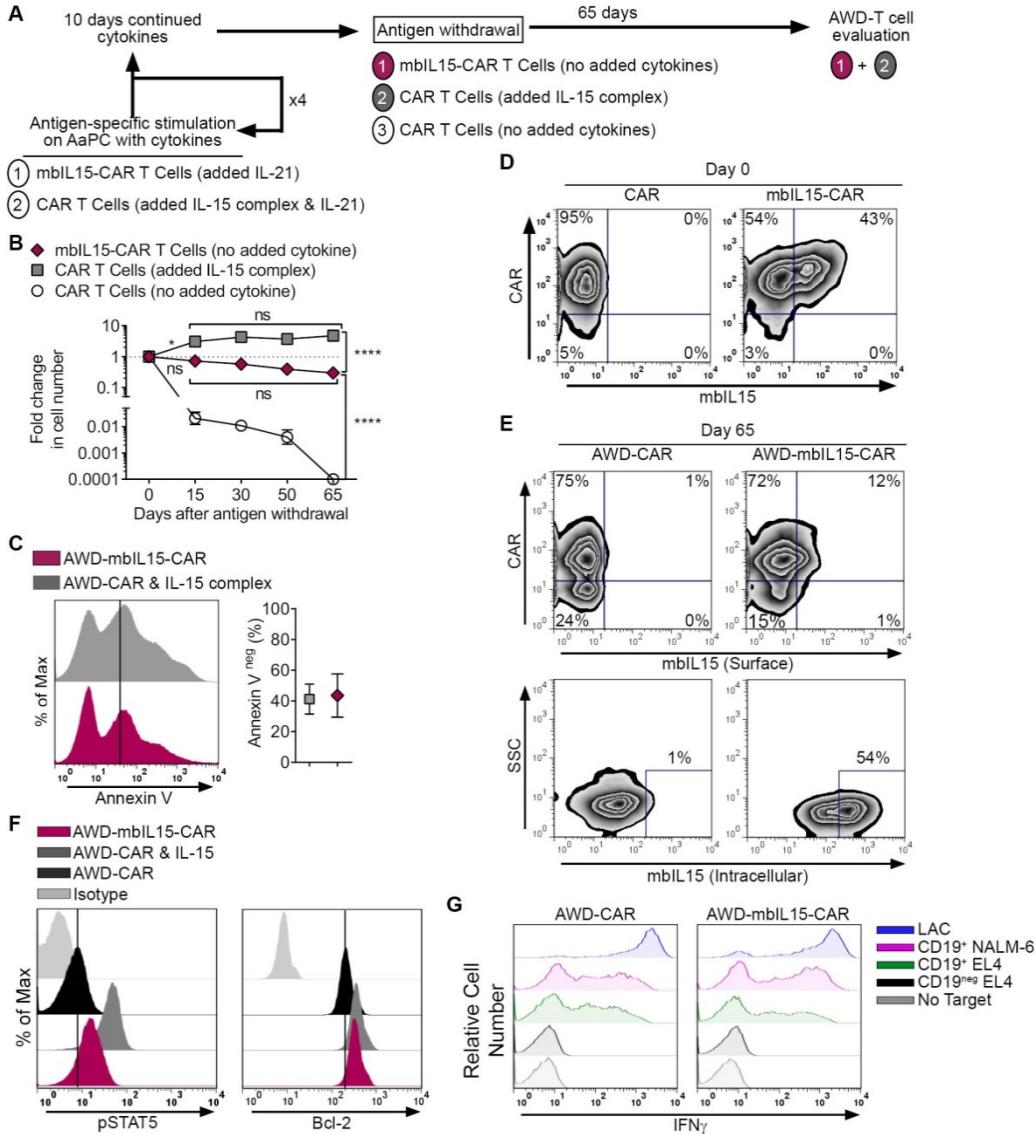


Fig. S3. Prolonged *in vitro* persistence of mbIL15-CAR T cells without CAR activation maintains IL-15 signaling and antigen-specific response. (A) Schematic summarizing the experimental design. T cells persisting at the end of the assay are termed AWD-T cells. (B) Fold change in T cell number under antigen and cytokine withdrawal conditions. *P < 0.05, ***P < 0.001, ns = not significant, two-way RM ANOVA (Tukey's post-test). Data displayed as mean ± SD. (C) Representative histogram showing Annexin V staining (*left panel*) and plotted live T cell (Annexin V^{neg}) frequencies (*right panel*). (D, E) Representative flow plots showing CAR and mbIL15 expression on T cells on day 0 (D) and day 65 (top panel) (E) with FLAG staining detecting intracellular localization of mbIL15 (bottom panel). (F) Representative histograms showing fluorescence intensity of pSTAT5 (*left panel*) and Bcl-2 (*right panel*). AWD-CAR T cells cultured with IL-15 complex were left without cytokine or treated with IL-15 to serve as positive and negative controls, respectively. Vertical bar references median fluorescence of the AWD-CAR negative control. (G) IFN γ production of T cells stimulated with CD19⁺ or CD19^{neg} targets, Leukocyte Activating Cocktail (LAC), or media alone. Histograms represent cells selected for expression of CD3 and CAR. All data are representative of three donors.

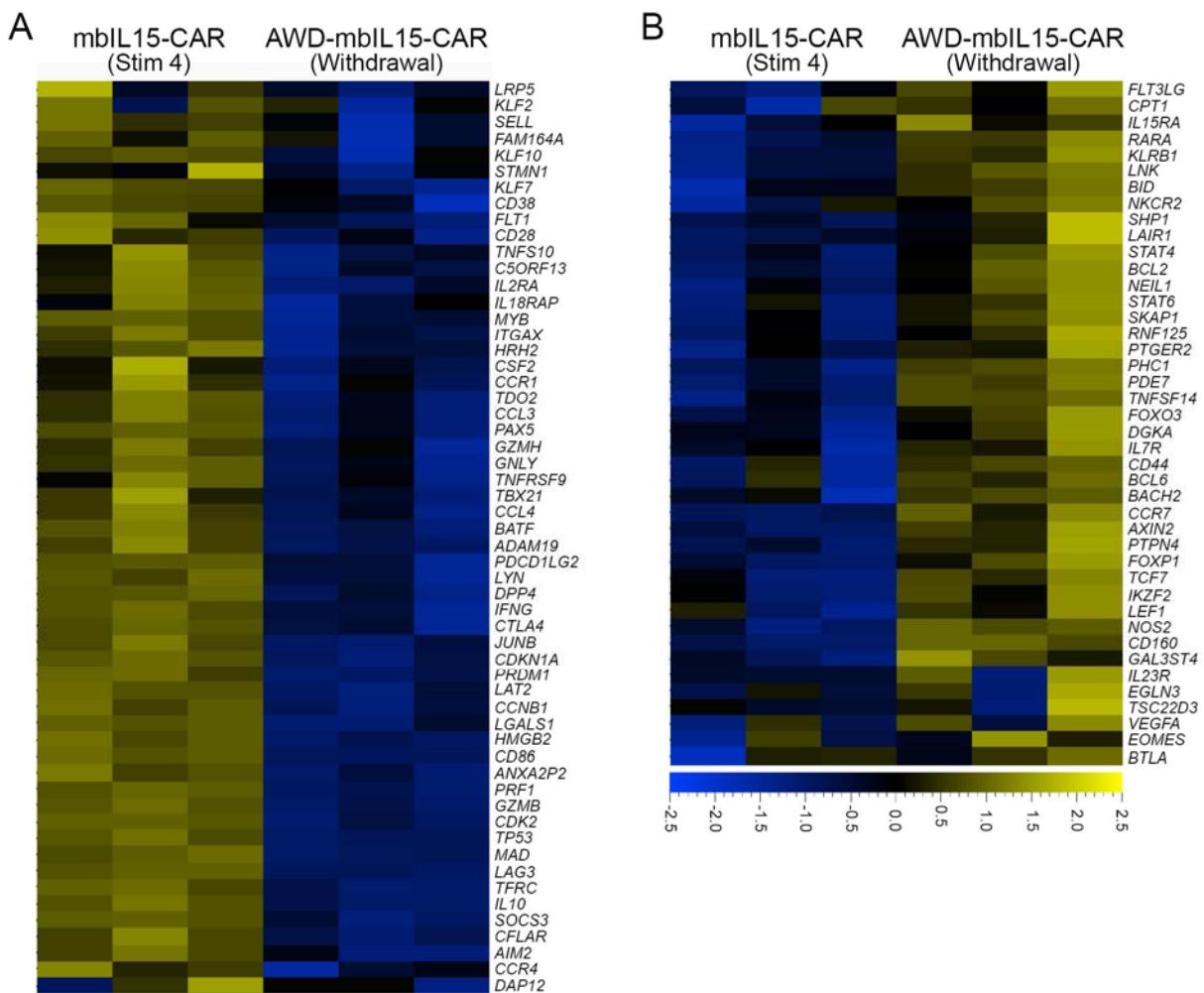


Fig S4. Differentially expressed genes among stim 4 and antigen withdrawal mbIL15-CAR T cells. Heat maps of significantly differentially expressed genes (2-fold cutoff, $P < 0.01$, FDR q value < 0.05) that were (A) downregulated and (B) upregulated in AWD-mbIL15-CAR T cells relative to mbIL15-CAR T cells, $n = 3$.

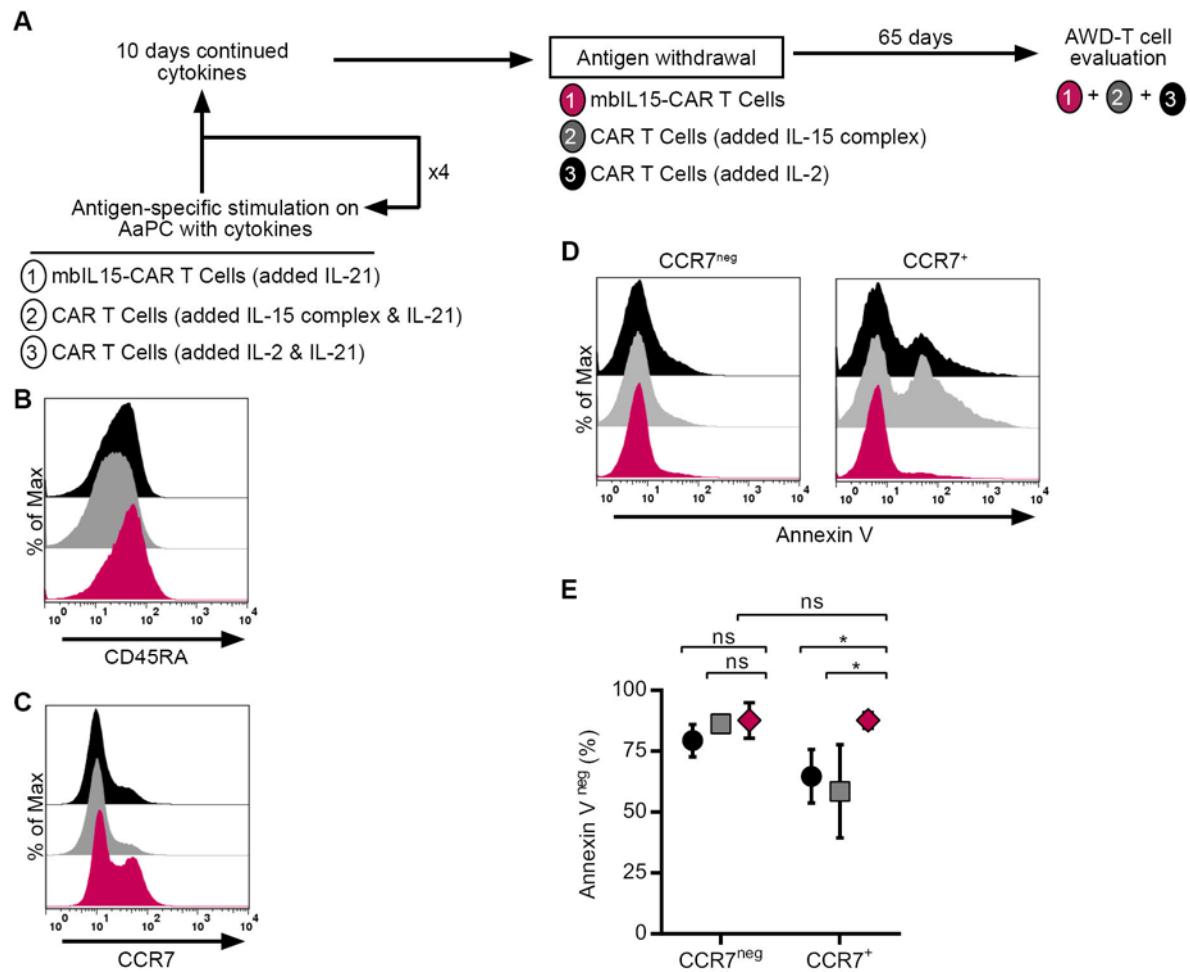


Fig. S5. Tethered IL-15 supports survival of CCR7⁺ T cells. (A) Schematic summarizing the antigen withdrawal experiment comparing the impact of IL-2, IL-15/IL15R α complex, and mbIL15 on CD45RA and CCR7 phenotype. During the 65-day antigen withdrawal, mbIL15-CAR T cells received no exogenous cytokine supplementation, while CAR T cell control treatments received either IL-2 (50 U/ml) or IL-15/IL15R α complex (5 ng/ml A representative histogram of (B) CD45RA and (C) CCR7 expression on AWD-T cells. (D) Representative histograms showing Annexin V levels in CCR7^{neg} and CCR7⁺ AWD-T cell subsets. Histograms display the lymphocyte population and respective CCR7 subsets selected by gating. (E) Graph plotting frequencies of live (Annexin V^{neg}) AWD-T cell treatment cohorts in the CCR7^{neg} and CCR7⁺ subsets. Data are represented as mean \pm SD. *P < 0.05, ns = not significant, two-way ANOVA (Tukey's post-test). All data are representative of three donors. Color legend: CAR T cells supplemented with IL-2 (black), CAR T cells supplemented with IL-15/IL15R α complex (grey), and mbIL15-CAR T cells (magenta).

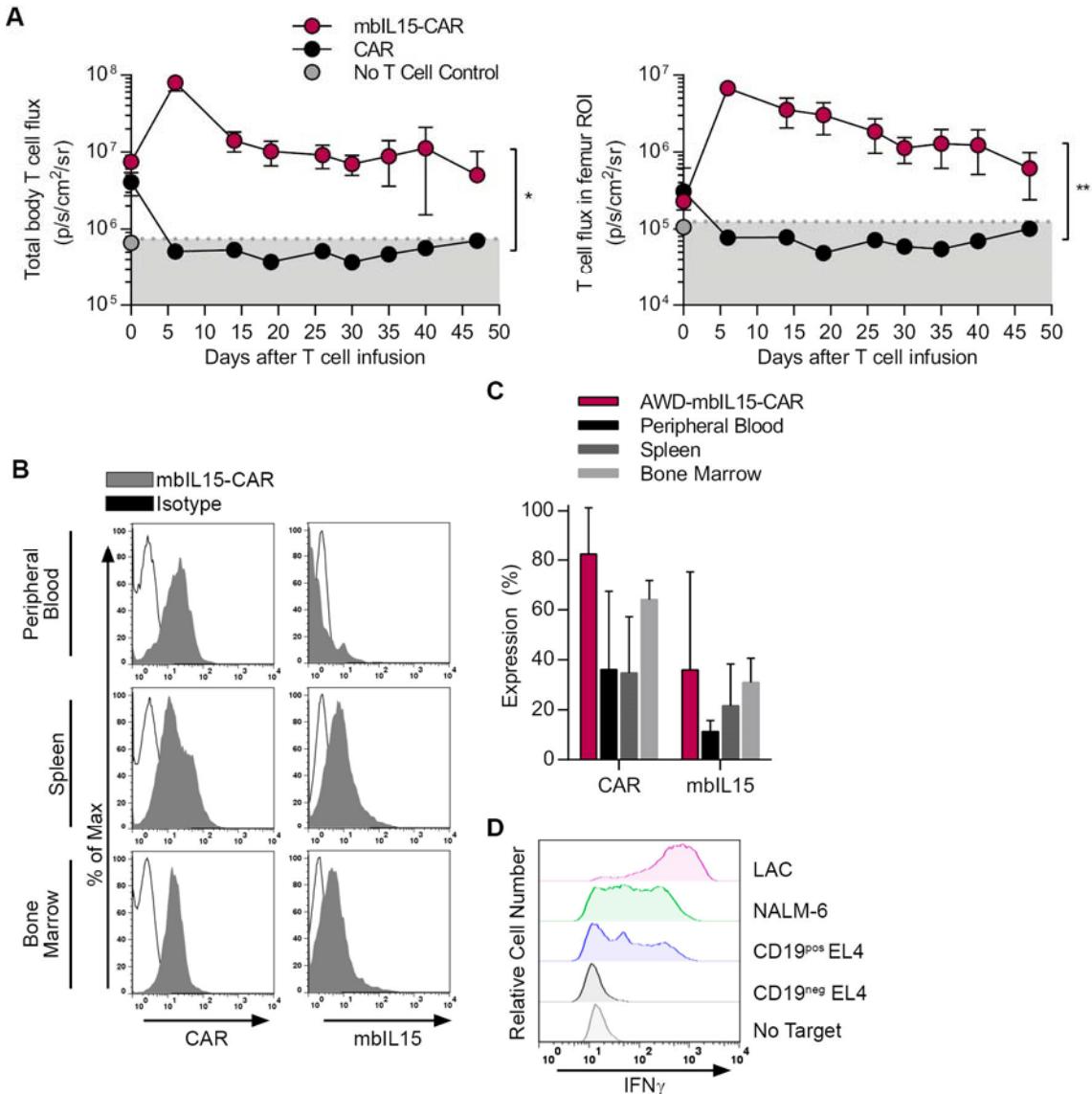


Fig. S6. Long-term *in vivo* engraftment, phenotype, and functionality of mbIL15-CAR T cells independent of CAR signaling. NSG mice were injected i.v. with $2 \times 10^7 \text{ ffLuc}^+ \text{CAR T cells}$ with or without co-expression of mbIL15. Infused T-cell number based on CAR T cells. (A) Longitudinal plotting of T cell flux. Background BLI (gray shaded) was derived by imaging mice without ffLuc^+ T cells that were treated with D-luciferin. Data are represented as mean \pm SD, $n = 5$, and are representative of two experiments. (B) Peripheral blood, spleen, and bone marrow were analyzed after 48 days *in vivo*. Representative histograms show expression of CAR and mbIL15 and (C) plotted frequencies of marker expression represented as mean \pm SD. Plots and graphs (B–C) show T cell frequencies after gating on human CD3^+ and mouse CD45^+ cells in the peripheral blood ($n = 5$), spleen ($n = 9$), and bone marrow ($n = 9$). (D) Cells isolated from spleen, liver, or bone marrow were *ex vivo* expanded on AaPC to generate cells for assessing intracellular IFN γ production in response to CD19 $^-$ and CD19 $^+$ targets, $n = 7$ from three tissue sources and four mice. Histograms represent CD3^+ cells.

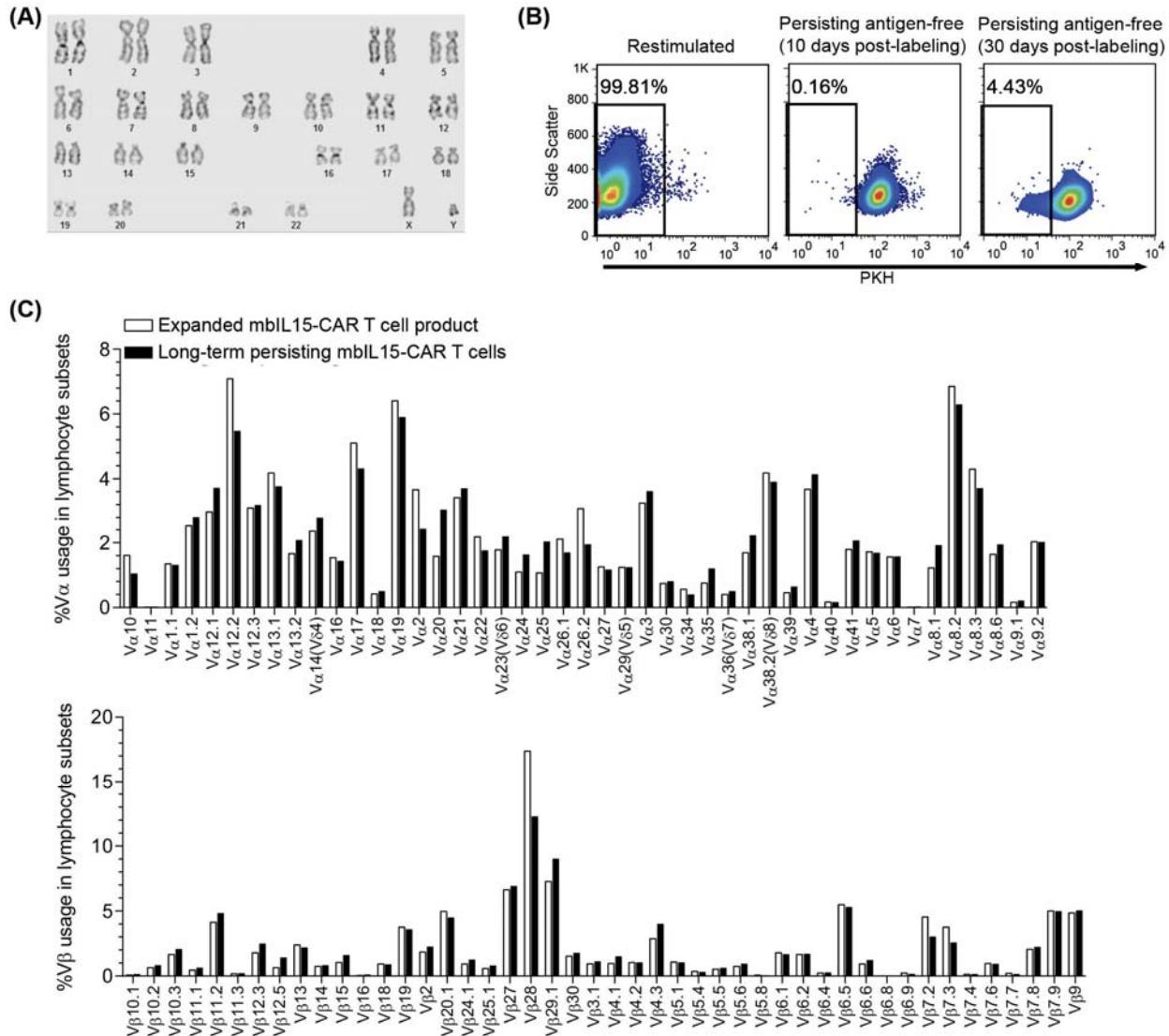


Fig. S7. Prolonged *in vitro* persistence of mbIL15-CAR T cells does not induce aberrant autonomous proliferation or evidence of T-cell transformation. (A) G-banding cytogenetic evaluation showing normal karyotype of *ex vivo* propagated mbIL15-CAR T cells persisting *in vitro* long-term (1 year for 3 donors and 2 years for one donor) under antigen-free conditions and without exogenous cytokines. A representative donor is shown. (B) Homeostatic proliferation, as determined by dilution of the PKH red fluorescent dye, of mbIL15-CAR T cells persisting in long-term continuous culture under antigen-free conditions and without exogenous cytokines ($n = 3$). The proliferation control was the restimulation of the resting T cells with γ -irradiated AaPC clone #9 showing PKH dilution 10 days after labeling (*left panel*). Gates show the percentage of PKH diluted cells. (C) Assessment of TCR polyclonality of the long-term persisting mbIL15-CAR T cells by quantification of mRNA species coding for V α and V β alleles using a designer TCR expression array as previously described (14). Control TCR V α and V β repertoires are the expanded mbIL15-CAR T cells prior to antigen withdrawal. Bars represent the average percentage of V α and V β allele usage in four normal donors.

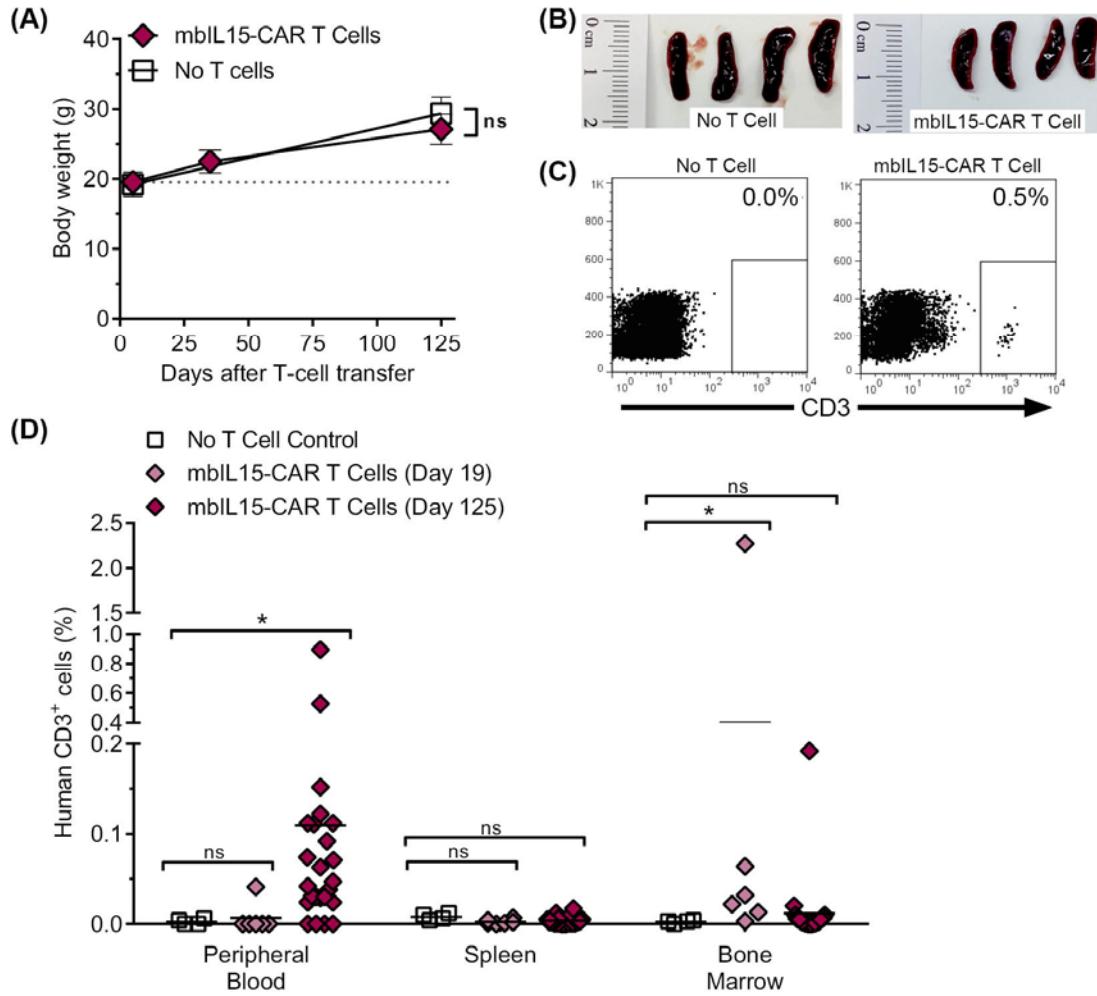


Fig. S8. mbIL15-CAR T cells persisting *in vivo* long-term do not show aberrant autonomous growth or toxicity to recipient mice. NSG mice (tumor-free and no exogenous cytokines) were injected i.v. with 5×10^6 mbIL15-CAR T cells to assess long-term safety of persisting T cells ($n = 8$, 3 donors). **(A)** Body weight to monitor for toxicity from adoptively transferred genetically modified T cells ($n = 8$, 3 donors). **(B)** Representative photos of spleens harvested from mice sacrificed at day 125. **(C)** Human CD3⁺ cell frequencies were assessed by flow cytometry, with representative flow plots from peripheral blood shown. **(D)** Human CD3⁺ cell frequencies observed in the peripheral blood, spleen, and bone marrow at days 19 and 125 of mice treated with mbIL15-CAR T cells. * $P < 0.05$, ns = not significant.

Table S1. nCounter Analysis System Lymphocyte-specific CodeSet

Gene Name	Accession	Target Region	Target Sequence
<i>ABCB1</i>	NM_000927.3	3910-4010	TATAGCACTAAAGTAGGAGACAAAGGAACACTAGCTCTGGTGGCCAGAAACAACGCATTGCCATAGCTCGTGC CTTGTAGACAGCCTCATATTTC
<i>ABCG2</i>	NM_004827.2	285-385	AGGTTAGGAACGACCGTGACATGCTGGTCTGTTAAGTGGAAACTGCTGCTTAGAGTTGTTGG AAGGTCGGGTGACTCATCCCACAT
<i>ACTB</i>	NM_001101.2	1010-1110	TGCGAGAGGAGACTGCCCCGGCACCCAGCACATGAAGATCAAGATCATTGCTCCTCTGAGCGCAAGTA CTCCGTGCGATGGCGGCTCATCT
<i>ADAM19</i>	NM_023038.3	1690-1790	GAGAAGGTGAATGGCAGGAGACACCTTGGAAACTGTTGAAAGGACATGAATGGTAACACAGGAAGTGCA ACATGAGAGATCGAAGTGTGGGAAGA
<i>AGER</i>	NM_001136.3	340-440	GAAAGGAGACCAAGTCCAACCTACCGAGTCCTGTCTACCAGATTCTGGGAAGGCCAGAAATTGAGATTG TCTGAACTCACCGCTGGTGTCCCAA
<i>AHNAK</i>	NM_001620.1	15420-15520	GGATTTGACCTGAATGCTCTGGGGTGAATTGATGCCAGCCTAAGGCTCCGGATGAGATGTCACATCG CAGGGCCGGATGCTGCACTCAAAGTC
<i>AIF1</i>	NM_032955.1	315-415	AAAAGCGAGAGAAAAGGAAAAGGACACAGGCCCGAGCCAAGAAAGCTATCTGAGTTGCCGTGATTGAA GGGAAAAGGGATGATGGGATTGAGGG
<i>AIM2</i>	NM_004833.1	607-707	ACGTGCTGCACCAAAAGTCTCTCATGTTAACGCTGAAACAGAAACAGATGGTGGCCAGCAGGAATCTATCA GAGAAGGGTTCAAGCGCTGTTG
<i>AIMP2</i>	NM_006303.3	507-607	CCCTCTCCCTGCTTGTGCTGCACAGGCTGCTCTGTGAGCAGTCAGGTCTGTCACGGTGCACACGCACTC CTCGGTCAAGAGCTGCTGAAACCT
<i>AKIP1</i>	NM_020642.3	570-670	GAACATCTCAAGGACCTACATAGAAGTATATCAGGGACCTATTCTGCACTGTGGCTCAAATGACTTAAC CAAGAAGACTCATGTGGTAGCAGTT
<i>AKT1</i>	NM_005163.2	1772-1872	TTCTTTCGGGTATCGTGGCAGCACGTGACAGAGAAGACTCAGCCCACCCCTCAAGCCCCAGGTACGT CGGAGACTGACACCAGGTATTTGATG
<i>ALDH1A1</i>	NM_000689.3	11-111	ATTGCTGAGCCAGTCACCTGTGTCAGGAGCCGAATCAGAAATGTCATCCTCAGGCACGCCAGACTACCTG CTCAACTACCGGATTGAGATTCAAT
<i>ANXA1</i>	NM_000700.1	515-615	GAAATCAGAGACATTAACAGGGTACAGAGAGGAACCTGAAGAGAGATCTGCCAAAGACATAACCTCAGACA CATCTGGAGATTTCGGAACGCTTGC
<i>ANXA2P2</i>	NR_003573.1	257-357	ATATTGCTCTCTTACAGAGAGGACAAAAGGAACCTGCACTCAGCACTGAAGTCAGCCTTATGCCAC CTGGAGACGGTGGATTGGGCCATT
<i>APAF1</i>	NM_181869.1	1160-1260	TTCTGATGAAACTGCAAACTTGCACACGGTGGATCAGGATGAGAGTTTCCCAGGGCTTCAACTTAATA TTGAAGAGGCTAAAGACCGTCTCG
<i>ARG1</i>	NM_000045.2	505-605	AAGGAACTAAAAGGAAAGATCCCGATGTGCCAGGATTCTCTGGTACTCCCTGTATATGCCAAGGATAT TGTGTATATTGGCTTGGAGACGTGG
<i>ARRB2</i>	NM_004313.3	1652-1752	CATTAATTTTGACTGCTGCTCTCCAGCCCCGCCGTGGTGGCAAGCTGTGTTCATACCTAAATTTC TGGAGGGCACAGTGGAAAGAGGAG
<i>ATF3</i>	NM_001030287.2	600-700	GGCTCAGAATGGGAGGACTCCAGAAGATGAGAGAAACCTCTTATCCAACAGATAAAAGAAGAACATTGAGA GCTAACGACTCGTGTGTTGGGGCGA
<i>ATM</i>	NM_000051.3	30-130	ACGCTAACTGCTGGCATTGGACATGGCGCAGGCGCTTGCTCGACGGGCGAATTTGGG GTGTTTGAGCCGGAGACCGCTGATA
<i>ATP2B4</i>	NM_001684.3	7640-7740	CTTCCCATACTATCATCTGCTCTGGAATGACTCTCTGTCCCTAAAGGGTTAAGAGAGAGATCCTAGAA ATCCCTCTGGACACTGTGGTTCTT
<i>AXIN2</i>	NM_004655.3	1035-1135	CTTGTCCAGAAACACTCTGAGGGCCACGGCGAGTGTGAGGTCCACGGAAACTGTTGACAGTGGATAAGGTCC TTCAGAGGAGCGATCTGTAACTCT
<i>B2M</i>	NM_004048.2	25-125	CGGGCATCTGAAGCTGACAGCATTGGGCCAGATGTCCTCGCTCGTGGCCATTGCTGCTCGCGTACT CTCTCTTCTGGCCTGGAGGCTATCCA
<i>B3GAT1</i>	NM_018644.3	145-245	CTGGACAGCGACCCCTCTGAGACTCCAGTGGGCCGACTCTCAAACCTGCTCCGAATGGTGGTTGT GAGTGTGGTAATGAGGAGCCGGTGG
<i>BACH2</i>	NM_021813.2	3395-3495	TGTGGCAGTGTGATCTGCTGCTGGAGAAACCGAGAACACATTGGTGCACACTACAGCGCTTAGCAGCA ATACTGTCCCGAAGTATCTCTCTC
<i>BAD</i>	NM_004322.2	195-295	CAGCTGTGCCCTGACTACGTAACATCTGCTCTCACAGCCCAGAGCATGTTCCAGATCCCAGAGTTGAGCCGA GTGAGCAGGAAGACTCCAGCTGCA
<i>BAG1</i>	NM_004323.3	1490-1590	CTCTTGTGATCGTAGTCCCCTAGCTGTAACCAAGAGAACACAGGAGTTGCACCTAGTCAGGAATATTGGG AATGGCCATAGAACACGGTGTGGCA
<i>BATF</i>	NM_006399.3	825-925	CACTGTGGGTTGAGGCCAACTGAGAACAGATTAAGAAAGATGCTCAAGTCCATGGCACAGAGCAAGGCC GGCAGGGACGGTATTCTCTAAATA
<i>BAX</i>	NM_138761.2	694-794	ATTTTCTGGGGGGGGGGGATTGGGGACATGGCATTTCCTACTTTGTAATTATTGGGGGTGTGGGG AAGAGTGGTCTTGAGGGGGTAATAAA
<i>BCL10</i>	NM_003921.2	1250-1350	TGAAAATACCATCTCTCAACTACACTTCCCAGACCTGGGACCCAGGGCTCCCTTGCACCAAGATC TACAGTTAGAAGAAGGAACCTGT
<i>BCL11B</i>	NM_022898.1	3420-3520	GAGATGTAGCAGTCATGCTCCCGAGTCAGCGCCCTTCTGTGTTGATTGGCTTCATATTACATAAGG GAAACCTTGAGGTGGTGTGGGG
<i>BCL2</i>	NM_000633.2	1525-1625	CCAAGCACCGCTTGTGGCTCCACCTGGATGTTCTGTGCTGAAACATAGATTGCTTCCATGTTGG CCGGATCACCATCTGAGAGCAGAC
<i>BCL2L1</i>	NM_138578.1	1560-1660	CTAAGAGCCATTAGGGGCCACTTGTGACTAGGGATTGAGCTGGCTGTTGGATAAGATGCAAGGACAGGACT CCCTCTCACCTGTGACTGGCTAGAG
<i>BCL2L1</i>	NM_001191.2	260-360	ATCTGGCTTGGATCTAGAAGAGAACACTAACCAACAGAGACGAGACTCAGTGAGTGAGCAGGTGTTGGACA ATGGACTGGTGGAGCCCATCCATT

<i>BCL2L11</i>	NM_138621.2	2825-2925	TGTTGGCACCGAACCTAAAGCGATGACTGGATGTCCTGTACTGTATGTTCTGTTCAAGATGCCCTGTG CAGAAAGTATGCCCTCCGTGGTAT
<i>BCL2L11</i>	NM_138621.4	257-357	CGGACTGAGAACGCAAGAAAAAAAGACCAAATGGCAAGCAACCTCTGATGTAAGTCTGAGTGTGACCGAG AAGGTAGACAATTGCAGCCTCGGGAG
<i>BCL6</i>	NM_001706.2	675-775	GTTGTCGACACTTGCGGAAGTTATTAGGCCAGTGAAGCAGAGATGGTTCTGCCATCAAGCCTCTCGTA AGAGTCTCAACAGCGGATGCTGA
<i>BCL6B</i>	NM_181844.3	2135-2235	CTTTATTGTTCTAGGGCAGCTGGACATGCCGGATTGGAATGGGTAGGAACCCCTCTGGTATTCT GGATGTTGAGAGTCTAGCAGTCT
<i>BHLHE41</i>	NM_030762.2	655-755	CGGCCATTGCGACTGGATGCCCTCACTCGGATTCAACATGCCAAAGAAGTCTGCAATACCTC CCCCGTTGAGAGCTGGACACCCAG
<i>BID</i>	NM_197966.1	2095-2195	GCTTAGCTTAAACAGTCACACTGGTCTGCTGTTCCAGTGGTAAGCTATGCCAGGAATCAGTTAAA GCACGACAGTGGATGCTGGTCCATA
<i>BIRC2</i>	NM_001166.3	1760-1860	TGGGATCCACCTAAGAACATGCTCCAATGAGAAACAGTTTGCAACATTATCTCCACCTGGAAACATA GTAGCTTGTCAAGTGGTCTACTC
<i>BLK</i>	NM_001715.2	990-1090	AGCTTCTGCTCCAATCAACAGGCCGGCTCTTCTATCAGAGAGGTGAACCAACAAAGGTGCCTCTCC CTGCTGAAAGATGTCACCCACCA
<i>BMI1</i>	NM_005180.5	1145-1245	CCTGGAGAAGGAATGGTCACTTCCATTGAAATACAGAGTTCGACCTACTTGAAAAGAATGAAGATCAGTCAC CAGAGAGATGGACTGACAAATGCTGG
<i>BNIP3</i>	NM_004052.2	325-425	CACCTCGCTCGCAGACACCACAAAGAACCAACAGGGCTCTGAAACAGAACCATGATTGGAGAGAAA AGCTCACAGTCTGAGGAAGATGATAT
<i>BTLA</i>	NM_001085357.1	890-990	GCACCAACAGAATATGCATCATATGTTGAGGAGTTAAGTCTGACTCCAACAGGGACATTGAATG TCAGCATGTTGACATCATTGTCGGG
<i>C21orf33</i>	NM_004649.5	1340-1440	TTGAGTAACTACGCTAAGGGGATTCTAAAGCAGGAATCCCTGAGCCGAGAGATAAACGCCCTCCAAA ATGCCAACCTCCACAGCCACATTTC
<i>CA2</i>	NM_000067.2	575-675	AGCTGTGAGCAACCTGATGGACTGCCGTTCTAGGTATTTTGAAAGGTTGGCAGCGCTAACCGGGCTTC AGAAAGTTGATGTCGGATTC
<i>CA9</i>	NM_001216.2	960-1060	CAGGTCAGGACTGGACATATCTGCACTCCTGCCCTGACTTCAGCCGCTACTTCAAATGAGGGCTCT GACTACGCCCTGCCCAGGGTG
<i>CARD9</i>	NM_052813.2	1850-1950	CGCTGACTTGGCTGGAACGAGGAATCTGGTCCACTGAAAGGCCAGCCGACTGCCGGATTGGGCCGT TTGTTAACGGGCACTATTGCGAGG
<i>CASP1</i>	NM_033292.2	575-675	ACAGGCATGACAATGCTCTACAAATGGGTACAGCGTAGATGAAAAAAATCTACTGCTTGGACAT GACTACAGAGCTGGAGGCAATTG
<i>CAT</i>	NM_001752.2	1130-1230	ATGCTTCAGGGCCCTTTGCCCTATCTGACACTCACGCCATGCCGGACCCATTATCTCATATACC TGTGAACCTGCCCTACCGTCTCGAG
<i>CBLB</i>	NM_170662.3	3195-3295	TAATGTCGAAGTGGCCGGACATCTCCGAGAAATTGCCCTCCCTCCAGTATCCCCACGCTAAATCTATA GCAGCCAGAACATGAGACACAAAA
<i>CCBP2</i>	NM_001296.3	1345-1445	GAACAGATGGGAACCAAGCTCAATTGGGTGTCCACTCAAAGTCTCTCCAGGGCCTCAGTACTGTTGC TAAACCCAGTGGTCAGTCTCAGTCT
<i>CCL3</i>	NM_002983.2	681-781	CTGTGTAGGCAGTCATGCCACCAAAGCCACCAACTGACAATGTTATGGATGCTTTGTTCAAGGCTGTGA TCGGCCTGGGAAATAAAAGATGC
<i>CCL4</i>	NM_002984.2	35-135	TTCTGCAGCCTCACCTCTGAGAAAACCTTGTGCCACCAATACCATGAAGCTCTCGTGAUTGCTCTC CTCATGCTAGTGTCTCTGCTC
<i>CCL5</i>	NM_002985.2	280-380	AGTGTGTGCCAACCCAGAGAACGAAATGGGTGGAGTACATCAACTCTGGAGATGAGCTAGGAGAG TCCCTGAACTGAACTTACAAACATT
<i>CCNB1</i>	NM_031966.2	715-815	AACTGGAGGAAGAGCAAGCAGTCAGACCAAAATACCTACTGGTCGGAAGTCACTGAAACATGAGAGCCAT CTAATTGACTGGTAGTACAGGTCA
<i>CCND1</i>	NM_053056.2	690-790	TTGAAACACTCCCTCCAAAATGCCAGGGCGAGGAGAACAAACAGATCATCCGAAACACGCCAGACCTT CGTGGCCCTCTGTCACAGATGTGA
<i>CCR1</i>	NM_001295.2	535-635	CATCATGGGCTGCCATCTGGCTTCCATGCCAGGTTACTTCCAAGACCCATGGAAATTCACT ACCACACCTGCCAGCCTTCACTT
<i>CCR2</i>	NM_001123041.2	20-120	ACATTCTGTTGCTCATATCATGCAAAATTACTAGTAGGAGAGCAGAGATGGAAATGTTCCAGGTATAAG ACCCACAAGATAAAAGAAGCTCAGAG
<i>CCR4</i>	NM_005508.4	35-135	GGTCCTCTTAGCATGTCCTCTGAGCAAGCTGCCATTGCCACAGACCTCTCAGAGCCGTTTCA AAAGCAAGTGTCTGGTGGGCC
<i>CCR5</i>	NM_000579.1	2730-2830	TAGGAACATACTCAGCTCACACATGAGATCTAGGTGAGGATTGATTACCTAGTAGTCATTCTAGGGTTGG GAGGATTCTAGGAGCAACCAAG
<i>CCR6</i>	NM_031409.2	935-1035	CTTAACTGCGGGATGCTCTGACTTGCATTAGCATGGACCGGTACATGCCATTGACAGCGACTAAGT CATTCGGCCTCCGATCAGAACACTA
<i>CCR7</i>	NM_001838.2	1610-1710	TTCCGAAACACAGGCCCTATCTCAAGACAGAGATAGTGGGAGACTTCTGGCTGGTGGAGAACAGCGGA CATCAGCTGGTCAAACAAACTCTGA
<i>CD160</i>	NM_007053.2	500-600	TTGATGTTCACCTAAAGCCAAGTCACACCGTGCACAGTGGACCTACAGTGTGTGCCAGAACCCAGAAGTC AGGTATCCGCCTCAGGGCATT
<i>CD19</i>	NM_001770.4	1770-1870	AGATTCAACACTGACTGAAATCTGAAGACCTCGAGCAGATGATGCCAACCTCTGGAGCAATGTTCTAGGA TGTGTGACTGTGTAAGTGTGTG
<i>CD19R-scfv</i>	SCFV013.1	204-304	GGCACCGACTACAGCCTGACCATCTCAACCTGGAGCAGGAGACATGCCACCTACTTGTGCCAGCAGGGCA ACACACTGCCCTACACCTTGGGGCG
<i>CD19RCD28</i>	MDA_00002.1	2-102	CAGGTGTTCTGCAAGATGAACAGCCTGCAACGCCATGGACTACTGTGCCAAGCAACTACTA CGGCGCAGCTACGCCATGGACTACT
<i>CD2</i>	NM_001767.2	1400-1500	TGGGTCTCACTACAAGCAGCCTATCTGCTTAAGAGACTCTGGAGTTCTATGTGCCCTGGTGACACTGCC ACCATCTGTGAGTAAAGTGAATA

<i>CD20-scfv</i> (ruxolimab)	SCFV002.1	8-108	GCTGCCCCAGAGCCCCGCCATCTGAGCGCCAGCCCTGGCGAGAAGGTGACCATGACCTGCCGGCAGCA GCTCTGTGAGCTACATGCAGTGGTATCG
<i>CD226</i>	NM_006566.2	163-263	TAACAGGATACGATAAAAGCTTAACCAAGACGCAGATGGGAAGAACGTTAGGCAGCACACTCACAT CTCAAGAACAGCCTTCAGACAGT
<i>CD244</i>	NM_016382.2	1150- 1250	AAGAGAACACAGCCCTCCTCAATAGCACTATCTATGAAGTGATTGGAAAGAGTCACCTAAAGCCAGAA CCCTGCTGATTGAGCCAAAGAGC
<i>CD247</i>	NM_198053.1	1490- 1590	TGGCAGGACAGGAAAACCCGTAATGTAAGGACTGTGCGTGTACAGGGCACAGGCCATGGATGGA AAACGCTCTGCTCTGCTTTTCT
<i>CD27</i>	NM_001242.4	330-430	CCAGATGTTGAGCCAGGAACATCCTCGTAAGGACTGTGACCAGCATAGAAAGGCTGTCAGTGTATCCT TGATCACGGGGTCTCTCTCCT
<i>CD274</i>	NM_014143.2	684-784	TAGGAGATTAGATCCTGAGGAAAACCATACAGTGAATTGGTATCCCAGAACACTCTGGCACATCCTCAA ATGAAAGGACTACTTGGTAATTCTG
<i>CD276</i>	NM_001024736.1	2120- 2220	ACATTCTAGGACACAGTACACTGACCACATCACCACCTCTCTCCAGTGCGTGGACCACATGGCTG CTTTTCTCCAAAAGATGCAATAT
<i>CD28</i>	NM_006139.1	305-405	GCTTGTAGCGTACGACAATGCGTCACCTTAGCTGAAGTATTCTACAATCTCTCAAGGAGTCCGG CATCCCTCACAAGGACTGGATAGT
<i>CD300A</i>	NM_007261.2	0-100	CGGGGAAGTGGAGTCGGGGATCAGTCTGCAAGCTACGGAGTCACTACAGGAGGGTCTCATCAGAA ATAGCCGAAGAACCTGAGCCCTAACCA
<i>CD38</i>	NM_001775.2	1035- 1135	CCTGACTCCTGTGGTTATGTATCATACATGACTCAGCATACTGCTGGTGCAGAGCTGAAGATTGGAG GGTCTCCACAATAAGGTCATGCC
<i>CD3D</i>	NM_000732.4	110-210	TATCTACTGGTGAAGTCCCGTGGAGATGGAACATAGCACGTTCTCTGGCTGTGGTACGGTACCCCT CTCGAAGTGAAGCCCTTCAGATAC
<i>CD3E</i>	NM_000733.2	75-175	AAGTAACAGTCCCAGTAAACAAAGATGCACTGGGACTCACTGGAGAGTTCTGGCCTCTGCCTTATCAGT TGGCGTTGGGGCAAGATGGTATG
<i>CD4</i>	NM_000616.3	835-935	AGACATCGTGGTCTAGCTTCCAGAAGGCCTCAGCATAGTCTATAAGAAAGAGGGGAAACAGGTGGAGTTC TCCTCCCACTCGCTTACAGTGAA
<i>CD40LG</i>	NM_000074.2	1225- 1325	GCATTGATTATCAGTGAAGATGCAAGAGGGAAATGGGGAGCCTCAGCTCACATTAGTTGACTCTG GGTCCCTATGGCTTGTGGAGGGG
<i>CD44</i>	NM_000610.3	2460- 2560	GTGGGAGAAAAGCTAGTGTACACAGTGGCAATGGAGCTGTGGAGGACAGAAAGCCAAGTGGACTCAA CGGAGGGCAGCAAGTCTAGGAAT
<i>CD45R-scfv</i>	SCFV006.1	222-322	TTACCCCTGAACATCACCCTGGCTCCGGCACCAAG CCCTTCACCTTCGGCTCCGGCACCAAGC
<i>CD47</i>	NM_001777.3	897-997	GCCATATTGGTATTAGGTGATAGCCTATCTCGCTGTGGTGGACTGAGTCTGTATTGGCGTGTAT ACCAATGCGATGCCCTCTCTGATT
<i>CD56R-scfv</i>	SCFV008.1	197-297	ATTCAAGCGGCTCTGGCTCCGACCTCAGTCTGATGATCTCGGGTGGAGGACCTGGCG GTACTACTGCTTCAGGGCAGCCACGT
<i>CD58</i>	NM_001779.2	478-578	GTGCTTAGTCTTCCATCTCCACACTAAGTGTGACTAATGGAAGCATTGAAGTCCATGCGATA CAAGACATTACACAGCCATGAG
<i>CD63</i>	NM_001780.4	350-450	GTCATCATCGCAGTGGTGTCTCTCTCCGGCTTTGGCTGTGCTGGGGCCTGCAAGGAGAACT ATTGCTTATGATCACGTTGCCATCT
<i>CD69</i>	NM_001781.1	460-560	AGGACATGAACCTTAAACGATACGCAGGTAGAGAGGAACACTGGTTGGACTGAAAAGAACCTGGCA CCCATGGAAGTGGTCAATGCCAAAGA
<i>CD7</i>	NM_006137.6	440-540	CCTACACCTGCCAGGCATCACGGAGGCTAATGTCAGGCTCCGCACCTGGTGTGACAGAGGAAC AGTCCAAGGATGCCACAGATCTCGGA
<i>CD80</i>	NM_005191.3	1288- 1388	AAAGATCTGAAGGCTCCACCTCCATTGCAATTGACCTCTCTGGGAACTTCCTCAGATGGACAAGATTACCC ACCTGCCCTTACGTATGCTCTT
<i>CD86</i>	NM_006889.3	146-246	TATGGGACTGAGTAACATTCTTTGTATGGCCTCTGCTCTGGTGTGCTCTGAAGATTCAAGCTTA TTCAATGAGACTGAGACCTGCCA
<i>CD8A</i>	NM_001768.5	1320- 1420	GCTCAGGGCTCTTCCCTCACACCTCAGGTCTTCTCCAGGCCCCCTGTCAGGGTGAGGTGCTTGAGT CTCCAGGCGCAAGGAAACAAGTACTT
<i>CDH1</i>	NM_004360.2	1230- 1330	CGATAATCTCCGATCTCAATCCACACCGTACAAGGGTCAAGGTGCTGAGAACGAGGCTAACGTGTAATCA CCACACTGAAAGTGAATGCTGCTGAT
<i>CDK2</i>	NM_001798.2	220-320	TCGCTGGCGCTCATGGAGAACCTTCAAAGGGGAAAGTCGGAGAGGGCACGTACGGAGTTGTGACAAA GCCAGAAACAAGTGAAGGGAGAGGT
<i>CDK4</i>	NM_000075.2	1055- 1155	ACTTTAACCCACAAGCGAACCTGCGCTTCAGCTGAGCTCTGCAGCACTTATCATAAGGATGAAGTAAT CGGGAGTGAAGCAATGGAGTGGTGC
<i>CDKN1A</i>	NM_000389.2	1975- 2075	CATGTGCTGGTCCCTCCAGTCTCCACCTAGACTGTAACCTCTGAGGGCAGGGACCACACCCTGACTGTT TGTGCTTACAGCTCCCTCCACAA
<i>CDKN1B</i>	NM_004064.2	365-465	GCTTCCGAGAGGGGTTGGCCCGTGAAGGGCGTTGTTGTTGGTTGTTGGAGAGTGCAGAG AGGGCGTGTGCAAGACCCGGAGAACAG
<i>CDKN2A</i>	NM_000077.3	975- 1075	AAGCGCACATTGAGGGCTTCTGCGAGGCTCGAGCCTCCGGAAGCTGCACTCATGACAAGCATT TGTGAAGTGGAGGACTCAGGGGG
<i>CDKN2C</i>	NM_001262.2	1295- 1395	ATAATGTAACGTCATGCACAAAATGGATTGGAAGGACTGCCCTGCAGGTATGAAACTGGAAATCCGAG CTGCCAGGAGACTGCTACTAGAGG
<i>CEBPA</i>	NM_004364.2	1320- 1420	GAGCTGGGACCCGCAACTCTAGTATTAGGATAACCTTGCCCTGGAAATGCAAACCTACCGCTCCAATGC CTACTGAGTAGGGGAGCAAATCGT
<i>CFLAR</i>	NM_003879.3	445-545	CAAGACCCCTGTGAGCTCCCTAGTCTAAGAGTAGGATGCTGCTGAGTCATCCATCAGGTTGAAGAAGCACT TGATACAGATGAGAAGGAGATGCTG
<i>CFLAR</i>	NM_001127183.1	653-753	TAGAGTGTGATGGCAGAGATTGGTGAAGGATTGGATAATCTGATGTGTCCTCATTAATTCCTCATGAAGGA TTACATGGGGAGCAAGATAAGC

<i>CHPT1</i>	NM_020244.2	1303-1403	GATATGGTATATACTTTAGTGTGCTGCAATTCAAGACACCTCATCTAAATATTCAGACTGCAT GTCATCAAGCACCTGAACAGGTC
<i>CIITA</i>	NM_000246.3	470-570	GCCTGAGCAAGGACATTCAAGCACATAGGACCAGATGAAGTGATCGGTAGAGATGGAGATGCCAGCAGA AGTTGGGAGAAAAGTCAGAAAAGAC
<i>CITED2</i>	NM_006079.3	965-1065	AGGAGCTGCCGAACTCTGGCTGGGCAAACAGAGTTGATTTATGACGGACTCGTGTGCAAACAGCAGCC CAGCAGAGTGAGCTGTTGACTGATCG
<i>CLIC1</i>	NM_001288.4	310-410	GTATGGGCAAGATTGGAACTGCCATTCTCCAGAGACTTGTACTGTGGCTAAGGGAGTCAC CTTCATGTTACACCACCGTGAACACAA
<i>CLNK</i>	NM_052964.2	1108-1208	GAAGGAGAACAGGATGGTAGTTCTGGTCCAGATTGTTCACAAAATCCAAGGAAGAGCCATGTTGG CTGTGTTTATGAGAACAAAGTAC
<i>c-MET-sfv</i>	SCFV004.1	138-238	CTATCTACGCCACAGCTGAAGAGCGCGTCCCAGCCGTTAGCGCTCTGGCTTGGCGCAG TTCACCTGACCATCAGCAGCTGCAGC
<i>CREB1</i>	NM_004379.3	4855-4955	TTTATGGTAGGTACAGCAGCTGCTAGTCTCTGAAAGCACAATACCACTGAGCAGCCTATCCATCAGATGT CATCTGGCTGAATTATCTCTGCT
<i>CREM</i>	NM_001881.2	260-360	CTCCACCTCCCGCGTAAGTACGTGACGAGGTCCGCTACGTAATCCCTTGCAGGACAAATGACCAT GAAACAGTGAATCCACAGATGATGG
<i>CRIP1</i>	NM_001311.4	269-369	CAACCCCCCTGCTACGCAGCCATTGGCCTAAAGCTTGGCGGGGGAGCCAGAGCCACACTT CAAGTAAACACAGGTGGAGACCCCAT
<i>CRLF2</i>	NM_022148.2	1420-1520	CAAGGCAGCACGTCAAAATGCTGAAACCATCTCCACTCTGTAGTCCCAGTCCGTACCTGT TCCATAGCATTGGATCTCGAGGAT
<i>CSAD</i>	NM_015989.4	205-305	TCAAATTCTGCTAGCCTAGCATTAGAGAGGGCTGTCAAAGATGGACTGCAAATGCCTGATGGA AGGAGATGTCATTCCACTGAAGATCC
<i>CSF2</i>	NM_000758.2	475-575	AGATGAGGCTGCCAACCTACATGATGGGAATGAGTTGACCTCTAGATGATTCCATCTGACGGCACCTCA AGGGACCAAGGGTGGCCACAGCAT
<i>CSNK2A1</i>	NM_177559.2	1930-2030	CACTCCACCATTTGCTCCACCGTCCCACATTAGGGGTTGGATCTCGTCTCTCCAGAGATTAC AAAAATGAGCTTCAGGGAGGCA
<i>CTGF</i>	NM_001901.2	1100-1200	ACCACCTGCGGGTGAGCTCAAGTGCCTGACGGCGAGGTCAATGAGAACATGATGTTCAAGACCT GTGCGTGCATACACTGTCGGAG
<i>CTLA4</i>	NM_005214.3	405-505	AGTCTGCGGCCAACCTACATGATGGGAATGAGTTGACCTCTAGATGATTCCATCTGACGGCACCTCA GTGAAATCAAGTGAACTCATCT
<i>CTNNA1</i>	NM_001903.2	75-175	TCGCCCAGCTAGCCGAGAAATGACTGCTGCATGCAGGCAACATAACTCAAGTGGATCTAAAGCTA GAGATCAGGACTCTGGCAGTTGAGAG
<i>CTNNB1</i>	NM_001098210.1	1815-1915	TCTTGCCTTGTCCCGAAATCATGCACCTTGCCTGAGCAGGGGCCATTCCACGACTAGTTGCTT TTCGTGACATCAGGATACCCAGCGC
<i>CTNMBL1</i>	NM_030877.3	855-955	TGATGCCAACAAACTGTATTGCACTGAAGTGCCTGCATATTGCCAGGACAATGATGAAAACAGGGAAATTG TGGGGAGCTGGATGGAATCGATGTG
<i>CTSC</i>	NM_001114173.1	260-360	TGCTCGGTTATGGACCACAAGAAAAAAAGTAGTGGTGTACCTTCAGAAGCTGGATACGCATATGATGACCT TGGCAATTCTGCCATTTCACCATCA
<i>CTSD</i>	NM_001909.3	1495-1595	GAAGCCGGCGGCCAACCGCCGACTGTGTTGTTCTGCCATCTGGTTCCCTCCGGTCAGAAATGCTGCC TGCGTGTCTCTCCATCTGTTGG
<i>CX3CL1</i>	NM_002996.3	140-240	AGCACCCAGGTGACGAAATCCAACATCACGTGCAGCAAGATGACATCAAAGAACCTGTAGCTTGCATC CACTAACACAAACAGGCATCG
<i>CX3CR1</i>	NM_001337.3	1040-1140	GGCGCTCAGTCCACAGTGAATCACAAGGAGCAGGCATGAAAGTGTCTGAGCAGCA ATTITACTTACACACAGTGTAGTGGAG
<i>CXCL10</i>	NM_001565.1	40-140	GCAGAGGAACCTCAGCTCACCCATTAGACCTGATCAAACCTGCGATTGATTGCTGCCTATCTTGACTCTAA GTGGCATTCAAGGAGTACCTCTC
<i>CXCL12</i>	NM_199168.2	505-605	GGGCCTGAGGTTGCCAGCATTAGACCCCTGCATTATAGCATACGGTATGATATTGAGCTTATATTCCAT GCCCTGTACCTGTGACCTTGGAA
<i>CXCL9</i>	NM_002416.1	1975-2075	CACCATCTCCCATGAAAGAACGGGAACGGTAAGACTAACGCCTAGAGGAAGCAGCCAAGTCGGTAGTGGAA GCATGATGGTGGCCAGTGGCTC
<i>CXCR1</i>	NM_000634.2	1950-2050	GCAGCCACCGTCATTGGGCAAGCAGATGTCCTAATAAGCTCTGTCCTGCTGTTGGAGTAT CTTGGTGTGACAGACTGAGGTGT
<i>CXCR3</i>	NM_001504.1	80-180	GTGAGTGCACCCAAGTGTAAATGACGCCAGGGTGCCTCCCTGGAGAACCTCAGCTTCCATGACT ATGGAGAAACAGAGTACTGCT
<i>CXCR4</i>	NM_001008540.1	135-235	GTCACATGGAAAAGATGGGAGGAGGTTGAGGATTCTACATTAATTCTTGTGCCCTAGCCCACT TCAGAATTCCCTGAAGAACAGCAAC
<i>DAPL1</i>	NM_001017920.2	190-290	CGAGAAAACAGTGCCTACATTGCAATGTCAGGAAATACAGACACTGGATGCCCTGAATGACGCACTGGAGAAG CTCAACTAATTTCAGCAACAGT
<i>DEC1</i>	NM_017418.2	190-290	AGGCCCTACTTCCAGATCCAGATCTGTGCTACACTGACTTGTGGGTAGGCTGCAGAAAAATCAG CTAGAACAGCCTGGGGTAGTGGCA
<i>DECTIN-1R</i>	SCFV010.1	270-370	CTGAAGATGCACAGCAGCAACGAGCTGGCTCATGTGAAGCAGGTGTCAGCCAGCCGACAACCTCT GGATCGGCCTGAGCAGGCCAGACCG
<i>DGKA</i>	NM_001345.4	1375-1475	TTCTAACACCCACCACTCTCTTGTCAATCTAAGAGTGGGGAAAGCAGGGCAAGGGTGTCTG GAAGTGGCAGTATATTAACCCCTG
<i>DOCK5</i>	NM_024940.6	630-730	TGGCAGAGTACAATGGGAACTCTCTAGACCCCTGACGAAACAGCAGCATTGCCCTTCAAGGCCATGAGGT GCCCTCAAAAGGAGTTGAGGAAAGAT
<i>DOK2</i>	NM_003974.2	650-750	GCCAGGGACCCAGCTGTACGACTGGCCCTACAGGTTCTGCGCGCTTGGGGGAGCAAGGTAACCTT TTGAGGAGCAGGCGTGCCTCT
<i>DPP4</i>	NM_001935.3	2700-2800	CAGCAGTCAAGCTCAGATCTCAAAGCCCTGGTCATGTTGGAGTGGATTCCAGGCAATGTT AGACCATGGAATAGCTAGCAGCACAG

<i>DUSP16</i>	NM_030640.2	615-715	ATGGGTTAACTCTCTTGCAGTCACCACGCCCTACACTTTAGTACAATGGAGTGGCTGAGCCTGACCATCACCTGGCCGTCAGCC
<i>EGFR-scfv_(NIMO CAR)</i>	SCFV015.1	7-107	AGATGACCCAGAGCCCTAGCAGCCTGAGCGCCAGCGTGGGCACAGAGTGACCATCACCTGGCCGTCAGCC
<i>EGLN1</i>	NM_022051.1	3975-4075	AGCAGCATGGACGACCTGATACGCCACTGTAAAGGGAAAGCTGGGCAGCTACAAAATCAATGGCCGGACGAAA
<i>EGLN3</i>	NM_022073.3	800-900	CAGACCTCTGGGATCCTGGGATATTCCAGAGGGAAATCATTCA TAGCAGATGTGGAGGCCATTNTGA
<i>EIF1</i>	NM_005801.3	869-969	CCTGAACAGTCCTCGGTGAATCTGAGAGGAGGGATGGGTAAGGCAGAACAGCATGCTATTACCTACAGCCCAGT
<i>ELF4</i>	NM_001421.3	335-435	GAGCTTGGAGGGCTCTGATAATCCCGTTGACGCTCTGAAAAGACAGCATGCTATTACCTACAGCCCAGT
<i>ELOF1</i>	NM_032377.3	125-225	AGACCCAGTCACCTGCCCTCTGCAACCACAGAGAACTCTGTGATGTGAAATGGACCGTGCACAC
<i>ENTPD1</i>	NM_001776.4	225-325	CGGAGTCATCTTGTACCGTGTGCT
<i>EOMES</i>	NM_005442.2	1670-1770	TTCGAGTAACTTAGGAAATAGCCTGCTGGACTCCTCAGTCATCTGCCTTCTAGTCATGAAAAAGACAGG
<i>EPHA2</i>	NM_004431.2	1525-1625	ATCCCATGCCCTGGGATTACCCAGACCCAACCTTCTGCAATGGCAGGGTGGGAGGTGAGGTTCTAC
<i>EPHA4</i>	NM_004438.3	20-120	CGAGGGTTCTCGTGCACCTGAGC
<i>EPHB2</i>	NM_017449.2	785-885	CAAGCAGGCTCGAGGGCTTGAAGATGGCACCGTCTGCCGAGGTTGTCATCTGGACTTTCAAGGCCAAC
<i>ETV6</i>	NM_001987.4	3840-3940	CAAGGGGATGGGCTGTACCCACTG
<i>FADD</i>	NM_003824.2	1560-1660	TGAGACTGCTAAGTAGGGCAGTGATGGTGCAGGACGAATTGAGATAATATGTCAGTCTGGCTCTTACCCA
<i>FAM129A</i>	NM_052966.2	3526-3626	ATTGACACACACTCTAAATCTT
<i>FANCC</i>	NM_000136.2	2130-2230	TGCCCCATAGATCAAGAGAAGCTAACCGGAATGGAGGGTGAAGGTGATCTGGACTGTCTGGCCT
<i>FAS</i>	NM_000043.3	90-190	GTACTCATGAGATTTGTCAGTCAATGACTCAAGTGAGCTTCGCTCTGATCTCGCGAAGAGTGACACACAGGTCAAAGACGTTCTGG
<i>FASLG</i>	NM_000639.1	625-725	GGAGTGAGGGAGCGGTTACAGGTGA
<i>FCGR3B</i>	NM_000570.3	73-173	TCCATGCTCTGAATGGGAAGACACCTATGGAATTGCTCTGCTTCTGGAGTGAAAGTATAAGGGTGGCCT
<i>FGL2</i>	NM_006682.2	250-350	TGTGATCAATGAAACTGGGCTACT
<i>FLT1</i>	NM_002019.2	5615-5715	GTGAGACTGCTTGAAGTCTGCTGGGCTGAGCATGATGGAAAGGGAGACAGGGTAGGAAAGGGCGCTA
<i>FLT3LG</i>	NM_001459.2	927-1027	CTCTCGAGGTCTAAAGATCAAGTGG
<i>FOS</i>	NM_005252.2	1475-1575	CCCATGAAATTGATATAAATCATCCT
<i>FOXO1</i>	NM_002015.3	1526-1626	ACTCAAGTCCTAACCTCTCCGGAGATGAGCTGAAAGCAGCTGGAGTGATTTGTCAGCTCAGAG
<i>FOXO3</i>	NM_001455.2	1860-1960	TCTCATACCAACATCATTAAGTGTGACCCAGTCCTCACCTGGCACCATGATGCAGCAGCCGTCTAC
<i>FOXP1</i>	NM_032682.5	6758-6858	GGGGCATCTGGAGGCTCCAGAGAACGGCGACACTCAATGAGATCTACCACTGGTTCACACCGATGTTGC
<i>FOXP3</i>	NM_014009.3	1230-1330	TCTTCAGAACCATCTGCCACCTG
<i>FYN</i>	NM_002037.3	765-865	GTCTTGGAGGTGTGAAGTCTCTCATACGGGGACCTGCGTACGAGAGGAACAGGAGTGACACTCT
<i>FZD1</i>	NM_003505.1	2430-2530	TTGTGGCCCTTATGACTATGAAC
<i>G6PD</i>	NM_000402.2	1155-1255	GTGCAACTCTGACATCTCGAGGTTCTCACTAGACAACTCTTCGAGGCTCTTGAACAACTCAGCTC
<i>GABPA</i>	NM_002040.3	1160-1260	CTGAAAAGCTCTCGCTTGCAGCA
<i>GADD45A</i>	NM_001924.2	865-965	ATTCGGGATCATCCGGACGTGATGCA
<i>GADD45B</i>	NM_015675.2	365-465	TTGTCAGCGTGGCTCTGCCCTTGCCATTGACGAGGAGGAGGAGGATGACATGCCCTGCA
<i>GAL3ST4</i>	NM_024637.4	1140-1240	AATCCACTTCAGCCTCATCCAGCTTC
<i>GAS2</i>	NM_005256.3	915-1015	CGAGCCCAAACCCATCCAAATGCCCTCATCCATCTGTTCCACTGTACTGATCATCGCAGCCAGATATCA
			ACTACTTGGTGGCTCTGCCAGTT

<i>GATA2</i>	NM_032638.3	1495-1595	GAAGAAGGAAGGGATCCAGACTCGAACCGGAAAGATGTCCAACAAGTCCAAGAAGAGCAAGAAAGGGCGGA GTGCTTCAGGAGCTGTCAAAGTGCATG
<i>GATA3</i>	NM_001002295.1	2835-2935	AAGAGTCGGCGGCATCTGTCTGCCATTCTGCAGCCTGTGCTGAGGGTAGCAGTGTATGAGCTACAG CGTCATGTCAAGCGACCCCTGGCCGAC
<i>gBAD-1R-scfv</i>	SCFV001.1	1-101	AGACAGACACCCCTGCTCTGGGTGCCACCTGTGGCGACATCGTGTGAGCAGAAGCCCCAGCAGCC TGGCGTGTCCGGCGAGAAAGTGAC
<i>GEMIN2</i>	NM_001009182.1	537-637	ACAAGCAACAGTAACTAGTGTCTGGAATATCTGAGTAATTGGTTGGAGAAAGAGACTTACTCCAGAATTGGG AAGATGGCTTATGCTTATTGGCT
<i>GF11</i>	NM_005263.2	2235-2335	TCATCACTGGAGGTAAGGACAAAGCAATGCCGTGACAAGATGTCATTCACTCAGCAAATGTTCATGG ATCACCGGCTTACAGGTACAGGCA
<i>GLIPR1</i>	NM_006851.2	255-355	CTGCTTCGAATCATAACAACTCGATCAGGGTAAACCAACAGCCAGTGTATGCTATACATGACTTGGG ACCCAGCACTAGCCAAATTGCAAA
<i>GLO1</i>	NM_006708.1	1240-1340	GGAAATGATATGGTACCCAGACACTGGGCTAGGCTGCAACTTATCTCATTTAATACTCCAGCTGTATGTA GAAAGAAAAGCAGGCTAGGCATGTGAA
<i>GNLY</i>	NM_006433.2	305-405	CAGGAGCTGGCCGACTACAGGACCTGTGACGATAGTCCAAAAGTGAAGAAGATGGTGATAAGCCCA CCCAGAGACTTCCAATGCTGCGA
<i>GSK3B</i>	NM_002093.2	925-1025	ACTGATTATACCTCTAGTAGATGTATGGCTGCTGGCTGTGTTGCTGAGCTTACTAGGACAACCAATA TTTCAGGGGATAGTGGTGTGGATC
<i>GZMA</i>	NM_006144.2	155-255	AGACCCCTACATGGTCTACTTAGTCTGACAGAAAACCATCTGTGCTGGGCTTGTGAAAGACTGGG GTTGACTGCAGCTACTGTAACTGA
<i>GZMB</i>	NM_004131.3	540-640	ACACTACAAAGAGGTGAAGATGACAGTGCAGGAAGATGCAAAGTGCATACTGACTACGCCATTATTACAGAC TACCATGGTGTGCGTGGGGAC
<i>GZMH</i>	NM_033423.3	705-805	AAAAAGGGACACCTCCAGGAGTCTACATCAAGGTCTCACACTCCTGCCCTGGATAAAGAGAACATGAAGCG CCTCTAACAGCAGGCTGAGACTAC
<i>HCST</i>	NM_001007469.1	132-232	ATCCTCTCCCTGCTTGTCCCAGTGGCTGCGACTCAGACGACTCCAGGAGAGAGATCATCACTCCCTGCC TTACCCCTGGCACTTCAGGCTCTGTT
<i>HDAC1</i>	NM_004964.2	785-885	CAAGCCGGTATGTCAAAGTAATGGAGATGTTCCAGCCTAGTGCCTGGCTAGCTGAGACTCC CTATCTGGGATCGGTTAGGTGCTTC
<i>HDAC2</i>	NM_001527.1	930-1030	AAGCCTTATTCCTAAAGGTGATGGAGATGTATCACCTAGTGTGTTACAGTGTGGTATTACAGTGTGGTCA CTGGTGTGAGACTCATT
<i>HER2-scfv</i>	SCFV014.1	64-164	CCTGCAGGCCAGCAGCAGCTGCTTACATGCAGTGGTACAGCAGAAGTCCGGCACTAGCCCCAAGCGT GGATCTACGACACCTACAAGCTGGCCAG
<i>HERV-K 6H5-scfv</i>	SCFV012.1	137-237	CGGCGGCCACAGCTACAACAGAACAGTTCAAGGACAAGGCCATCTGACCGTGACAAGAGCAGCAGCACCG CTACATGGAACCTCGGGCTGACCGAC
<i>HLA-A</i>	NM_002116.5	1000-1100	GGAAAGGCTCAGATAAAAAGGAGGGATTACACTCAGGCTGCAAGCAGTGCAGTGCAGTGCCTGATGT GTCCTCACAGCTTGTAAAGTGTGAGA
<i>HMGB2</i>	NM_001130688.1	125-225	CTGTCACATGGTAAAGGAGACCCAAACAGCCGGGGCAAATGTCCTGTCAGCCTTCTCGTCAGAC CTGGGGGGAAAGACAAAGAACAA
<i>HOPX</i>	NM_001145460.1	1117-1217	AAACATAGGAAGCTATGTATCTGTGTAAGCAGTGGCTACTGGAAAAATGGTGTGGCTAGCATTT CTTGAGTCATGATGACAGATGGTGT
<i>HOXA10</i>	NM_018951.3	1503-1603	TTCCTATAGAGATAGATAATGTCCTAAGTGTCAAGCTGACTGGGCTGGTTGCTGTCTGGGCTCCACTGC TCGAATGGCCCTGCTTCCGGCCGA
<i>HOXA9</i>	NM_152739.3	1015-1115	GGCTCTAACCTCAGGCCACATCTTCCAAGGCAAACCTGTTCAAGGCTGGCTGCTAGGCCTGCCCTTGAT GGAGGAGGTTGTAAGCTTCATT
<i>HOXB3</i>	NM_002146.4	60-160	TGTCTGTTAAATGCTGCTGGAGACTCGTAAAGGAAACCTGACTGCCAGATAGTGCAGTGTGGCTT ATTCGGTCGGATTGCGGTGTGGTGT
<i>HOXB4</i>	NM_024015.4	1340-1440	CCTTCTTGTCCCCACTCCCACACCCGAAACGCCCTGACTGCCAGATAGTGCAGTGTGGCTT CGGTAAACACACACACTCTCCCTCA
<i>HPRT1</i>	NM_000194.1	240-340	TGTGATGAGGAGATGGAGGACATCACATTGAGCCCTCTGTGCTCAAGGGGGCTATAATTCTTGCTG ACCTGCTGGTATACATCAAGCAGTC
<i>HRH1</i>	NM_000861.2	3055-3155	GTGGCAGCTAAATGATATTTGAGTAGACGAACAGCTGACATGGAGTCCCGTGCACCTACCGAAGGGGA CGCTTGAAGGAACCAAGTGCATT
<i>HRH2</i>	NM_022304.1	600-700	GCGGCCCTCATCTCATACCGGCTGCTGCAATGTGGCTCTGTGCTGGCCGGCTTGAACCGCCGGCTCC GCAACCTGACCAATTGTTCATGTC
<i>Human CD19R-scfv</i>	SCFV009.1	215-315	CTTCACCATCAGCAGCCTGCAAGCCCAGGGACATGCCACCTACTACTGCCAGCAGTACCAAGCCTGCC ACCTCGGCCAGGGCACCAAGTCAG
<i>ICOS</i>	NM_012092.2	640-740	AACTCTGGCACCCAGGCATGAAGCAGTTGGCCAGTTCCCTCAACTGAAGTGAAGATTCTTTCCGG GACCAAGGGAGAGCTGACTAACAC
<i>ICOSLG</i>	NM_015259.4	1190-1290	CTGCTGGCGTTGGCTGTGATCTGGATGAGGCCCTTCAAAAGCGTCATCCACACCAAGGCAAATGCCCC AAAGTGAATGGCTCCCGCTGTCACTG
<i>ID2</i>	NM_002166.4	505-605	CGGATATCAGCATCTCTGCTTGCAGGCTTCTGAATTCCCTCTGAGTTAATGTCATAAGCAGCAA GCTGTCGAATAAGCGGTGTTCATGA
<i>ID3</i>	NM_002167.3	195-295	AGGAAGCCTTGTGCAATTAAAGGGGCTGTGAACGCCAGGCCGGGGCAGGGCCAGGGCG TTTGAATAAGAGCGCTGCTTCCAGGC
<i>IDO1</i>	NM_002164.3	50-150	CTATTATAAGATGCTCTAAACACTCTCAGACACTGAGGGCACCAGAGGAGCAGACTACAAGAATGG CACACGCTATGGAAAACACTCTGGACAATCAGT
<i>IFNA1</i>	NM_024013.1	585-685	ATCCCTCTTATCAACAAACTTGAAGAAAGTAAAGGAGGAGGAATAACATCTGGTCAACATG TTCTTATTGACTCATACACCAGTC
<i>IFNG</i>	NM_000619.2	970-1070	ATACTATCCAGTTACTGCCGGTTGAAAATGCTGCAATCTGAGCCAGTGCTTAATGGCATGT CTTGAATGTGTCAGGTGACCTGAT

<i>IFNGR1</i>	NM_000416.1	1140-1240	CCGGGGCAGCCATCTGACTCCAATAGAGAGAGAGTTTACCTTAAGTAGTAACCAGTCTAACCTGGCA GCATCGCTTAAACTCGTATCACCTC
<i>IGF1R</i>	NM_000875.2	455-555	TGGGGGGGCCATCAGGATTGAGAAAAATGCTGACCTCTGTTACCTCTCCACTGTGGACTGGCCCTGATCCTG GATCGGGTGTCCAATAACTCATGTG
<i>IKZF1</i>	NM_006060.3	4485-4585	CCGCTGTGACTACTGTGTCCTAGATCCATGCACTCTCGTTGTTGAAGTAATATTGGAGACGGAGGG TAACAGGTTGGCTGTGATTACAGC
<i>IKZF2</i>	NM_001079526.1	945-1045	CCATGTACCTCTATGGAAGATTGTAAGGAACAAGACCTATTATGGACAACAATTCTCTGGTCCTTGAAG GAGACTGTGTCATAGAGAACCTC
<i>IL10</i>	NM_000572.2	230-330	AAGGATCAGCTGGACAACCTTGTGTTAAAGGAGTCCTTGCTGGAGGACTTAAGGGTACCTGGTTGCAAGC CTTGTCTGAGATGATCCAGTTTAC
<i>IL10RA</i>	NM_001558.2	150-250	TGCCAGCCCCCGCTGTGTTGAAGCAGAATTTCACACATCCTCACTGGACACCCATCCAAAT CAGTCTGAAAGTACCTGTATGAAGT
<i>IL12A</i>	NM_000882.2	775-875	CTTCTAGATCAAACATGCTGGCAGTTATTGATGAGGCTGATGCAAGGCCGTAATTCAACAGTGAGACTGTGC CACAAAAATCCTCCCTGAAGAACCG
<i>IL12B</i>	NM_002187.2	1435-1535	GCAAGGCTGCAAGTACATCAGTTTATGACAATCAGGAAGAATGCACTGTTCTGATACCAGTGCAC TTGTATGATGGAAACCGAACAGAT
<i>IL12RB1</i>	NM_005535.1	1292-1392	AGGAAAAGTGTACTACATTACCCATTGCGCACCCCGAGAAGCTCACCTGTGGTACCGTCTG TCCACCTACCACTTGGGGCAATGC
<i>IL12RB2</i>	NM_001559.2	1315-1415	CCTCGTGGGACATTAGAATCAAATTCAAAGGCTCTGTGACGAGATGTACCTTATTGGAGAGATGAGGG ACTGGTACTGTAAATCGACTCAGAT
<i>IL13</i>	NM_002188.2	516-616	TTCTTTCTGATGTCAAAATGCTTGGTAGGCGGGAGGAGGGTTAGGAGGGTAAATTCTTAG ACCTCAGCTGTGTCCTTC
<i>IL15</i>	NM_172174.1	1685-1785	AGGGTGTAGTCAAATTATGTTGGGGCTGGTACCAATGTCAGGTCAACAGCTATGCTGGTAGGCT CCTGCAGTGTGAAACACTGACT
<i>IL15RA</i>	NM_002189.2	39-139	CGCTGCCCGGGAGTCCAGGGTGTCTGTGGAGCTGCCCATGGCCCCCGGGCGCGCGCGCTGC CGGACCTCGGTCTCCGGCGCTGACTG
<i>IL17A</i>	NM_002190.2	240-340	TAATACAACCGATCCACCTCACCTTGAATTCACCGCAATGAGGACCTGAGAGATATCCCTGTGATCTG GGAGGCAGAATGCGCCACTTGGG
<i>IL17F</i>	NM_052872.3	210-310	GCCCCCTGTGGCAGGAGGTAGATGAAGCTGACATTGGCATCATCAATGAAACACAGCGCTTCCATGTCA CGTAACATCGAGAGCCGCTCACATC
<i>IL17RA</i>	NM_014339.4	3020-3120	CTACTATGTGGGGCATTTGGATACCAAGATAATTGATGCGATGGCCATGGCCCAGCCATGAAGGAAC CCGCTAGTGTGGAGACAGTAAACG
<i>IL18</i>	NM_001562.2	48-148	GACAGTCAGCAAGGAATTGTCCTCCAGTGCATTGCCCCCTGCGCAACTCTGGCTGCTAAAGCGGCTG CCACCTGCTGCACTCACAGCTCG
<i>IL18R1</i>	NM_003855.2	2025-2125	GAATGAGGGATTAAAGTCTGAAGAGGCATTCTAGGGACAGTGGGTACTGAGTAACGAAATGCTG TTTCACTCCCTAACACATGGATCTG
<i>IL18RAP</i>	NM_003853.2	2412-2512	GCTTGATGGACAATGGAGTGGGATTGAGACTGTGGTTAGAGCCTTGATTCCTGGACTGACGGCGA GTGAAATTCTAGACCTTGGTACTTT
<i>IL1A</i>	NM_000575.3	1085-1185	ACTCCATGAAGGCTGATGGCATCTGTCTCTGAGTATCTGAAACCTCTAAACATCCAAGCTTAC CAAGGAGAGCATGGTGGTAGTGC
<i>IL1B</i>	NM_000576.2	840-940	GGGACCAAAGGCCAGGATAACTGACTTCACCATGCAATTGTGCTTCTAAAGAGAGCTTACCCAGA GAGTCTGTGCTGAATGTGGACTCAA
<i>IL2</i>	NM_000586.2	300-400	AGGATGCAACTCTGTGACTGCACTAAGTCTGCACTTGTCACAAACAGTCACCTACTCAAGTTCTACA AAAGAAAACACAGCTAACACTGGAC
<i>IL21R</i>	NM_021798.2	2080-2180	CGTGTGTTGTCACAGATGACAACAGCCGCTCCCTCCCTAGGGCTTGTGTCAGTTGGCCACAGCAT CTCCGGGGCTTGTGGATCAGGGCA
<i>IL22</i>	NM_020525.4	319-419	CTATCTGATGAAAGCAGGTGCTGAAACTTCACCCCTGAAAGAAGTGTGTTCCCTCAATCTGATAG ATATGCAAGGAGGTGGCTCCCTCTG
<i>IL23A</i>	NM_016584.2	411-511	CAGGGACACAATGCACTGCTTGCACAGGATCCACCCAGGGCTGATTTTATGAGAAGCTGCTAGGATCG ATATTTCACAGGGGAGCCTCTCG
<i>IL23R</i>	NM_144701.2	710-810	ACTGCAATTACCTGGATGATATGATACCTCTGCAGCCGTATTCAGGGCTGAGACTATAATGCTA CAGTGCCTAACAGGATAATTATG
<i>IL27</i>	NM_145659.3	143-243	CAGGAGCTGCCAGGGAGTCACAGTCAGCCTGCATCTGCCAGGAAGCTGCTCCAGGTTGGGGCCAG GCCCAACCGCTTGGGAATCTCACCTC
<i>IL2RA</i>	NM_000417.1	1000-1100	CTTGTGAAAGCAGGGAAACAGACAAAGAGTCATGAAGCCAAAGTGAATCAAAGGTCTAAATGGTC CAGGAGACATCGCTGTGCTGCTGC
<i>IL2RB</i>	NM_000878.2	1980-2080	GTCTGCTGCCAGGCAAGGACTGTGTTGAGGGGGCAGTAACCTCCAACTCCCTGTTAATCACA GGATCCACGAAATTAGGCTCAGAAC
<i>IL2RG</i>	NM_000206.1	595-695	CCACAGCTGACTGAAACATCAGTGGATTAGACATAAGTCTCTGCTAGTGTGGATGGCAGAACGCT ACACGTTCTGTTGGAGCCGCTT
<i>IL4</i>	NM_000589.2	625-725	GACACTCGTGCCTGGGTGCGACTGCACAGCAGTCCACAGGACAAGCAGCTGATCCGATTCTGAAAC TCGACAGGAACCTCTGGGGCTGGCG
<i>IL4R</i>	NM_000418.2	705-805	ATCATCTCACCTATGCACTAACATTGGAGTAAAACGACCCGGCAGTTCAAGATCTATAAC TAGAACCTCCCTCCGATCGCAGC
<i>IL5</i>	NM_000879.2	105-205	CCACAGAAATCCCACAAAGTGCATTGGTAAAGAGACCTGGACTGCTTCTACTCATGAACTCTG GATAGCCATGAGACTCTGAGGATTCTGT
<i>IL6</i>	NM_000600.1	220-320	TGACAAACAAATTGGTACATCTCGACGGCATCTAGCCCTGAGAAGAGAGACATGTAACAAG TAGTGTGGAGCAAGTCAGCAAACCTTCAAGGGT
<i>IL6R</i>	NM_000565.2	993-1093	CTTCTACATAGTGTCCATGTGGTGCCTGGAGCAGTAGTGTGGAGCAAGTCAGCAAAC TCAACCTTCAGGGTGTGGAATCTGAGCTGCG

IL7R	NM_002185.2	1610-1710	TTGCTTGGACCACTTCCGTAGTTAGTCAGTGGACTAACATGAGTCAGAGCATCCTGCTTACCATGTGGATTGGTCACAAGGTTAACGGTACCCA
IL9	NM_000590.1	300-400	AAGTACTAAAGAACACAAGTGTCATATTTCCTGTGAACGCCATGCAACCAAACCACGGCAGGCCACCG
IRF1	NM_002198.1	510-610	CTGTCGGAGTGTACCGGTCTCCACCTCTCACCAAGAACAGAGAAAAGAAAGAACAGTCGAAGTCAGCCGAGAGAA
IRF2	NM_002199.2	1375-1475	CAGTACTGGAGCTCTTAACTCAGGACTCCAGGCCATTGGTAGACGTGTTCTAGGCCGCTGGATCTCCAGGG
IRF4	NM_002460.1	325-425	GGGCAGTAAAGAACAGTCCGAGAAGGCATCGACAAGCCGCCCTCCACCTGGAAGACGCCCTGC
ITCH	NM_031483.4	155-255	GGTGCCTTGAAACAAGAGCAATGACTTGCACCTGAAATCACAGCTTCAG
ITGA1	NM_181501.1	1875-1975	AAGTGGCAAGACTAAAGAACAGAGATGACAACGTATTCCATCAGGTGGGATGGTAAGACACTGAAATTTCAGGCAATGGGTA
ITGA4	NM_000885.4	975-1075	GCCCACTGCCAACCTGGCTGCCAACGCCAGCTGCGAACAGCTC
ITGA5	NM_002205.2	925-1025	AGAAGACTTGTGCTGGTGTGCCAACAGGAACTCACTACGGCTATGTCACCACCTTAATGGCTCAGACA
ITGAL	NM_002209.2	3905-4005	TTGATGCCCTCTACAACTTCTCAGGGGTGCTTACCCAGCTCCTTGGTTCTCCCTGGAAAGAGAAATGTCGATCTA
ITGAM	NM_000632.3	515-615	AATGTGGAGAACTGAGTCAGGAGTTTGACTTCAGGA
ITGAX	NM_000887.3	700-800	CCCCCTCAGCCTGTGGCTCTGTCACCAGCTGCAAGGGTTACATACAGGCCACGCCATCAAATGTCGTCACCCACA
ITGB1	NM_033666.2	2000-2100	GCACCGATTGTCATGCCCTCATATG
ITGB7	NM_000889.1	1278-1378	TTTTAACATTACCAAGGTAGAAAAGCTGGGACAAATTACCCAGCCGTCACCTGATCTGTCAGGATCTTCACCTGGTAAACACTCTTCACTCCC
ITK	NM_005546.3	3430-3530	CAACCTGGTACAGCTCATGGATGCTTATAATAGCCTGCTTCCACCGTGACCTGAAACACTGTTGATGGCTCTGGTAGCATCCAC
JAK1	NM_002227.1	285-385	GGAGAACACCAAGCTCTGGTATGTCACCAATCGCACCATCACCCTGATGACAAGATGCCCTCGGCTCCACTA
JAK2	NM_004972.2	455-555	CCGGATGAGGTTCTATTCACCAATT
JAK3	NM_000215.2	1715-1815	CTCCCTCCGGACGCCAATTGTCGAAAGACTCTGATGGCAACAGCAGCTGTTGATGAGCTGTCGAGCTGATGAAACATCCCTCTCCGGCTCCACTA
JUN	NM_002228.3	140-240	AGGAAAGCATCCACCTCTTATATAC
JUNB	NM_002229.2	1155-1255	GGGCGCTGGAGGACAAAGGTGAGACGCTCAAGGCCGAGAACGCCGGCTGCGAGTACCGCCGGCTCCT
KIR2DL1	NM_014218.2	881-981	CCGGAGCAGTCAGGCTGTGGCTCC
KIR2DL2	NM_014219.2	814-914	GGGAGGACATCCCTCTTACACAGAGAAAA
KIR2DL3	NM_015868.2	741-841	TCTCCCTCATCGCTGGTCTCAACAAAAAAATGTCGCGTAATGGACCAAGAGTCAGGAAACAGAC
KIR2DL4	NM_002255.5	15-115	CGAATAGCGAGGACTCTGATGAAACA
KIR2DL5A	NM_020535.3	1451-1551	CTCCGAACCGGTAAACCCAGAACCTGCGATGTTGATGGACCTCAGTGGTCATCCTCTTCCATCTCC
KIR2DS1	NM_014512.1	698-798	CTCTGGCTGGAGGACTCTGTCGAGCTGTCACCCACGGTACATCTGGCATGTTCTGATGGACCTCAGTGGTC
KIR2DS2	NM_012312.2	856-956	CTCCCTCATCGCTGGTCTCAACAAAAAAATGTCGCGTAATGGACCAAGAGTCAGGAAACAGAC
KIR2DS3	NM_012313.1	693-793	CTCCCTCATCGCTGGTCTCAACAAAAAAATGTCGCGTAATGGACCTCAGTGGTCATCCTCTTCCATCTCC
KIR2DS4	NM_012314.3	1427-1527	CTCCCTCATCGCTGGTCTCAACACAGCACTAGACACGCTGCTTCCACCTCAGACTATCTTCAGC
KIR2DS5	NM_014513.2	204-304	CTCCCTCATCGCTGGTCTCAACACAGCACTAGACACGCTGCTTCCACCTCAGTGGTCATCCTCTTCCATCTCC
KIR3DL1	NM_013289.2	1054-1154	CTCCCTCATCGCTGGTCTCAACACAGCACTAGACACGCTGCTTCCACCTCAGTGGTCATCCTCTTCCATCTCC
KIR3DL2	NM_006737.2	884-984	CTCCCTCATCGCTGGTCTCAACACAGCACTAGACACGCTGCTTCCACCTCAGTGGTCATCCTCTTCCATCTCC
KIR3DL3	NM_153443.3	508-608	CTCCCTCATCGCTGGTCTCAACACAGCACTAGACACGCTGCTTCCACCTCAGTGGTCATCCTCTTCCATCTCC
KIR3DS1	NM_001083539.1	1000-1100	CTCCCTCATCGCTGGTCTCAACACAGCACTAGACACGCTGCTTCCACCTCAGTGGTCATCCTCTTCCATCTCC
KIT	NM_000222.1	5-105	CTCCCTCATCGCTGGTCTCAACACAGCACTAGACACGCTGCTTCCACCTCAGTGGTCATCCTCTTCCATCTCC

<i>MAPK14</i>	NM_001315.1	450-550	TGGGCTCTGGCGCTATGGCTGTGCTGCTGTTGACACAAAAACGGGTTACGTGTCAGTGAAGAA GCTCTCCAGACCATTCAGTCATCAT
<i>MAPK3</i>	NM_002746.2	580-680	AACGTGCTCCACCGAGATCTAAAGCCCTCAACCTGCTCATCAACACCACCTGCGACCTAAGATTGTGATT CGGCCCTGGCCCCGATTGCCGATCCTG
<i>MAPK8</i>	NM_139049.1	945- 1045	TCTCTGTAGATGAAGCTCTCAAACACCCGTACATCAATGTCGGATGATCCTTGAAGCAGAACGCTCCACCA CAAAGATCCCTGACAAGCAGTTAGA
<i>MBD2</i>	NM_003927.3	2015- 2115	ATTTACATTCAACTCTGATCCCCTGGGCCTAGGTTGACATGGAGGTGGAGGAAGATAGCGCATATAATTGCG TATGAACTATTGCCCTGAGCTTG
<i>MCL1</i>	NM_021960.3	1260- 1360	GCTGTAACCTCTAGAGTTGACCCCTAGCAACCTAGCCAGAAAAGCAAGTGGCAAGAGGATTATGGCTAACAG AATAAATACATGGGAAGAGTGCCTG
<i>MIF</i>	NM_002415.1	319-419	TCCTACAGCAACGTGCTGTGGGCCGCTGGCGACGCGCCTGCGCATCAGCCCGACAGGGTACATCAC TATTACGACATGAACGCCAATGTGG
<i>MMP14</i>	NM_004995.2	1470- 1570	GACAAGATTGATGCTGCTCTTCTGGATGCCAATGGAAAGACCTACTTCTCGTGGAAACAAGTACTACCG TTTCAACGAAGAGCTCAGGGCAGTGG
<i>MPL</i>	NM_005373.2	895-995	CAGTGGCACTTGGACTGCAATGCTTACCTTGACCTGAAAGATGTTACCTGCAATGGCAGAACAGGACCAT CTAGCTCCAAAGGCTTCTCATCA
<i>MTOR</i>	NM_004958.2	5095- 5195	TTAGTGTGCTCTGGAGTTGATCCGTCGGCAACTGACCATCCTGCAACAGTACCCCTAGGTGAC CTATGCCATACATGAAAAACATGTGGA
<i>MXD1</i>	NM_002357.2	880-980	GAGAATAAAGCTGCAGGACAGTCACAAGGGCTGCTTGGTCTCAAGAGAGTGGGACTGCGCTGTCCTT GAAGGTTCTCCCTGTTGTTGATTA
<i>MYB</i>	NM_005375.2	3145- 3245	AACTGTTGATGGATCCTGTTGCAACTGGGAGACAGAAACTGTTGATAGCCAGTCACTGCCCTAAAGA ACATTGATGCAAGATGGCAGCACT
<i>MYC</i>	NM_002467.3	1610- 1710	TCGGACACCGAGGAGAATGTCAGAGGCGAACACAAACGTCTGGAGCGCCAGAGGAGGAACGAGCTAAA CGGAGCTTTTGGCCCTGCGTGAACAGA
<i>MYO6</i>	NM_004999.3	6655- 6755	AAGTTGGGAGATGGCACCTCTCAGAGGATTGAAAATGAGGAAGAAACAAAACAGTCATGTAGGAGC ACAGGGCCACAAAGGCATTCTATTG
<i>NANOG</i>	NM_024865.2	1100- 1200	CTACTCCATGAACATGCAACCTGAAAGACGTGTAAGATGAGTGAACAACTGATATTACTCAATTTCAGTCTGGACAC TGGCTGAATCTCCCTCTCCCTCC
<i>NBEA</i>	NM_015678.3	8645- 8745	CTGAGAGCCCTTAAGGACCAAGAAAAGTCTTACCCACGCTTGTATCTGCTCCAGCGAAGGCCACTGTAT CATATACTGAAACAGGGCATTCA
<i>NCAM1</i>	NM_000615.5	1620- 1720	GGTATTGCTATCCCAGTGCAAGCATCTATGGTTGGATGCCAGCTGCTGCCAAGCTCCAATTACAGCA ATATCAAGATCTACAACACCCCTCT
<i>NCL</i>	NM_005381.2	1492- 1592	GAACAGAGATCGATGGCGATCTTCCCTGACTATACTGGAGAGAAAGGTCAAATCAAGACTATAGAGGT GGAAAGAATAGCACTTGGAGTGGTGA
<i>NCR1</i>	NM_001145457.1	145-245	TTTCATGTTCCAAAGGAAAGCAAGTGAACCATCTGTCAGGGAAATTATGGGCTGTTGAATACAGCTGC ACTTGAAGGAAGCCTTTCGCGT
<i>NCR2</i>	NM_004828.3	798-898	CTTCAACAGGTACCGACCTCCCTGGACCTAGTTCCCTCACCTGTAGAGAGAGAAATTATCACACTGTT GCAAGGACTAAGATAAGCGATGATG
<i>NCR3</i>	NM_147130.1	50-150	GCATCTGCTCTCTCCTCAGGGAGGCAAGCATTGATGCTCGAGGTCCCTGGCAGTTGTTGCTTGGCAAG TGATGTGAGTCCCCTGTCATAGG
<i>NCRNA00185</i>	NR_001544.2	143-243	GAGGCTGCTGCCAACATCTTCACTCTGCTGCAACTATGAAAATTAGTTCTAAAAATGCAACCTGC TAAATTGAGTACTAATAGGATTGGT
<i>NEIL1</i>	NM_024608.2	1675- 1775	TTAGCAGGGCTCTCTGACTCACCCTTCTTATTGCTTGCCTGCATCTGGGGCTGAATTITG GGAGCAGGAATATCTGAAGGTGCA
<i>NEIL2</i>	NM_145043.2	2570- 2670	GCCCCGGTGGTGTAGAGAAAAGCTGCTTACTCCTAACGCAATGTAATGTTGGACTGTTGATTGTAAC AATTCAAGGAATCAAGGGCTGAGG
<i>NFAT5</i>	NM_173214.1	3290- 3390	CCCTGACAACATTAAACCCAGGACATCTCACAGCCTGGACTTCCAGCAGTTCTGCTTCACTG CCAACAGCGATCACTATTGAGCA
<i>NFATC1</i>	NM_172390.1	2510- 2610	CCAGTACCAAGCGTTACCTCTCCGCCAACGTAACGCCATCTTCAACCGTAAGCGTGAACATGAGC GCCGGGGTGTCTTCTAAAGCGC
<i>NFATC2</i>	NM_012340.3	1815- 1915	GACGGACATTGGAAGAAAGAACACGCGGGTAGACTGGTTCCAGTTACATCCCAGAGTCAGTGGCAGA ATCGTCTTCTACAGACTGCATCAAC
<i>NFATC3</i>	NM_004555.2	2190- 2290	GTCCTTGAAAGTTCCTCCATATCATAACCCAGCAGTTACAGCTGCACTGCAAGGACTTTATCTT GCAATGGCAGAGGAAAAAAAGCCAGTCAC
<i>NFKB1</i>	NM_001165412.1	2305- 2405	CTTGGGTAACCTGTTGACCTAGCTGCCAAGAAGGACATGATAAGTTCTAGTATCTACTCAAGC AAAGGCAAGCAGTACTTCTGACAC
<i>NOS2</i>	NM_000625.4	605-705	TTGCTGGGTCATTATGACTCCAAAAGTTGACAGAGGCCAGGGACAAGCCTACCCCTCCAGATGAG CTTCTACCTCAAGGCTATGCAATTGTC
<i>NOTCH1</i>	NM_017617.3	735-835	CTGGCAGGCTTACCGGCCAGAACGTGAGGAAAATATCGACGATTGTCAGGAAACA CTGCAAGAACGGGGGTGCGCTGAGCACCT
<i>NR3C1</i>	NM_001018077.1	1665- 1765	GCTTCTCTCTGGGGAGAAGACGATTCTTCTGGAGGAAACTCGAATGAGGACTGCA TTTACGGGACACTAAACCCAAAATT
<i>NR4A1</i>	NM_002135.3	155-255	CGGGCGGGTAGGGTCAAGCTGAGGCTTGTCA TGAAGCCGGATTCTCCCACTGCTCT
<i>NREP</i>	NM_001142474.1	990- 1090	AAACTCATGTTCTTGTGGTAAGTGACCGAGATGCTGCCACAGGACCTGAGACACTG TTGACTTCAACATGCTCTTGGCG
<i>NRIP1</i>	NM_003489.2	335-435	TGACTCATGGAGAAGAGCTGGCTGATGTC TGCACTGAGCAGCAGGGGATCAGG
<i>NRP1</i>	NM_003873.5	370-470	GCCTCGCTGCTTCTTCTCAAGACGGGCTGAGGATTG CGCTTAGATTCTCTTGTGCAATT

<i>NT5E</i>	NM_002526.2	1214-1314	ATTGGGTTTGAATGGATAAACTCATCGCTAGAAAGTGAGGGGTGGACGTCGTGGAGGACACTC CAACACATTCTTACACAGGCATCC
<i>OAZ1</i>	NM_004152.2	313-413	GGTGGGCAGGGAAATGTCAGAGGGATCACAACTTCACTAATTCTACTCCGATGATGGCTGAATGT AACAGAGGAACTAACGTCACAGACA
<i>OPTN</i>	NM_001008211.1	625-725	TGAAGCTAAATAATCAAGCCATGAAAGGGAGATTGAGGAGCTTCGGCCTGGACAGAGAACAGAAGGAAGA ACGCCAGTTTGAGATACAGAGCAA
<i>P2RX7</i>	NM_002562.4	340-440	AGTTGGTCACAGTGTTCGACACCGCAGACTACACCTCCCTTGAGGGAACTCTCTCGTGTGACA AACTTCTCAAAACAGAACGGCAAGA
<i>PAX5</i>	NM_016734.1	2288-2388	CTCCAAGAGGAGCACACTTGGGGAGATGTCCTGGTTCTGCCATTCTGGACCGATGAGTACAG CAGCTCTTCCAGATCAAAGAAC
<i>PDCD1</i>	NM_005018.1	175-275	CTTCTCCAGGCTCGTGTGACCGAAGGGACAAACGCCACCTCACCTGAGCTCTCAAACACATCG GAGAGCTCGTGTAAACTGGTACCGC
<i>PDCD1LG2</i>	NM_025239.3	235-335	TGTGGAGCTGGCAAGTCCTCATCAAAACAGAACATGATCTCCTCTGCTAATGTTGAGCTTGAATTG CAGCTTACCAAGATAGCAGCTTATT
<i>PDE3A</i>	NM_000921.3	3010-3110	CTGGCCAACCTTCAGGAATCTCATCTCTCACATTGAGGGCTCTGCAACTCTATGATTCA GAGCAGACTTGGTGAAGACA
<i>PDE4A</i>	NM_001111307.1	3855-3955	ATAATGGTGTATACCCCTATTCTGGCAGCCCTCCACCTGGCACAAAATAATTCTCCTC CATCGTACCTGGCTAGCCTCTCC
<i>PDE7A</i>	NM_002604.2	2210-2310	GTAGCTCAACAGGAATAGAGGGAGGAGTGTAAATTGGTAGCTGGTGTGAATAGGGCTTGAGAAC TGAACACAGTGAATATGGCCAAA
<i>PDK1</i>	NM_002610.3	1170-1270	TGGATTGCCATATCACGCTTACGCAAACTTCAAGGAGACCTGAAGCTGTATTCCCTAGAGGGTAGC GGACAGATGCAAGTATCACATTAG
<i>PDXK</i>	NM_003681.3	580-680	TCCCCGGAGGACTCTTCCCCTACAAAGAAAAAGTGGTGCCTGAGCATTACGCCAACAGTT GAGGCCGAGTTACTGAGTGGCCGAA
<i>PECAM1</i>	NM_000442.3	1365-1465	ATCTGCACTCGAGGTATTGACAAGTGGTCAAGAAAGCACACAGTCAGATAGCTGTATGAA CTCAGCCCAGGATTCTTATGATGCC
<i>PHACTR2</i>	NM_001100164.1	8350-8450	GGCAGAAATGCCACTCACCCCTCAGGCAATTATGGTATGAAAGTCCAGTAATTTGTGCCACTGG CTCGGGGGAGGGCTTCTCC
<i>PHC1</i>	NM_004426.2	2905-3005	ATACAGCTCCACCTCACCGGAATTACATGGCATCACCCGTGTTCTGTCCAGTAATCCAG GTAGAGGAGGTGTACGAGTTATTG
<i>POLR1B</i>	NM_019014.3	3320-3420	GGAGAACCTGGCTTAGAATACTTGGTAGATGTTAAAGGCTGCTGGTACAATTCTATGG TATAGTGGCATCAGTGGCTAGAA
<i>POLR2A</i>	NM_000937.2	3775-3875	TTCCAAGAAGCCAAGACTCCTCGTTACTGTCTTCTGAGTGGCAGTCGAGATGCTGAGAG AGGATATTCTGTCGCTGAGCAT
<i>POP5</i>	NM_015918.3	560-660	GCTTCAGGCCACTTGTGAAACAGAACATCTGGTAGCAACAGCATCTCCACAGTT TGCCAACAGCAGACATTACCACT
<i>POU5F1</i>	NM_002701.4	1225-1325	AAGTCTTCACTTCAAGGAAGGAATTGGAAACACAAGGGGGAGTTGGGAACTGGT GAGGAAGGTGAAGTCAATGATGCTC
<i>PPARA</i>	NM_001001928.2	5220-5320	GGGTGTGTTGCTATACGAACATAATGGACGTGAAAGTGGGGAGAAACCC CAGGAGAGCTGCCCTGTTAAAGAG
<i>PPP2R1A</i>	NM_014225.3	1440-1540	AACTTAACCTTGTGCTATGGCTGGCTGTGGATCATGTATATGCCAT AAGAAGCTAGTGGAAAGTTGGGAA
<i>PRDM1</i>	NM_182907.1	310-410	CATCCCTGCCACCAGGAACCTTGTGTTATTGCGGACTTGCAGAAAG GAGCTGACAATGATGATCACA
<i>PRF1</i>	NM_005041.3	2120-2220	ACTGTTTCAGGGAGGTGGCTGGTTACACGCTAATCCGATT CCATTCTCAAGCCCTGAGTCACAGC
<i>PRKAA2</i>	NM_006252.2	975-1075	ATAGTGGTACCCCTCAAGACCACTGAGCTGGCTTATCAT GCCAGTGAATGCGAGAATAATGAACCAA
<i>PRKCQ</i>	NM_006257.2	1325-1425	GATGACGATGAGTGGAGCTGACGATGGTAGAGAAGAG ACCGCATGTTGTACATCCGAC
<i>PROM1</i>	NM_006017.1	925-1025	AGCTCGGGTCACTCTCAATGACCTCTGTGCTGGCAT GTCTCTAAGCCAGCTGAATGCAAC
<i>PTGER2</i>	NM_000956.2	1410-1510	GTCAGAAGGAGCTACAAACCTACCCCTAGTGAGCAT AGTGGACTGGCTTGAGGAAACATCGCTGCATTG
<i>PTK2</i>	NM_005607.3	1005-1105	GGTCAAGCTGGATTATTCAGTGGACTGGCAAT CGGCCAGAAGAAGGAATCAGTACCTAACGGACAAG
<i>PTPN11</i>	NM_002834.3	4650-4750	TAGCTCACTGGTCTACGGCTTACATGTGCT ATGAGATGGAATCAGGAGAGGC
<i>PTPN4</i>	NM_002830.2	705-805	TCGAGGCTTCTCCAGCCAGAGGAGC ACGTTCCGATTGCCGTGGCAGAA
<i>PTPN6</i>	NM_002831.5	1734-1834	TGGTGCAGCGAGGGCGCAGTACAAGT CTACGTGGCCATGCCAGTT GCTGGAGGTCTCGAGTCGAGAAGGG
<i>PTPRK</i>	NM_001135648.1	4315-4415	GTGATCAACCGGATT TGTAGGATGGCTTCTCATCGAG GTACCTAGGATGGCTTCTCATCGAG
<i>RAB31</i>	NM_006868.3	3800-3900	TTTGTAAAGAGCT CTGGCTGGACCCAGTTCTGCAC ATACAAGACACCAG GAGGAGGTGAGCAGAC TTGCTCTCATGAGT
<i>RAC1</i>	NM_198829.1	1250-1350	AAAGACCTCGT TGTGAGAAGACGG TAGCTTCAG GAGCAGACT TGCTCTCATGAGT
<i>RAC2</i>	NM_002872.3	1069-1169	GCTGCCACAACT TGTGACCT CAGGGAT GGGGCTT ACTCCCTCT GAGGCCAG CTGCT TAATATCGATGG TCCTGCT GCCAGAG AGTCCTAC

<i>RAF1</i>	NM_002880.2	1990-2090	CCTATGGCATCGTATTGTATGAACTGATGACGGGGAGCTCCTTATTCTCACATCAACAACCGAGATCAGATC
<i>RAP1GAP2</i>	NM_015085.4	4140-4240	CCACGGCTGAAAGAGGCCCTGACGTTCTGGACCGTTGGCTGGCTGGAGGACTGGCAAGG
<i>RARA</i>	NM_000964.2	115-215	AGCCACCTAGCTGGGGCCCCTAGAGGTGGCATTTTGTCAGGGCTGAAGGCCAGCTGGACCTTCCAAGAAAAGTGCAGCTCACAGAACAGTT
<i>RBPMS</i>	NM_001008710.1	842-942	AAACAGCTGTAGGTTGTCAGTTGACAGTCGCTCAGAAGCAGAGGCTGCAAAGAACATGTTGAATGGCATCGCTTCGATCTGAAATTCCGCAA
<i>RHOA</i>	NM_001664.2	1230-1330	GGTACTCTGGTAGCTGACCACTCAGGGCTTACTCGAACAGATTTGTTGCATAGCTGGGGCTGGAGG
<i>RNF125</i>	NM_017831.3	790-890	GCAAGGTGTATGTCCTTTGTCAGAGGAACGTATGAAGAACAGCTGCTGGATCATGATTACTCATCACAGATCGGAACGGAGGCCGTGTTCT
<i>RORA</i>	NM_134261.2	1715-1815	AAAATAACCAGAACACTTATGGCCCTGCACAGACCTGGAGGCCACACACTGCACATCTGGTGATCGGGTACGGTACGGAAAGGAAACAT
<i>RORC</i>	NM_001001523.1	1350-1450	CTCATCATGCCATGGCCAGGGCTCCAAGAGAAAAGGAAAGTAGAACAGCTGCAGTACAATCTGGAGCTGGCTTCATCTGCACAGAAC
<i>RPL27</i>	NM_000988.3	23-123	GGGCCGGTGGTTGTCGCCAAATGGCAAGTTCATGAAACCTGGAGGTGGCTTGCTGGAGCCTACTCCGACGAAAGCTGTCATGTC
<i>RPS13</i>	NM_001017.2	331-431	GCATCTTGAGAGGAACAGAACAGATAAGGATGCTAAATTCCGCTGATTCTAATAGAGAGCCGATTACCGTTGGCTCGATATTAAAGACCAAGCGA
<i>RUNX1</i>	NM_001754.4	635-735	CAGCCATGAAAGAACAGCTGTCAGATTAAATGACCTCAGGTTGTCGGAAGTGGAAAGGGAAAGCTTCACTCTGACCTCACTGTTCCACAA
<i>RUNX2</i>	NM_004348.3	1850-1950	GAAGCCACAGCAGTGGCTTGAATTCTAGTGGCAGAACGGATGAATCTGTTGGCGACCATATTGAAATTCTCAGCTGGCCAGTGGTA
<i>RUNX3</i>	NM_004350.1	2085-2185	GTGGTCTCATATACTTGGAGAGAACAGAGGGCCAGATAGATAGGTCTAGCAGAACGGATTGAGGTGAGGATCATTTGGGTACGACATCAA
<i>S100A4</i>	NM_002961.2	263-363	CAGGGACAACGAGGTGGACTTCAAGAGTACTGTGCTCCCTGATCGCCATGATGTGAAACGAAATTCTTCACTGGCTTCCAGATAACG
<i>S100A6</i>	NM_014624.3	539-639	TTCTGGGGGCTTGGCTTGTACTAACATGAAGCCCTCAAGGGCTGAAATAAATAGGGAGATGGAGACACCTCTGGGCTCTCTGACTCAAAT
<i>SATB1</i>	NM_001131010.1	1335-1435	TTCCGAAATCTACCACTGGTACGGCATGAACTGAAACGAGCAGGAATCTCCAGGCCGTATTGACGTGTTG
<i>SCML1</i>	NM_001037540.1	925-1025	GCAACGTATGGTCTCTTCAGGGCTCTGCCCTGGCAACCCCTGGCTGACAGCATCCAAACACTTAAC
<i>SCML2</i>	NM_006089.2	360-460	ATTGGAAACCCCTGACCCCTGAACTGCCACTTCAGTATGTTGCTACGGTTATTGAAATTACTGGGGCAGGTACGGTTACGCTGTTGAC
<i>SEL1L</i>	NM_005065.4	980-1080	GGGCAATCTAATGCCACATGGTTGGTTACAGATACTGGCTGGCATCGCGTCCAGAGTTGTAAT
<i>SELL</i>	NM_000655.3	110-210	CTCCCTTGGGCAAGGACCTGAGACCCCTGTGCTAAGTCAGAACGGCTCAATGGCTGCAGAACAGAGAGAA
<i>SELPLG</i>	NM_003006.3	2297-2397	CATGGCTGTAGTTGACTTCAGTTGGCTCTGGACAACAGGGGCTTGACATCCTGGGACAGGAA
<i>SERpine2</i>	NM_006216.2	240-340	CGCTGCCCTCATCTGCTCCACCTGCTCTGAGGAACAGGCTCAACACGGGATCCAGGTTTCAT
<i>SH2B3</i>	NM_005475.2	4285-4385	CCTCCAGGAAAGTAAACATCTGGATATGACGTTCTCATGCCAGGGCACTCATTCTAGCAGCCTCTACATACATCTCAGGTGGTGC
<i>SH2D2A</i>	NM_001161443.1	341-441	TGCTGGAGCCAAGCCTCAGGGGTGACTTGGTGCAGCGAGAGCGCCGTGACCTCGTGACTTACAGGAGCCGACTTGCC
<i>SIT1</i>	NM_014450.2	720-820	GCCCCAGCCCCCTGAGCAGGGCATGACTGTTCCAACACGACCCAAAGACGGGCCATTGCCAAC
<i>SKAP1</i>	NM_003726.3	1360-1460	ACAGGATGTGATCTACCCGGACTTCATGAACTACACACATCTGCTT
<i>SKAP2</i>	NM_003930.3	3374-3474	TTTACAGTTAATCAGGAGAGGGAGTCTTGCCTTCAACTGATGACCAACAGCTCAAGCCAGATAGTCG
<i>SLA2</i>	NM_032214.2	1640-1740	AAAGGAAAGTCAAGATGATGTTCTACCGTAGCAGCAGATCTGGATGGCCAGGCTATGTGACCTCCAGAGCAAGAGAACACTGGACGCTAG
<i>SLAMF1</i>	NM_003037.2	580-680	GTGCTCTTGATCCATCGGAAGCAGGCCCTCCACGTTACTGGAGATCGCTACAAGTTATCTGGAGAATCTACCCGGGATACGGGAAGCAGGA
<i>SLAMF7</i>	NM_021181.3	215-315	GGGCACTATCATAGTACCCAAATCGTAATAGGGAGAGTAGACTTCCAGATGGAGGCTACTCCCTGAAGCTCAGAACTGAA
<i>SLC2A1</i>	NM_006516.2	2500-2600	AGGCTCCATTAGGATTGCCCCCTCCCATCTTCTCACCAACCAACTCAAAATTCTTCTTACCTGAGACCA
<i>SMAD3</i>	NM_005902.3	4220-4320	TTAAGGACAGTTGAAAGGCAAGAGGAACCCAGGGCAGTCTAGAGGAGTGTGGTACTGAGCTT
<i>SMAD4</i>	NM_005359.3	1370-1470	AGGTGCACTAGGCAAAGGTGTCAGTTGAATGAAAGGTGAGTGTGGCAGGTAGCTG
<i>SNAI1</i>	NM_005985.2	63-163	GACCACTATGCCGCTTCTCGTCAGGAAGCCCTCGACCCAACTCGAACGCTAACTACAGCAGCTGCAAGCTTACATACCCAGTATCTTGC
<i>SOCS1</i>	NM_003745.1	1025-1125	TAACTGTATCTGGAGCCAGGACCTGAACCTCCACCTTACGTTACATACCCAGTATCTTGCACAAACCAGGGTGGGGAGGTC

<i>SOCS3</i>	NM_003955.3	1870-1970	GGAGGTGAGGAGACGGGACATCTTCACCTCAGGCTCTGGTAGAGAACAGGGATTACTCTGCC TCCTGACTATGCTGGCTAACAGAGATTC
<i>SOD1</i>	NM_000454.4	35-135	GCCTATAAAGTACTCGCGAGACGGGGTGCCTGGTGCCTGAGTCCTGCAGCGCTGGGTTCCGTTG CAGTCCTCGAACCGACCTCGCGCT
<i>SOX13</i>	NM_005686.2	3039-3139	ATTTATTGAGTCCCCACTACGTGCCAGGCACTGTTGCTGAGTTCTGTGGGTGTCTCGATGCCACTCTG CTTCTCTGGGGCTTCTGTGCT
<i>SOX2</i>	NM_003106.2	151-251	CTTAAGCCTTCAAAAATAATAAACATCATCGCGGCCAGGATCGGCAGAGGAGGGAAAGCGC TTTTTTGATCTGATTCAGTTGCC
<i>SOX4</i>	NM_003107.2	3040-3140	GTTCACGGTCAAACGAAATGGATTGACGTTGGGAGCTGGCGGGCGGCTGCTGGCCTCCGCCTCT TTTCTACGTGAATCAGTGAGGTGAGAC
<i>SOX5</i>	NM_152989.2	1885-1985	TAGCCATGAACTGATGATTCAATCTGAGTGGAGATTCTGATGGAAGTGCTGGAGTCTCAGAGTCAGAAATT ATAGGGAATCCGAGGGCGTGTAGC
<i>SPI1</i>	NM_003120.1	730-830	CTCCGCAGCGGCCAGATGAAGGACAGCATCTGGTGGTGGACAAGGACAAGGGCACCCTCCAGTTCTGCC AAGCACAAAGGAGGGCGTGGCGCACCGC
<i>SPN</i>	NM_001030288.1	2798-2898	AAGCCAGGCTCATGGAAAGATCGTATGTTGACCCAATATGAGTTCTCAGCTAGCCATGGTAATCCCTC CTTGAAGTCTCATCTTGAGTACA
<i>SPRY2</i>	NM_005842.2	85-185	AAAGAGGAAATACTCCGCGTGGCTGTAGAAGGGAGTCGTCAGCTCCAGCTCGAACCCGGAGTGTTCATCG CGGGGAATCTGGCTCCGAATTCTCTT
<i>STAT1</i>	NM_007315.2	205-305	TTTGTCTATGCCATCTCGAGAGCTGTCTAGGTAACGTTGCACTCTGTATATAACCTCGACAGTCAGTCTGGC ACCTAACGTGCTGTGCTAGTGTCT
<i>STAT3</i>	NM_139276.2	4535-4635	AGACTTGGCTTACATTGGTTAAATCATAGGGACCTAGGGCAGGGTCAGGCTCTGTGAGCAGATA TTGTCAGTTCATGGCTTAGGTAGCA
<i>STAT4</i>	NM_003151.2	789-889	AGACAATGGATCAGAGTACAAGAATAGTGCATGGTAATCAGGAAGTTTGACACTGCAGGAATGCTAAC AGCCTCGATTCAAGAGAAAGGAGC
<i>STAT5A</i>	NM_003152.2	3460-3560	GAGACAGAGAGAGAAAGAGAGTGTGTTGGTATGTAAATGCATCTGTCTCATGTGTTGATGTAACCGA TTCATCTCTCAGAAGGGAGGCTGGG
<i>STAT5B</i>	NM_012448.3	200-300	AAGGAGAAGCCCTTCATCAGATGCAAGCGTTATATGGCAGCATTTCCATTGAGGTGCGGCAATTATTC CAGTGGATTGAAAGCCAAGCTGGGA
<i>STAT6</i>	NM_003153.3	2030-2130	AGAACATCCAGCATTCTCTGCCAAAGACCTGTCCTCGCTCACTGGGGACCGAATCCGGATCTGCTCA GCTAAAAATCTCATCCAAAGAACCC
<i>STMN1</i>	NM_203401.1	287-387	CGTGGTGGCGGCAGGACTTCTTATCCAGTTGATTGCGAGAATACACTGCCGTGCGCTGTCTTCTATT ACCATGGCTTCTCTGATATCCAGGT
<i>SYK</i>	NM_003177.3	1685-1785	CGGACTCTCAAAGCACTGCGTGTGATGAAAACACTACAAGGCCAGACCCATGGAAAGTGGCTGTCAAG TGGTACGCTCCGAATGCATCAACTAC
<i>TAL1</i>	NM_003189.2	4635-4735	ACAGCATCTGAGTCAAGCACAACATTTCGCCCTTTGGGGTGGCTGGCGTACTTGTGATTTGATGG TACGTGACCTCTGTGAGACTTC
<i>TBP</i>	NM_003194.3	25-125	CGCCGGCTGTTAACTTCGCTTCCGCTGGCCATAGTGTATTTGAGCTGACCCAGCAGCATCACTGTTCTG GCGTGTGAAGATAACCCAAGGAATTG
<i>TBX21</i>	NM_013351.1	890-990	ACACAGGAGCGCACTGGATGCCAGGAAGTTCTTGGAAACTAAAGTCACAAACAAACAGGGGGCGTC CAACAATGTGACCCAGATGTTGTCT
<i>TBXA2R</i>	NM_001060.3	385-485	CACACGGCGCTCTCTTCCACCTCCCTGCGGCCCTGCTCTACCGACTTCTGGGCTGCTGGTACCG GTACCATCTGTGTTCCCAGCACCGC
<i>TCF12</i>	NM_207037.1	1105-1205	CACATGACCGCTTGAGTTATCCCAACTCAGTTCACCAACAGACATAAACACGAGTCCTCCACCAATGTCCA GCTTTCATCGCCGGCAGTACCGACAG
<i>TCF3</i>	NM_003200.2	4325-4425	ATACGTGCAACACAGCTGGCTGTGATTGGACTTTAAACGACCCCTTTCAAGTGGATTAGAGACCTGT CTGTATATAACAGCACTGTAGCAAT
<i>TCF7</i>	NM_003202.2	2420-2520	ATTCATTTCCAGTTCATCTATGGCAGTCAGCCAGCTCTGGCAGCTGAGAGGGCAACCCAAAACCTCAT GACAGCCAGAGCTGTCTTCAGCAT
<i>TDGF1</i>	NM_003212.2	1567-1667	AAGGAAGAAAACATCTTAAAGGGAGGAACCAAGAGTCTGAAGGAATGGAAGTCCATCTGCGTGTGCGAG GAGACTGGTAGGAAAGAGGAACAA
<i>TDO2</i>	NM_005651.1	0-100	AAGGTCAATGATAGCATCTGCCCTAGAGTCAGCCAGCTCTGGCAGCTGAGAGGGCAACCCAAAACCT GCCCATTTAGAAACAACATTGGA
<i>TEK</i>	NM_000459.2	615-715	CGAGTTCGAGGAGGGCAATCAGGATACGAACCATGAAGATGCGTCAACAGCTCCTTCTACCAAGCT AACTATGACTGTGACAAGGGAGATA
<i>TERF1</i>	NM_003218.3	1037-1137	CTGAAGCAGAAACCTGTTCAACAGAGTCAGCCGTAACCTCTGAAAACATCGAGCTGAAAGAACAGGCA TGGCTTGGGAAAGAACAGAACATT
<i>TERT</i>	NM_198253.1	2570-2670	GGCTTCAAGGCTGGAGGAACATGCGTCGCCAAACTCTGGGGCTTGCAGCTGAAGTGTACAGCCTGTT TGGATTGCGAGTGAACAGCCTCCAGA
<i>TF</i>	NM_001063.2	640-740	CTGCTCCACCTTAACCAACTTCCGGTACTCGGGAGCCCTCAAGTGTCTGAAGGATGGCTGGGATGTG GCCTTGTCAAGCACTGACTATATT
<i>TFRC</i>	NM_003234.1	1220-1320	CAGTTTCCACCATCTGGTCAGGATTGCCATAATACCTGTCAGACAACTCCAGAGCTGCTGCAGAAAA GCTTTGGAAATGGAAGGAGACT
<i>TGFA</i>	NM_003236.2	780-880	TGCCACAGACCTCTACTTGGCTGTAATCACCTGTCAGCCTTGTGGCCTTCAAAACTCTGCAAGAAC TCCGCTCTGCTTGGGGTATTCAAGTGT
<i>TGFB1</i>	NM_000660.3	1260-1360	TATATGTTCTCAACACATCAGAGCTCGAGAACGGTACCTGAACCCGTGTTGCTCCGGCAGAGCTGC GTCTGCTGAGGCTCAAGTAAAAGTGG
<i>TGFB2</i>	NM_003238.2	1125-1225	AAGCCAGAGTGGCTGAACACGGATTGAGCTATCAGATTCTCAAGTCCAAAGATTAAACATCTCAACCCAG CGCTACATCGACAGCAAAGTTGAA
<i>TGFB1</i>	NM_004612.2	4280-4380	GGGAAATACGACTTGTGAGGCATAGACATCCCTGGCCATCCTTCTGTCTCCAGCTGTTCTGGAACCTG CTCTCTGCTGTGCTGGCCCTGACG

<i>Thymidine Kinase</i>	SCFV007.1	100-200	TCTACGTACCCGAGCGCATGACTTACTGGCAGGTCTGGGGCTCCAGACAATCGCAACATCTACACCAC ACAACACGCCCTCGACCAAGGGTGAGAT
<i>TIE1</i>	NM_005424.2	2610-2710	CATCGGGGAGGGAACTTCGGGCAGGTCTCCGGGCATGATCAAGAAGGACGGCTGAAGATGAACGCAGC CATCAAAATGCTGAAAGAGTATGCTCT
<i>TLR2</i>	NM_003264.3	180-280	CTGTTCAACTGGTAGTTGGTTGAAGCACTGGACATACTGGGATGGCATCGACACACTTGTGGATGGTGGGCTTGG GGTCATCATCACGCCCTCCAAGGAAG
<i>TLR8</i>	NM_138636.3	2795-2895	GACAAAAACGTTCTCTTGTCTAGAGGAGAGGGATTGGGATCCGGATTGGCATCATCGACACCTCATGCA GAGCATCAACCAAAGCAAGAAAACAG
<i>TNF</i>	NM_000594.2	1010-1110	AGCAACAAGACCACTTCAAGAACCTGGGATTCAAGGAATGTGTCCTGCACAGTGAAGTGCTGGCAACAC TAAGAATTCAACACTGGGCCCTCAGAA
<i>TNFRSF14</i>	NM_003820.2	916-1016	CTAGGGGAGCCTCTCATCGTCACTGTTGCTCACAGTTGGCTAATCATATGTGAAAGAAGAAAGCCAA GGGGTGATGTAAGTCAAGGTGATCGTC
<i>TNFRSF18</i>	NM_004195.2	445-545	AGGGGAAATTCACTTTGGCTTCCAGTGTACTGACTGTGCCTGGGGACCTTCTCCGGGGGCCAGAAGGCCA CTGCAAACCTGGACAGACTCACCCA
<i>TNFRSF1B</i>	NM_001066.2	835-935	CCAGCTGAAGGGGACACTTGCCTCGCTCTCCAGTTGGACTGATTGGGTGACAGCCTGGCTA CTAATAATAGGAGTGTGAACTGTGTC
<i>TNFRSF4</i>	NM_003327.2	200-300	CCGTGCGGGCCGGGCTTCAACAGACGCTGGCAGCTCAAGCCGTGCAAGCCCTGACGTGGTAAACCTC AGAAGTGGGAGTGAAGCGGAAGCAGCTG
<i>TNFRSF9</i>	NM_001561.4	255-355	AGATTGCACTCCCTGCTCTCCAAATAGTTCTCCAGCGCAGGGGACAAGGACCTGTGACATATGCAGGAG TGTAAGGTTTTCAAGGACAGGAAG
<i>TNFSF10</i>	NM_003810.2	115-215	GGGGGGACCCAGCCTGGGACAGACCTGCGTGTGATCGTGTACCTCACAGTGCCTGCACTCTGTG GCTGTAACCTACGTGACTTTAACCAAC
<i>TNFSF11</i>	NM_003701.2	490-590	TACCTGATTCTAGTAGGAGAATTAAACAGGCCTTCAAGGAGCTGTGAAAGGAATTACAACATATCGTGGAT CACAGCACATCAGCAGAGAAAGC
<i>TNFSF14</i>	NM_003807.2	270-370	ATTTTCAAGACCTCTGGAAAGTCGTGACAGCCCAGGAGTGTGAGCAATTGGTTCTCTGAGGTTGAAG GACCCAGGCGTGTCAAGCCCTGTC
<i>TOX</i>	NM_014729.2	3950-4050	AATGAGCAGCTTGACTTGTACAGCGGTTGTGCAAGGAAAGCAGTCAGTGCCTGTTACAGCTTCTAGA GCAGCTGCGACAGGGTAGAGAGT
<i>TP53</i>	NM_000546.2	1330-1430	GGGGGAGCAGGCTCACTCCAGGACCTGAAGTCAAAAGGGTAGTCTACCTCCGCCATAAAAACATG TTCAAGACAGAAGGGCTGACTCACAC
<i>TRAF1</i>	NM_005658.3	3735-3835	CGAGTGATGGCTAGGCCCTGAAACTGTGCTTAGCAATAACCTCTGATCCCTACTCACCGAGTGGC CCAAGGGGGGATTGTAACAAGCC
<i>TRAF2</i>	NM_021138.3	1325-1425	GTGGCCCTTCAACCAGAAGGTGACCTTAATGCTGTCGACCAGAATAACCGGGACACGTGATTGACGCC AGGCCCAGCTGACTTCATCCTCTT
<i>TRAF3</i>	NM_145725.1	1795-1895	ATATGATGCCCTGCTCCCTGGCGTTAACAGAAAGTACACTCATGCTGATGGATCAGGGCTCTCGAC GTCAATTGGGAGATGATTCAAGCC
<i>TSC22D3</i>	NM_198057.2	1400-1500	TTAACGCAGGCAACCTCTCTCTCTCTGTTCTGTAAGGCAGGGACACAGATGGGAGAGATTGAGCCA AGTCAGCCTCTGTTGTTAATGTT
<i>TSPL</i>	NM_033035.3	395-495	CCGTCTCTGTAATCGGCCACATTGCCACTGAACTTCAACCTCAATCCACGCCGGCTGC GGTCGCTCGCAAAGAAATGTTG
<i>TXK</i>	NM_003328.1	800-900	ATGACTCTCTCGGATATCCAGTGGGCTGATGGCAGTTGTTACAGCAGCTGGTTAGCTACGAAAA GTGGGAGATAGATCCATCTGAGTT
<i>TYK2</i>	NM_003331.3	485-585	TCTACGCTGACAGCTGAGGAAGTCTGCATCCACATTGACATAAGTTGGTACTCCTCTGCTCAATCTC TTGCCCTCTTCGATGCTCAGGCC
<i>TYROBP</i>	NM_003332.2	457-557	CTGACCTCTTCAACTCTACCGCGATACAGACCCACAGAGTGCATCCCTGAGAGACCAAGCGCTCCCC AAATACTCTCTAAATAAACATGAAAGC
<i>UBASH3A</i>	NM_001001895.1	1970-2070	GAGATGCTGTTCCAGAGGGCTTCTAGTCTACCCAATGTGATTGTTAGAACGACGAGACGACTTTATA TCCCCGAATATTCCTCCCGCTTC
<i>VAX2</i>	NM_012476.2	871-971	CAGCGCCAGCGCTGCAAGAACACTAACACTTAAGACTCCCACCTGTGACACTGAGTCCCAGCACAGCACC TTCCAGCTCCCTGCCCCAGGGCAG
<i>VEGFA</i>	NM_001025366.1	1325-1425	GAGTCCAACATCACCATGCAAGATTGCGGATCAAACCTACCAAGGCCAGCACATAGGAGAGATGAGCTTCT ACAGCACAACAAATGTGAATGCAAGC
<i>WEE1</i>	NM_003390.2	5-105	TGCGTTTGAGTTGCCCGAGCCGGGCAATGGTTGCCAACGCGATGCCACGTGCTGGCGAACAAATGTA AACACGGAGATCGTGTGGCGACTT
<i>XBP1</i>	NM_005080.2	440-540	GGAGTTAACGACGCCCTGGGATGGATGCCCTGGTTGCTGAAGAGGAGGCCAGCAAGGGAATGAAGT GAGGCCAGTGGCCGGTCTGCTGAGTCC
<i>XBP1</i>	NM_001079539.1	935-1035	ATTCAATTGCTCTAGTGAAGGAAACCTGTAGAAGATGACCTCGTCCGGAGCTGGTATCTCAAATCTGCTT CATCCAGCGCACTGCCAAAGCCATCT
<i>YY1AP1</i>	NM_139118.2	755-855	ATGGGAGCTATGCGATGTTGAAGACTTCAGCACACATGTCAGCATTGACTGCAGCCCTCATAAAATGTC AAAGACTGCCATGAATTCCCTGTT
<i>ZAP70</i>	NM_001079.3	1175-1275	GGAGCTCAAGGACAAGAAGCTTCTCTGAAGCCGATAACCTCTCATAGCTGACATTGAACTTGGCTGCC AACTTGTGATGAGCTGCGG
<i>ZBTB16</i>	NM_006006.4	1585-1685	TCCCTGGATAGTTGCCGTGAGAACACTTACTGGCTATTAGCGGGTGCCAAAGCCTTGTGATG TGGCGTGCACAGTTGAGAGGA
<i>ZC2HC1A</i>	NM_016010.2	665-765	ACGATTACCGCAGCCAAGTGGCCTGGCAAACACTGTTAGGTGTTCTCAGGTAATGCAATGCGAGTTTG GGTGGGGTTAAGGCCAT
<i>ZEB2</i>	NM_014795.2	20-120	TCCCAGAGAAACACTGGCGATCACGTTTACATGATGCTCACGCTCAGGCCCTCAATTATCCCTCCCC AAAGATAGTGGCGCGTGTTCAGGG
<i>ZNF516</i>	NM_014643.2	4830-4930	GGTGGGGACGGCTCATATACCTCTCTCAGTAATGCAATGCGAGTTTG GGTGGGGTTAAGGCCAT

Table S2. Fold change in mRNA abundance of mbIL15-CAR T cells relative to CAR T cells

GENE	logFC	PValue	FDR	SIP1	-0.5	3.7E-01	1.0E+00
IL7R	-2.4	1.9E-05	6.7E-03	ITGA1	-0.5	3.7E-01	1.0E+00
NRP1	-2.7	1.6E-04	2.9E-02	KIR3DL2	-0.8	3.7E-01	1.0E+00
DAP12	1.9	3.1E-04	3.7E-02	TBXA2R	0.6	3.8E-01	1.0E+00
CD4	-1.9	5.5E-04	4.9E-02	CCR7	-0.5	3.9E-01	1.0E+00
TNFRSF4	-1.7	2.5E-03	1.5E-01	RNF125	-0.5	3.9E-01	1.0E+00
TIE1	-1.9	2.6E-03	1.5E-01	ITGA4	-0.4	4.1E-01	1.0E+00
HRH2	1.6	4.6E-03	2.2E-01	IL17RA	0.4	4.1E-01	1.0E+00
CPT1	-1.6	5.0E-03	2.2E-01	NOTCH1	0.4	4.1E-01	1.0E+00
PDCD1	-1.6	8.6E-03	3.4E-01	CD28	-0.4	4.2E-01	1.0E+00
ICOSLG	-1.3	1.3E-02	4.2E-01	BIM	-0.5	4.2E-01	1.0E+00
CMRF35H	-1.7	1.4E-02	4.2E-01	STMN1	-0.4	4.3E-01	1.0E+00
CCR4	-1.4	1.4E-02	4.2E-01	LYN	0.4	4.3E-01	1.0E+00
RORC	1.4	2.5E-02	6.4E-01	KIR2DS1	0.4	4.3E-01	1.0E+00
GILZ	-1.1	2.6E-02	6.4E-01	NKp30	0.4	4.4E-01	1.0E+00
CD86	-1.3	2.7E-02	6.4E-01	TNFRSF18	-0.5	4.4E-01	1.0E+00
TSLPR	-1.5	3.1E-02	6.8E-01	RUNX1	0.4	4.4E-01	1.0E+00
BCL6	-1.1	3.8E-02	7.9E-01	SH2D2A	0.4	4.5E-01	1.0E+00
PAX5	1.1	4.2E-02	8.3E-01	PDK1	-0.4	4.5E-01	1.0E+00
IL23R	1.1	4.8E-02	8.9E-01	SELL	-0.4	4.6E-01	1.0E+00
KIR2DS2	1.1	5.2E-02	9.2E-01	POU5F1	-0.5	4.7E-01	1.0E+00
CD40LG	-1.1	6.3E-02	1.0E+00	EGLN1	-0.4	4.7E-01	1.0E+00
IKZF2	-1.0	7.6E-02	1.0E+00	TRAF1	-0.4	4.7E-01	1.0E+00
BCL2	0.8	9.2E-02	1.0E+00	SOCS1	0.4	4.7E-01	1.0E+00
SOX5	1.3	1.1E-01	1.0E+00	KIR2DL1	-0.7	4.8E-01	1.0E+00
SOX13	-0.9	1.4E-01	1.0E+00	IL23p19	0.4	4.8E-01	1.0E+00
BNIP3	-0.8	1.5E-01	1.0E+00	LNK	0.3	4.9E-01	1.0E+00
GZMH	0.7	1.5E-01	1.0E+00	SKAP1	-0.4	4.9E-01	1.0E+00
CD11B	-0.8	1.6E-01	1.0E+00	BIRC2	-0.3	4.9E-01	1.0E+00
LEF1	0.9	1.6E-01	1.0E+00	STAT1	-0.2	5.0E-01	1.0E+00
SMAD3	-0.7	1.6E-01	1.0E+00	LAIR1	-0.3	5.0E-01	1.0E+00
FLT1	0.7	1.6E-01	1.0E+00	TNFRSF9	-0.5	5.0E-01	1.0E+00
KLRB1	-0.7	1.7E-01	1.0E+00	MYB	-0.4	5.0E-01	1.0E+00
CCL5	-0.6	1.7E-01	1.0E+00	CDK4	0.4	5.0E-01	1.0E+00
C50RF13	-0.8	1.8E-01	1.0E+00	XBP1	0.4	5.0E-01	1.0E+00
KLF2	-0.6	2.0E-01	1.0E+00	TFRC	-0.4	5.0E-01	1.0E+00
CD276	-0.9	2.0E-01	1.0E+00	SLA2	0.4	5.1E-01	1.0E+00
LRP5	0.7	2.0E-01	1.0E+00	CD69	0.3	5.1E-01	1.0E+00
P2RX7	-0.7	2.1E-01	1.0E+00	FAS	-0.4	5.1E-01	1.0E+00
NCAM1	0.6	2.1E-01	1.0E+00	CCL3	0.2	5.1E-01	1.0E+00
ATF3	0.8	2.2E-01	1.0E+00	EGLN3	-0.3	5.1E-01	1.0E+00
ANXA1	-0.7	2.2E-01	1.0E+00	TNF	0.2	5.2E-01	1.0E+00
EOMES	-0.8	2.3E-01	1.0E+00	IGF1R	-0.3	5.2E-01	1.0E+00
NFATC1	-0.6	2.3E-01	1.0E+00	BACH2	0.3	5.2E-01	1.0E+00
MMP14	-0.8	2.5E-01	1.0E+00	CD94	-0.4	5.2E-01	1.0E+00
PHACTR2	-0.7	2.6E-01	1.0E+00	BCL2L1	0.3	5.2E-01	1.0E+00
GATA3	-0.7	2.6E-01	1.0E+00	TXK	0.3	5.2E-01	1.0E+00
CD44	-0.6	2.6E-01	1.0E+00	IL10RA	-0.3	5.3E-01	1.0E+00
AIM2	-0.6	2.7E-01	1.0E+00	RARA	-0.4	5.3E-01	1.0E+00
AGER	-0.6	2.7E-01	1.0E+00	XBP1	0.3	5.3E-01	1.0E+00
TGFBR1	0.6	2.8E-01	1.0E+00	IL12RB2	0.4	5.3E-01	1.0E+00
CSF2	-0.7	2.9E-01	1.0E+00	DPP4	0.3	5.3E-01	1.0E+00
CD80	-0.6	3.0E-01	1.0E+00	RUNX3	0.4	5.4E-01	1.0E+00
IL10	0.4	3.2E-01	1.0E+00	HVEM	-0.3	5.4E-01	1.0E+00
FGL2	-0.6	3.3E-01	1.0E+00	FLT3LG	-0.3	5.4E-01	1.0E+00
MYC	-0.5	3.4E-01	1.0E+00	ITGAL	-0.3	5.5E-01	1.0E+00
PHC1	-0.6	3.5E-01	1.0E+00	EPHA4	-0.3	5.5E-01	1.0E+00
LOC282997	-0.5	3.5E-01	1.0E+00	IL2RA	-0.4	5.5E-01	1.0E+00
ITGA5	-0.6	3.6E-01	1.0E+00	PTGER2	-0.3	5.6E-01	1.0E+00

ID2	0.3	5.6E-01	1.0E+00	RAC2	0.2	7.4E-01	1.0E+00
DNAM1	-0.2	5.7E-01	1.0E+00	NFATC2	0.2	7.5E-01	1.0E+00
CD7	0.3	5.7E-01	1.0E+00	STAT6	-0.2	7.5E-01	1.0E+00
SLAMF1	-0.4	5.7E-01	1.0E+00	GNLY	0.2	7.5E-01	1.0E+00
BCLXL	0.2	5.9E-01	1.0E+00	CCL4	0.0	7.5E-01	1.0E+00
SATB1	0.3	5.9E-01	1.0E+00	CD11C	-0.2	7.5E-01	1.0E+00
TP53	0.2	5.9E-01	1.0E+00	CSAD	-0.2	7.5E-01	1.0E+00
MAD1L1	0.3	6.0E-01	1.0E+00	ABCB1	-0.2	7.6E-01	1.0E+00
FOS	0.3	6.0E-01	1.0E+00	CD8A	0.2	7.6E-01	1.0E+00
NKG2D	-0.3	6.0E-01	1.0E+00	BMI1	-0.1	7.6E-01	1.0E+00
CDK2	0.2	6.0E-01	1.0E+00	RAC1	0.2	7.6E-01	1.0E+00
GZMB	0.2	6.0E-01	1.0E+00	LAT	-0.1	7.6E-01	1.0E+00
IFNGR1	0.4	6.0E-01	1.0E+00	PNK	-0.2	7.6E-01	1.0E+00
CTNNA1	0.2	6.1E-01	1.0E+00	AP1	0.2	7.6E-01	1.0E+00
ICOS	-0.2	6.2E-01	1.0E+00	STAT5B	-0.2	7.6E-01	1.0E+00
SNAI1	-0.3	6.2E-01	1.0E+00	CD19RCD28CAR	-0.3	7.7E-01	1.0E+00
KIR2DL3	0.2	6.2E-01	1.0E+00	CITED2	0.2	7.7E-01	1.0E+00
LAG3	0.2	6.2E-01	1.0E+00	DGKA	-0.2	7.7E-01	1.0E+00
HES1	-0.3	6.2E-01	1.0E+00	CD274	-0.2	7.7E-01	1.0E+00
CD85	-0.3	6.3E-01	1.0E+00	YAP	-0.1	7.7E-01	1.0E+00
PRF1	0.2	6.4E-01	1.0E+00	IL2RG	-0.2	7.7E-01	1.0E+00
CDH1	-0.3	6.5E-01	1.0E+00	POP5	0.2	7.7E-01	1.0E+00
VEGFA	-0.2	6.5E-01	1.0E+00	TRF	0.2	7.7E-01	1.0E+00
mTOR	0.3	6.6E-01	1.0E+00	SLC2A1	-0.2	7.8E-01	1.0E+00
AKT1	0.2	6.6E-01	1.0E+00	NFATC3	-0.2	7.8E-01	1.0E+00
CD38	-0.2	6.6E-01	1.0E+00	CFLIP	-0.1	7.9E-01	1.0E+00
TRAF3	0.3	6.6E-01	1.0E+00	SERPINE2	-0.3	7.9E-01	1.0E+00
GLIPR1	-0.2	6.6E-01	1.0E+00	PDCD1LG2	-0.1	8.0E-01	1.0E+00
Dock5	0.2	6.6E-01	1.0E+00	CTLA4	-0.2	8.0E-01	1.0E+00
BCL2L11	-0.3	6.6E-01	1.0E+00	SLAMF7	0.1	8.0E-01	1.0E+00
LAT2	-0.3	6.6E-01	1.0E+00	HOPX	0.1	8.1E-01	1.0E+00
ANXA2P2	0.2	6.7E-01	1.0E+00	TERT	0.2	8.1E-01	1.0E+00
p38	0.3	6.7E-01	1.0E+00	NFAT5	-0.1	8.1E-01	1.0E+00
ETV6	-0.2	6.7E-01	1.0E+00	SCAP2	0.1	8.1E-01	1.0E+00
ATM	0.3	6.7E-01	1.0E+00	TNFRSF1B	-0.1	8.1E-01	1.0E+00
KIR3DL3	0.3	6.8E-01	1.0E+00	FANCC	0.1	8.1E-01	1.0E+00
CBLB	-0.3	6.8E-01	1.0E+00	JUNB	0.1	8.1E-01	1.0E+00
IL15RA	-0.2	6.9E-01	1.0E+00	NT5E	-0.1	8.1E-01	1.0E+00
CATHEPSINC	0.2	6.9E-01	1.0E+00	LGALS1	-0.2	8.1E-01	1.0E+00
MAPK8	-0.2	6.9E-01	1.0E+00	ITK	0.1	8.1E-01	1.0E+00
FYN	-0.2	6.9E-01	1.0E+00	BAX	0.2	8.1E-01	1.0E+00
ITGB1	-0.3	6.9E-01	1.0E+00	CCR6	-0.1	8.2E-01	1.0E+00
RUNX2	-0.2	7.0E-01	1.0E+00	APAF1	-0.2	8.2E-01	1.0E+00
CCR1	0.1	7.0E-01	1.0E+00	BTLA	0.2	8.2E-01	1.0E+00
PTPN4	-0.2	7.0E-01	1.0E+00	GADD45A	0.1	8.2E-01	1.0E+00
FASLG	0.2	7.0E-01	1.0E+00	B2M	-0.1	8.2E-01	1.0E+00
CD19Rscfv	-0.3	7.0E-01	1.0E+00	LDHA	-0.2	8.2E-01	1.0E+00
PDE4	-0.2	7.1E-01	1.0E+00	CD43	-0.1	8.2E-01	1.0E+00
STAT5A	-0.2	7.1E-01	1.0E+00	KLRC1	0.0	8.3E-01	1.0E+00
SHP2	0.2	7.1E-01	1.0E+00	STS2	-0.2	8.3E-01	1.0E+00
E2A	-0.2	7.2E-01	1.0E+00	HOXB3	-0.2	8.3E-01	1.0E+00
CREM	0.2	7.2E-01	1.0E+00	IL15	0.2	8.3E-01	1.0E+00
LTA	-0.3	7.3E-01	1.0E+00	RAP46	-0.1	8.3E-01	1.0E+00
FOXO3	-0.2	7.3E-01	1.0E+00	CDKN2C	0.1	8.3E-01	1.0E+00
CASP1	0.2	7.3E-01	1.0E+00	MCL1	-0.1	8.4E-01	1.0E+00
SHP1	0.2	7.3E-01	1.0E+00	CAT	-0.1	8.4E-01	1.0E+00
BATF	0.2	7.4E-01	1.0E+00	HPRT1	0.1	8.4E-01	1.0E+00
MAD	-0.1	7.4E-01	1.0E+00	IKZF1	0.1	8.4E-01	1.0E+00
SOCS3	-0.2	7.4E-01	1.0E+00	JAK2	-0.1	8.4E-01	1.0E+00
NKG2F	-0.3	7.4E-01	1.0E+00	NEIL2	-0.2	8.4E-01	1.0E+00
IRF4	-0.2	7.4E-01	1.0E+00	NFKB	-0.1	8.5E-01	1.0E+00

CXCR4	0.1	8.5E-01	1.0E+00	PRDM1	0.0	9.4E-01	1.0E+00
KLF10	0.2	8.5E-01	1.0E+00	ITCH	0.0	9.4E-01	1.0E+00
DUSP16	-0.1	8.5E-01	1.0E+00	CLA	0.0	9.4E-01	1.0E+00
MAP2K1	-0.1	8.6E-01	1.0E+00	LGALS3	0.1	9.4E-01	1.0E+00
IL21R	0.1	8.6E-01	1.0E+00	CATHEPSIND	0.0	9.4E-01	1.0E+00
RAF1	0.1	8.6E-01	1.0E+00	IL18RAP	0.0	9.4E-01	1.0E+00
FOXO1	0.1	8.6E-01	1.0E+00	IL4R	0.0	9.5E-01	1.0E+00
MIF	-0.1	8.6E-01	1.0E+00	TNFSF10	0.0	9.5E-01	1.0E+00
FADD	0.1	8.6E-01	1.0E+00	CD58	0.0	9.5E-01	1.0E+00
GADD45B	0.2	8.6E-01	1.0E+00	BID	0.1	9.5E-01	1.0E+00
FOXP1	0.1	8.7E-01	1.0E+00	JAK1	0.0	9.5E-01	1.0E+00
HLAA	-0.1	8.7E-01	1.0E+00	CD247	0.0	9.5E-01	1.0E+00
NR3C1	-0.1	8.7E-01	1.0E+00	SOD1	0.0	9.6E-01	1.0E+00
PRKCQ	0.1	8.8E-01	1.0E+00	CIITA	0.1	9.6E-01	1.0E+00
TCF12	0.1	8.8E-01	1.0E+00	CRIP1	0.1	9.6E-01	1.0E+00
NR4A1	-0.1	8.8E-01	1.0E+00	ARRB2	0.1	9.6E-01	1.0E+00
BCL10	-0.1	8.8E-01	1.0E+00	NEIL1	-0.1	9.6E-01	1.0E+00
ITGB7	0.0	8.8E-01	1.0E+00	SMAD4	0.0	9.6E-01	1.0E+00
MAPK14	0.1	8.8E-01	1.0E+00	SELL	0.0	9.6E-01	1.0E+00
HDAC2	0.1	8.8E-01	1.0E+00	C11ORF17	0.0	9.6E-01	1.0E+00
CD244	0.0	8.9E-01	1.0E+00	TNFSF14	0.0	9.6E-01	1.0E+00
IL18R1	0.1	8.9E-01	1.0E+00	RHOA	0.0	9.6E-01	1.0E+00
S100A4	0.1	8.9E-01	1.0E+00	CCR5	-0.1	9.6E-01	1.0E+00
HOXB4	-0.1	8.9E-01	1.0E+00	NKG2C	-0.1	9.7E-01	1.0E+00
CSNK2A1	0.1	8.9E-01	1.0E+00	S100A6	0.0	9.7E-01	1.0E+00
BCL11B	0.0	9.0E-01	1.0E+00	SIT1	0.0	9.7E-01	1.0E+00
CTNNBL1	0.1	9.0E-01	1.0E+00	KIR2DL4	-0.1	9.7E-01	1.0E+00
NCL	0.1	9.0E-01	1.0E+00	CD3E	0.0	9.7E-01	1.0E+00
ELF1	0.1	9.0E-01	1.0E+00	GZMA	0.0	9.7E-01	1.0E+00
MBD2	0.1	9.0E-01	1.0E+00	TBX21	0.0	9.7E-01	1.0E+00
DAP10	-0.1	9.0E-01	1.0E+00	GLO1	0.1	9.7E-01	1.0E+00
ELF4	0.1	9.0E-01	1.0E+00	IFNG	-0.1	9.7E-01	1.0E+00
GSK3B	-0.1	9.1E-01	1.0E+00	IL12RB1	0.0	9.7E-01	1.0E+00
IRF1	0.0	9.1E-01	1.0E+00	CCNB1	-0.1	9.7E-01	1.0E+00
CDKN1A	-0.1	9.1E-01	1.0E+00	IRF2	0.0	9.7E-01	1.0E+00
CREB1	0.0	9.1E-01	1.0E+00	PDE7	0.0	9.7E-01	1.0E+00
TDO2	0.1	9.1E-01	1.0E+00	TNFRSF7	0.0	9.8E-01	1.0E+00
STAT3	-0.1	9.1E-01	1.0E+00	CLIC1	0.0	9.8E-01	1.0E+00
AHNAK	0.1	9.1E-01	1.0E+00	JAK3	0.0	9.8E-01	1.0E+00
CD63	0.0	9.1E-01	1.0E+00	OPTN	0.0	9.8E-01	1.0E+00
LCK	0.0	9.1E-01	1.0E+00	MAPK3	0.0	9.8E-01	1.0E+00
KLF6	0.1	9.2E-01	1.0E+00	RORA	0.1	9.8E-01	1.0E+00
IL2RB	0.0	9.2E-01	1.0E+00	KLF7	0.0	9.9E-01	1.0E+00
NKp46	0.0	9.2E-01	1.0E+00	CDKN1B	0.0	9.9E-01	1.0E+00
EIF1	0.1	9.2E-01	1.0E+00	PPP2R1A	0.0	9.9E-01	1.0E+00
TRAF2	0.1	9.2E-01	1.0E+00	ATP2B4	0.0	9.9E-01	1.0E+00
DOK2	0.1	9.2E-01	1.0E+00	CD2	0.0	9.9E-01	1.0E+00
ADAM19	-0.1	9.2E-01	1.0E+00	CXCR3	0.0	9.9E-01	1.0E+00
GFI1	0.0	9.2E-01	1.0E+00	C10RF24	0.0	9.9E-01	1.0E+00
NKG2E	-0.1	9.3E-01	1.0E+00	TGFB1	0.0	9.9E-01	1.0E+00
AIF1	0.0	9.3E-01	1.0E+00	STAT4	0.0	1.0E+00	1.0E+00
HMGB2	0.0	9.3E-01	1.0E+00	ZAP70	0.0	1.0E+00	1.0E+00
HDAC1	0.1	9.3E-01	1.0E+00	CD3D	0.0	1.0E+00	1.0E+00
CTNNB1	0.0	9.3E-01	1.0E+00	GABPa	0.0	1.0E+00	1.0E+00
TYK2	0.1	9.3E-01	1.0E+00	CCR10	-0.1	1.0E+00	1.0E+00
IAP	0.1	9.3E-01	1.0E+00	PECAM1	-0.1	1.0E+00	1.0E+00
C80RF70	0.2	9.3E-01	1.0E+00	SOX4	0.1	1.0E+00	1.0E+00
PPARA	0.0	9.3E-01	1.0E+00				