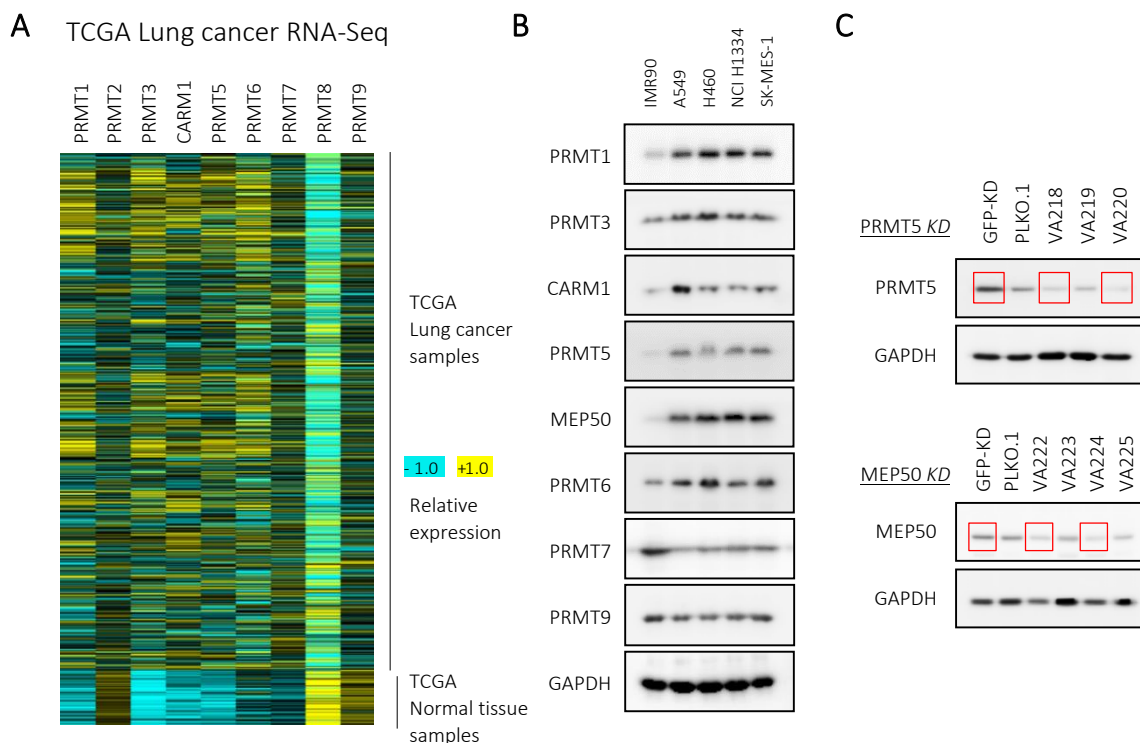


A TGF β -PRMT5-MEP50 Axis Regulates Cancer Cell Invasion through Histone H3 and H4 Arginine Methylation Coupled Transcriptional Activation and Repression

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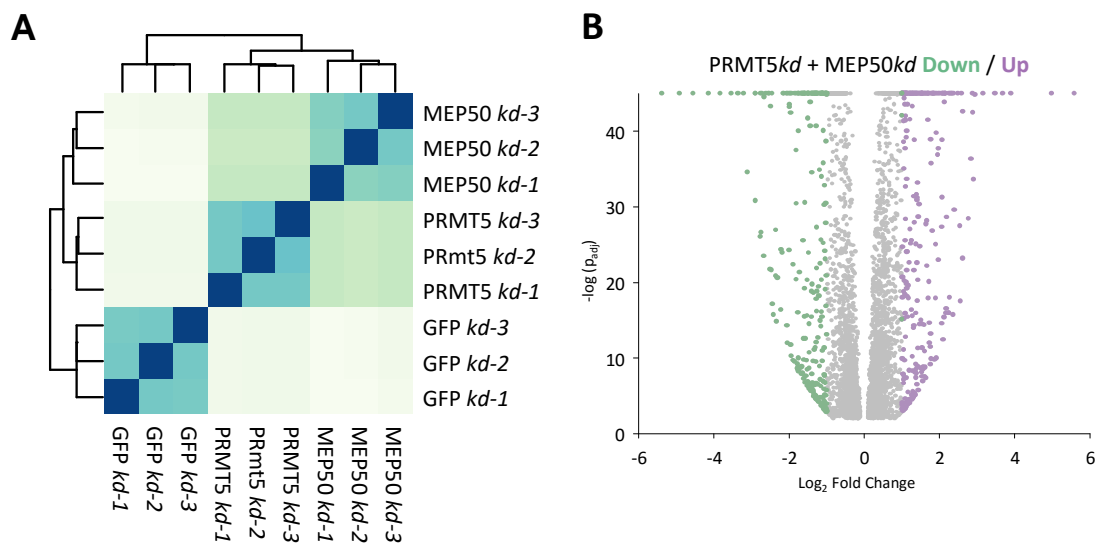


Supplementary Figure S1, Related to Figure 1. PRMTs expression profile in lung cancer cell line and PRMT5 and MEP50 shRNA knockdown.

A. Heatmap of gene expression of PRMT 1-9 from TCGA lung cancer RNA-Seq data in normal and patients with lung adenocarcinoma (LUAD) or squamous cell carcinoma (LUSC), ranked by sample type (tumor or normal tissue).

B. Whole cell lysate immunoblots for PRMTs and MEP50 in the lung fibroblast cell line (IMR90) and corresponding lung cancer cell lines. GAPDH is a loading control.

C. Whole cell lysate immunoblots for PRMT5 and MEP50 in A549 cells (Lung adenocarcinoma). GAPDH is a loading control. Different shRNAs against PRMT5 or MEP50 are indicated. shRNAs against GFP or empty vector (PLKO.1) were used as control.



C Gene Set Enrichment Analysis

Input dataset = Cufflinks FPKM for PRMT5kd+MEP50kd vs. GFPkd

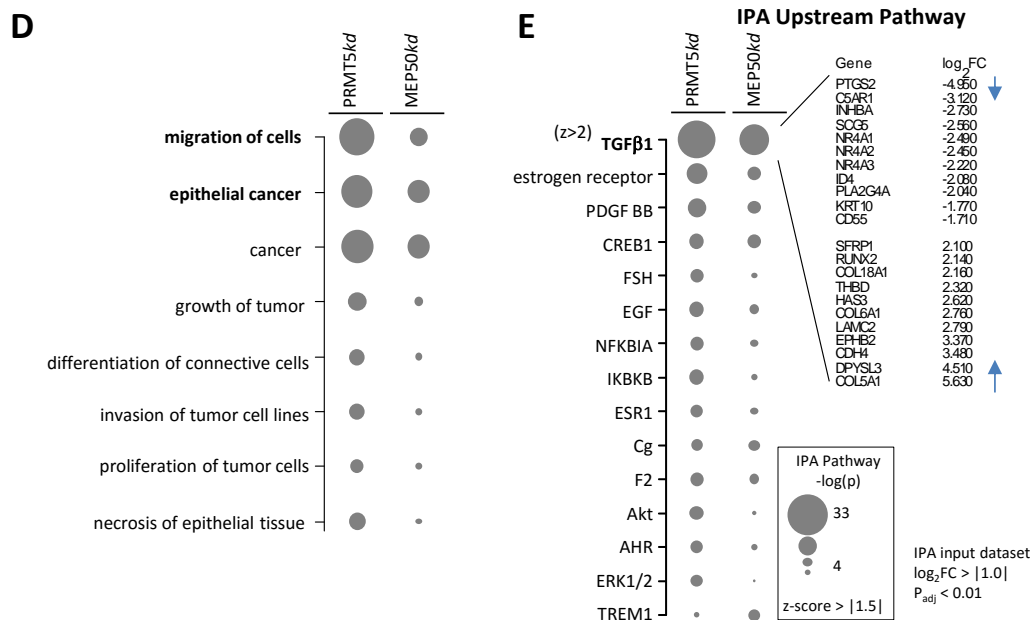
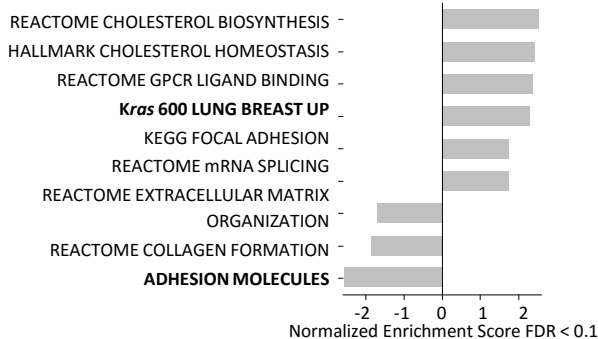


Figure S2, Related to Figure 2. Transcriptome is dramatically altered upon PRMT5-MEP50 knockdown.

- A.** The replicates of the PRMT5 and MEP50 knockdown RNA-Seq are highly correlated as illustrated by DESeq2 clustering.
- B.** Volcano plot of Spearman's correlation coefficient for the differentially regulated genes altered in both PRMT5 and MEP50 knockdowns versus the corresponding significance score ($-\log_2(p_{adj})$).
- C.** Functional annotation by gene set enrichment analysis of genes changes upon PRMT5 and MEP50 knockdown in A549 cells. Enriched groups are ranked by the significance p -value.
- D.** Dotplot of gene ontology terms enriched on PRMT5 and MEP50 knockdown upregulated genes are shown in dots scaled by $-\log(p)$.
- E.** Dotplot of Ingenuity Pathway Analysis (IPA) diseases and functions enriched in both knockdowns are shown in dots scaled by $-\log(p)$.

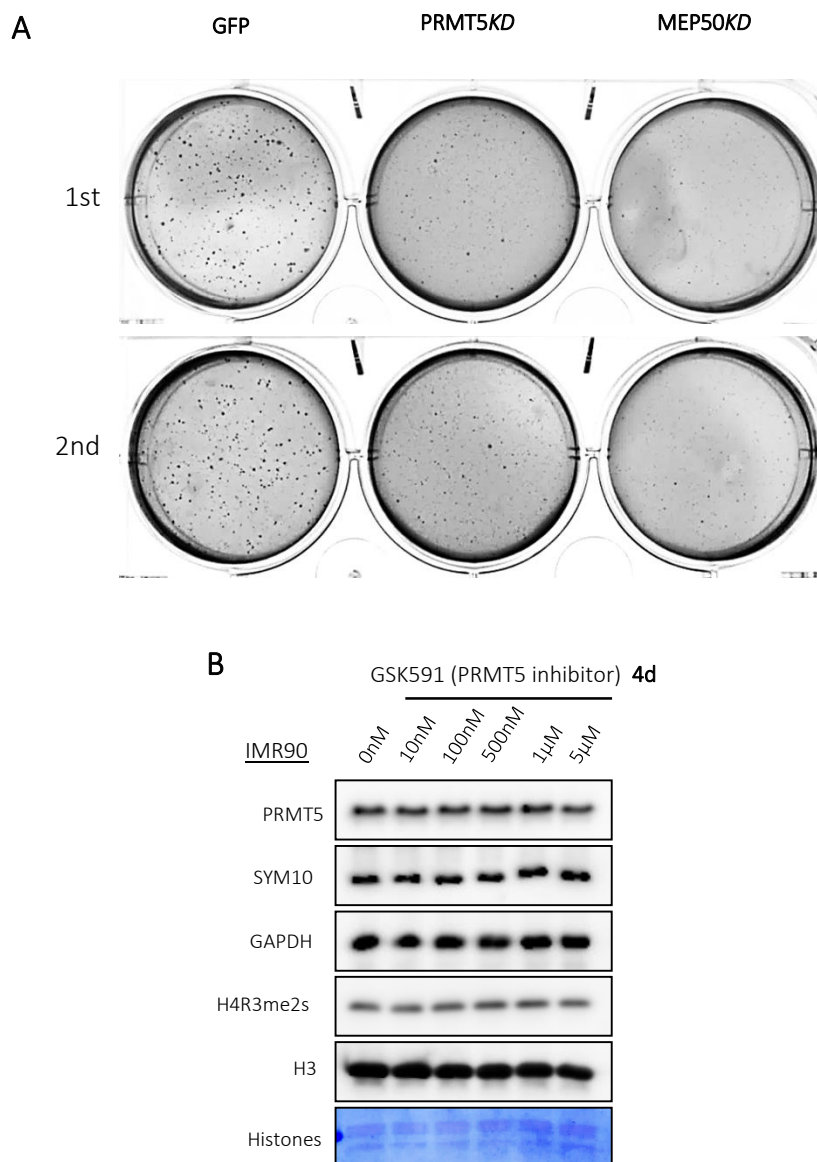


Figure S3, Related to Figure 3. PRMT5-MEP50 knockdown can prevent lung cancer cell mobility.

A. Representative crystal violet staining for colonies after 14d cell culture of A549 cells expressing shRNA targeted against GFP as a control (*GFPkd*), against PRMT5 (*PRMT5kd*) or against MEP50 (*MEP50kd*). 1st and 2nd represent 2 independently biological experiments.

B. Normal lung fibroblast cell line (IMR90) was treated with various indicated dosages of GS591 for 4 days and lysates or extracted histones were blotted for PRMT5, SYM10 (methylated SmD3), GAPDH (control), H4R3me2s, and H3 as indicated. DB71 stain of extracted histones is also shown.

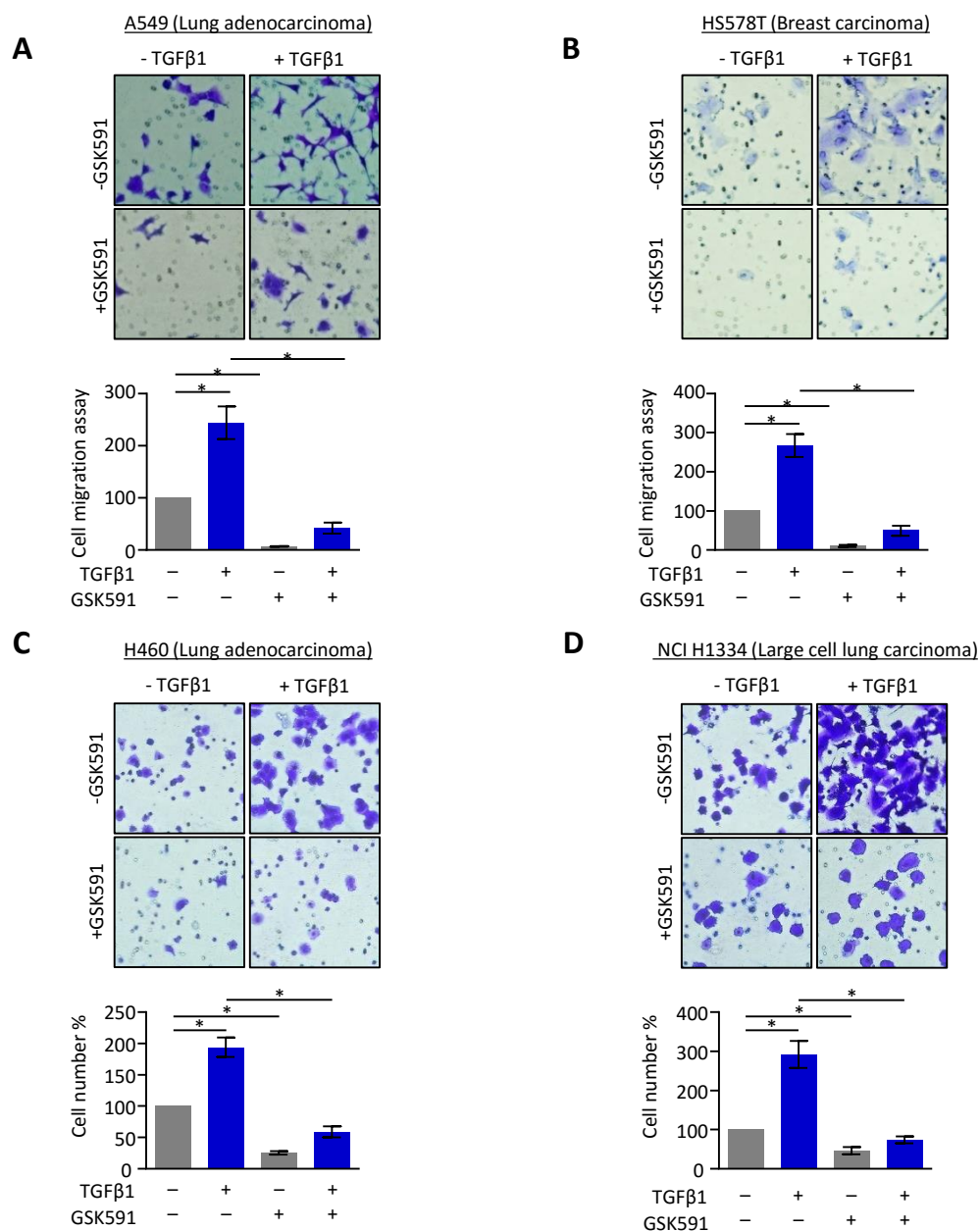


Figure S4, Related to Figure 4. PRMT5 inhibitor GSK591 blocked both lung and breast cancer cell lines TGFβ-stimulated cell mobility.

A. Cell migration assay for A549 and **(B)** HS578T cell lines. Migration of indicated cells treated with (+) or without (-) 500 nM GSK591 for 4 days and (+) or without (-) 10 nM TGFβ1 for 2 days.

C. Cell invasion assay for H460 and **(D)** NCI H1334 cell lines treated with (+) or without (-) 500 nM GSK591 for 4 days and (+) or without (-) 10 nM TGFβ1 for 2 days. Top: Representative crystal violet staining of invaded cells on the underside of the porous polycarbonate membrane through a phase-contrast microscope (20X) is shown. Bottom: Quantification of the invaded cells shown in. Values are mean \pm S.E.M. of three independent experiments. $*p < 0.05$ from one-way ANOVA test.

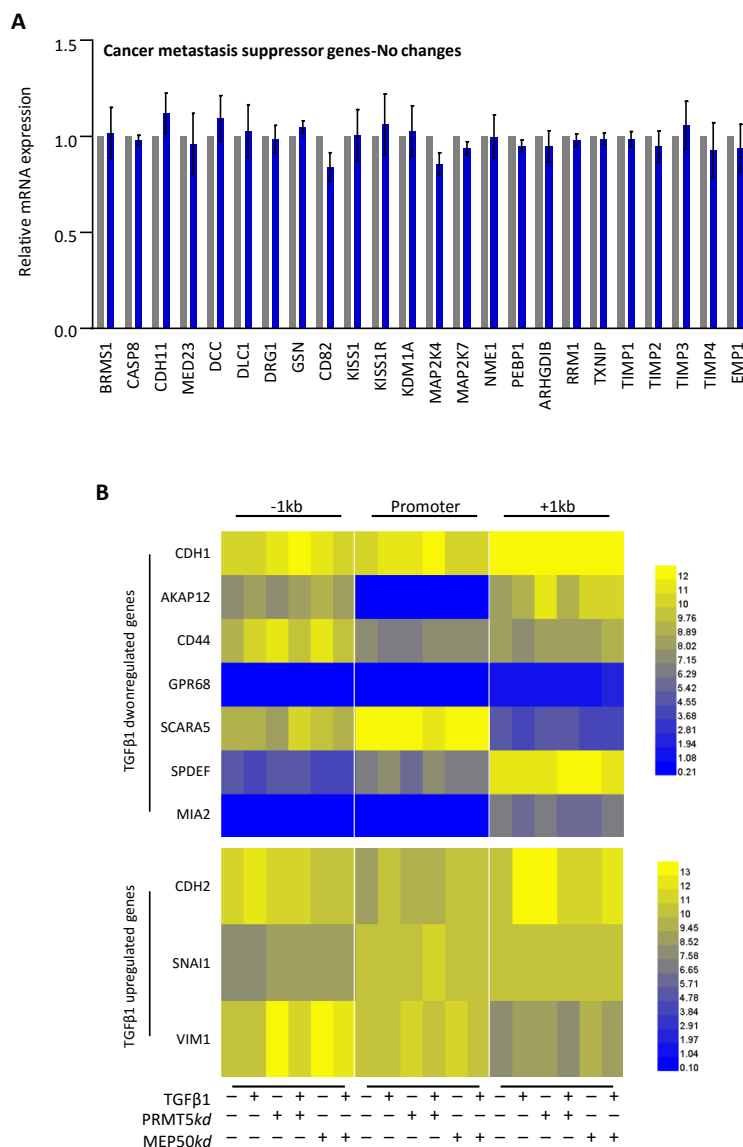


Figure S5, Related to Figure 6. Controls for qRT-PCR and ChIP experiments

A. Relative mRNA levels of indicated cancer suppressor genes in A549 cells treated with 10 nM of TGF β 1 for 2 days were determined by qRT-PCR. β -Actin was used as an internal control. Values are means \pm S.E.M. of three independent experiments. To normal cells, there has no significant gene expression changes for the ones treated with 10 nM of TGF β 1 for 2 days for the genes listed. p value are measured with one-way ANOVA test.

B. Heatmap generated by ChIP-qPCR values for histone H3 values at -1kb, the promoter, or +1kb of the indicated genes demonstrate the H3 was pulled down evenly as the ChIP control for the various histone PTMs in Figure 6. ChIP-qPCR of TGF β 1-downregulated genes are arrayed on the top and TGF β 1-upregulated genes are on the bottom. The heatmap is arrayed from blue (no enrichment) to yellow (maximal enrichment). Between different groups there is no significant H3 enrichment changes. p value are measured with one-way ANOVA test.

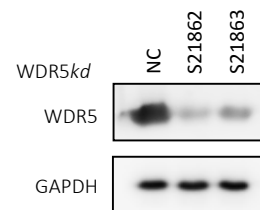


Figure S6, Related to Figure 7. Confirmation of depletion of WDR5 in A549 cell line.

Immunoblots for WDR5 and GAPDH as control, from A549 cells infected with siRNA targeted against negative control (NC) or against 2 independent WDR5 siRNA (ID s21862 and s21863) for 3 days.

Table S1. Antibodies used in this study.

Factor or PTM	Vendor	Cat Number	Dilution For blotting	Vol. for ChIP (μl)
PRMT1	Millipore	07-404	1:5,000	
PRMT3	Gene Tex	GTX116478	1:5,000	
CARM1	Cell Signaling	4438	1:5,000	
PRMT5	Millipore	07-405	1:5,000	
MEP50	LPBio	AR-0145-S	1:5,000	
PRMT6	Cell Signaling	14641	1:5,000	
PRMT7	Pierce	PA5-30748	1:10,000	
PRMT9	Homemade	N/A	1:5,000	
E-cadherin	Cell signaling	3195	1:3,000	
Vimentin	Cell signaling	5741	1:3,000	
Snail	Cell signaling	3879	1:2,000	
SYM10	Millipore	07-412	1:5,000	
SNRPD3	Abcam	ab157118	1:10,000	
WDR5	Millipore	07-706	1:5,000	
GAPDH	Abcam	ab9484	1:10,000	
H3R2me1	Abcam	ab15584	1:5,000	8
H3R2me2s	Abcam	Ab194684	1:5,000	6
H3R8me2s	Novus Biologicals	NB21-1063	1:5,000	10
H4R3me1	Novus Biologicals	NB21-2011	1:10,000	10
H4R3me2a	Active Motif	39705	1:10,000	8
H4R3me2s	Abcam	ab5823	1:10,000	3
H3K4me3	abcam	Ab8580	1:10,000	5
H3	Abcam	ab1791	1:100,000	4
HRP-conjugated anti-rabbit secondary antibody	GE	NA834V	1:30,000	
HRP-conjugated anti-mouse secondary antibody	GE	NA931V	1:30,000	

Table S2. ChIP primers used in this study.

The ChIP primers with red bold are the ones used for genes indicated in Figure 6.

Primer set name	Location in the genome	Forward sequence	Reverse sequence
AKAP12 -1kb	chr6:151238642+151238737	TCAAATGGGGGAAATGTGT	CACGAAGGCCAATTTCTCTG
AKAP12 P	chr6:151239706+151239799	TCCGGTAACAGCCTCATTTTC	ATCCTCCGGAACAAGTGATG
AKAP12 +1kb	chr6:151240812+151240886	AGAGAACTTCCCAGCCCATC	ACCCTGAAAAGCCTCCAAAG
CD44-1kb	chr11:35138161+35138345	ATGGTGGATGGTTGTGGTTT	CATCCTCCTGTCCATCCACT
CD44 P	chr11:35138742+35138891	AACCCAGAGATCTTGCTCCA	GAGACGCACTGGCTTTCTTC
CD44 +1kb	chr11:35139817+35140003	TGGTCACAGAAGGGATCACA	TCATGGAGACCCAAGACCTC
MED23-1kb	chr6:131585753+131585847	TTTTTCCCCTCTGCTGTATTCT	AATTTCCCTGGATAGGCTTTT
MED23 P	chr6:131586621+131586745	TAGGCCAATGTCAGTCACCA	TGATTGTTTCCCGACTCCAT
MED23 +1kb	chr6:131587945+131588058	GGAGTGTCTGGGGTAGATG	GGGTGTGTGTGAGGGAGAAC
GPR68-1kb	chr14:91231464+91231541	CAGATGAATTCCAGGGATGG	TCTTGGCTCGTTGCTTTTCT
GPR68 P	chr14:91232216+91232332	CTGGGCTCTGTCTCAAAG	AGAGCCTTCTCCTCCAC
GPR68 +1kb	chr14:91233661+91233767	CACAACTCTGCAGGAAGCA	GAATAAGCCGTGCCTCTGC
SCARA5-1kb	chr8:27869187+27869300	GCTCTTCTCCATGGAACCTG	GGAAATCCACAAAGGAAGCA
SCARA5 P	chr8:27869627+27869703	TCTTGTAAGATGGGCCTTGG	TGGCTCAAGCCTTATTTGGT
SCARA5 +1kb	chr8:27870841+27870986	AGAGGCTTGAAGATGCAAA	GGAGGAGAGCTGATGTGGAG
SPDEF-1kb	chr6:34536714+34536828	CAAGGGAGTGAGACACAGCA	TGGCAGTGAGCAAATAGCAC
SPDEF P	chr6:34537854+34537970	AACTCAGGGGTGCAGATGTC	AGGGCAGTGACTCGACAAAG
SPDEF +1kb	chr6:34538618+34538715	GGTGTAGTTGCGGTGAGGTT	TACCGGGAGGTTTGTGACTC
MIA2-1kb	chr14:39233121+39233300	CCAACGGAAATCACACTGTA	TCTGCCATTGAAAGAGGTCA
MIA2 P	chr14:39234123+39234200	ATTTGGCGTTCACAGAATCC	AGGTCTGCCAGCAGTTTTGT
MIA2 +1kb	chr14:39234900+39235017	TGCAAAATAAATTACATCCCAAT	AAGCAATTTTTGATTCTGCTGAG
CDH1 -1kb	chr16:68735940+68736048	TGCTGGCCCTATTGTTACT	CCCTATGCTGTTGTGGGACT
CDH1 P	chr16:68737215+68737321	GTGAACCCTCAGCCAATCAG	TCACAGGTGCTTTGCAGTTC
CDH1 +1kb	chr16:68738469+68738561	AGAAATTGCACTCCACACC	GATCCCCAAATCTGCGTAAA
CDH2 -1kb	chr18:27950018+27950116	CAGCATGGAGGCACAGTCTA	GAGCTTGCCTCTGGGAATTT
CDH2 P	chr18:27951910+27952010	GGCACATAAAATCCCAGTGC	TGGGCTCAGAGGGAATATCA
CDH2 +1kb	chr18:27951682+27951787	TGGTCTCATCCCCAAGATA	TGCTTCAACACGCTTTTGT
VIM -1kb	chr10:17226963+17227105	ATGCCTTGTCTCCTTTTCC	GTGTGCCTGGAACCTTAGA
VIM P	chr10:17228395+17228494	GGCCAGCTGTAAGTTGGTA	CCTAGCGGTTTAGGGGAAAC
VIM +1kb	chr10:17229220+17229290	GAGGGACCCTCTTTCTAA	GAGAGTGGCAGAGGACTGGA
SNAI1 -1kb	chr20:49981560+49981683	CGGCACCAAGTGACTAAACA	CACAGGTCTCACCGTTCTTG
SNAI1 P	chr20:49983118+49983246	GCGAGCTGCAGGACTCTAAT	GTGACTCGATCCTGGCTCA
SNAI1 +1kb	chr20:49983626+49983698	GGGTCTACGTGTGAGAGA	TCCACAGGACAGACCAGGTT

Table S3. RT-qPCR primers used in this study.

Gene name	Forward sequence	Reverse sequence
ACTB	AGCTACGAGCTGCCTGAC	AAGGTAGTTTCGTGGATGC
PRMT5	TTGCCGGCTACTTTGAGACT	ACAGATGGTTTGGCCTTCAC
MEP50	AGCACTGCCTCTCTCACC	ACACGGCCAATTCCTCATAG
BRMS1	GAAGGCACCTCTGGTTTCTG	CTGCCCTAGCCTTTTTGATG
CASP8	TCCCCAACTTGCTTTATGC	GACCCCAGAGCATTGTTAGC
AKAP12	TGGCAGGAAGACATTCTGTG	GCGGGTGAATTTAAACAAA
CDH1	CAAGTGCCTGCTTTTGTGA	GTTTTCTGTGCACACCTGGA
CDH2	TGCTTCAACACGCTTTTGT	TGGTCTCATCCCCAAGATA
CDH11	CACGGCTCCTCCTTATGACT	CGAGGTCCCCAGTTCTGTAG
CD44	AAGGAACCTGCAGAATGTGG	TCCAACGGTTGTTTCTTTCC
MED23	AGCCATGAACAGTGGGTCTC	AATTTGGAACCCTTGCTGTG
DCC	GGGACCACTTTGGAAATGAA	AGAAGTGGCGATGATGGAAC
DLC1	ATCATTCCAAGGCCAAACTG	GAGAATCTCCGTGCTTCTG
DRG1	GCTGGATGCTGAACTGTGA	TGTTTCTTCCACCACATCA
GAS1	AGATTGTGGCCAGTGAGGAT	GGCGCAGATACAAACAGTGA
GSN	ACGGCTGAAGGACAAGAAGA	TGAGCTCACCAGGAACCTCT
CD82	GATGGTCTGTCCATCTGCT	TCAGTACTTGGGGACCTTGC
KISS1	GACCTGCCTCTTCTCACCAA	TGGGTCTAGAATTCCCCACA
KISS1R	TTGGTCTCTTGTGACGTTCCG	TGATCCAGAAAAGTCTGTGTTG
KLF17	GGGAGAGGAAGGGACATAGC	TCAGGAACCTGGAAGTCACC
KDM1A	TCTTCTTTGCGGGAGAACAT	CCCAAAACTGGTCTGCAAT
MAPK14	TGTATTTGGCCAAGGTGTT	CACCCTGTCCCTCTGGAGTA
MAP2K4	CCCCACGGTATCCTAAACT	CAGATGGGAAGTTGACAGCA
MAP2K7	GGGCTGCCTGGTTTTATTTT	AGGGCTCCCCACTTAACACT
NME1	AGAAAGGATTCCGCCTTGTT	GGCCCTGAGTGCATGTATTT
PEBP1	TGTTGGGACATGGCAATCTA	CATTCAGCAACTCCAGACCA
ARHGDI8	CTCGGCCTGAGGAGTATGAG	TGGTCTTGCTTGTATCGTC
RRM1	CATCCACATTGCTGAGCCTA	GATTAGCCGCTGGTCTTGTC
TXNIP	CTTGCCCACTGTGACTTCAA	TCCTAACACAGGGCAGGAAG
GPR68	CTGGGTCACTGACATTGGTG	TGGGAAGCCAGTGTTTAAGG
TIMP1	TGACATCCGTTTCGTCTACA	TGCAGTTTCCAGCAATGAG
TIMP2	CTCTCCATTTGGCATCGTTT	TTTGAGTTGCTTGCAGGATG
TIMP3	GGAGAGCTGCAGAGTGCCT	AGCTAGGGAAAGGGAACCAA
TIMP4	GCCAGGACTATTCCCTTTCC	ATGACATTGCCATTTCTCC
MMP2	AGCGTGAAGTTTGAAGCAT	CCTCCGGTCTTCTCTAGT
MMP9	AAGCTGGACTCGGTCTTTGA	CCTGTGTACACCCACACCTG
SCARA5	GGAGGAGAGCTGATGTGGAG	AGAGGCTTGAAGATGCAAA
SPDEF	GACCAGTGAGGAGAGCTGGA	CATAGCTGTGGGGCTTGAGT
MIA2	CAGAGCACATCCCAAACCT	GCCCTGTATCCTCATCTCCA
EMP1	GGCTCCTAGGCTCAGTGGTA	GCCTGTTGTTTGGTTTTGGT
ARHGAP29	TAAACCACATGCTCCCATCA	CTTTGTCTGGGTCTGGCATT
SNAI1	GCTCCACAAGCACCAAGAGT	ATTCCATGGCAGTGAGAAGG