

Title: *Bifidobacterium breve* UCC2003 metabolises the human milk oligosaccharides lacto-N-tetraose and lacto-N-neo-tetraose through overlapping, yet distinct pathways.

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Purpose	Primer	Sequence (5'-3')
Cloning of Bbr_1556 in pNZ8150	1556F 1556R	<i>tgc</i> atcc agct gatgatcaccatcaccatcaccatcaccatcacaacgggtcaaacatctctgca <i>tgc</i> catc ctag aggattggaagcgcagatggc
Cloning of internal 465bp fragment of Bbr_0530 in pOR119	IM530F IM530R	ctggtc aa gcttgaaggagaccggcatcaagg ctggtc ctctag agcttggaaaggcttggcgc
Cloning of internal 488bp fragment of Bbr_1554 in pOR119	IM1554F IM1554R	ctggtc aa gcttgaaggagaccggcatcaagg ctggtc ctctag agcttggaaaggcttggcgc
Cloning of internal 443bp fragment of Bbr_1556 in pOR119	IM1556F IM1556R	ctggtc aa gcttgaatacctcgccatccc ctggtc ctctag accagtgacttcatccagtcgc
Amplification of <i>tetW</i>	tetWFw tetWRv	tcagctgtcgacatgctcatgtacggtaaggaagca gcgacggtcgaccataacttctgattgtgccg
Confirmation of site specific homologous recombination	526confirm1 526confirm2 530confirm1 530confirm2 1554confirm1 1554confirm2 1556confirm1 1556confirm2	gcgtagctgttacaatggc gccatttccaaccctctc gaggaaaggagaacagacgatg gcgaaaggaaatggcttcatag ctgctgacgccaatgccgacaag ccgctactgcaggtcgccatcatcaagcac gcagctcagcagacatccgc gcgcgcgagtcggtgac
Cloning of 2086bp fragment containing Bbr_0529 into pNZ44	529pNZ44F 529pNZ44R	ctggtc ctg caggactccatttgcccacagtag cgagc ctctag agatttctctcgacactg
Cloning of p44-529 in pBC1.2	P44 Forward	ctggtcc ggatcc gagataatgccgactgtac
Cloning of 1637bp fragment containing Bbr_1554 into PBC1.2	1554PBC1.2F 1554PBC1.2R	ctggtccggatccgccggaacaacagtctgc cgagctctagaattcaccgtttggattattgg
Cloning of 2360bp fragment containing Bbr_1556 into PBC1.2	1556PBC1.2F 1556PBC1.2R	ctggtccggatcccccacgaggtggacaagt cgagctctagagaaatcacaatcgagcc

Restriction sites incorporated into oligonucleotide primer sequences are indicated in bold, and His-tag sequences incorporated into nucleotide primer sequences are indicated in italics.

Supplementary Table S1: Oligonucleotide primers used in this work.

Strain or plasmid	Relevant Features	Reference or Source
Strains		
<i>Escherichia coli</i> strains		
<i>E. coli</i> EC101	Cloning host, repA ⁺ km ⁺	1
<i>E. coli</i> EC101-pNZ-M.BbrII + M.BbrIII	EC101 harbouring pNZ8048 derivative containing <i>bbrIIM</i> and <i>bbrIIIM</i>	2
<i>E. coli</i> XL1-Blue	<i>supE44 hsdR17 recA1 gyrA96 thi relA1 lac F</i> [<i>proAB+ lacIq lacZAM15 Tn10(Tet^r)</i>]	Stratagene
<i>E. coli</i> XL1-blue-pBC1.2-529	XL1-blue containing pBC1.2-529	This study
<i>Lactococcus lactis</i> strains		
<i>L. lactis</i> NZ9000	MG1363, pepN:nisRK, nisin-inducible overexpression host	3
<i>L. lactis</i> NZ9700	Nisin-producing strain	3
<i>L. lactis</i> NZ9000-pNZ-lacZ2	NZ9000 conatining pNZ-010	4
<i>L. lactis</i> NZ9000-pNZ-IntA	NZ9000 conatining pNZ-529	4
<i>L. lactis</i> NZ9000-pNZ-lacZ6	NZ9000 conatining pNZ-1552	4
<i>L. lactis</i> NZ9000-pNZ-nahA	NZ9000 conatining pNZ-1556	This study
<i>L. lactis</i> NZ9000-pNZ44-IntA	NZ9000 containing pNZ44-529	This study
<i>Bifidobacterium</i> sp. Strains		
<i>B. breve</i> UCC2003	Isolate from nursling stool	5
<i>B. breve</i> UCC2003-lacZ2	Tn5-transposon mutant in Bbr_0010 of UCC2003	6
<i>B. breve</i> UCC2003-IntP1	pORI19-tet-bbr_0527 insertion mutant of UCC2003	4
<i>B. breve</i> UCC2003-IntA	pORI19-tet-bbr_0529 insertion mutant of UCC2003	4
<i>B. breve</i> UCC2003-lacS	pORI19-tet-bbr_1551 insertion mutant of UCC2003	4
<i>B. breve</i> UCC2003-lacZ6	pORI19-tet-bbr_1552 insertion mutant of UCC2003	4
<i>B. breve</i> UCC2003-InbP	pORI19-tet-bbr_1587 insertion mutant of UCC2003	7
<i>B. breve</i> UCC2003-IntS	pORI19-tet-bbr_0530 insertion mutant of UCC2003	This study
<i>B. breve</i> UCC2003-nahS	pORI19-tet-bbr_1554 insertion mutant of UCC2003	This study
<i>B. breve</i> UCC2003-nahA	pORI19-tet-bbr_1556 insertion mutant of UCC2003	This study
<i>B. breve</i> UCC2003-IntA-PBC1.2-IntA	pORI19-tet-529 insertion mutant of UCC2003 harbouring complementation construct pBC1.2-529	This study
<i>B. breve</i> UCC2003-nahS-PBC1.2-nahS	pORI19-tet-1554 insertion mutant of UCC2003 harbouring complementation construct pBC1.2-1554	This study
<i>B. breve</i> UCC2003-nahA-PBC1.2-nahA	pORI19-tet-1556 insertion mutant of UCC2003 harbouring complementation construct pBC1.2-1556	This study
<i>B. breve</i> UCC2004	Isolate from infant faeces	UCC
<i>B. breve</i> JCM7017	Isolate from human faeces	JCM
<i>B. breve</i> JCM7019	Isolate from infant faeces	JCM
<i>B. breve</i> NCFB2258	Isolate from infant intestine	NCFB
<i>B. breve</i> UCC2005	Isolate from nursling stool	UCC
<i>B. breve</i> 8815	Isolate from human faeces	NCIMB
<i>B. breve</i> 11815	Isolate from infant intestine	NCIMB
<i>B. breve</i> NIZO658	Isolate from human faeces	NIZO
<i>B. breve</i> UCC2006	Mother's milk	UCC
<i>B. breve</i> UCC2007	Mother's milk	UCC
<i>B. breve</i> UCC2008	Isolate from infant/adult faeces	UCC
<i>B. breve</i> LMG13208	Isolate from infant faeces	LMG
<i>B. breve</i> UCC2009	Isolate from infant/adult faeces	UCC
<i>B. breve</i> UCC2010	Mother's milk	UCC
<i>B. breve</i> UCC2011	Mother's milk	UCC
Plasmids		
pAM5	pBC1-puC19-Tet ^r	8
pORI19	Em ^r , repA ⁺ , ori ⁺ , cloning vector	1
pORI19-tet-IntS	Internal 465 bp fragment of <i>bbr_0530</i> and tetW cloned in pORI19	This study
pORI19-tet-nahS	Internal 443 bp fragment of <i>bbr_1554</i> and tetW cloned in pORI19	This study
pORI19-tet-nahA	Internal 443 bp fragment of <i>bbr_1556</i> and tetW cloned in pORI19	This study
pNZ8150	Cm ^r , nisin inducible translational fusion vector	9
pNZ-nahA	Cm ^r , pNZ8150 derivative containing translational fusion of Bbr_1556 encoding DNA fragment to nisin inducible promoter	This study
pNZ44	pNZ8048 containing constitutive p44 promoter from Lactococcal chromosome	10
pNZ44-IntA	pNZ44 harbouring Bbr_0529 downstream of p44 promoter	This study
pBC1.2	pBC1-pSC101-Cm ^r	8
pBC1.2-IntA	pBC1-pSC101-Cmr harbouring Bbr_0529 downstream of p44 promoter	This study
pBC1.2-nahS	pBC1-pSC101-Cmr harbouring Bbr_1554	This study
pBC1.2-nahA	pBC1-pSC101-Cmr harbouring Bbr_1556	This study

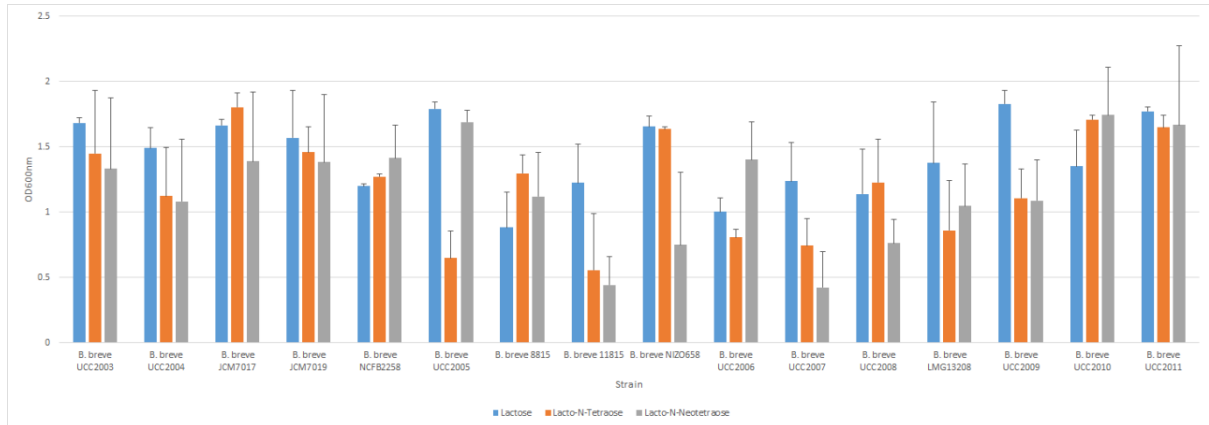
Cm^r, Em^r, Km^r and Tet^r, resistance to chloramphenicol, erythromycin, kanamycin and tetracycline, respectively.

Collection of the University of Goteborg; CIP, Collection de l'Institut Pasteur; DSM, German Collection of Microorganisms and Cell Cultures; JCM, Japan Collection of Microorganisms; LMG, Belgian Co-ordinated Collection of Microorganisms; NCDO, National Collection of Dairy Organisms; NCFB, National Collection of Food Bacteria; NCIMB, National Collection of Industrial and Marine Bacteria; NCTC, National Collection of Type Cultures; UCC, University College Cork Culture Collection.

Supplementary Table S2: Bacterial plasmids and strains used in this work.

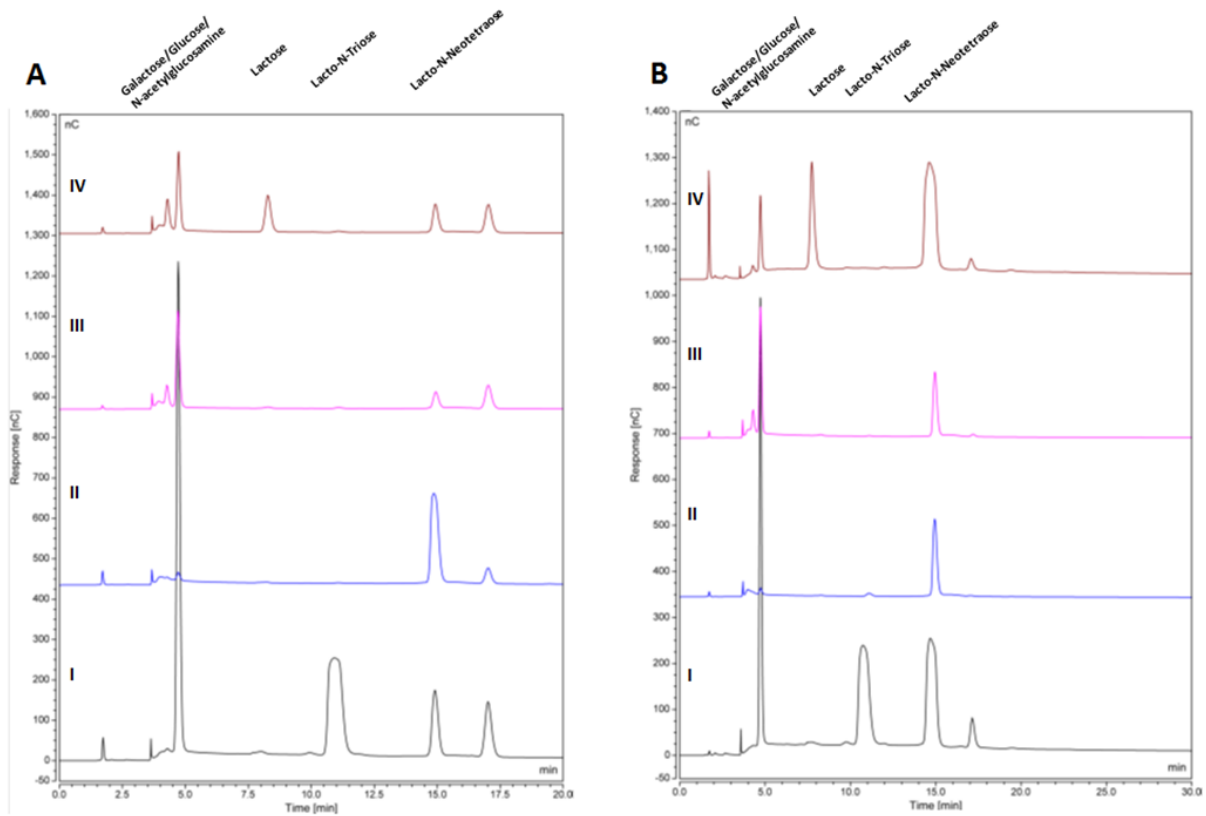
Gene ID	Gene Name	Location of Activity	Function in HMO Metabolism
<i>Bifidobacterium breve</i> UCC2003			
Bbr_1556	nahA	Intracellular	nagZ Beta-N-acetylhexosaminidase; hydrolyses Lacto-N-Triose at GlcNAc β 1-3Gal residue, releasing GlcNAc and galactose.
Bbr_1587	lnbP	Intracellular	Lacto-N-biose phosphorylase; hydrolyses LNB, releasing GlcNAc and phosphorylated galactose.
<i>Bifidobacterium bifidum</i> PRL2010			
BBPR_1438	lnbB	Extracellular	Lacto-N-biosidase; hydrolyses LNT at its GlcNAc β 1-3Gal residue, releasing LNB and lactose.
<i>Bifidobacterium longum</i> subsp. <i>longum</i> JCM1217			
BLLJ_1505	lnbX	Extracellular	Lacto-N-biosidase; hydrolyses LNT at its GlcNAc β 1-3Gal residue, releasing LNB and lactose.

Supplementary Table S3: Locus tags, names and product functions of *B. breve* UCC2003 and *B. bifidum* PRL2010 genes selected for bioinformatic analysis of LN(n)T/LNB utilisation-associated homologs across the *Bifidobacterium* genus.

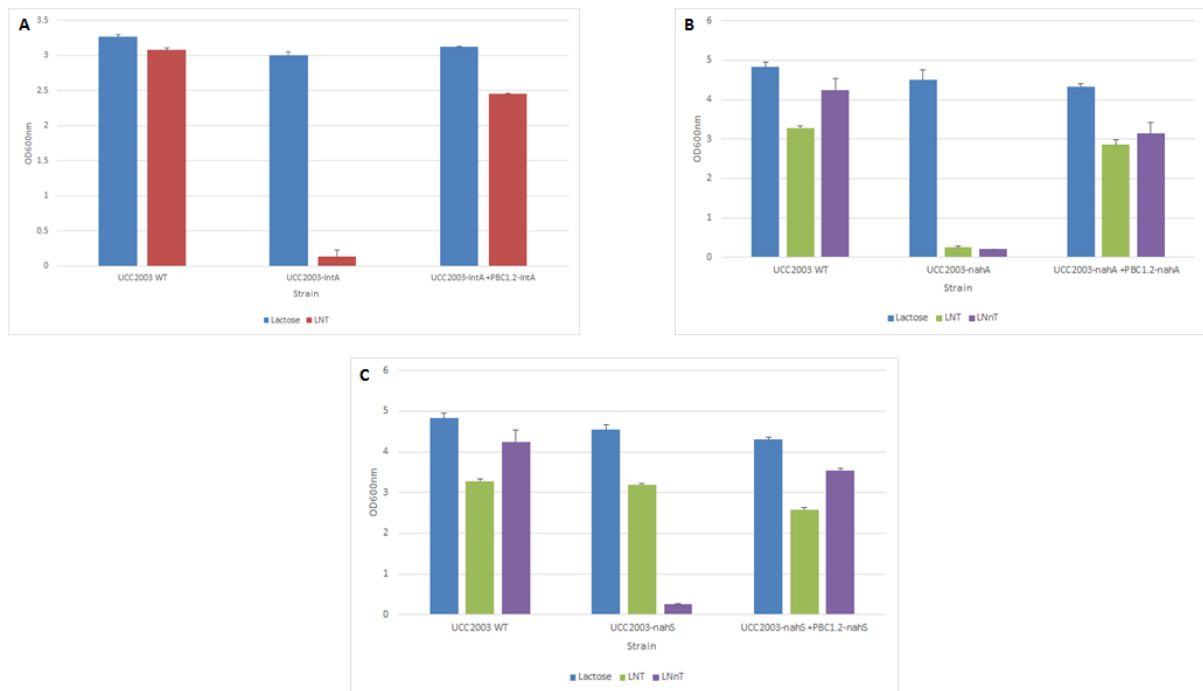


Supplementary Figure S1: Final OD_{600nm} values (measured using PowerWave microplate spectrophotometer) after 24 hours of growth of 16 *B. breve* strains in modified MRS containing 1 % (wt/vol) LNT, 1 % (wt/vol) LNnT or 1 % (wt/vol) lactose as the sole carbon source.

The results are the mean values obtained manually from two separate experiments (due to the limited availability of certain carbohydrates). Error bars represent the standard deviation.



Supplementary Figure S2: (A) HPAEC chromatogram profiles of LNnT, when incubated in MOPS buffer (pH7) with: (I) LacZ2 alone, (II) NahA alone, (III) LacZ2 and NahA together, and (IV) LacZ2, followed by a denaturation step and the subsequent addition of NahA. (B) HPAEC chromatogram profiles of LNnT, when incubated in MOPS buffer (pH7) with: (I) LacZ6 alone, (II) NahA alone, (III) LacZ6 and NahA together, and (IV) LacZ6, followed by a denaturation step and the subsequent addition of NahA.



Supplementary Figure S3: (A) Final OD600 values (measured manually) after 24 hours of growth of wild type *B. breve* UCC2003, the insertion mutant *B. breve* UCC2003-lntA, and the complementation strain *B. breve* UCC2003-lntA+PBC1.2-lntA in modified MRS containing 1 % (wt/vol) lactose or 1 % (wt/vol) LNT as the sole carbon source. The results are the mean values obtained from two separate experiments*. Error bars represent the standard deviation. (B) Final OD600 values (measured manually) after 24 hours of growth of wild type *B. breve* UCC2003, the insertion mutant *B. breve* UCC2003-nahA, and the complementation strain *B. breve* UCC2003-nahA+PBC1.2-nahA in modified MRS containing 1 % (wt/vol) lactose or 1 % (wt/vol) LNT as the sole carbon source. The results are the mean values obtained from two separate experiments*. Error bars represent the standard deviation. (C) Final OD600 values (measured manually) after 24 hours of growth of wild type *B. breve* UCC2003, the insertion mutant *B. breve* UCC2003-nahS, and the complementation strain *B. breve* UCC2003-nahS+PBC1.2-nahS in modified MRS containing 1 % (wt/vol) lactose or 1 % (wt/vol) LNT as the sole carbon source. The results are the mean values obtained from two separate experiments*. Error bars represent the standard deviation.

*Experiments were carried out in duplicate due to a limited amount of carbohydrate material.

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