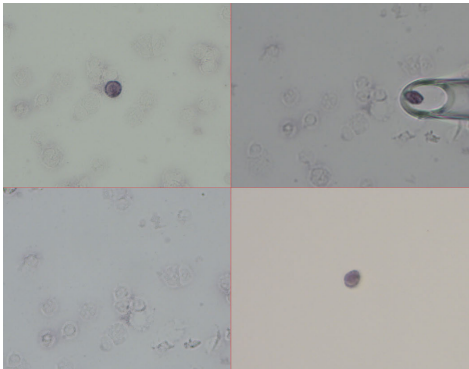


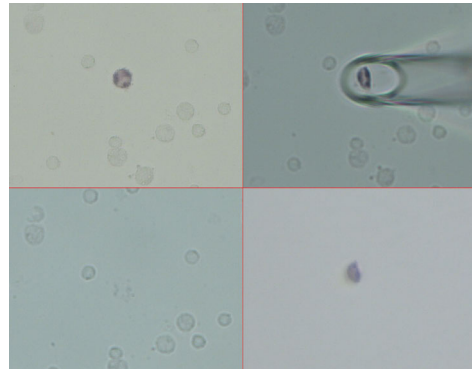
**Figure S1: Isolation of disseminated tumor cells from bone marrow aspirates.** Single cells isolated from patient MicMa003 (A), MicMa017 (B), MicMa019 (C), MicMa044 (D), MicMa083 (E), MicMa107 at diagnosis (F), and MicMa107 3 years post-diagnosis (G). Each panel documents the isolation process of the indicated single cell in four subsequent images taken before picking (top left), during micromanipulation (top right), of the area where the cell was picked from after micromanipulation (bottom left), and of the isolated cell in the lid (bottom right). The immunomorphological class of each cell is provided next to its name. Boxed names indicate control cells stained with an isotype control antibody.

**A**

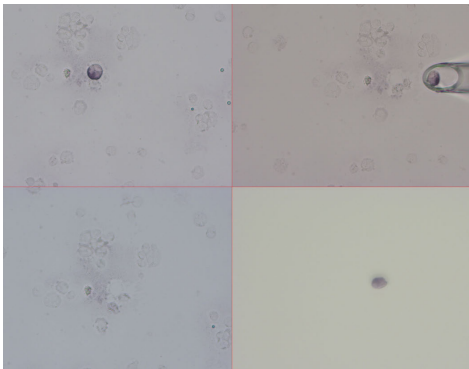
003A - TC



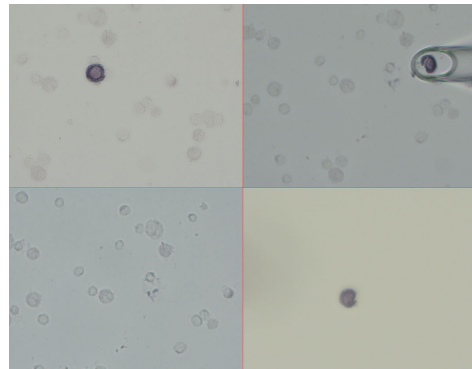
003B - TC



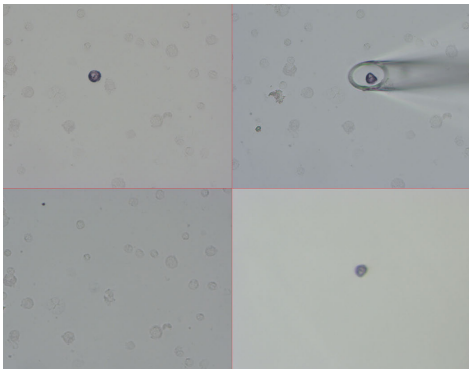
003C - TC



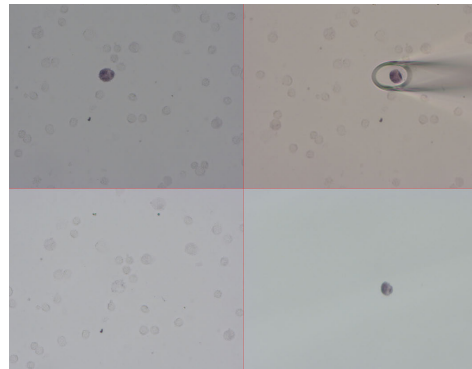
003D - TC



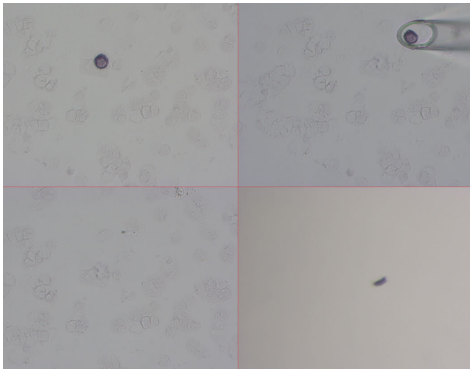
003E - TC



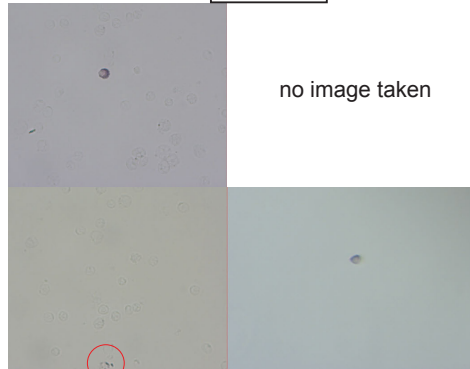
003F - PHC



003G - PHC

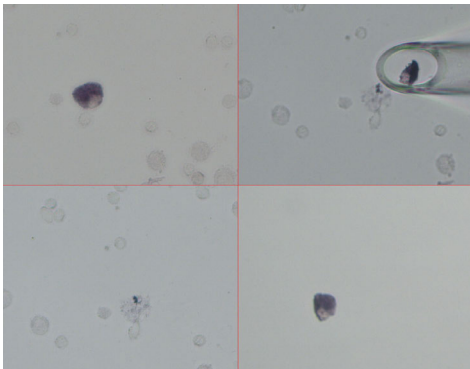


003H - HC

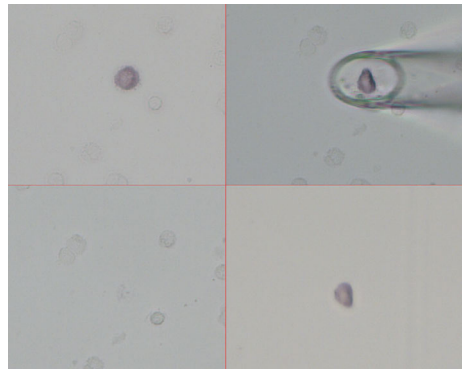


no image taken

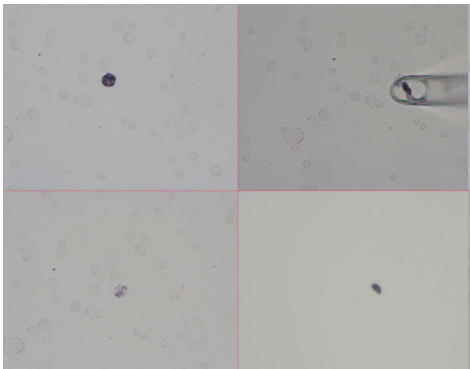
003I - HC



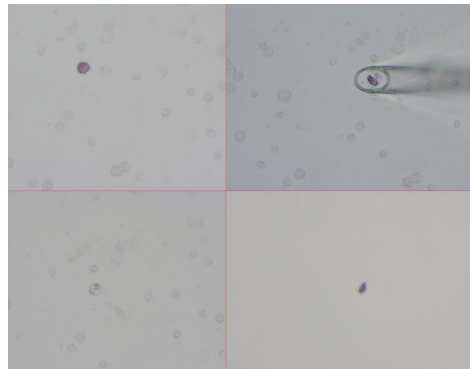
003J - uncertain



003K - uncertain

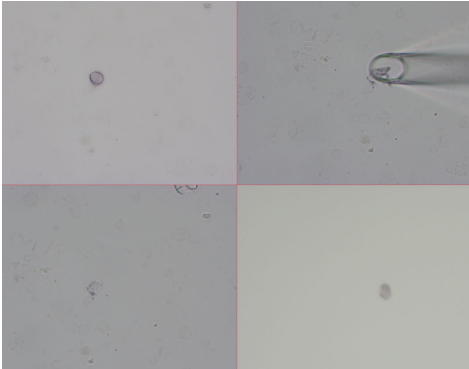


003L - PHC

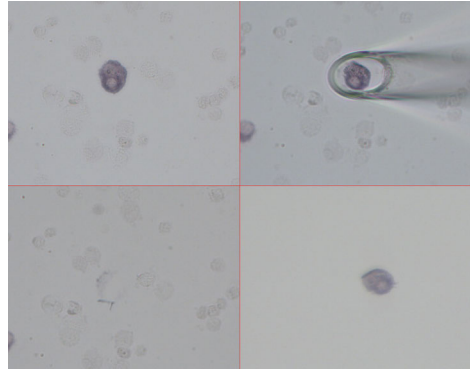


**B**

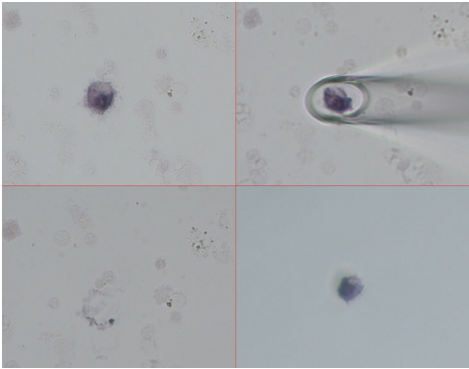
017A - uncertain



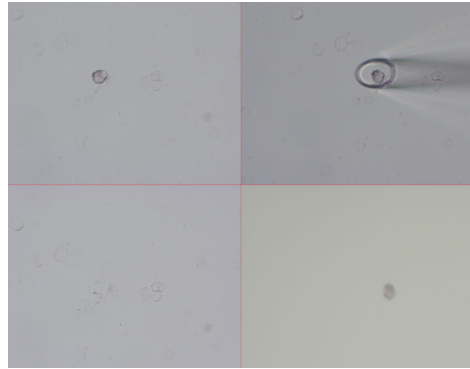
017B - uncertain



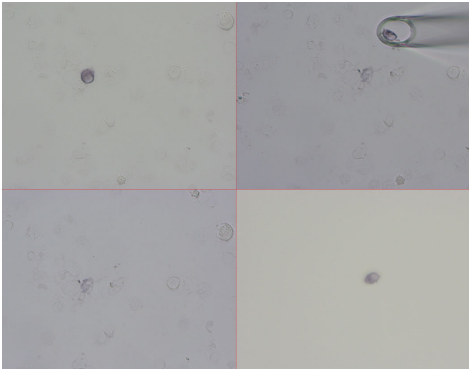
017C - uncertain



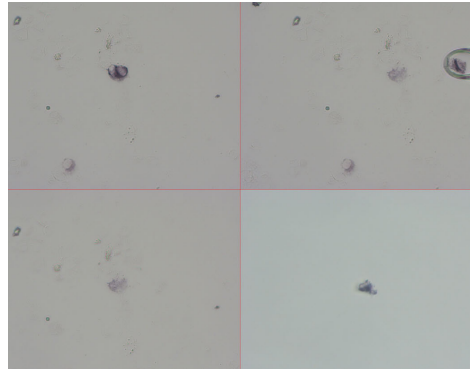
017D - uncertain



017E - PHC

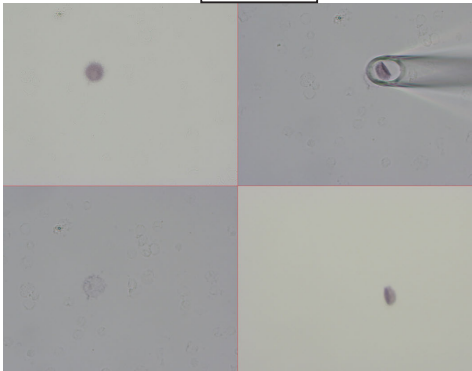


017F - uncertain

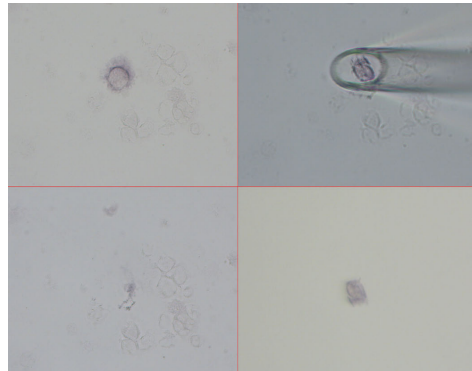




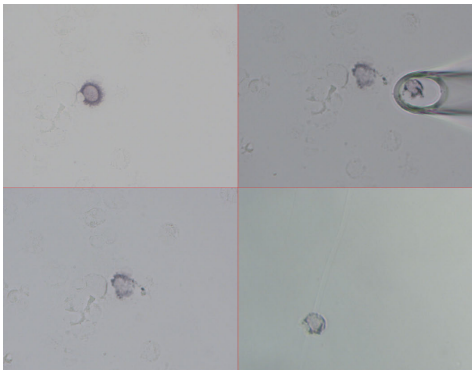
017G - HC



017H - PHC

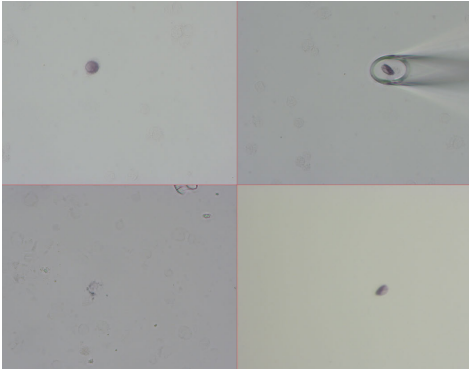


017I - PHC

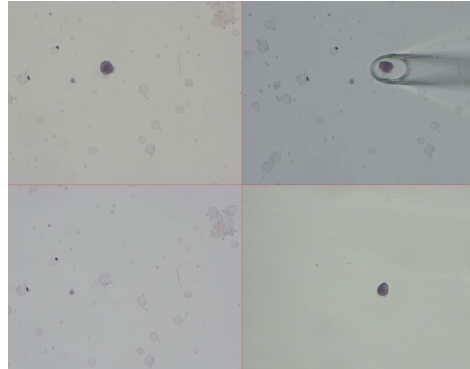


C

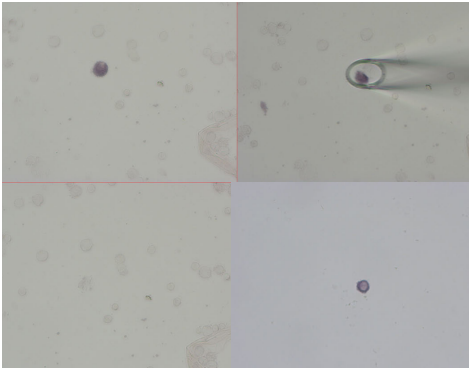
019A - uncertain



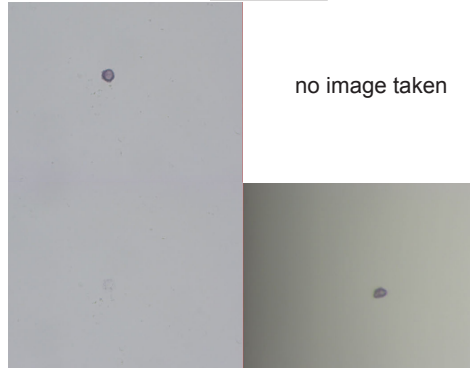
019B - PHC



019C - HC

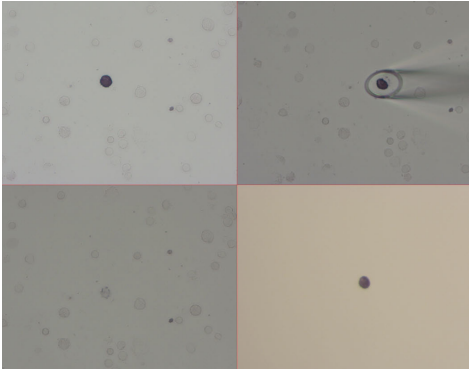


019D - HC

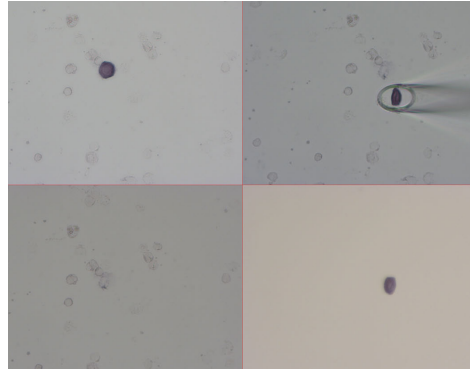


D

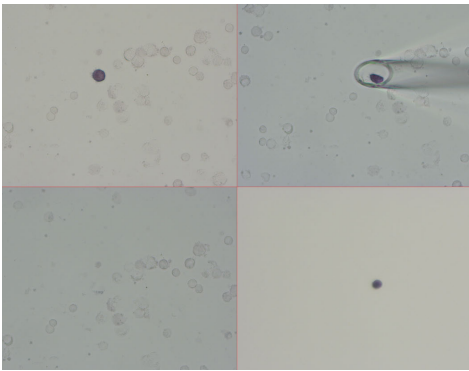
044A - TC



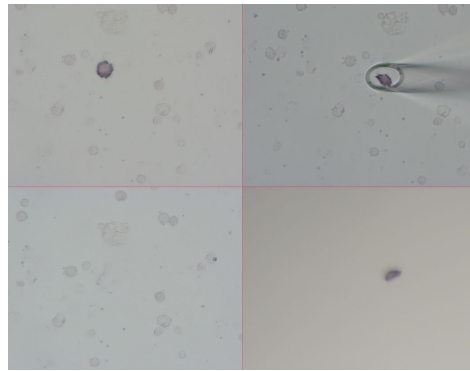
044B - TC



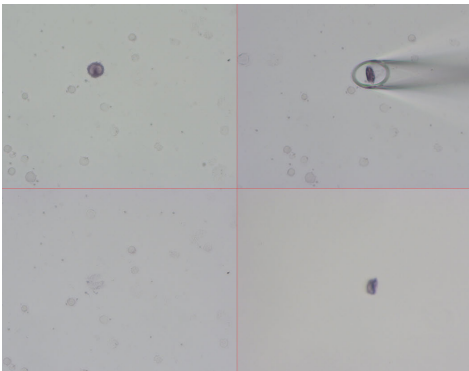
044C - uncertain



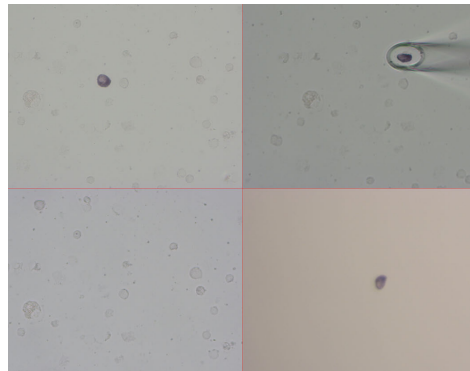
044D - PHC



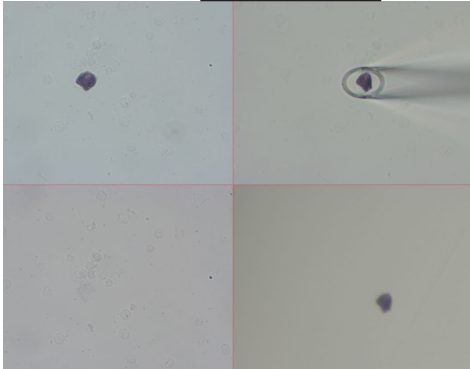
044E - TC



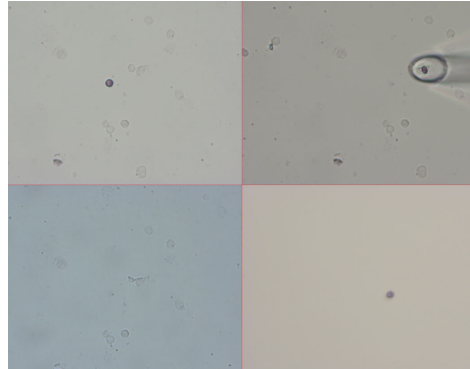
044F - PHC



044G - uncertain

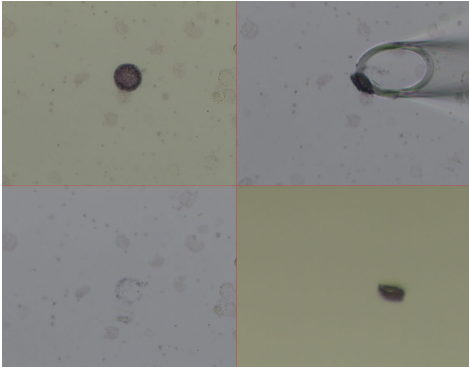


044H - PHC

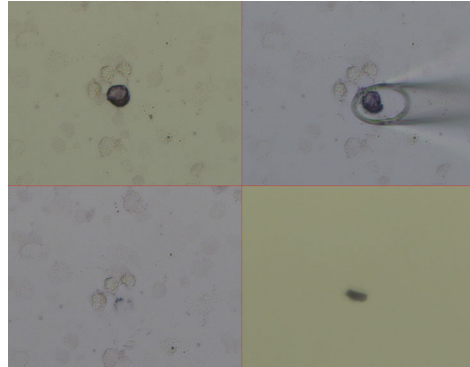


E

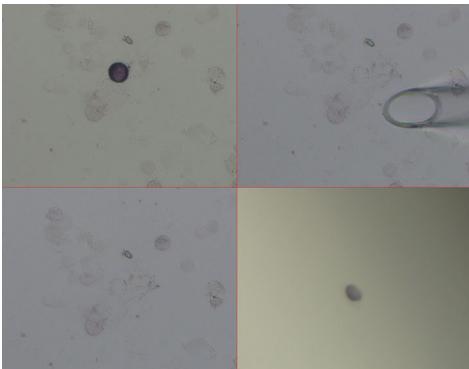
083A - TC



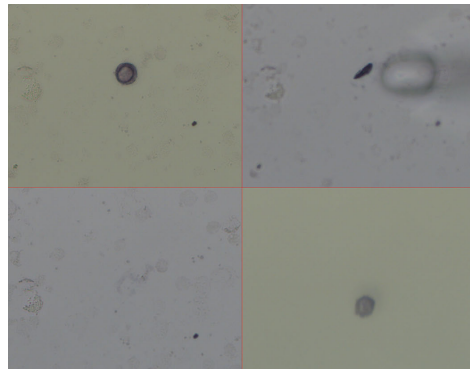
083B - TC



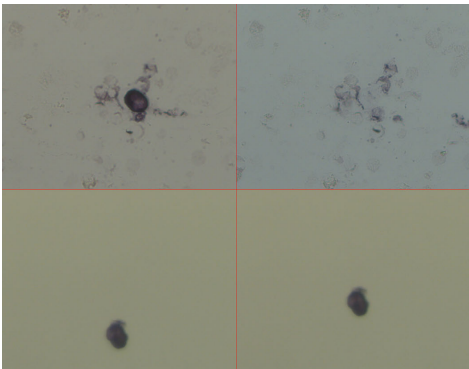
083C - TC



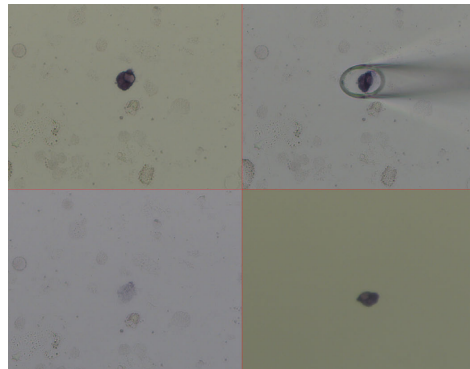
083D - TC



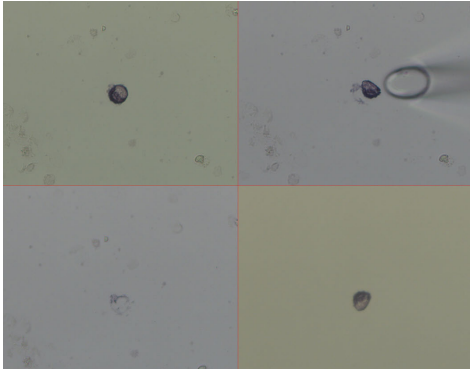
083E - TC



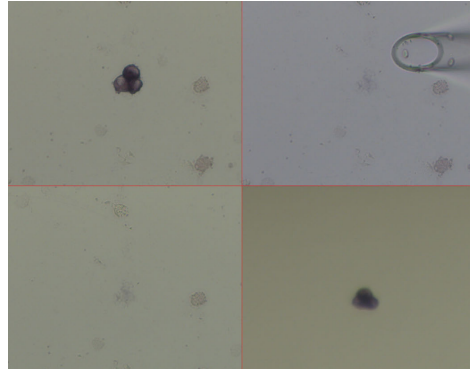
083F - PHC



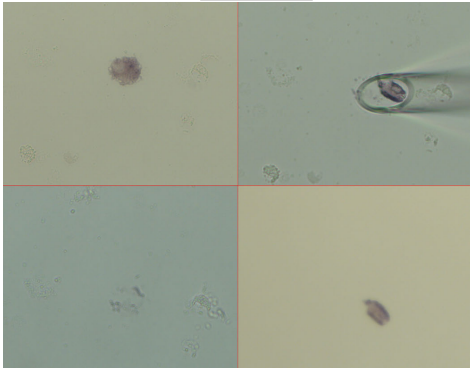
083G - TC



083H - TC

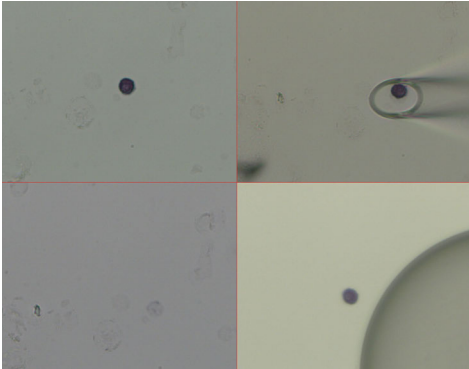


083I - HC

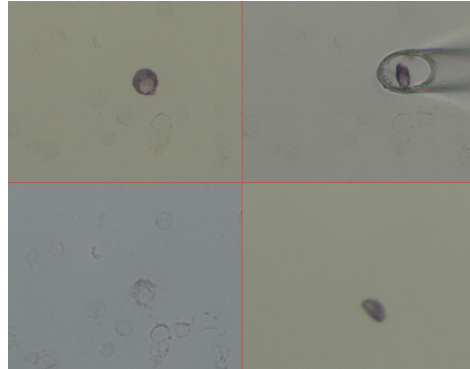


**F**

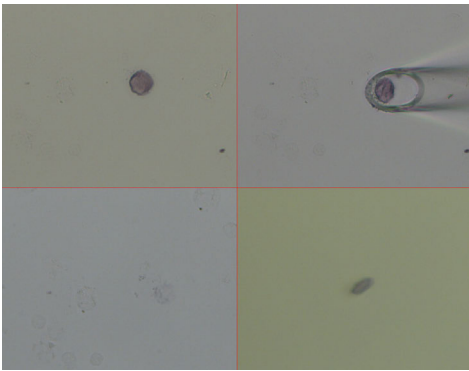
107A - TC



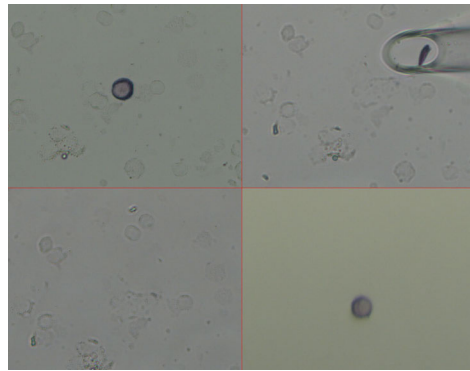
107B - uncertain



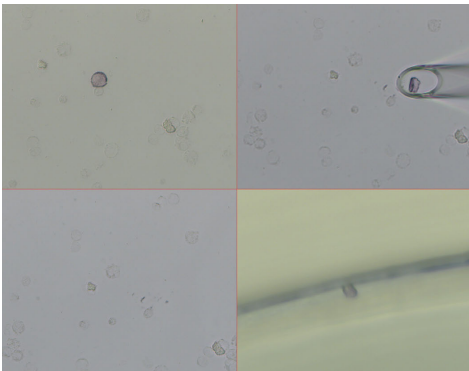
107C - uncertain



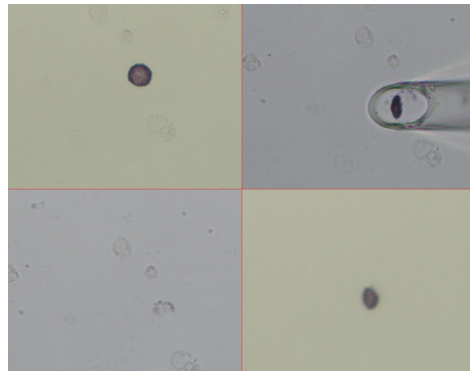
107D - TC



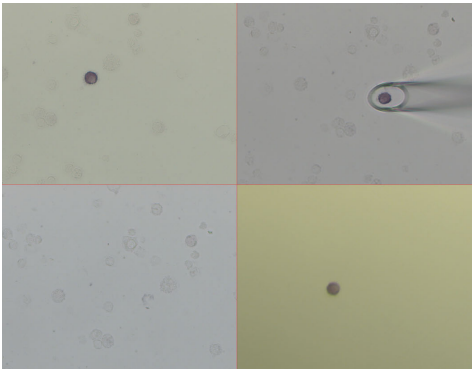
107E - TC



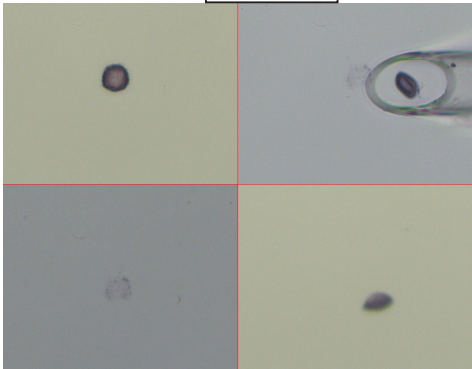
107F - PHC



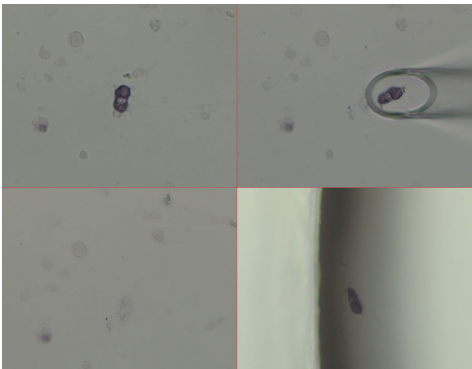
107G - PHC



107R - PHC



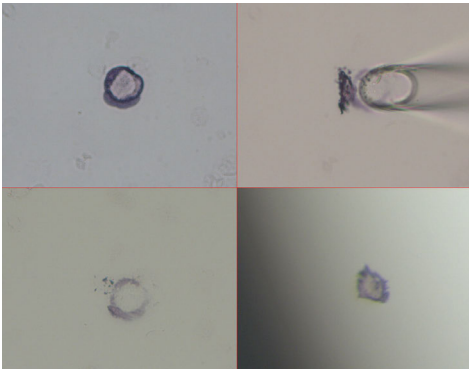
107S - uncertain



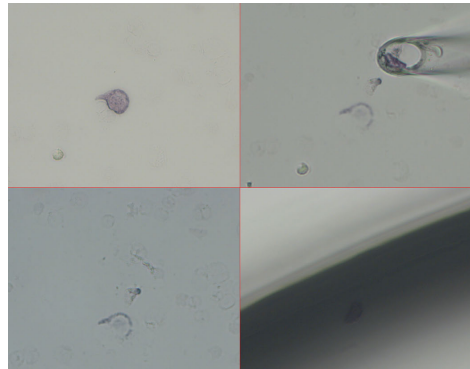


**G**

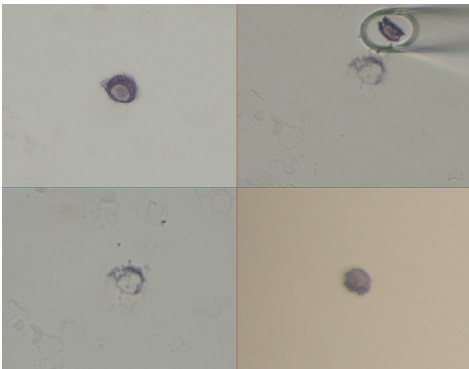
107H - TC



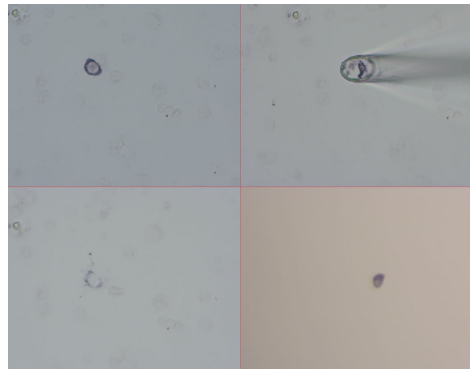
107I - TC



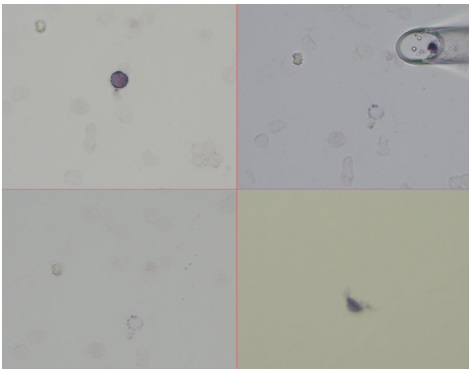
107J - HC



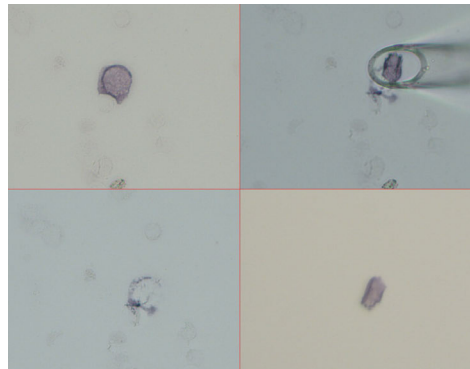
107K - TC



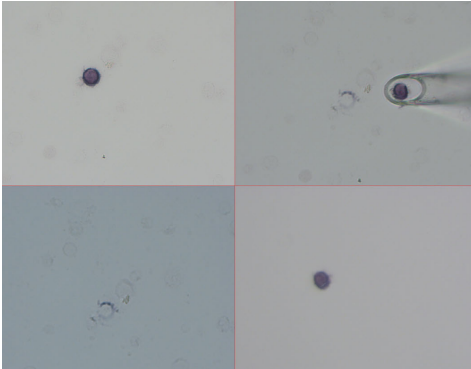
107L - TC



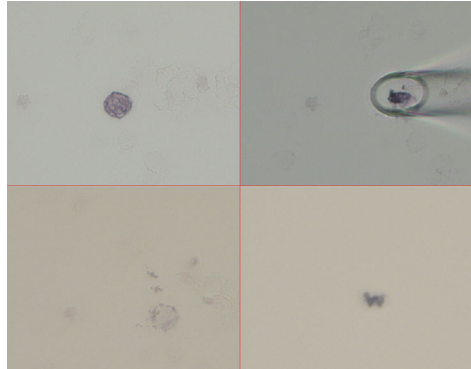
107M - TC



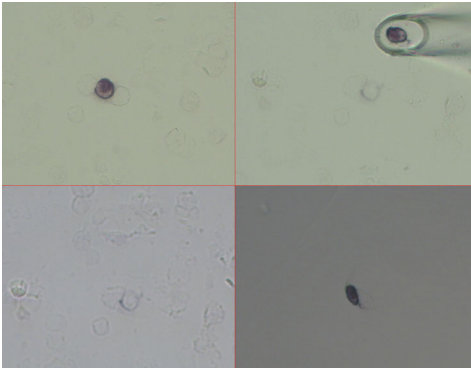
107N - TC



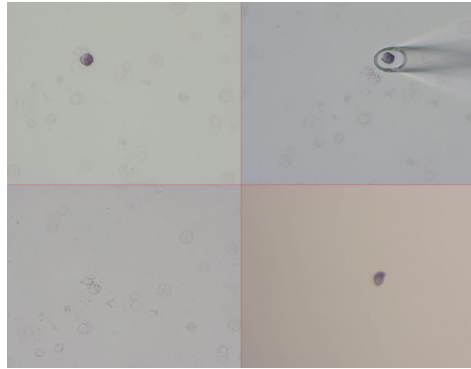
107O - TC



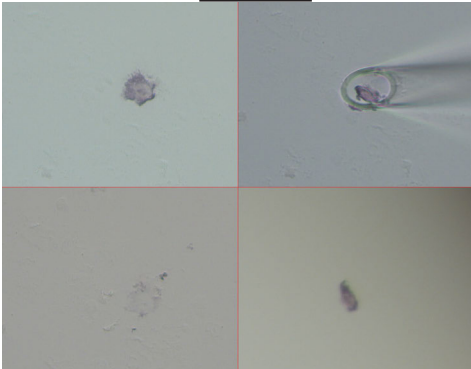
107P - TC



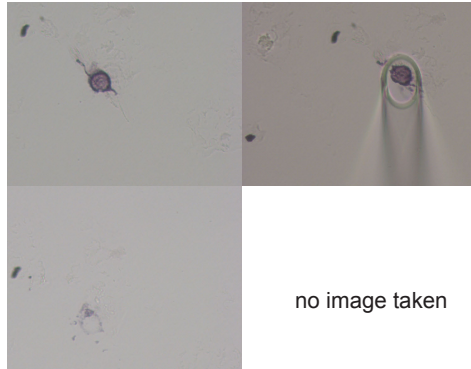
107Q - uncertain



107T - HC

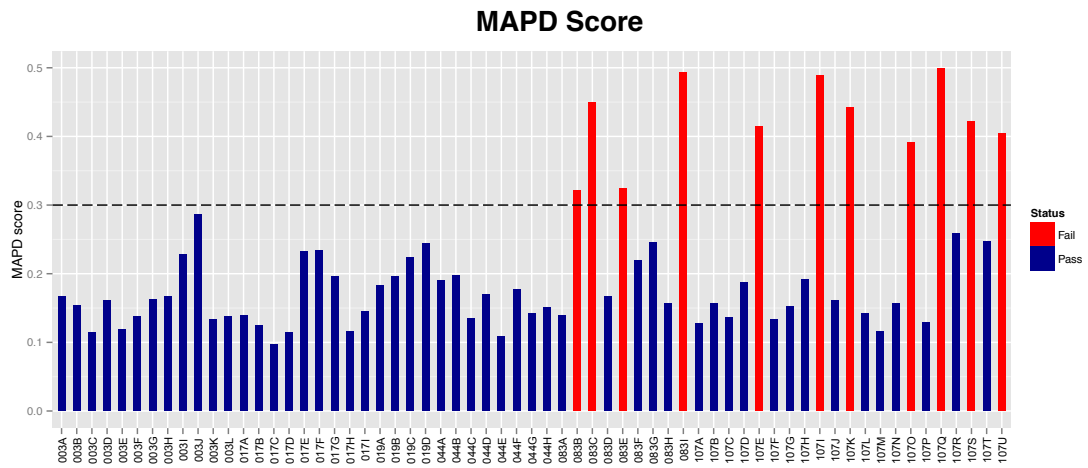


107U - TC



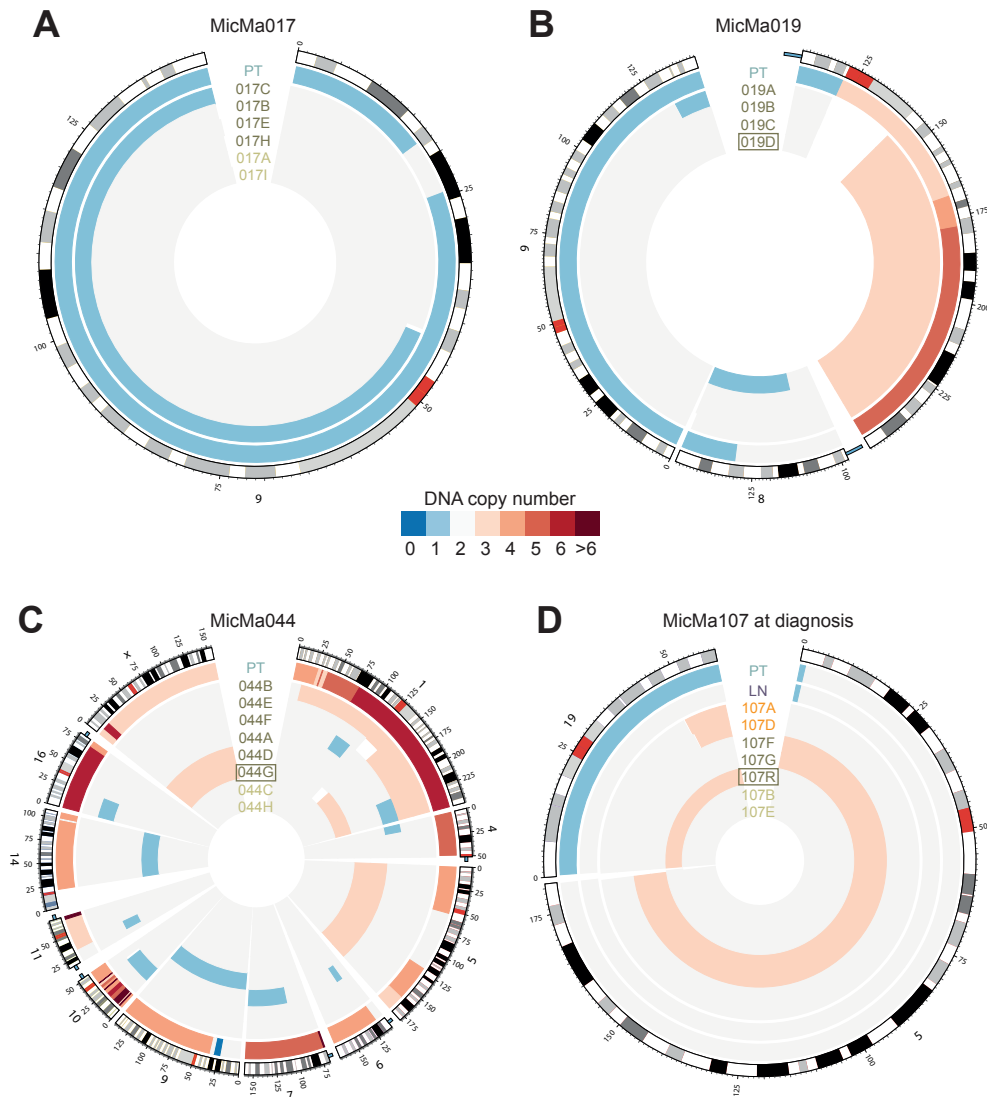
no image taken

**Figure S2: Assessment of noise in single-cell logR profiles.** Median Absolute Pairwise Difference (MAPD) values (Materials and methods) embody the noise in logR between two consecutive genomic bins genome wide. Red cells failed this QC filter.

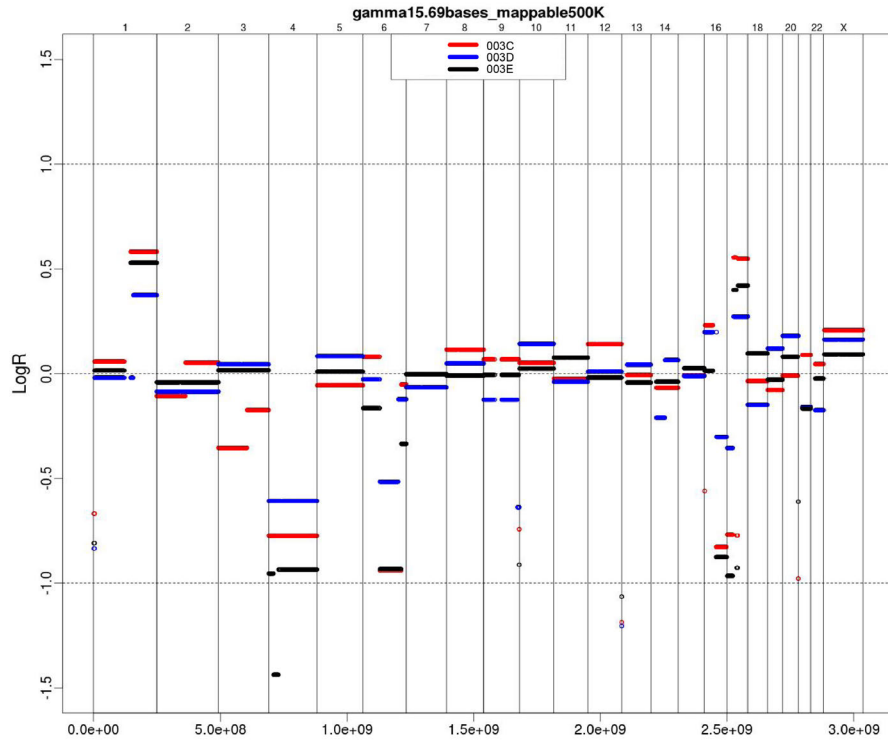


**Figure S3: Focus on DNA copy number events overlapping in AUs and bulk tumor.**

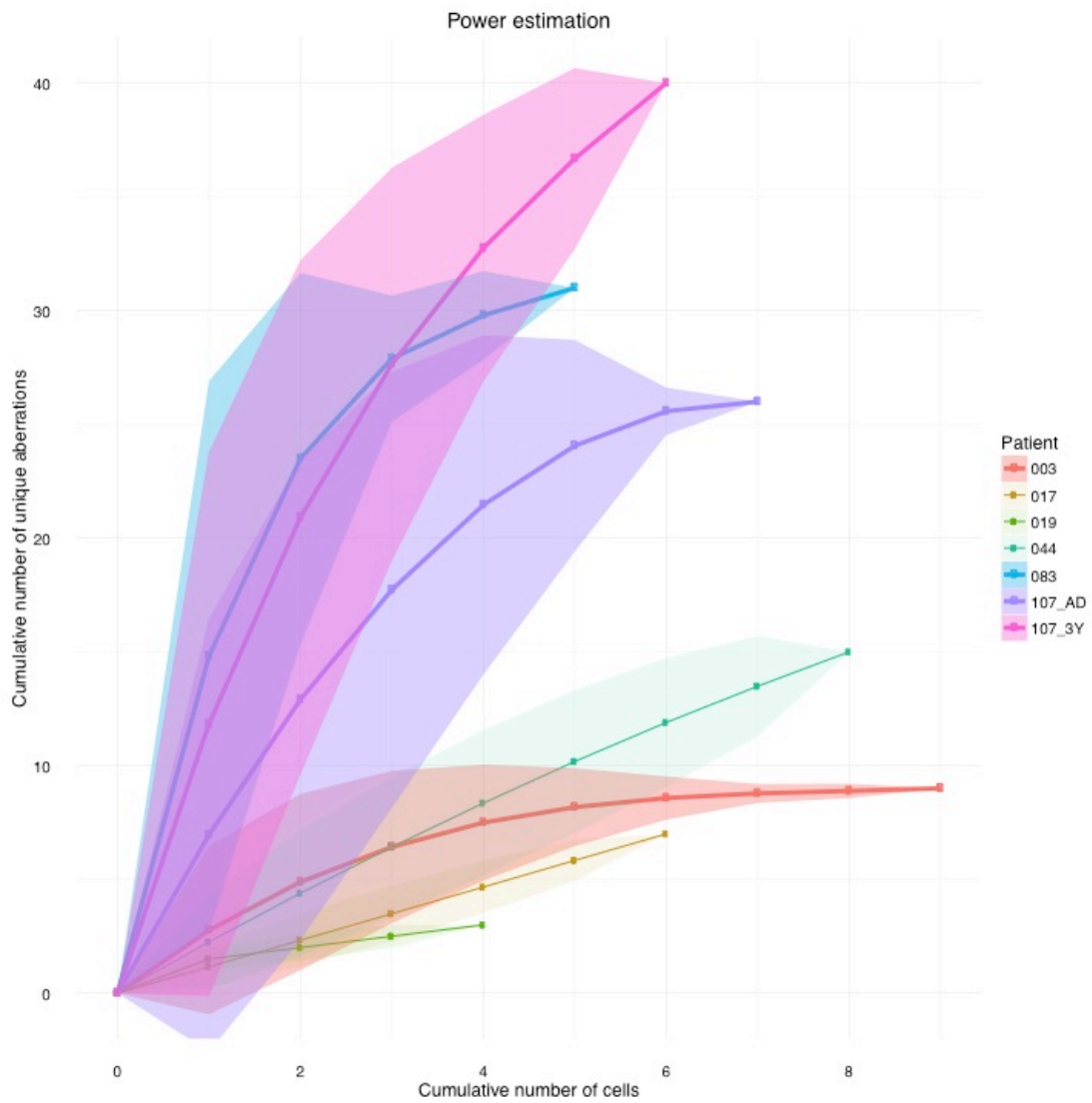
(A-D) Per patient profiles are shown as concentric circles inside the circular human karyogram. Only those patients and regions that have overlapping CNAs between AUs and the tumor bulk are depicted. Total copy number is represented as a heat map from blue to red as indicated. Tumor samples and single cells are labeled on the corresponding circles and are color-coded: primary tumor bulk (PT) in cyan, lymph node bulk (LN) in purple, DTCs in orange, aberrant cells of unknown origin in dark green and Normal cells in green. Control cells are boxed. Panels are shown for patient MicMa017 (A), MicMa019 (B), MicMa044 (C) and MicMa107 at the time of diagnosis (D).



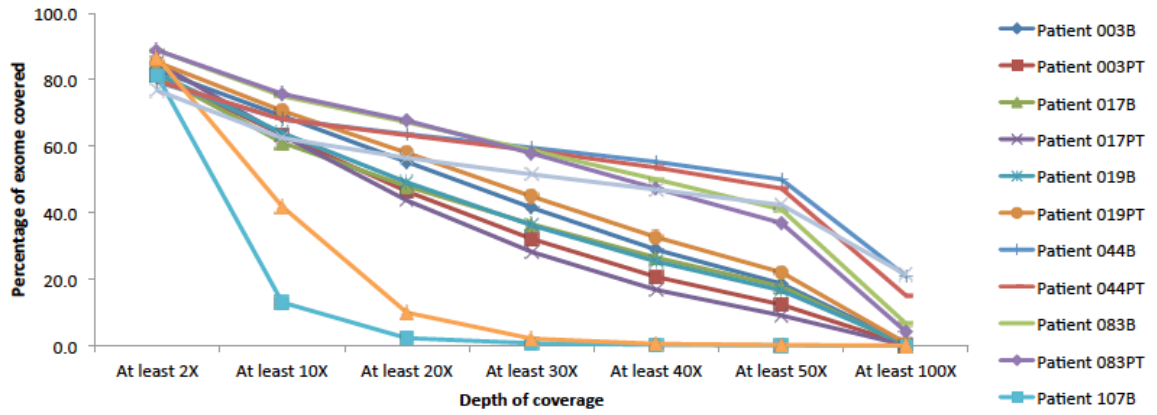
**Figure S4: LogR profile of a ‘Doublet’ and two single cells.** Genome-wide logR segments of 3 DTCs (003C, 003D, 003E) of patient MicMa003. Red and black depict logR segments obtained from single DTCs 003C and 003E, respectively, while blue indicates the logR segments of a doublet (003D).



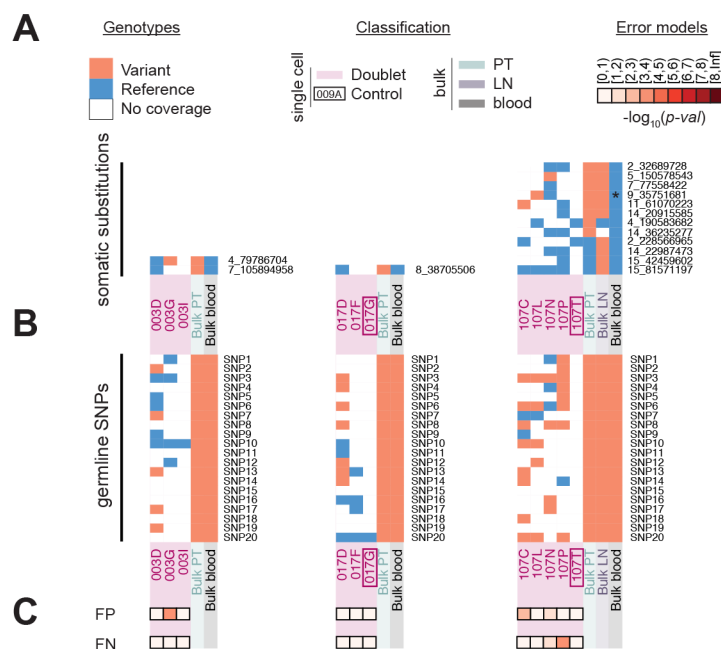
**Figure S5: Retrospective power analysis.** The cumulative number of unique CNAs is plotted as a function of the cumulative number of single cells for each sample. Mean and standard deviations of 100,000 randomized orders are shown. AD, at diagnosis; 3Y, 3 years after diagnosis.



**Figure S6: Coverage statistics of bulk exome sequences for all tumors and matched normal blood samples.** The breadth of coverage across the exome (Y-axis) is depicted for different sequence depth cutoffs (X-axis). Primary tumor (PT), lymph node metastasis (LN) and matched blood (B).



**Figure S7: Genotyping of single nucleotide variants from bulk tumor exome sequences in the Doublet sequences.** (A, B) Heat maps per tumor, where each row represents either a single somatic substitution called on the corresponding bulk exome and the matched blood (A), or a random heterozygous germline SNP (20 total, B), and columns represent the different Doublet or exome datasets obtained for that tumor. Tile colors reflect the presence of the mutant allele (orange), its absence (reference, blue), or whether there was no coverage at that position (white). (C) Modeled probability of observing an at least equally extreme pattern of somatic reference and variant alleles for that Doublet through false positives (none of the cells have any of the tumor's somatic mutations and hence derive from another lineage) or false negatives (at least one of the Doublet's cells derives from the tumor and hence contains these somatic mutations). Model results are encoded as heat maps of  $-\log_{10}(p)$ . For clarity, loci with zero coverage in all of the Doublets of that patient are omitted.



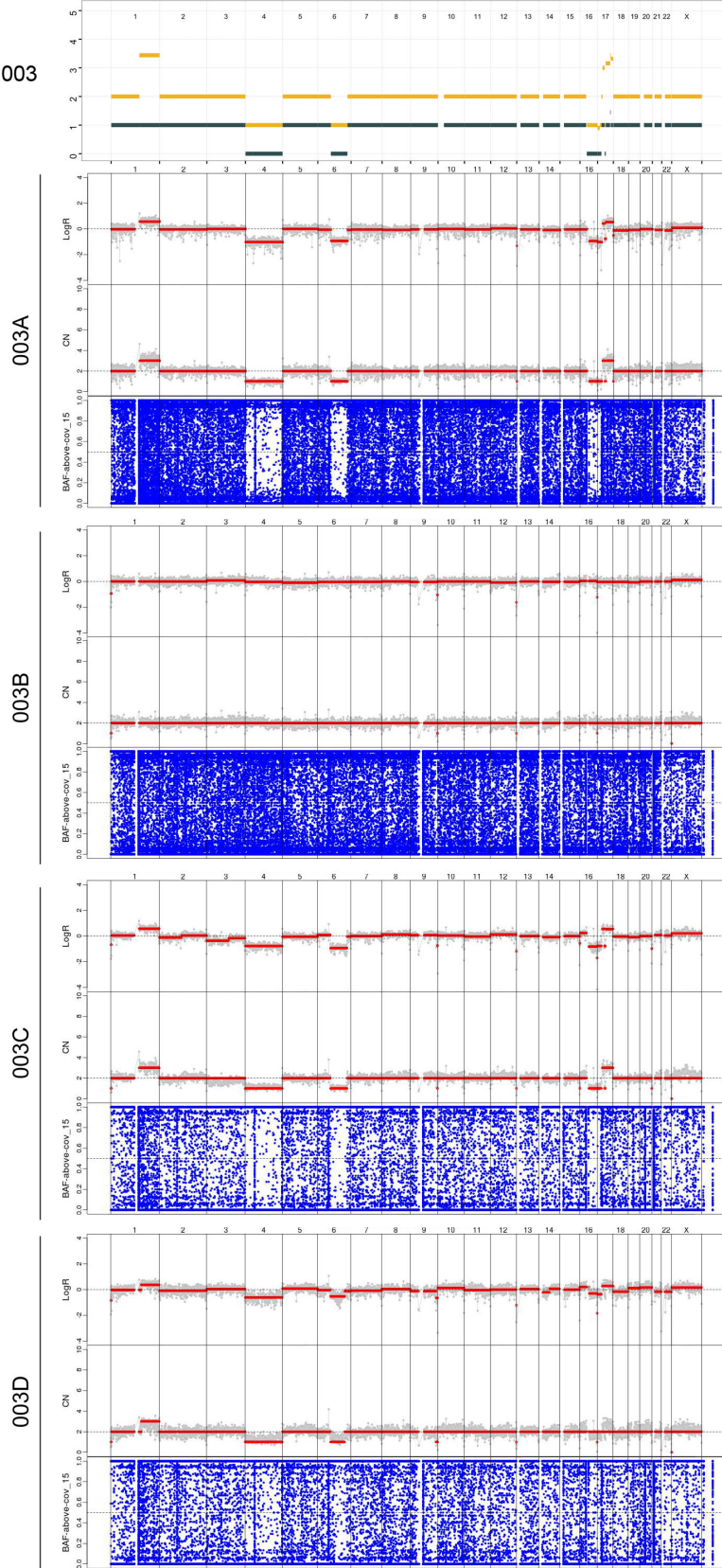


**Figure S8: Tumor bulk copy number profiles and logR, copy number and BAF data of single cells from the corresponding patient.** Each panel highlights the bulk copy number profile (on top) and single cell data (bottom) for a single patient/time point: patient MicMa003 (A), MicMa017 (B), MicMa019 (C), MicMa044 (D), MicMa083 (E), MicMa107 at diagnosis (F) and 3 years post-diagnosis (G). The copy number profile(s) are the output of the Battenberg algorithm based on the SNP-array data of bulk tumor DNA. Yellow indicates the total copy number, black the copy number of the minor allele. Regions with subclonal aberrations are evidenced by a non-integer copy number state. For each corresponding single cell, the logR, fitted copy number and BAF are plotted across the genome.

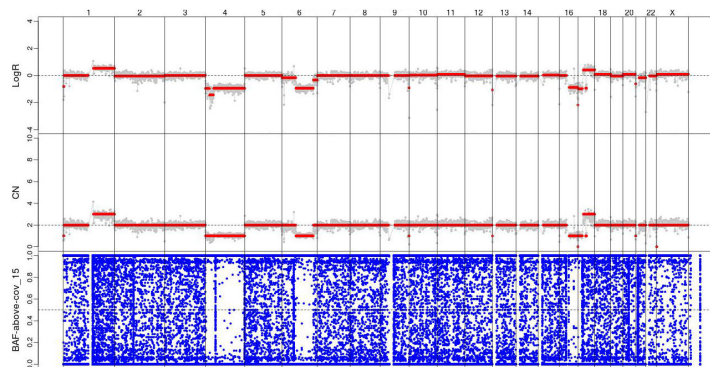
A

MicMa003

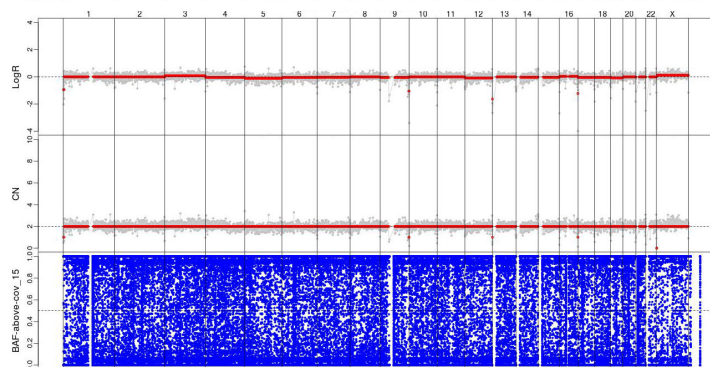
Ploidy: 1.94, aberrant cell fraction: 64%



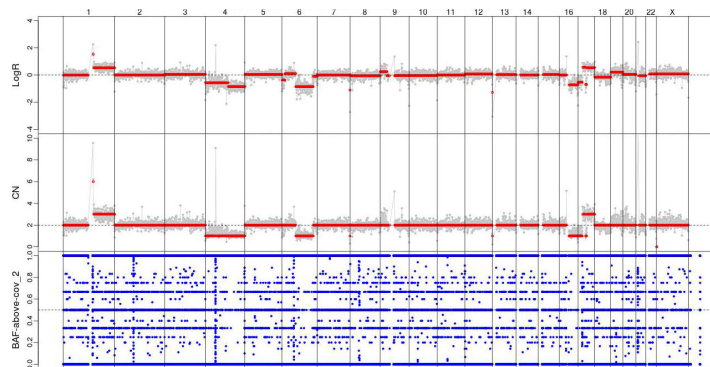
003E



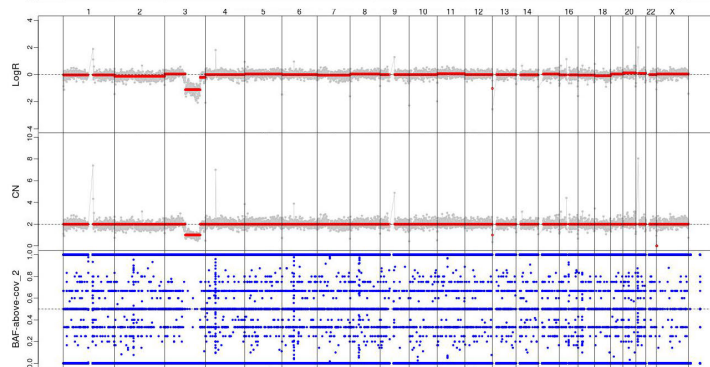
003F



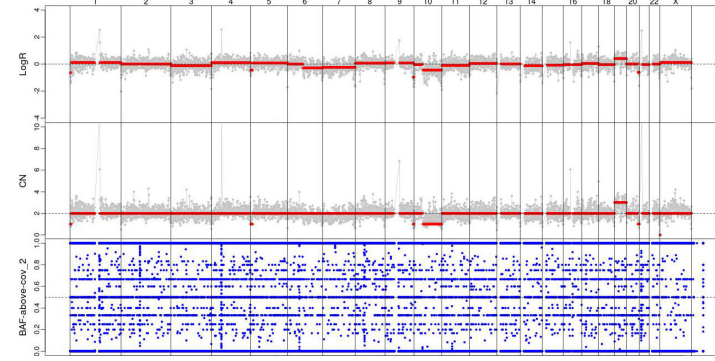
003G



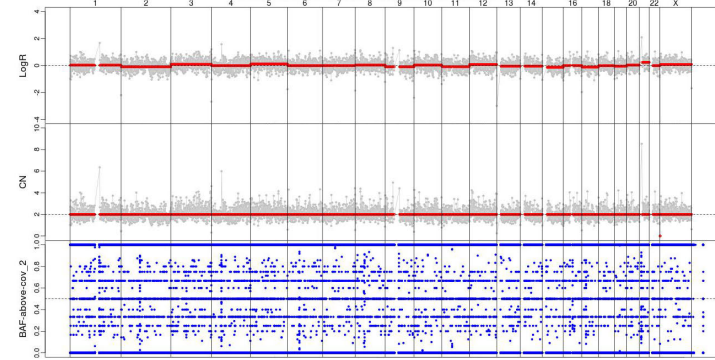
003H



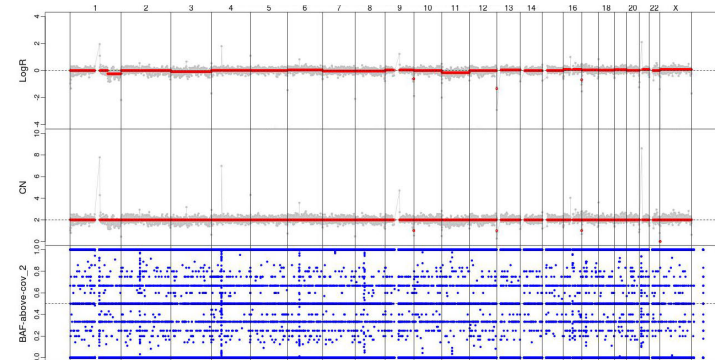
003I



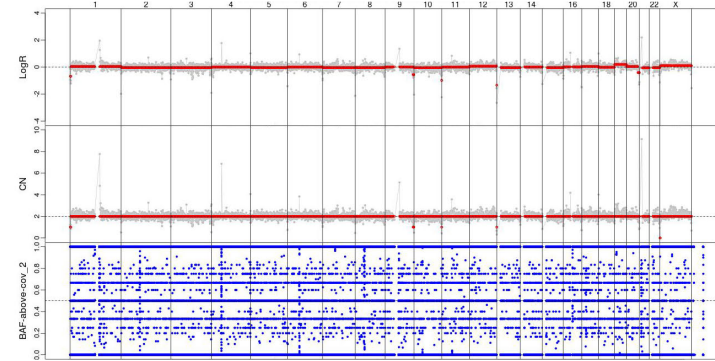
003J



003K



003L

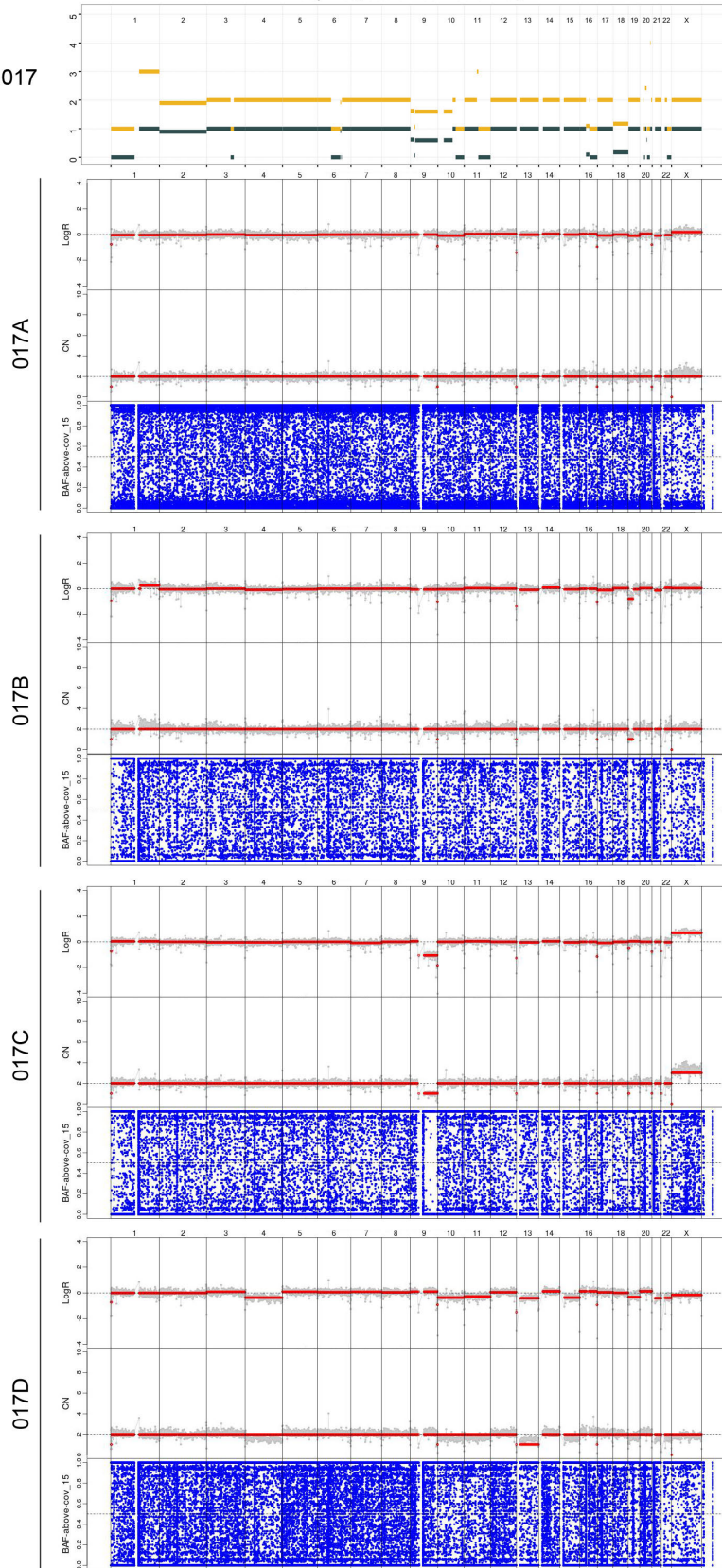




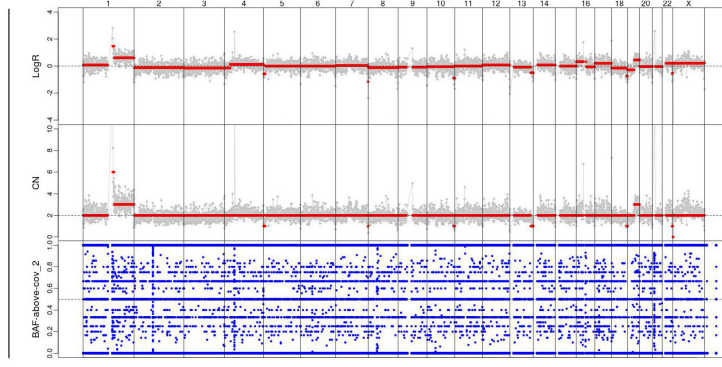
B

MicMa017

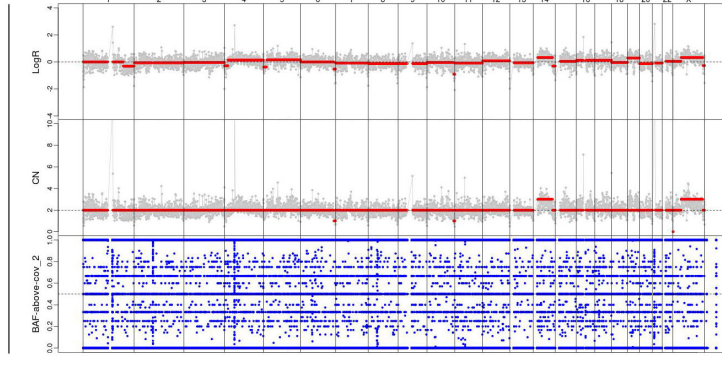
Ploidy: 1.95, aberrant cell fraction: 49%



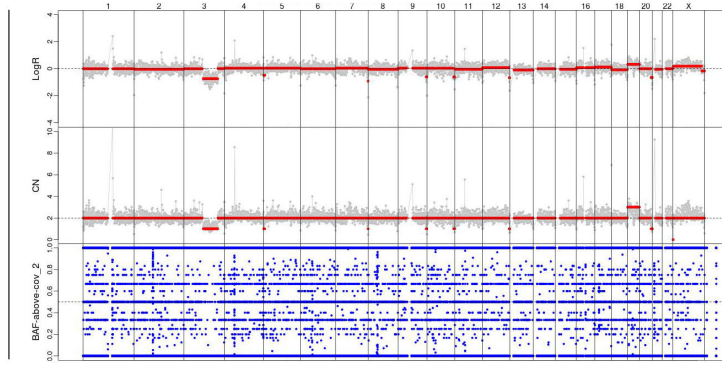
017E



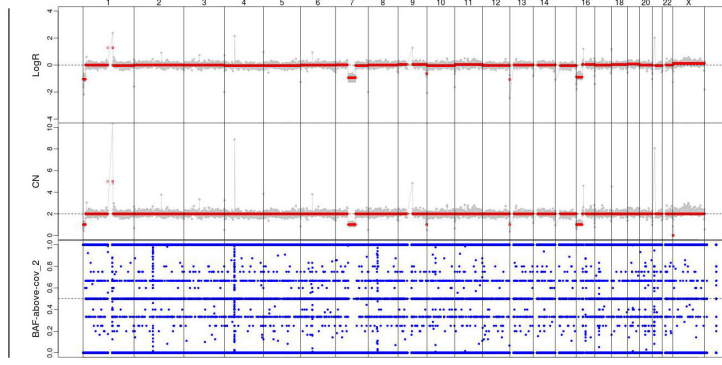
017F



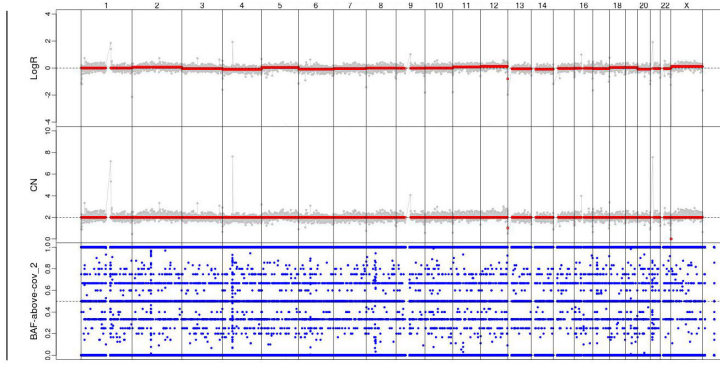
017G



017H



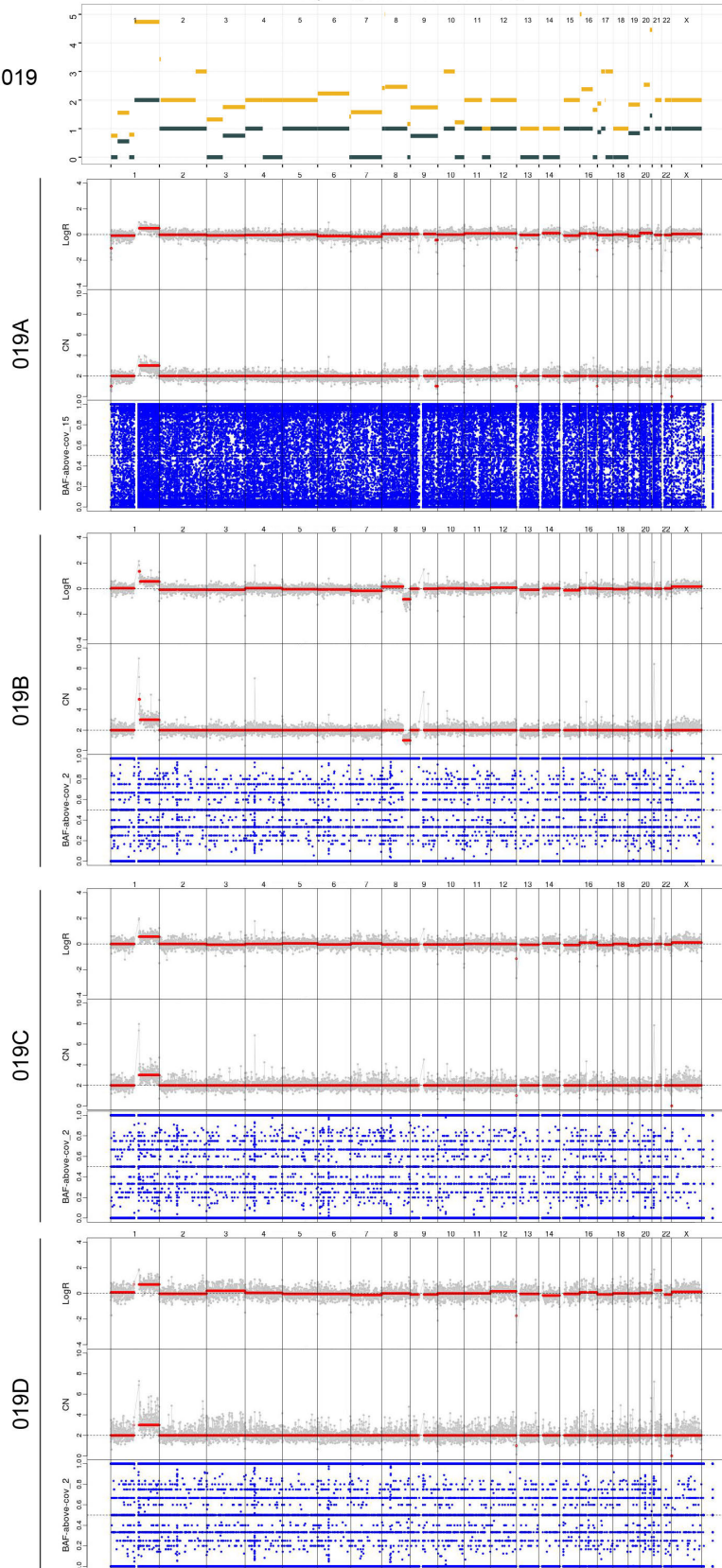
0171



C

MicMa019

Ploidy: 1.78, aberrant cell fraction: 36%

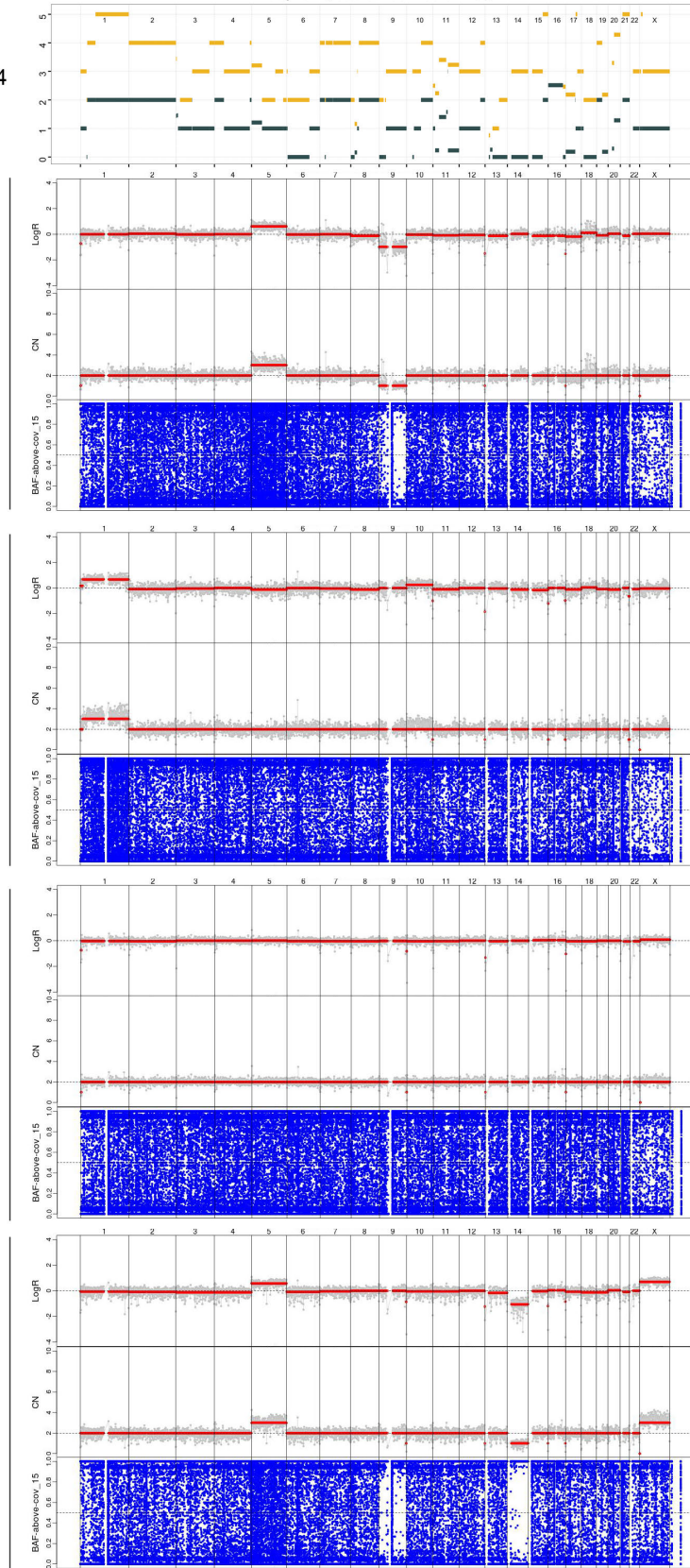




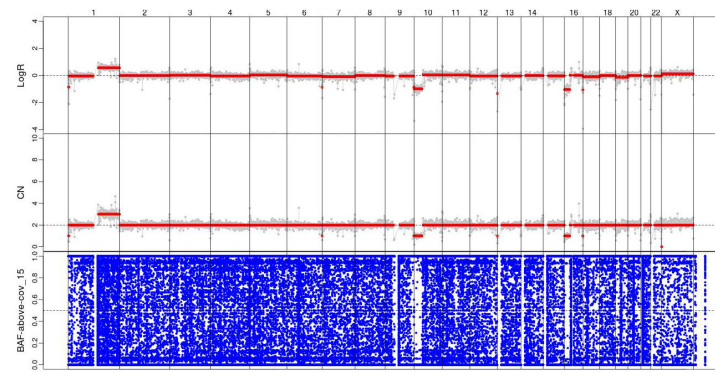
D

MicMa044

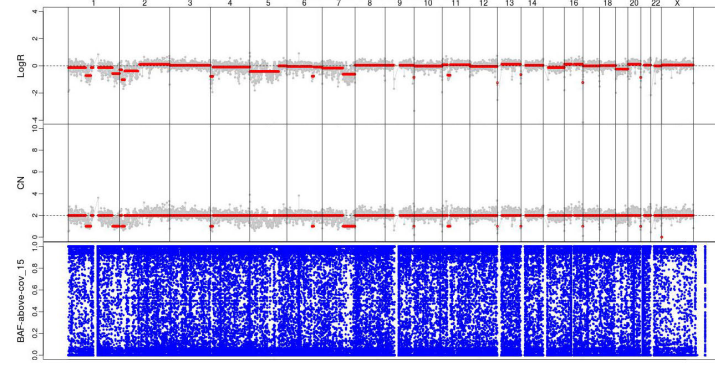
Ploidy: 3.22, aberrant cell fraction: 23%



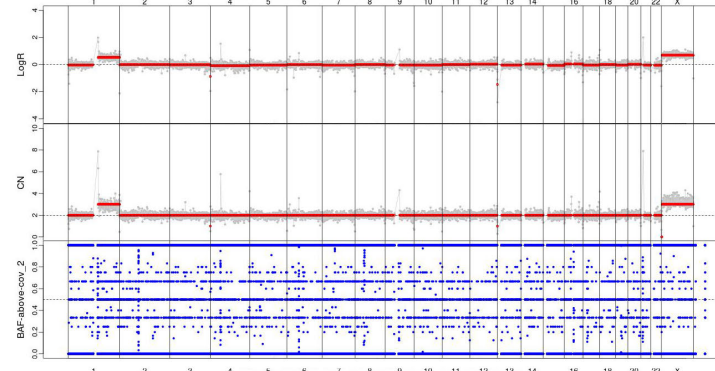
044E



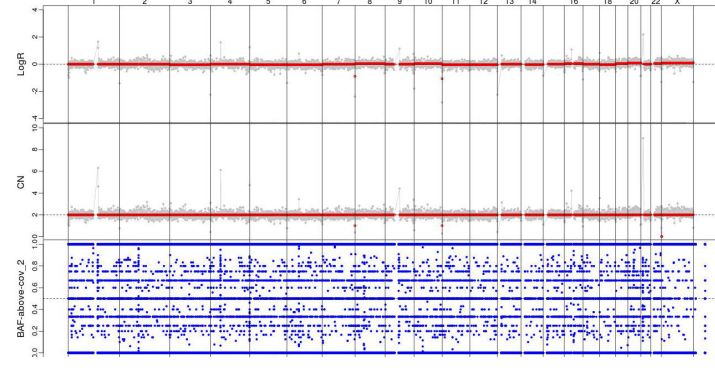
044F



044G



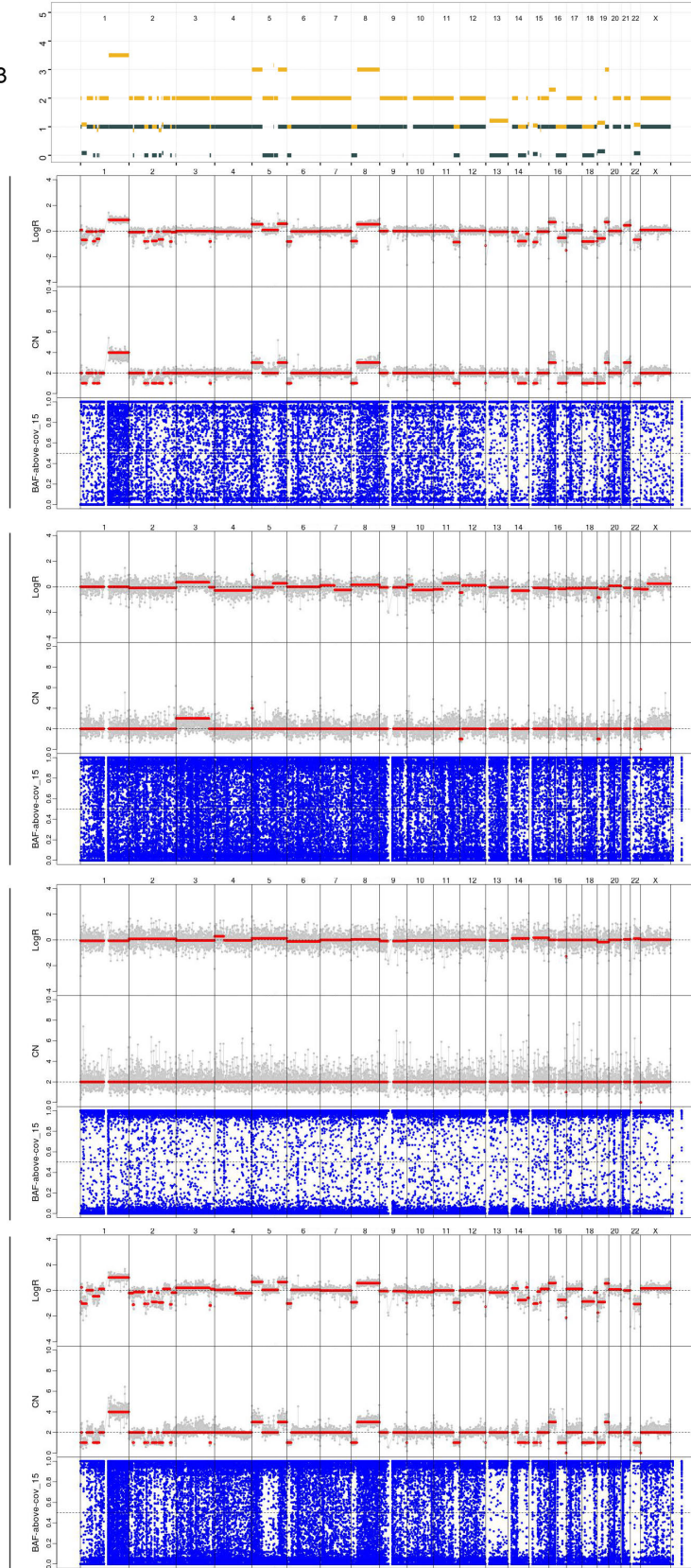
044H



E

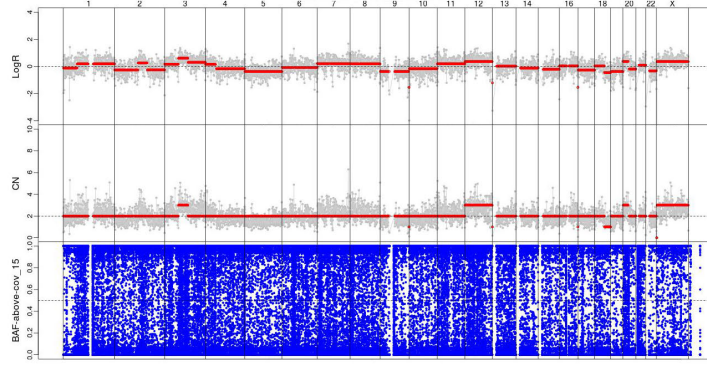
MicMa083

Ploidy: 1.97, aberrant cell fraction: 52%

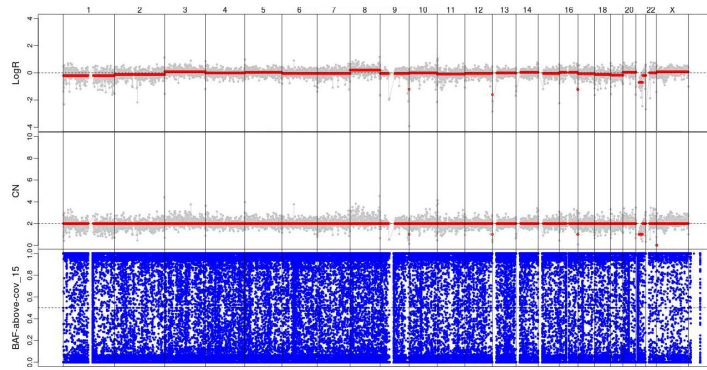




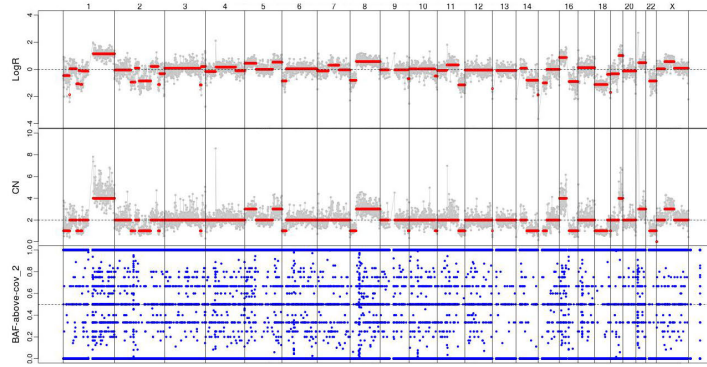
083E



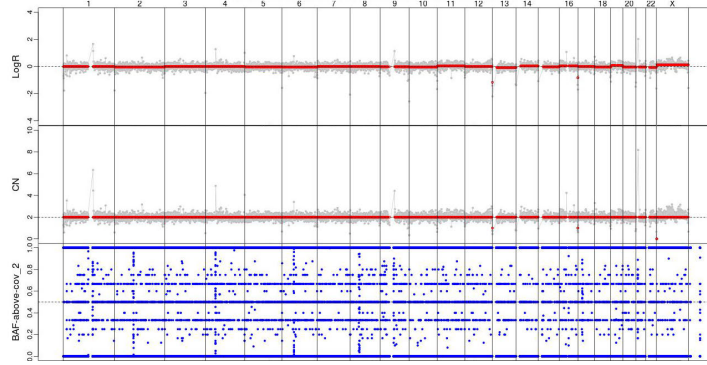
083F



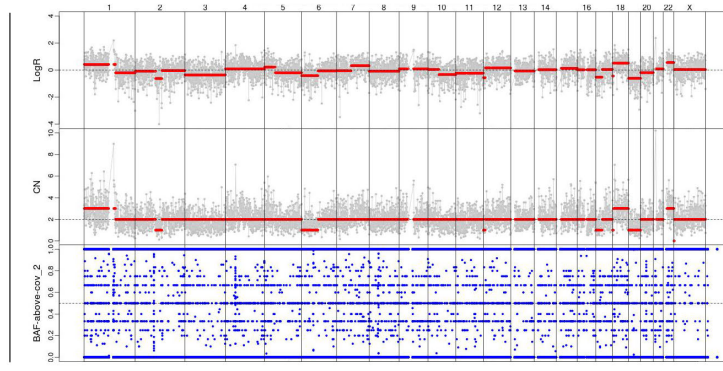
083G



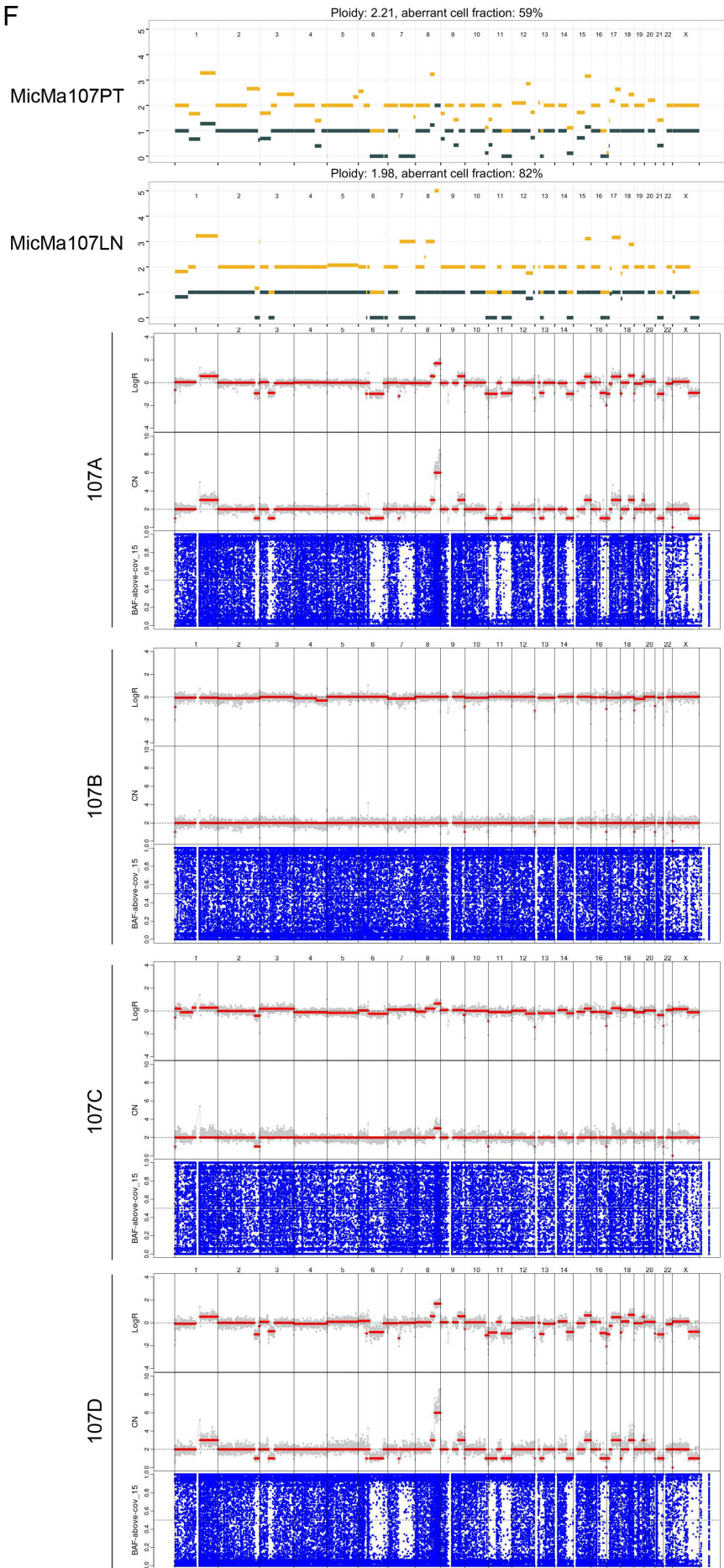
083H



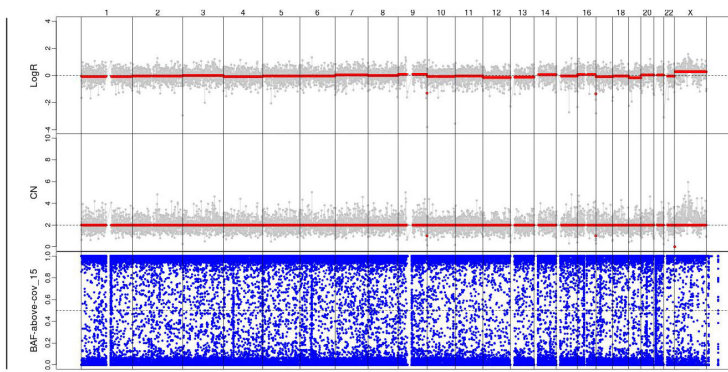
0831



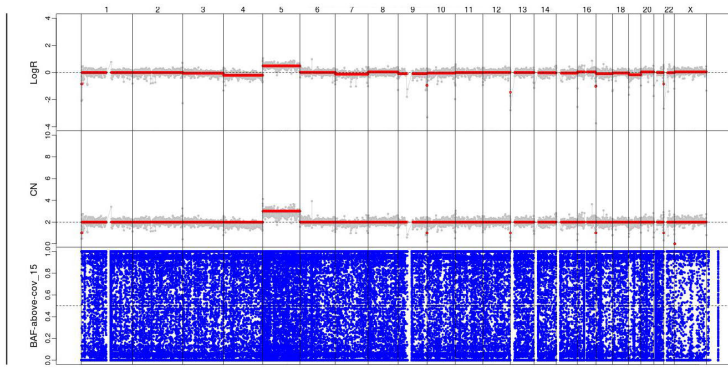
F



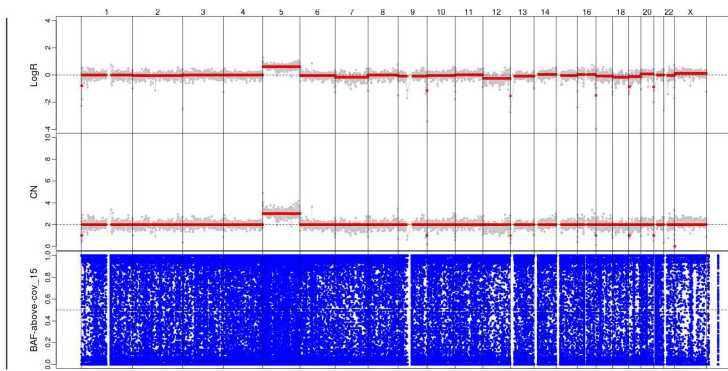
107E



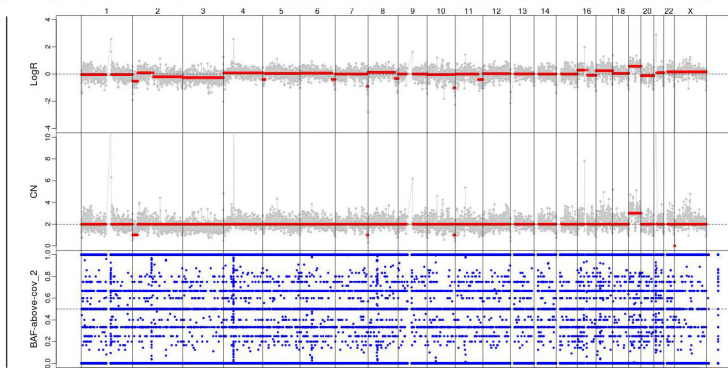
107F



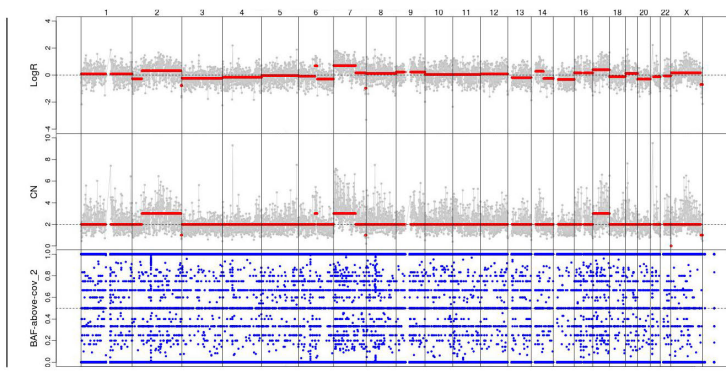
107G



107R

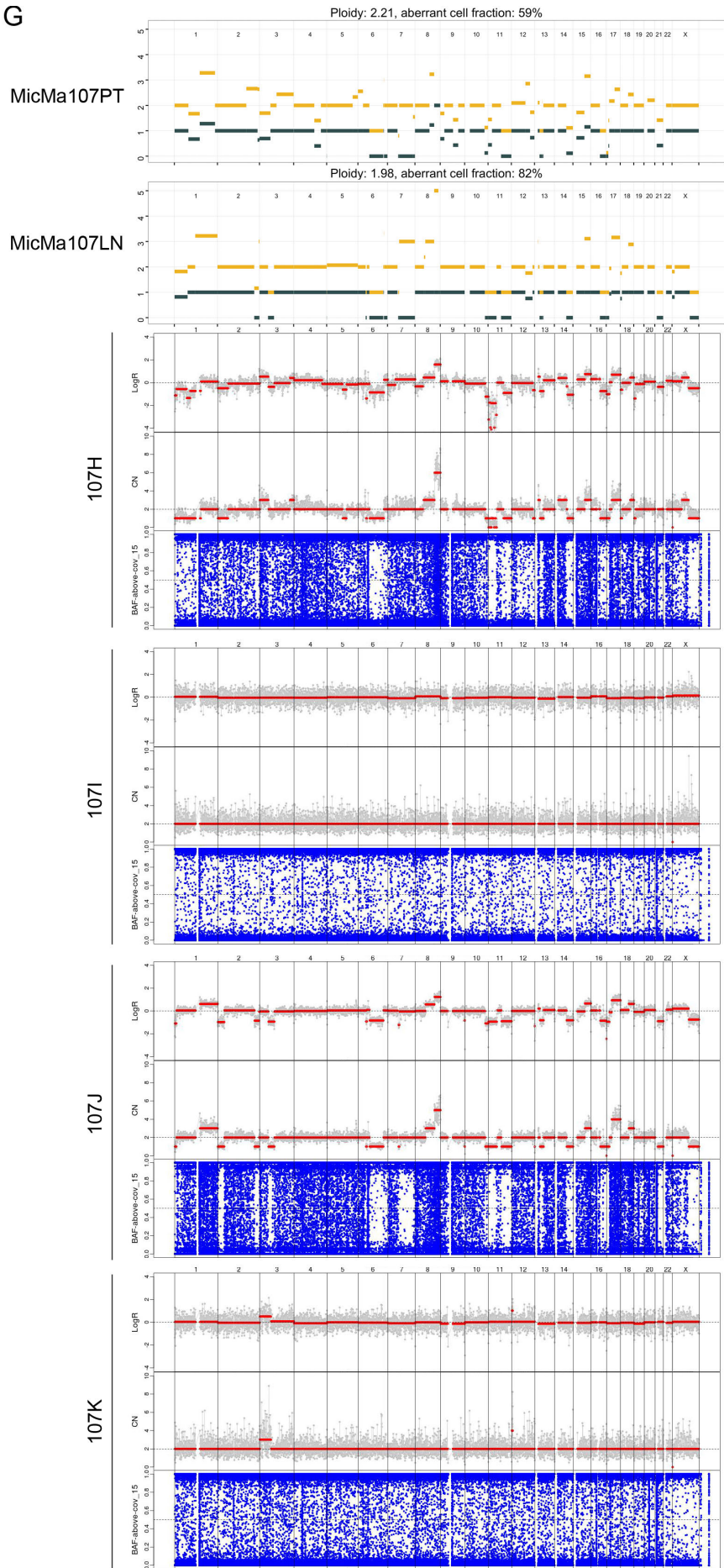


107S

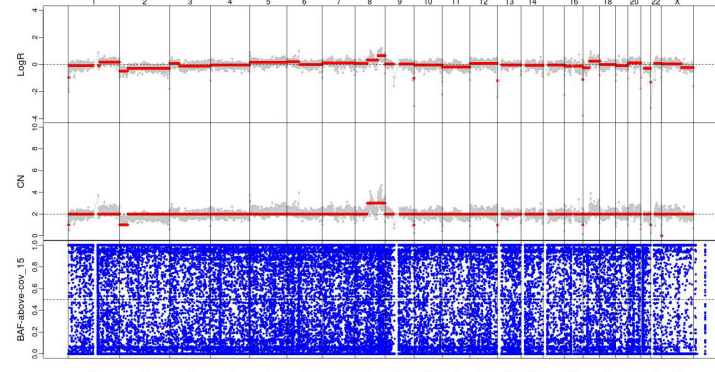




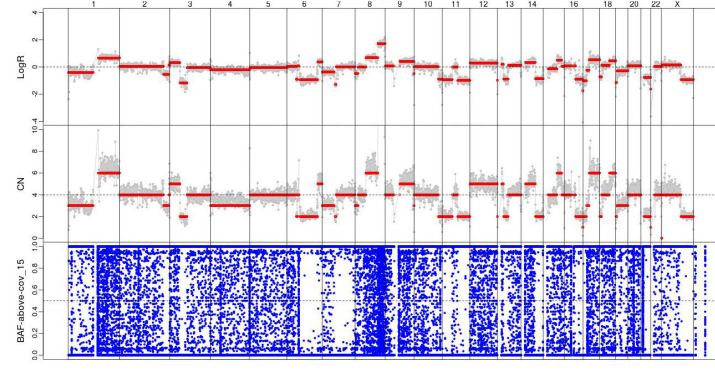
G



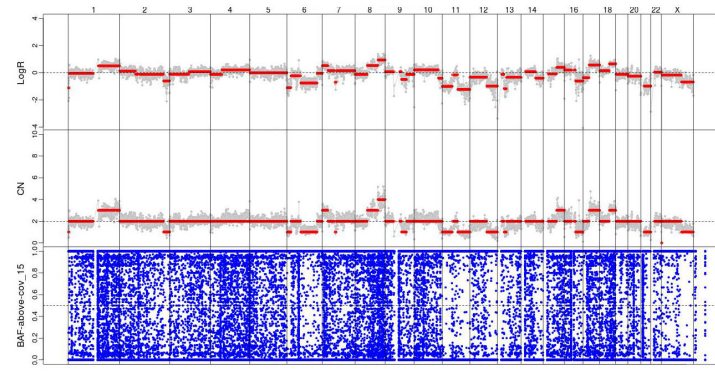
107L



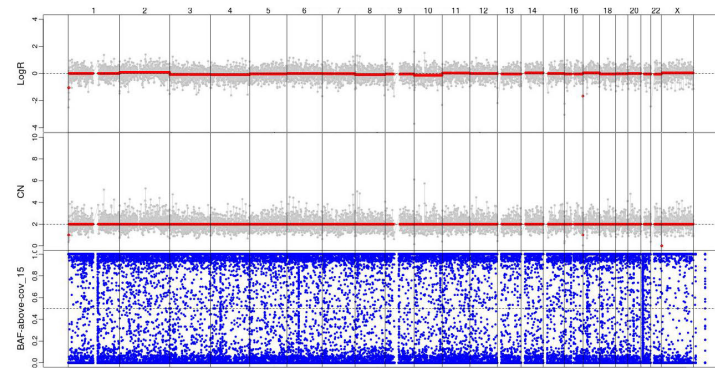
107M



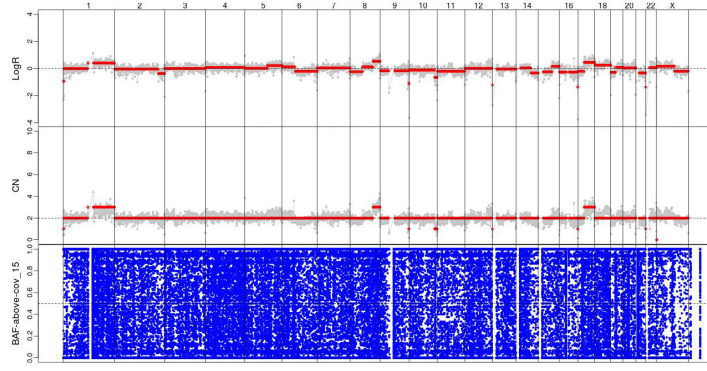
107N



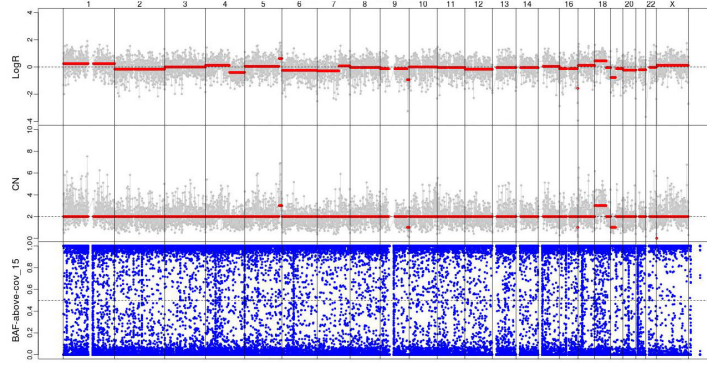
107O



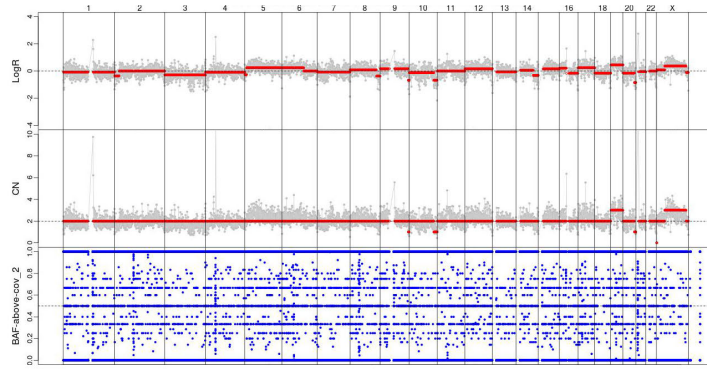
107P



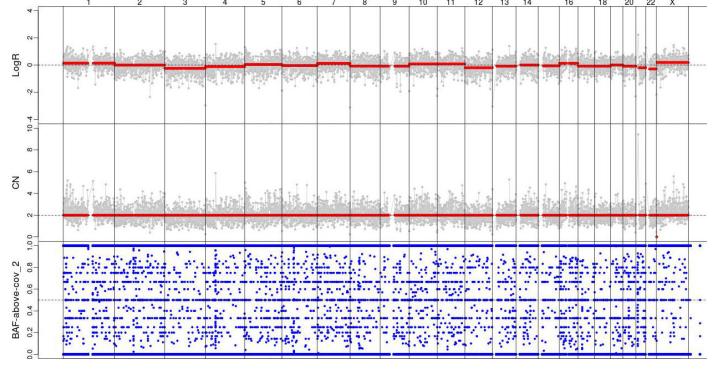
107Q



107T



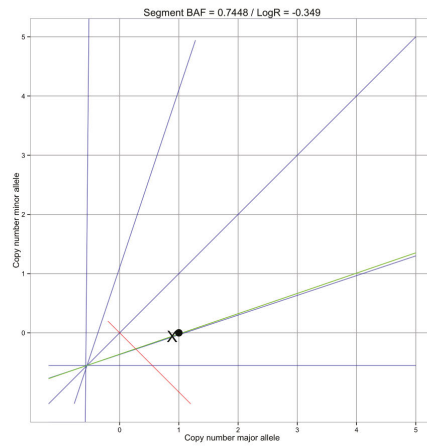
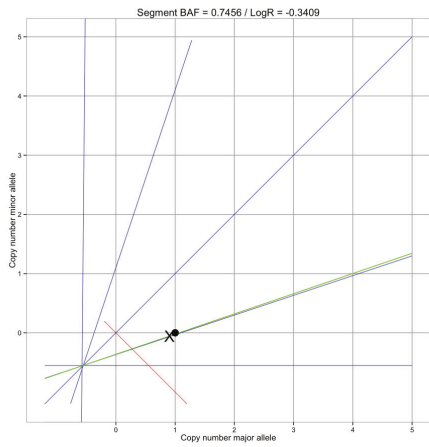
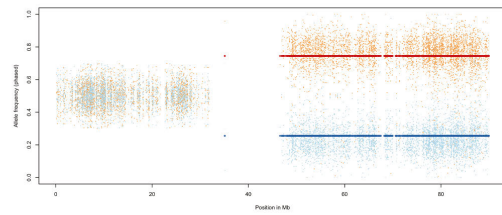
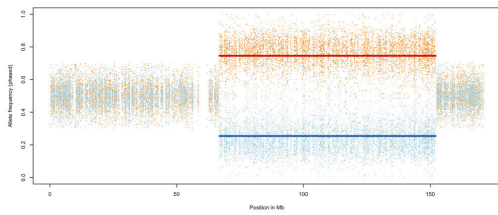
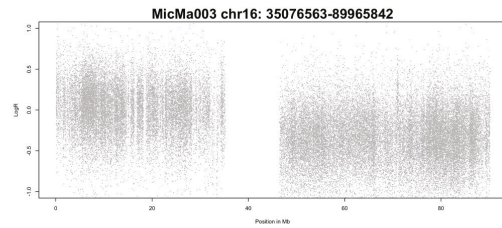
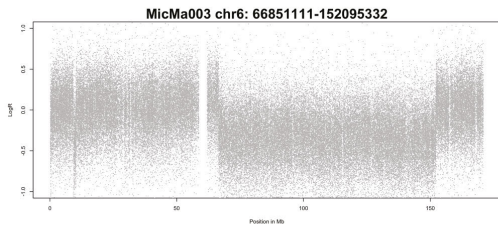
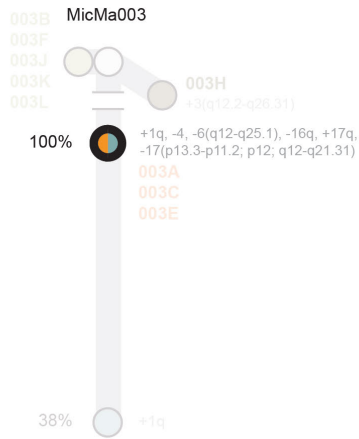
107U



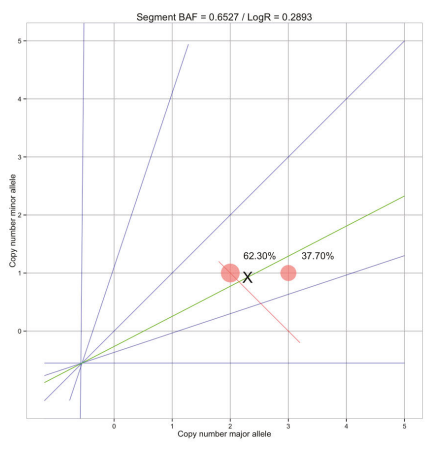
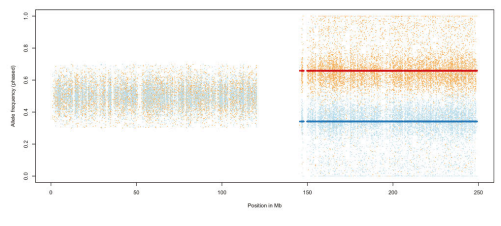
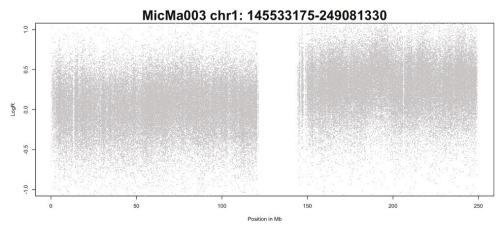
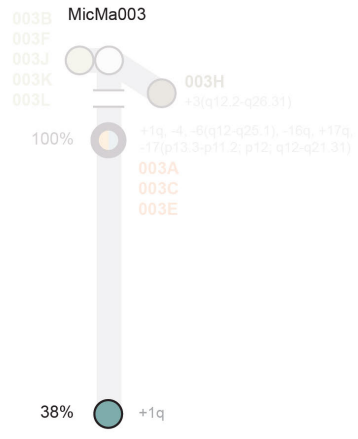
**Figure S9: Battenberg bulk tumor results for phylogenetically informative segments.** Each panel highlights a single node in the phylogenetic tree for the indicated patient (MicMa003, 083 or 107, top) and one or two evidencing segments as derived by Battenberg from the SNP6-array data from the primary tumor or lymph node metastasis (bottom). Battenberg plots represent the logR (top) and fitted segmented allele frequency for the relevant chromosome (middle), and the inferred (sub)clonal copy number state (bottom). The latter plot shows a grid of copy number states for the major ( $n_{Maj}$ ) and minor ( $n_{Min}$ ) alleles in the tumor. Blue lines are lines of equal BAF (0, 0.25, 0.5, 0.75 and 1) and red lines of equal logR in the copy number space. Using the estimated tumor purity ( $\rho$ ) and ploidy ( $\psi_t$ ) according to  $n_{Maj} = \frac{\rho - 1 + 2^{\frac{r}{\psi_t}} b (2(1-\rho) + \rho \psi_t)}{\rho}$  and  $n_{Min} = \frac{\rho - 1 + 2^{\frac{r}{\psi_t}} (1-b) (2(1-\rho) + \rho \psi_t)}{\rho}$ , the measured logR ( $r$ ) and B-allele frequency ( $b$ ) correspond to a single point (plotted as 'X') in this copy number space ([38], Van Loo et al, *Proc Natl Acad Sci U S A*, 2011). The distance from this point to a clonal solution (i.e. the closest grid point) is checked via a standard t-test (black dot if clonal). If  $p < 0.05$ , a subclonal solution (the two red points) is proposed along one side of the copy number square containing the 'X' ([3], Nik-Zainal et al, *Cell*, 2012). The iso-BAF line corresponding to the segment's BAF (green) crosses two edges of the square, one of which is chosen based on whether the segment's logR is smaller or larger than the square's diagonal of constant logR. The cancer cell fractions are determined by the point of intersection between this edge and the line of equal BAF and are reflected by the size of the red points. Panels detail evidence for the MRCA (A) or the 38% subclone (B) in MicMa003; for the MRCA (C), the 50% (D), the 30% (E) or the 21% (F) subclone in MicMa083; for the MRCA (G), the 88% (H) or other subclones (I) in the primary of MicMa107; and for the MRCA (J), the 89% (K), the 39% (L), the 25% (M), the 16% (N) or other subclones (O) in the lymph node metastasis of MicMa107.



A

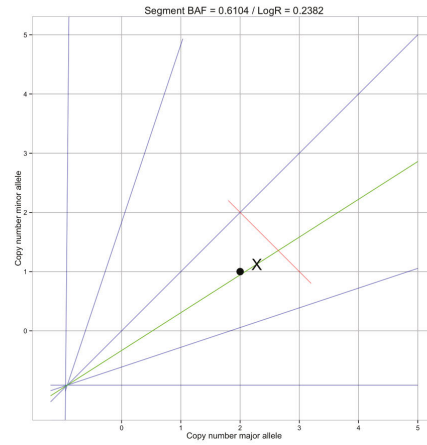
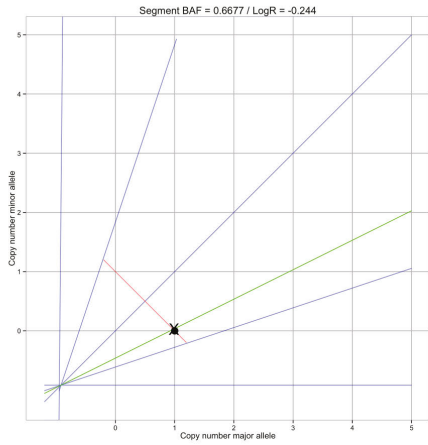
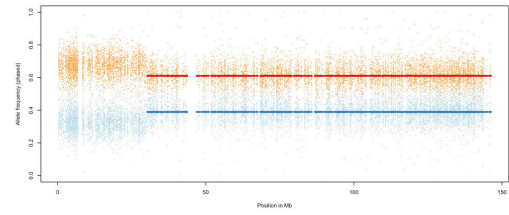
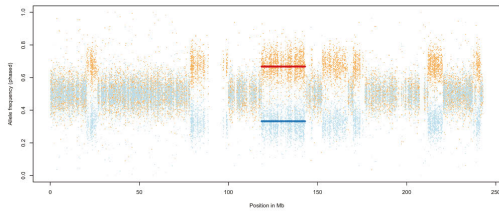
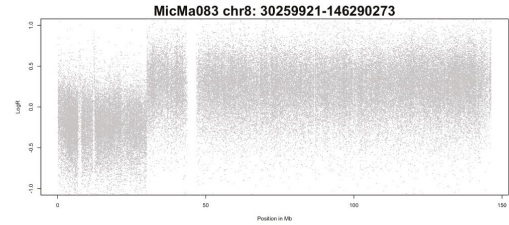
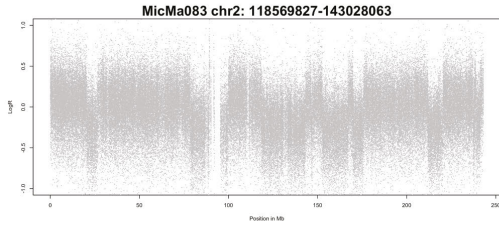
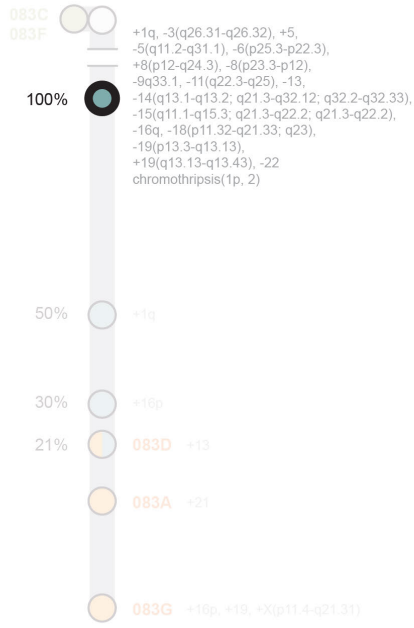


**B**



C

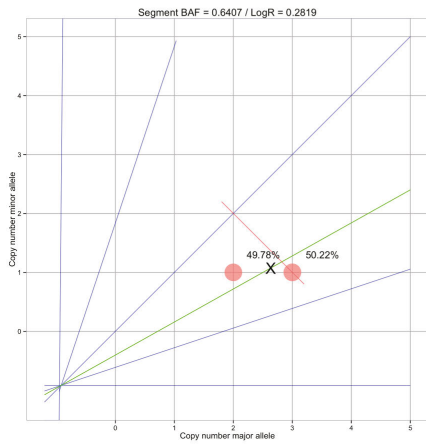
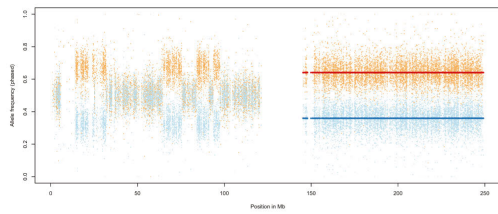
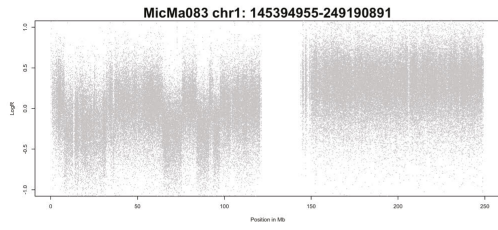
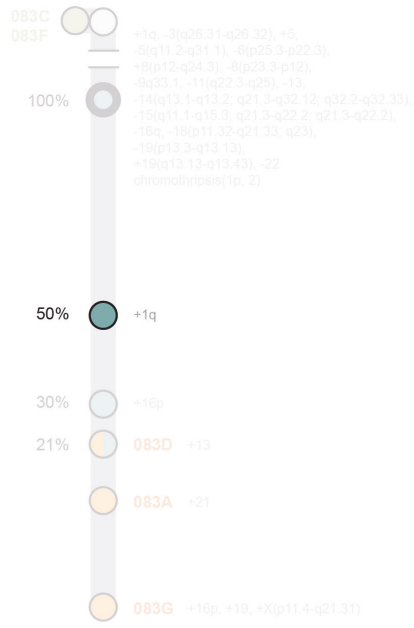
MicMa083





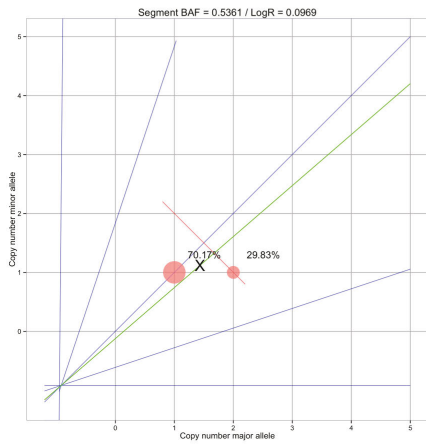
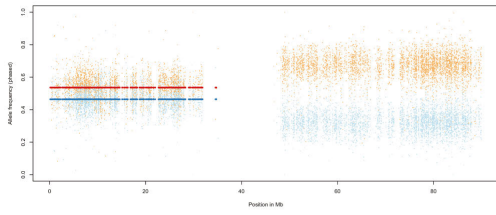
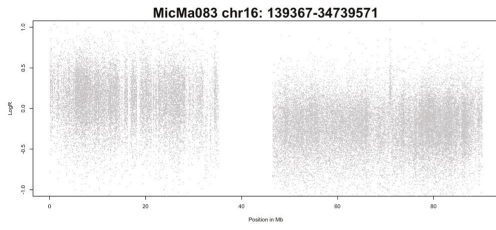
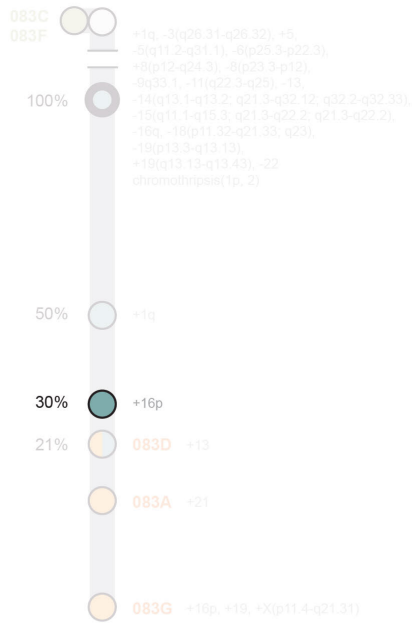
D

MicMa083



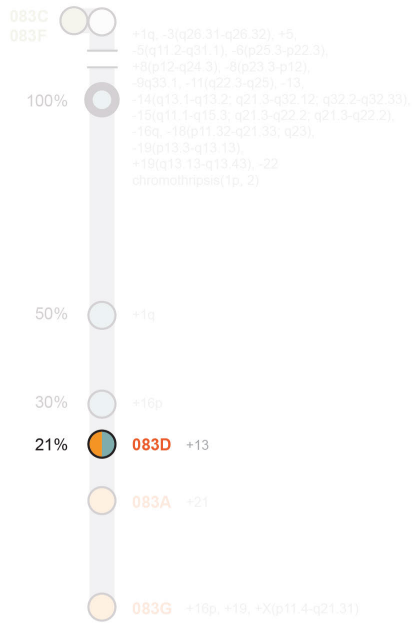
E

MicMa083

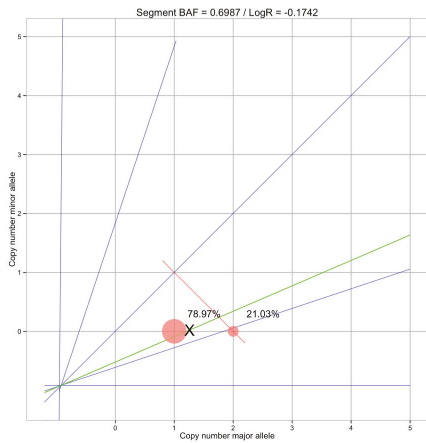
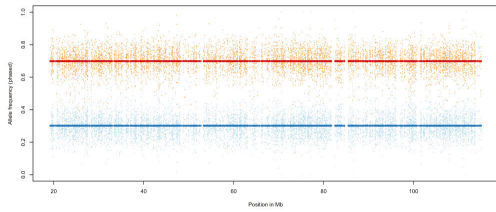
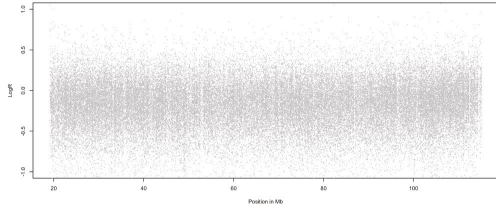


F

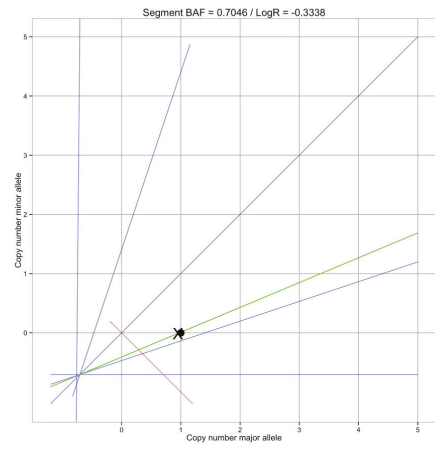
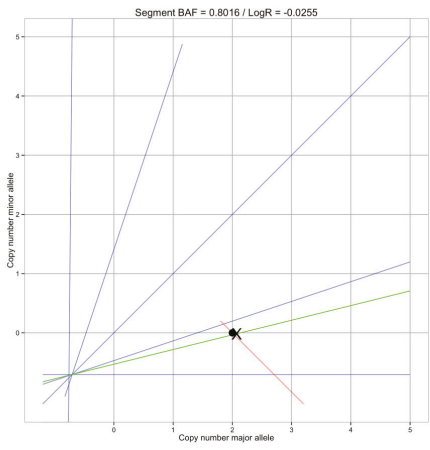
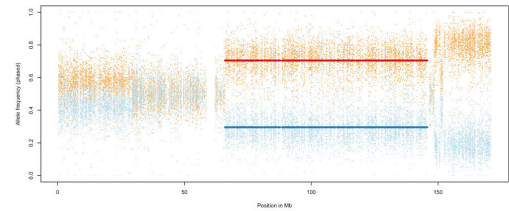
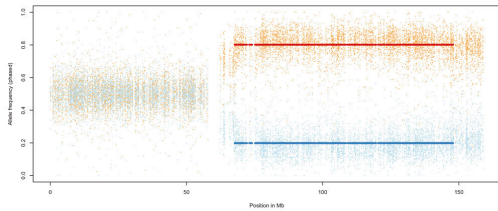
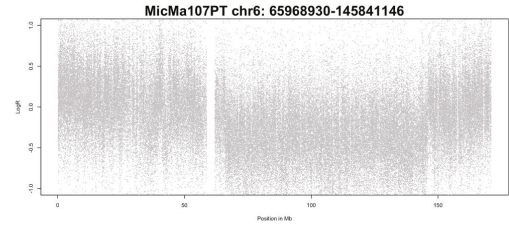
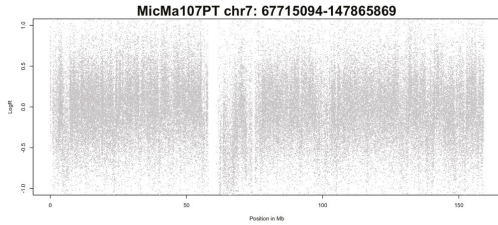
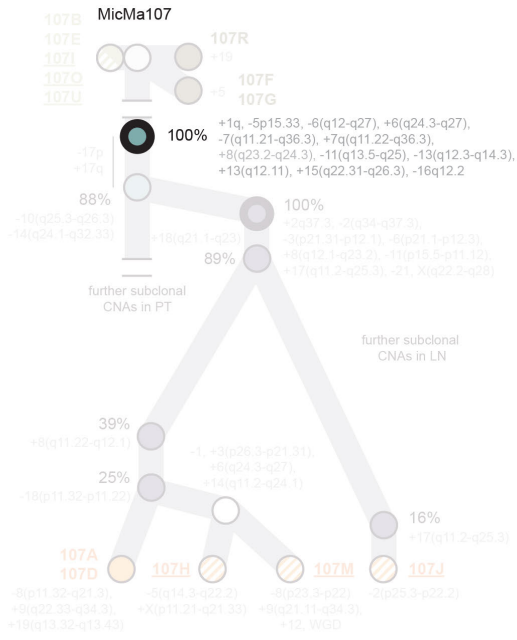
MicMa083



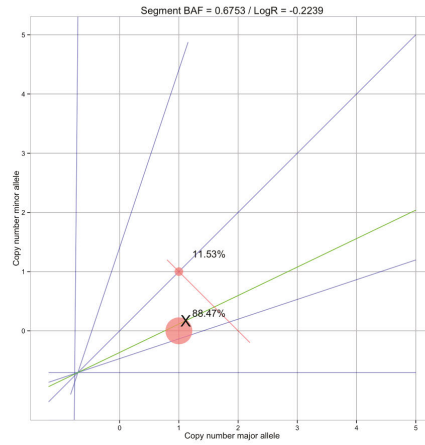
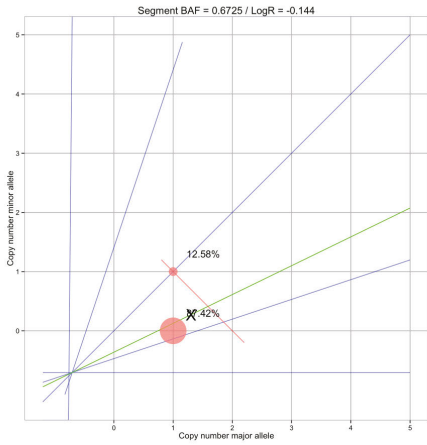
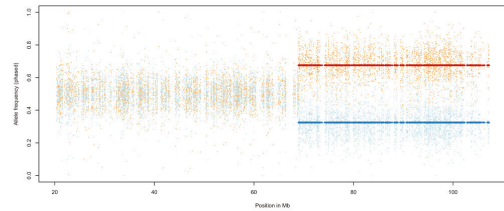
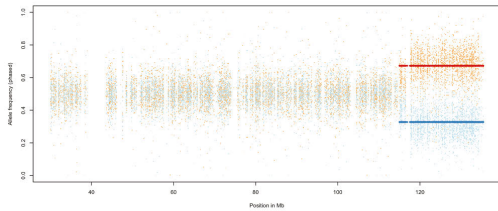
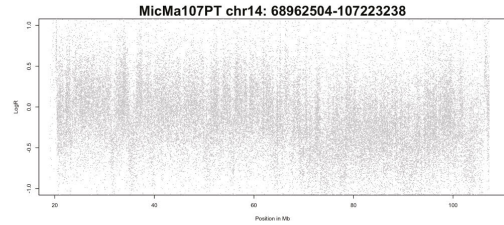
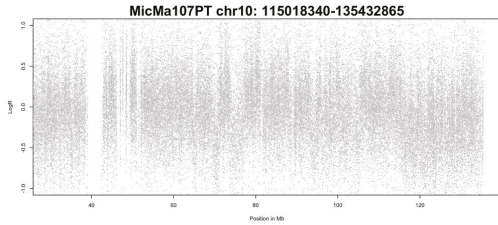
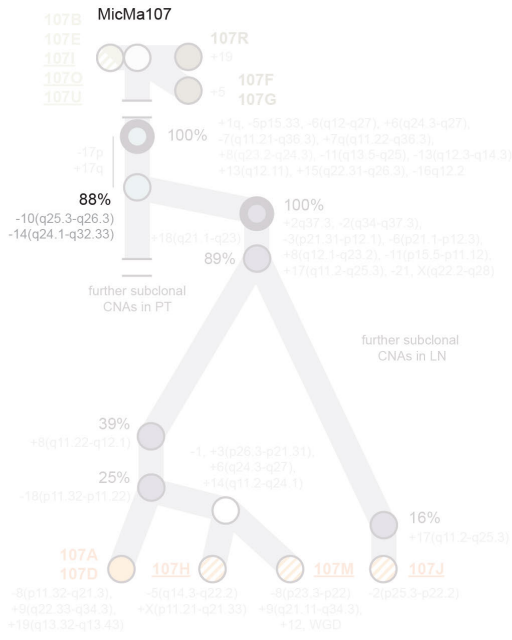
MicMa083 chr13: 19212262-114999838



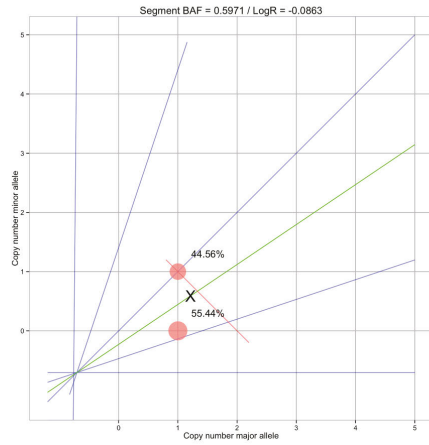
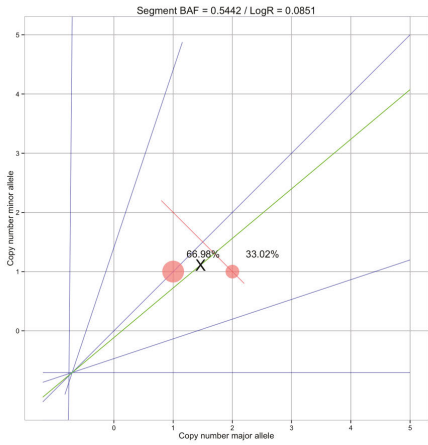
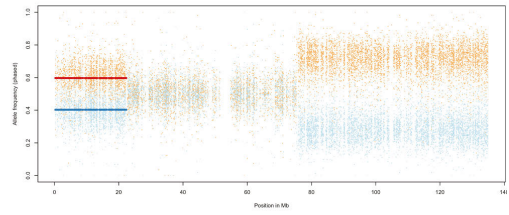
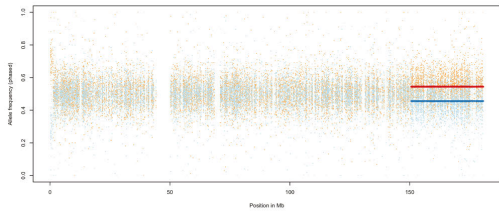
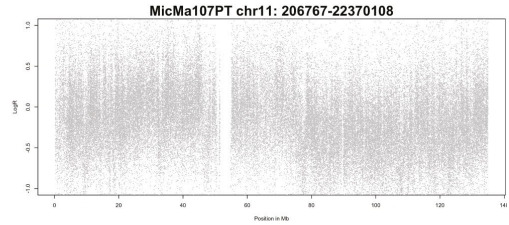
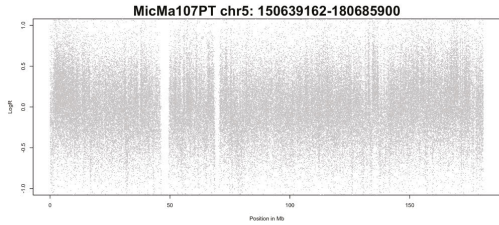
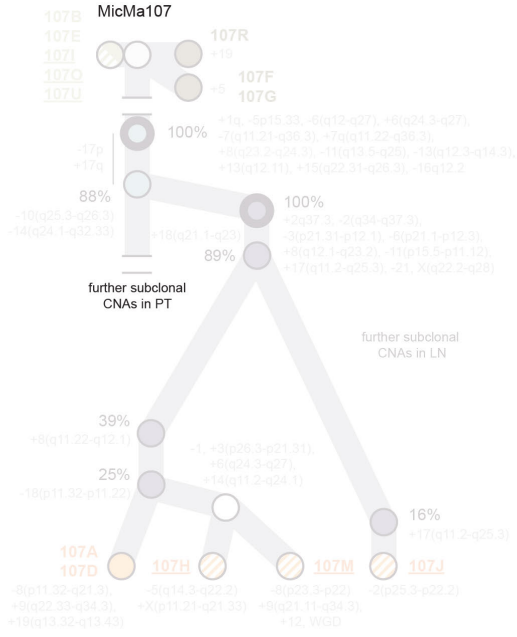
**G**



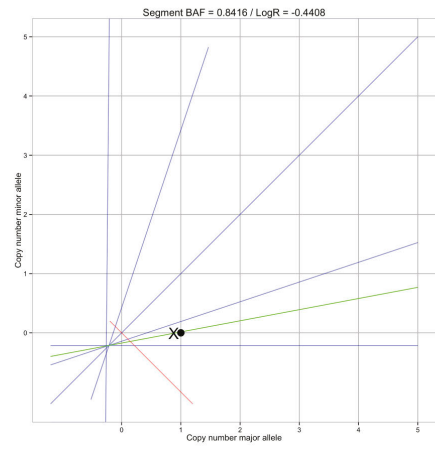
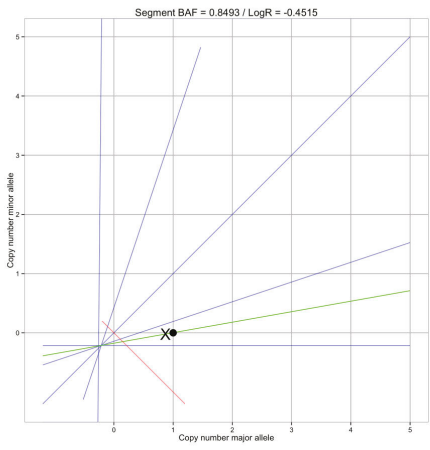
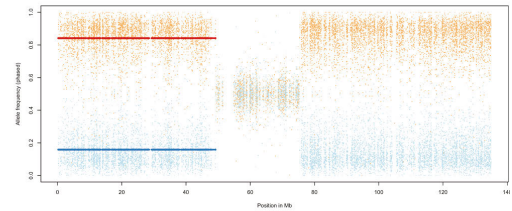
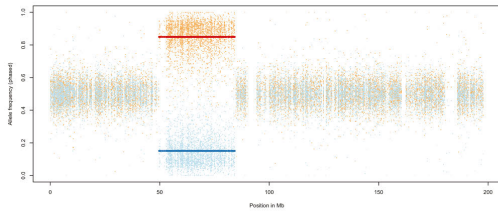
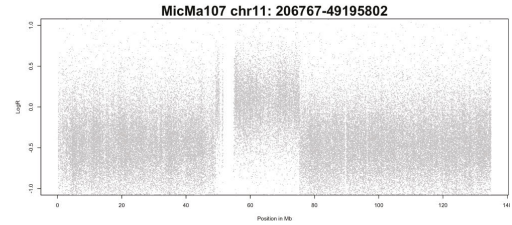
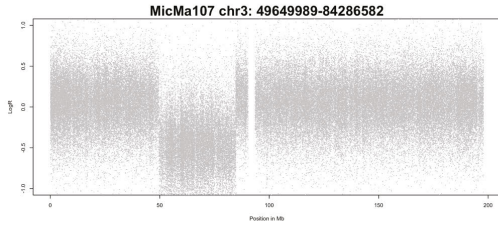
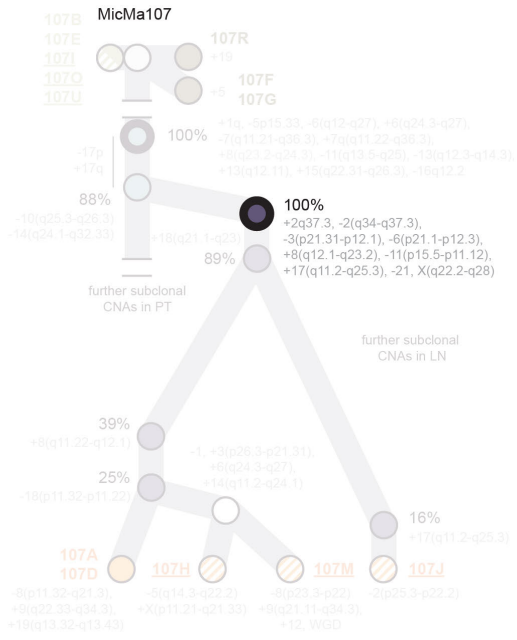
H



I

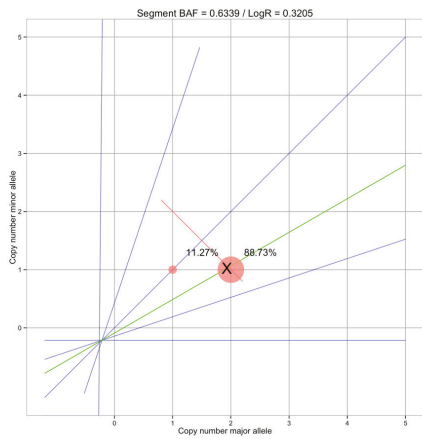
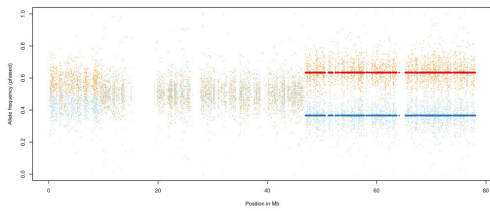
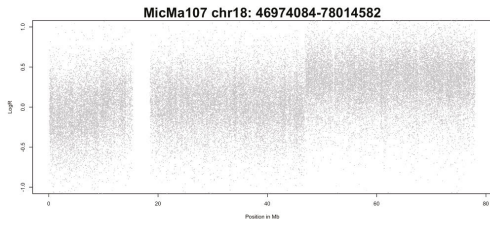
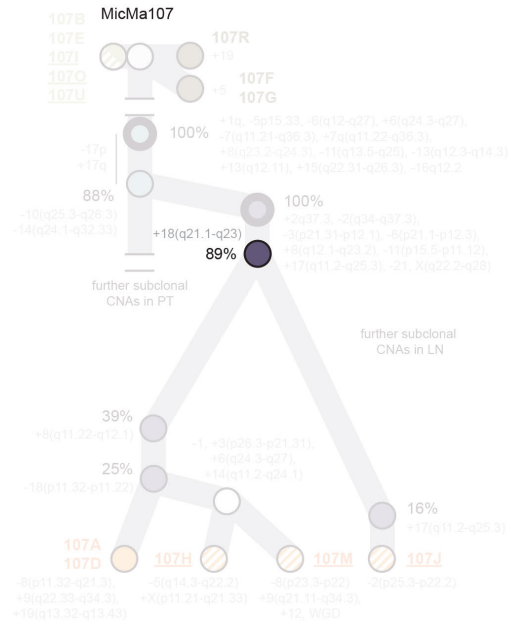


J

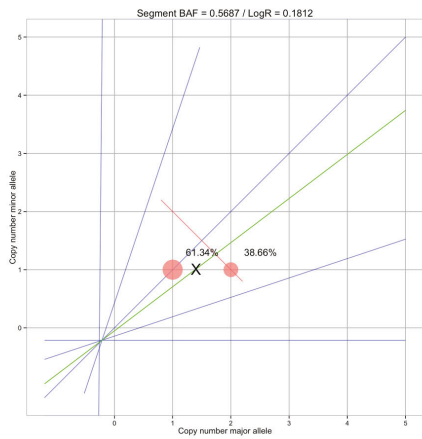
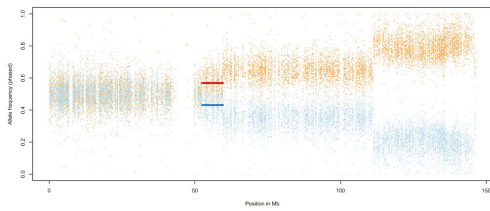
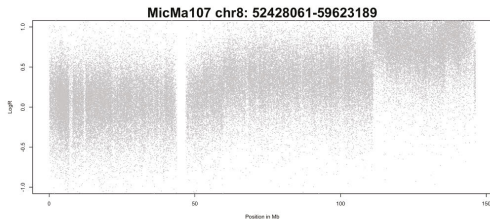
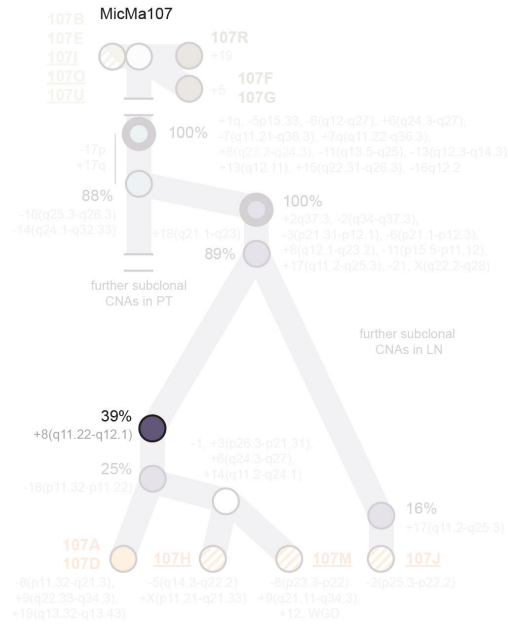




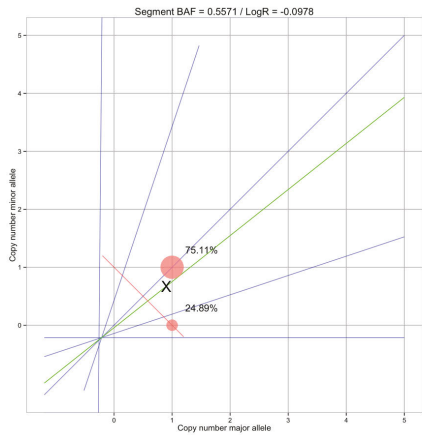
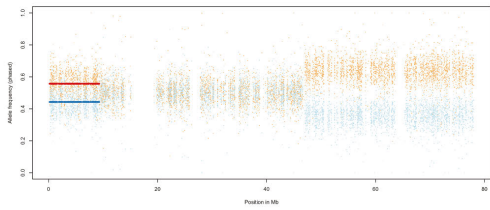
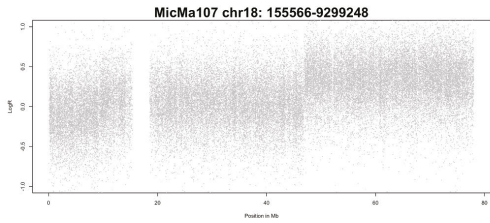
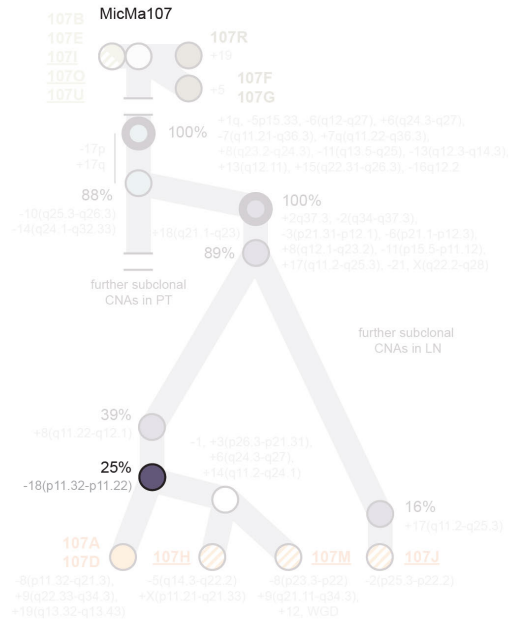
K



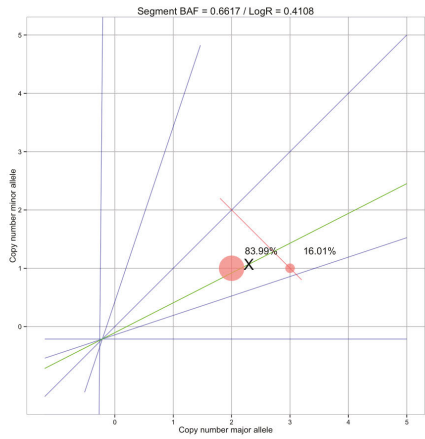
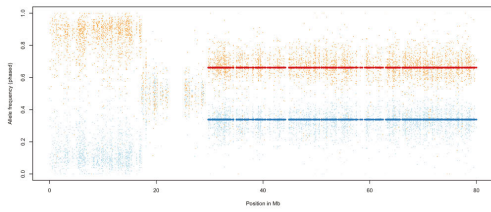
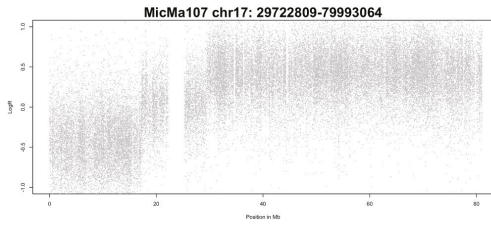
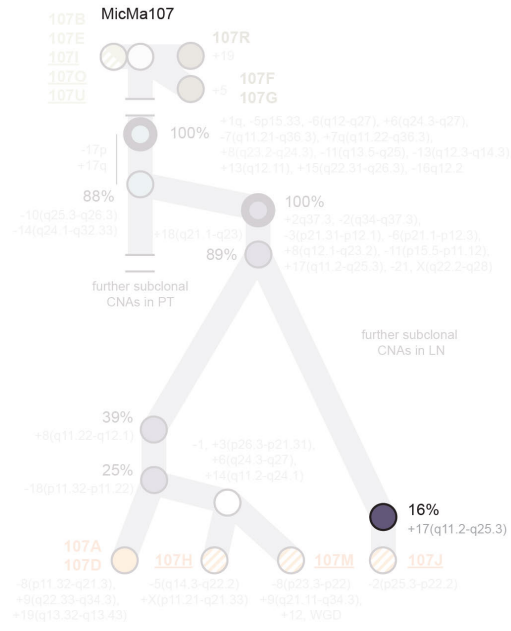
L



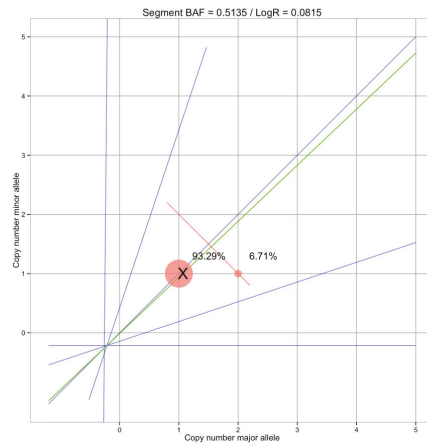
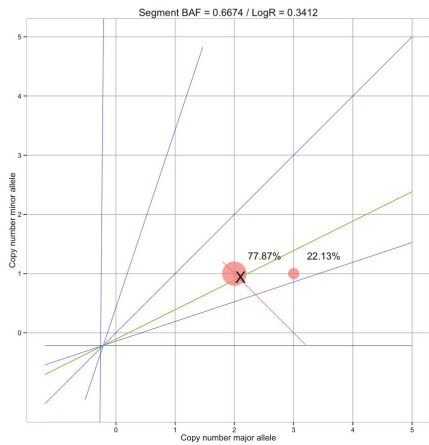
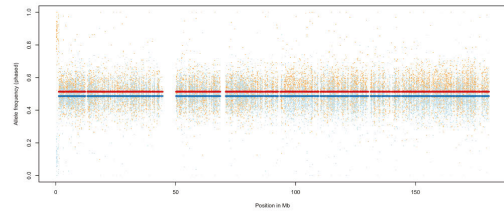
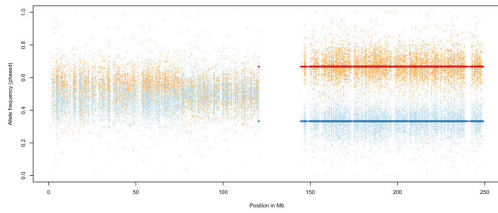
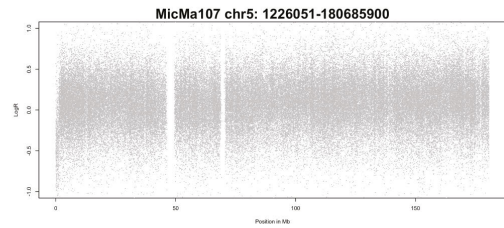
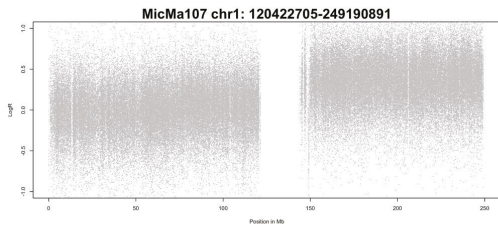
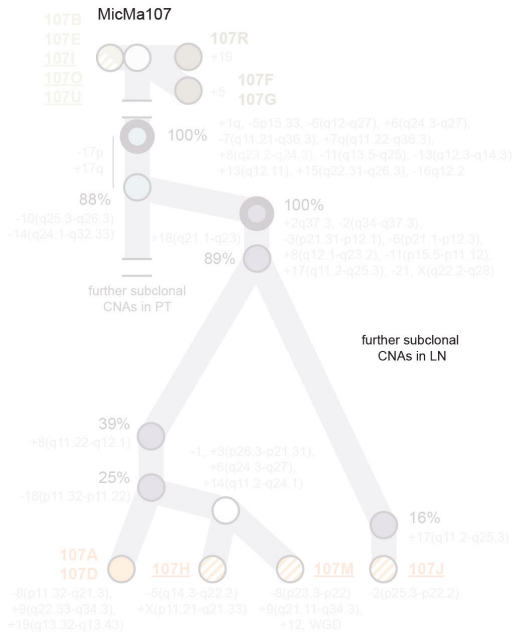
M



N



O



**Figure S10: Flowchart of bulk exome sequence analysis.** The steps schematically outline our workflow to detect somatic mutations in the bulk tumor exomes and filter out putative false positives.

