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Supplemental Information

IL-17 Receptor Signaling in the Lung Epithelium

Is Required for Mucosal Chemokine Gradients

and Pulmonary Host Defense against K. pneumoniae

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Supplementary Fig 1, related to Figure 1. Characterization of the IL17R flox mice in vitro. (A) Ear skin fibroblasts were derived from the *II17ra^{fl/fl}* mice and infected with adenovirus encoding CRE recombinase for 3 days. Cells treated with AdCRE or control virus were stimulated with TNF-a or the combination of TNF-a (1ng/ mL) and IL-17 (10ng/mL) for 24h. Gene expression was determined by real-time RT-PCR. (B) Homozygotes *II17ra^{fl/fl}* mice were sacrificed 5 days after receiving 10⁸ Ad-CMV or Ad-CRE though intravenous injection. PCR using primers detecting recombined allele in the spleens were performed. Bands with no Cre-mediated recombination and Post-Cre in the spleens were indicated by arrows. (C) Peripheral blood from the *II17ra^{fl/fl} E2a-Cre⁻* and *II17ra^{fl/fl} E2a-Cre⁺* mice were collected from the tails and stained with IL-17RA and Gr-1 antibodies after red blood cell lysis. 3 mice for each genotype were shown.

Supplementary Fig 2, related to Figure 1. Specific knock out of IL-17R in the flox mice. (A) Peripheral blood (PBL) from the *II17ra^{fl/fl} E2a-Cre⁻* and *II17ra^{fl/fl} E2a-Cre⁺* mice were collected from the tails and stained with IL-17RA and Gr-1 antibodies after red blood cell lysis. (B) Percentage of Gr-1+ cells in the PBL were graphed. (C) *II17rc^{fl/fl}* and *II17rc^{fl/fl} E2a-Cre⁺* mice were challenged with recombinant CXCL1 intranasally (1ug/ mouse) for 24h and ^{neutrophils} as well as macrophage numbers were determined by FACS.

Supplementary Fig 3, related to Figure 2. Scgb1a1-Cre mediated recombination.

(A) PCR using primers detecting recombined allele in the lungs from *ll17ra*^{fl/fl} *Scgb1a1-Cre*⁻ and *ll17ra*^{fl/fl} *Scgb1a1-Cre*⁺ mice were performed. Bands with no Cremediated recombination and Post-Cre were indicated by arrows. (B) *ll17ra* expression in the bronchial brushes from *ll17ra*^{fl/fl} *Scgb1a1-Cre*⁻ and *ll17ra*^{fl/fl} *Scgb1a1-Cre*⁺ mice were determined by real-time RT-PCR.

Supplementary Fig 4, related to Figure 4. Real-time RT-PCR validation of RNAseq findings. *Il17rc*^{fl/fl} *Scgb1a1-Cre*⁻ and *Il17rc*^{fl/fl} *Scgb1a1-Cre*⁺ mice were challenged with recombinant IL-17 intranasally (300ng/mouse) for 6h and bronchial brushings were harvested for RNA extraction and real-time RT-PCR analysis.

Supplementary Fig 5, related to Figure 5. Airway inflammation in CXCL5 rescued *II17ra^{fl/fl} x Scgb1a1-Cre*⁺ after *K. pneumoniae* infection. *II17ra^{fl/fl} Scgb1a1-Cre*⁻ and *II17ra^{fl/fl} Scgb1a1-Cre*⁺ mice were infected with 10⁴ KP43816 intranasally. 2h post infection, half of *II17ra^{fl/fl} Scgb1a1-Cre*⁺ the mice received 1ug recombinant CXCL5 (LIX). Mice were sacrificed 24h after infections. Airway inflammation was assessed by H&E staining.

Supplementary Fig 6, related to Figure 6. Role of IL-17RE in IL-17 mediated airway inflammation. RNAseq analysis was conducted on NHBE cells stimulated with recombinant human IL- 17 (100ng/mL) for 48h (N=3-4). Heat map of the expression for IL-17 family cytokines and IL-17 receptor family were shown (A). *II17c* expression in naïve unchallenged B6, WT or *II17re+/-* as well as littermates *II17re-/-* mice 24h post intranasal IL-17 challenge, and B6 mice 24h post KP infection were determined by real-time RT-PCR (B). BAL cell infiltration from control littermates and *II17re-/-* mice 24h post intranasal IL-17 challenge were mesearued by FACS (C). Gene expression of IL-17 downstream chemokines and cytokines were determined by real-time RT-PCR (D).

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