

The high osmotic response and cell wall integrity pathways cooperate to regulate morphology, microsclerotia development, and virulence in *Metarhizium rileyi*

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Table S1 The primers designed for genes analysis

Genes	Name of primer	Sequences (5'-3')
primers used for cDNA and genomic DNA sequences	<i>Mrhog1-F</i>	CATGGGAGCCTTCGGTCTGGTTT
	<i>Mrhog1-R</i>	TGCGCGAGATCAACTCACAAACC
	<i>Mrhog1-gF</i>	CATGGGAGCCTTCGGTCTGGTTT
	<i>Mrhog1-gR</i>	GTGCCTGTCGTATGCCGTCGTTG
	<i>Mrslt2-F</i>	ATGGCCGACCTTCACCTCA
	<i>Mrslt2-R</i>	CTCCTGGAAGCATCTAGGCCGG
	<i>Mrslt2-gF</i>	TTATCTCCTGGAAGCATCTA
	<i>Mrslt2-gR</i>	ATGGCCGACCTTCACCTCA
universal primers used for FPNI-PCR	FP1	GTAATACGACTCACTATAGGGCACGCGTGGTNTCGASTWTSGWGTT
	FP2	GTAATACGACTCACTATAGGGCACGCGTGGTNGTCGASWGANAWGAA
	FP3	GTAATACGACTCACTATAGGGCACGCGTGGTWGTGNAGWANCANAGA
	FP4	GTAATACGACTCACTATAGGGCACGCGTGGTAGWGNAGWANCAWAGG
	FP5	GTAATACGACTCACTATAGGGCACGCGTGGTNGTAWAASGTNTSCAA
	FP6	GTAATACGACTCACTATAGGGCACGCGTGGTNGACGASWGANAWGAC
	FP7	GTAATACGACTCACTATAGGGCACGCGTGGTNGACGASWGANAWGAA
	FP8	GTAATACGACTCACTATAGGGCACGCGTGGTGTNCGASWCANAWGTT
	FP9	GTAATACGACTCACTATAGGGCACGCGTGGTNCAGCTWSCTNTSCTT
	FSP1	GTAATACGACTCACTATAGGGC
	FSP2	ACTATAGGGCACGCGTGGT
special primers used for FPNI-PCR	<i>Mrhog1-F1</i>	TGTGCTTCCGACAGACAGCCTCCTT
	<i>Mrhog1-F2</i>	GTCGGTGCAGATGGAGATGGACGAG
	<i>Mrhog1-F3</i>	TGACTCGTCGTATCCGCAGAGCAAG
	<i>Mrhog1-R1</i>	TCGGTTCACGACAATGCCTACAGCG
	<i>Mrhog1-R2</i>	GGCAATGTATGTAGGAGTGCGGGGA

	<i>Mrhog1-R3</i>	TCTTGCGTTTTGGGAGACTTGGTG	
	<i>Mrslt2-F1</i>	ACCTTGAAAACCTTTCGTCCCTGAAG	
	<i>Mrslt2-F2</i>	GAGAGCCGAACGCAGTAGCTTGAATC	
	<i>Mrslt2-F3</i>	AGAAGAATGGGTCTGGGTATTTTATGG	
	<i>Mrslt2-R1</i>	ATTGTCACCTTGCGGGAAGGACGGTA	
	<i>Mrslt2-R2</i>	TCCGTAATACCCAGCACGGATAAGAC	
	<i>Mrslt2-R3</i>	AGAAGCCCTTTATGCCCAATGTCTG	
primers used for flanking sequences	HLF1	cgGAATTC <u>CACTTGGGATTTTCTGAACGCACATGG</u> EcoRI site is underlined	
	HLR1	ccgCTC <u>GAGAAATGGCGGGATGGCTGGGTTT</u> GAGA XhoI site is underlined	
	HRF1	gcTCTAG <u>AAGACAACGACGGCATA</u> CGACAGGCAC XbaI site is underlined	
	HRR1	cccAAGCTTTGTAGGGCGAACAGGCTCGGGCAAAA HindIII site is underlined	
	SLF1	cgGAATTCGAGGGCGCTTAAGGTTGACGGGTTC EcoRI site is underlined	
	SLR1	ccgCTC <u>GAGCGAGAAGAATGGGTCTGGTATTTT</u> ATGG XhoI site is underlined	
	SRF1	gcTCTAG <u>AGAAGGTAGGGAGGGACGGATGGAAAG</u> XbaI site is underlined	
	SRR1	cccAAGCTTAAACACACCCCTTTTCTCTCGCCCC HindIII site is underlined	
	primers used for PCR screening transformants	HF	GCTGTTTACTTCCGCCATCCATCCCT
		HR	GAGTTTCGAGACGTGACGATCCGCTA
Ho-OF		CCAGATATTCGACCTTCAGCCTGTG	
Ho-OR		CCAGCCTCTACATTATGGTAGTCAA	
SF		TGGCGAGAAGTGAAAACGCTGGTAT	

	SR	GGGCGCAGGTGCAGGTTCTTGAA
	SI-OF	CTCGGACAGGGTGCCTATGGTATTG
	SI-OR	CGCATCTCCCCGACATCGTCAATGA
	<i>hph</i> -F	GCTCTCGCTAAACTCCCAATGTCA
	<i>hph</i> -R	CATTGACTGGAGCGAGGCGATGTTC
primers used for	HosF	GTTGTCGTTTCTGTTTCATCCCATCC
Southern blotting	HosR	TTGACTCGTCGTATCCGCAGAGC
	SlsF	GCTCTGAGGTCTGTAGAAAAGTG
	SlsR	AGAATGGGTCGGGTATTTTATGGC
primers used for	<i>Mrhog1-qF</i>	GCATCGTGAAGCCTA
RT-qPCR analysis	<i>Mrhog1 -qR</i>	GGGCTCGGTAATATCGTGTT
	<i>Mrslt2-qF</i>	TGTGCGGCCTCAAGTATATC
	<i>Mrslt2-qR</i>	CGAGGCCAAAGTCACAGAT
	<i>Mrpbs2-qF</i>	GAGGTCCACATACTGGACGA
	<i>Mrpbs2-qR</i>	CTTGGCATGCTTGACTTTGT
	<i>Mrmsn2,4-qF</i>	AGAGGACCCTCCAAGACCT
	<i>Mrmsn2,4-qR</i>	GCTTCAGGTGTTCTTGACGA
	<i>Mrmkk1,2-qF</i>	GCCTCCGGTGTATAGCTTGT
	<i>Mrmkk1,2-qR</i>	GTGATCCTACACGAGTGCGT
	<i>Mrswi4,6-qF</i>	CCACCCAGATTCTCAAGGTT
	<i>Mrswi4,6-qR</i>	TGCTCGCCTGTTTGTATCTC
	<i>Mrtef-qF</i>	GTCATCGTCCTCAACCATC
	<i>Mrtef-qR</i>	CAGTCTCAACAGCCTTACC
	<i>Mrtub-qF</i>	GGCAAGGTCGCTATGAAG
	<i>Mrtub-qR</i>	CTGGATGGAGGTAGAGTTAC
	<i>MrpksP-qF</i>	AGGCCTATAAAGGCATGCAA

*MrpksP-qR*

GCAACTGTCCACCCACATAG

*Mrchs1- qF*

CATCCGTCAACACCAAAGAC

*Mrchs1- qR*

GCTGAATAAGGCGACCTCTC

*Mrchs2- qF*

AATCGGCGACAATTTCTACC

*Mrchs2- qR*

ATTCGTATCCTGCCTTCCAC

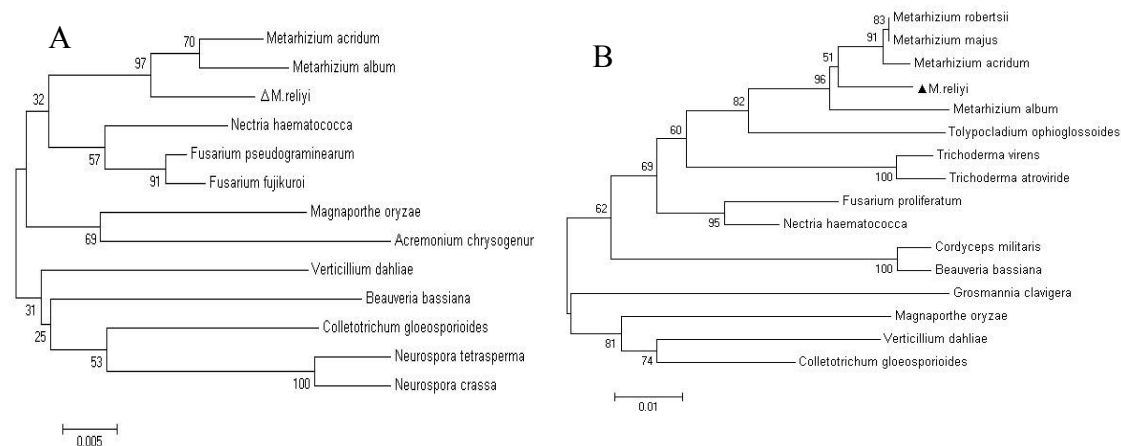
*Mrchs4- qF*

GTCAATGAGCTCGAAAGTCG

*Mrchs4- qR*

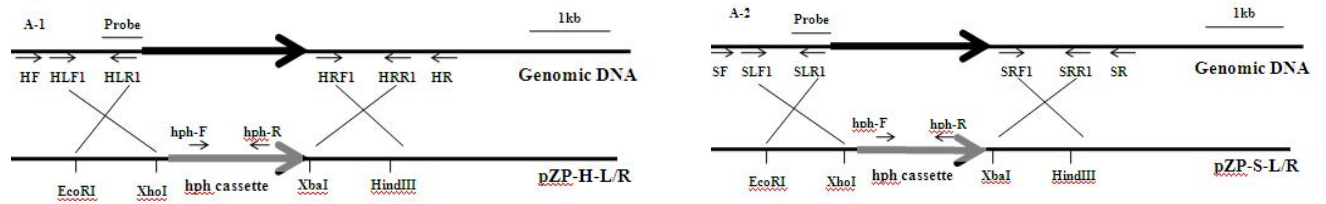
TCGACGTCATCTACAAAGGC

**Fig. S1** Phylogenetic tree inferred from Mrhoglp (A) and Mrs1t2p (B) protein sequences alignments



The numbers on the nodes represent the results of bootstrap analyses (1000 replicates) carried out using the neighbor-joining method. The aligned sequences of Mrhoglp protein are from *Acremonium chrysogenum* ATCC 11550 (XP KFH47112.1); *Neurospora tetrasperma* FGSC 2508 (XP 009849980.1); *Neurospora crassa* OR74A (XP 962163.2); *Verticillium dahliae* VdLs.17 (XP 009655080.1); *Colletotrichum gloeosporioides* Nara gc5 (XP 007280298.1); *Fusarium fujikuroi* IMI 58289 (XP CCT64307.1); *Beauveria bassiana* ARSEF 2860 (XP 008598528.1); *Fusarium pseudograminearum* CS3096 (XP 009257285.1); *Nectria haematococca* mpVI 77-13-4 (XP 003046679.1); *Magnaporthe oryzae* 70-15 (XP 003714838.1); *Metarhizium acridum* CQMa 102 (XP 007814424.1); *Metarhizium album* ARSEF (XP KHO01547.1). The aligned sequences of Mrs1t2p protein are from *Metarhizium robertsii* ARSEF 23 (XP 007819370.2); *Metarhizium majus* ARSEF 297 (XP 014574977.1); *Metarhizium acridum* CQMa102 (XP 007814364.1); *Metarhizium album* ARSEF 1941 (KHN93898.1); *Tolypocladium ophioglossoides* CBS 100239 (KDB88410.1); *Trichoderma virens* Gv29-8 (XP 013959852.1); *Trichoderma atroviride* IMI 206040 (XP 013937734.1); *Fusarium proliferatum* (ABD67163.1); *Nectria haematococca* mpVI 77-13-4 (XP 003050911.1); *Cordyceps militaris* CM01 (XP 006674833.1); *Grosmannia clavigera* kw1407 (XP 006674015.1); *Beauveria bassiana* ARSEF 2860 (XP 008596653.1); *Magnaporthe oryzae* 70-15 (XP 003712437.1); *Verticillium dahliae* VdLs.17 (XP 009651532.1); *Colletotrichum gloeosporioides* Nara gc5 (XP 007285039.1).

Fig. S2 The disruption of *Mrhog1* and *Mrslt2* gene

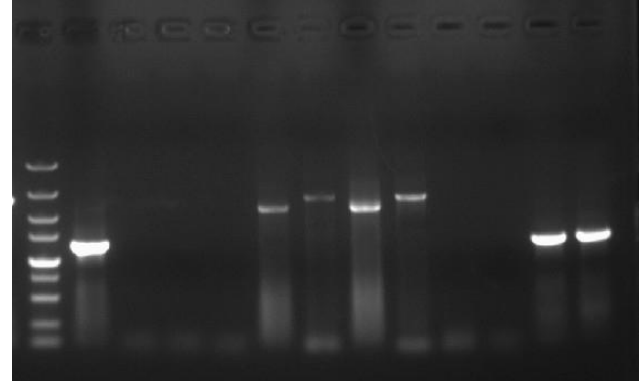
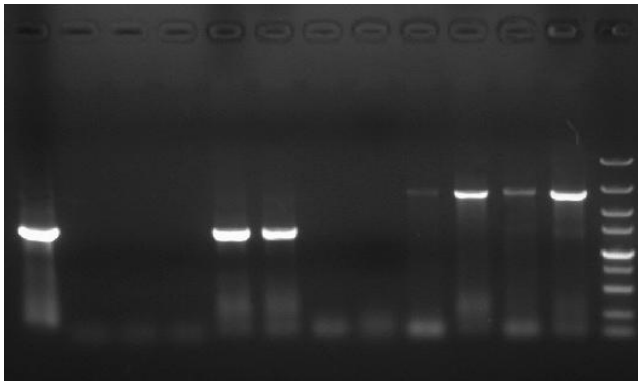


B-1

B-2

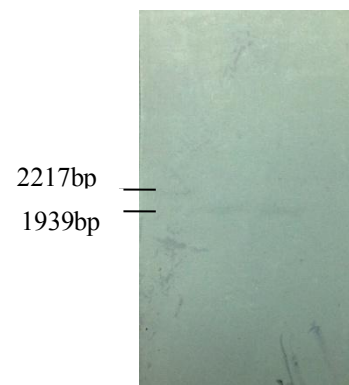
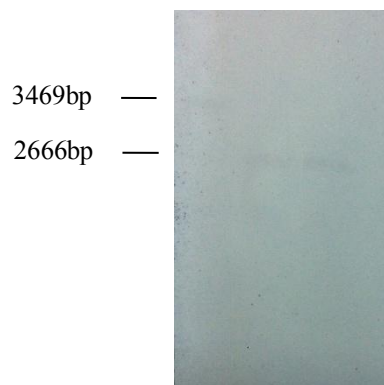
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M 1 2 3 4 5 6 7 8 9 10 11 12



C-1

C-2

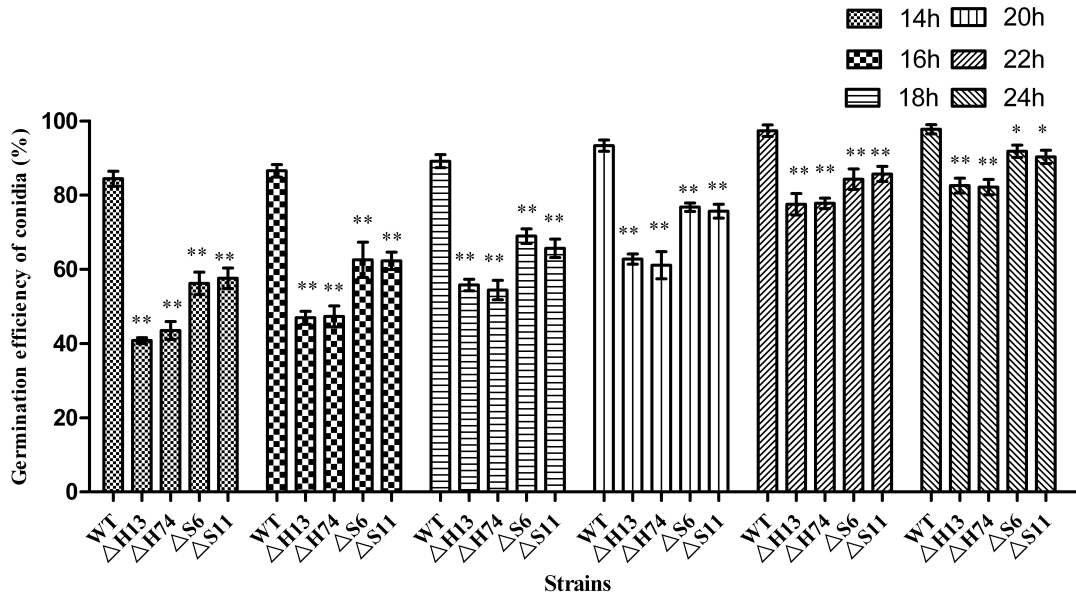


(A) The construct of *Mrhog1* (A-1) and *Mrslt2* (A-2) loci, disruption constructs with *hph* cassette and flanking sequences. Arrows indicate the positions of primers used for PCR screening of deleted mutants and generation of hybridization probes. (B) PCR characterization of the four disrupted mutants. (B-1) PCR characterization of  $\Delta MrHog1$  strains. PCR screen the ORF (Lane 1-3: 1-wild-type, 2- $\Delta H13$ , 3- $\Delta H74$ ); PCR screen the *hph* (Lane 4-6: 4- wild-type, 5- $\Delta H13$ , 6- $\Delta H74$ ); PCR screen the

*hph* and genomic sequence outside the flank regions (Lane 7-12: 7-LF-WT, 8-RF-WT, 9-LF- $\Delta H13$ , 10-RF- $\Delta H13$ , 11-LF- $\Delta H74$ , 12-RF- $\Delta H74$ ); (B-2) PCR characterization of  $\Delta MrSlt2$  strains. PCR screen the ORF (Lane 1-3: 1-WT, 2- $\Delta S6$ , 3- $\Delta S11$ ); PCR screen the *hph* and genomic sequence outside the flank regions (Lane 4-9: 4-LF-WT, 9-RF-WT, 5-LF- $\Delta S6$ , 6-RF- $\Delta S6$ , 7-LF- $\Delta S11$ , 8-RF- $\Delta S11$ ); PCR screen the *hph* (Lane 10-12: 10- wild-type, 11- $\Delta S6$ , 12- $\Delta S11$ ). M, DNA molecular size markers (DL 5000). (C) Southern blotting analysis of  $\Delta MrHog1$  (C-1) and  $\Delta MrSlt2$  (C-2) mutants. Genomic DNA was digested with BamHI/SaII and HindIII/XbaI in  $\Delta Mrhog1$ ,  $\Delta Mrslt2$  and WT isolates. The PCR product obtained with primers HosF/HosR and SlsF/SlsR was used as the probe.

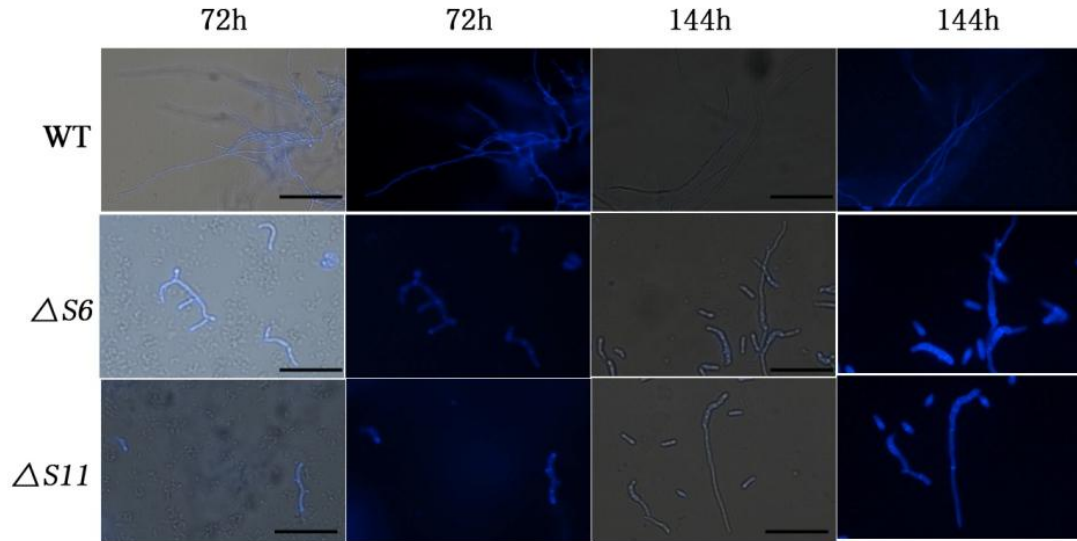


Fig. S3 Germination of conidia from WT,  $\Delta MrHog1$ , and  $\Delta MrSlr2$  isolates



The percentage of conidia germination was determined under a light microscope. Conidium was scored as germinated if the germ-tube length was half in size to that of the conidium. Values were the mean of three independent experiments. Standard error bars indicate variation in measurements. \*  $P < 0.05$ , \*\*  $P < 0.01$ , when compared with the results observed at WT.

**Fig. S4** The abnormal hyphal morphology of  $\Delta Mrsl2$  mutants



All strains were stained with Calcofluor for chitin distribution and fluorescence was mainly distributed on the apex of hyphae and septa. Mycelial morphology was determined under a microscope after the  $\Delta Mrsl2$  mutants and WT strains were grown in AM cultures for 3 days. Scale bar: 50  $\mu\text{m}$ .