

Supporting Information

Article title: Associative nitrogen fixation in nodules of the conifer *Lepidothamnus fonkii* (*Podocarpaceae*) inhabiting ombrotrophic bogs in southern Patagonia

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The following Supporting Information is available for this article:

Fig. S1 Relative abundances of 16S rRNA derived, classified operational taxonomic units (OTUs) in amplicon Illumina libraries retrieved from live roots of *Lepidothamnus fonkii* and peat material.

Table S1 Peat characteristics of the sampling sites Seno Skyring (SKY) and Seno Obstruccion (OBS)

Table S2 ^{15}N natural abundance of plant and peat reference material used for ^{15}N uptake calculations after conversion into ^{15}N atom %.

Table S3 Diversity of *nifH* genes and transcripts recovered from live roots of *Lepidothamnus fonkii* and peat material.

Table S4 Relative abundance of *nifH* gene and transcript (mRNA) defined OTUs retrieved from live roots of *Lepidothamnus fonkii* and peat material.

Fig. S1 Relative abundances of 16S rRNA derived, classified operational taxonomic units (OTUs) in amplicon Illumina libraries retrieved from live roots of *Lepidothamnus fonkii* and peat material. Approximately 13.000 quality-filtered reads were obtained per amplicon library. OTUs with a relative abundance of > 1% in one of the libraries are presented. *Beijerinckiaceae* include nitrogen fixing genera and are associated with *L. fonkii* roots rather than with peat material.

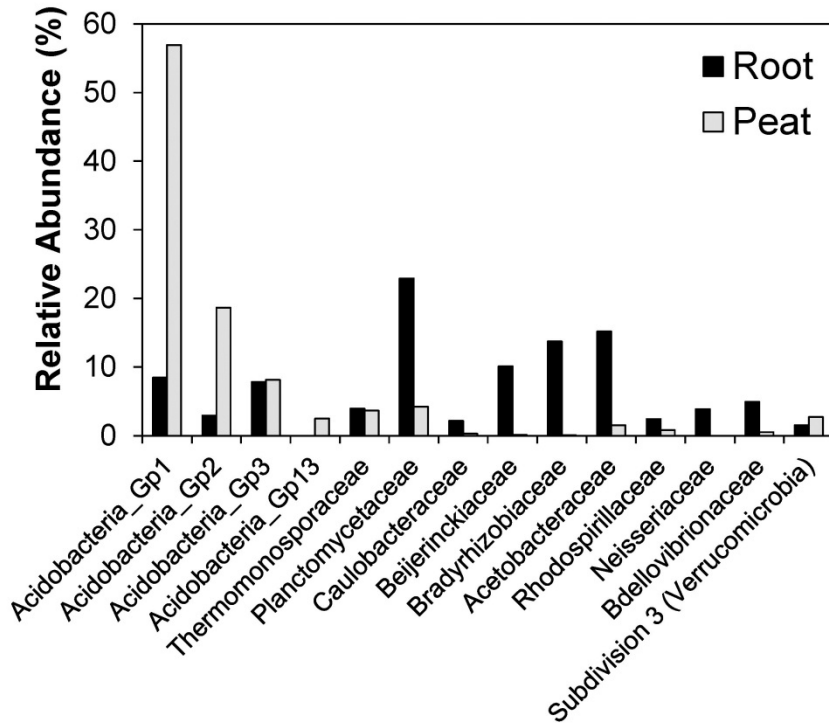


Table S1 Peat characteristics of the sampling sites Seno Skyring (SKY) and Seno Obstruccion (OBS): Content of nitrogen, molar ratios of carbon to nitrogen and ^{15}N isotopic signatures of bulk peat for the upper 80 cm.

Depth (cm)	OBS			SKY		
	N (%) by mass	Molar C/N ratio	$\delta^{15}\text{N}$ vs. air (‰)	N (%) by mass	Molar C/N ratio	$\delta^{15}\text{N}$ vs. air (‰)
0 - 10	0.69	93.9	-5.3	0.72	68.4	-4.4
10 - 20	0.70	92.0	-2.8	1.74	33.6	0.3
20 - 30	1.44	45.6	0.7	2.03	27.9	2.5
30 - 40	1.67	38.0	1.7	2.16	27.0	2.8
40 - 50	1.66	36.2	3.9	1.85	30.1	2.4
50 - 60	1.96	32.3	3.6	0.89	35.4	1.3
60 - 70	1.36	44.8	2.1	1.14	42.0	0.4
70 - 80	1.72	37.7	1.0	1.31	46.0	0.3

Table S2 ^{15}N natural abundance of plant and peat reference material used for ^{15}N uptake calculations after conversion into ^{15}N atom %. All reference plants and peat samples were obtained in close vicinity and similar microforms as of samples used for incubations. Peat samples were obtained as a representative sample from 0-10 cm depth.

	OBS		SKY	
	$\delta^{15}\text{N}$ (‰)	n	$\delta^{15}\text{N}$ (‰)	n
<i>L. fonkii</i>				
Leafs	-8.12 ± 0.83	3	-6.99 ± 0.77	3
Stems	-2.96 ± 1.44	3	-1.04 ± 0.75	3
Roots	-4.56 ± 0.95	3	-2.93 ± 0.65	3
Peat				
Root free bulk	-1.69 ± 0.21	6	$+0.22 \pm 1.71$	6
<i>T. repens</i>				
Roots	-0.73 ± 0.45	3		

Table S3 Diversity of *nifH* genes and transcripts recovered from live roots of *Lepidothamnus* sp. and peat material.

Sample	No. of sequences	Good's coverage (%) [*]	No. of OTUs observed	No. of OTUs estimated [†]	H [‡]	E [§]
Root						
Genes	20	86	7	8	1.1	0.7
Transcripts	76	99	3	3	0.3	0.3
Peat						
Genes	90	93	18	26	1.5	0.6
Transcripts	78	94	19	20	1.4	0.6

^{*}Percent library coverage $C = (1 - n_s/n_t) * 100$ (n_s = OTUs that occur only once, n_t = total number of sequences)

[†]Chao1 richness estimate with upper and lower 95% confidence intervals given in parentheses.

[‡]Shannon diversity index.

[§]Species Evenness.

Table S4. Diversity of *nifH* genes and transcripts recovered from live roots of *Lepidothamnus fonkii* sp. and peat material.

Phylogenetic affiliation of close relative (taxonomically validated)	OTU	NifH of close taxonomically validated relative (Accession number)	I^* (%)	NifH with highest I (Accession number)	I (%)	Relative abundance (%)				
						Root		Peat		
						Genes	mRNA	Genes	mRNA	
<i>Proteobacteria</i>										
<i>Alphaproteobacteria</i>										
<i>Rhizobiales</i>										
<i>Biejerinckiaceae</i>	3	<i>Beijerinckia indica</i> subsp. <i>indica</i> ATCC 9039 (AJ563938)	97	<i>Beijerinckia indica</i> subsp. <i>indica</i> ATCC 9039 (AJ563938)	97	55.0	93.5	0.0	1.3	
<i>Bradyrhizobiaceae</i>	7	<i>Bradyrhizobium elkanii</i> strain S 127 (DQ485702)	94	Uncultured bacterium clone JSS4-52 (HM750468)	96	10.0	0.0	7.8	15.4	
<i>Hyphomicrobiaceae</i>	19	<i>Rhodomicrobium vannielii</i> ATCC 17100 (NC_014664)	86	Uncultured bacterium clone P1_A05 (GU945902)	91	5.0	0.0	0.0	0.0	
<i>Xanthobacteraceae</i>	9	<i>Azorhizobium caulinodans</i> strain DSM 5975 (AJ563960)	92	Uncultured nitrogen-fixing bacterium clone SB-12 (EU090266)	95	0.0	0.0	6.7	3.8	
<i>Rhodospirillales</i>										
<i>Rhodospirillaceae</i>	2	<i>Azospirillum brasilense</i> Sp245 (NC_016617)	95	Unidentified bacterium <i>nif</i> gene cluster (AF194084)	96	0.0	0.0	3.3	1.3	
	8	<i>Azospirillum lipoferum</i> strain 4B (FQ311868)	95	<i>Azospirillum lipoferum</i> strain 4B (FQ311868)	95	5.0	0.0	1.1	9.0	
<i>Betaproteobacteria</i>										
<i>Burkholderiales</i>										
<i>Burkholderiaceae</i>	4	<i>Burkholderia vietnamiensis</i> strain MMi-302 (EF158809)	88	Uncultured bacterium clone Yushu-14 (AY601050)	89	0.0	0.0	8.9	10.3	

	5	<i>Burkholderia tropica</i> strain Aa1 (AY313947)	96	Uncultured bacterium clone Yushu-14 (AY601050)	96	15.0	1.3	17.8	10.3
<i>Incertae sedis</i>	6	<i>Leptothrix cholodnii</i> strain SP-6 (NC_010524)	95	<i>Leptothrix cholodnii</i> strain SP-6 (NC_010524)	95	0.0	0.0	0.0	1.3
<i>Gammaproteobacteria</i>									
<i>Methylococcales</i>									
<i>Methylococcaceae</i>	11	<i>Methylovulum miyakonense</i> HT12 (NZ_KB913025)	92	Uncultured bacterium clone pCOFnifE11 (EU156126)	94	0.0	0.0	3.3	2.6
<i>Thiotrichales</i>									
<i>Thiotrichaceae</i>	12	<i>Thiothrix nivea</i> DSM 5205 (NZ_JH651384)	87	Uncultured bacterium clone JSS16-2 (HM750519)	91	0.0	0.0	2.2	1.3
<i>Deltaproteobacteria</i>									
<i>Desulfarculales</i>									
<i>Desulfarculaceae</i>	20	<i>Desulfarculus baarsii</i> DSM 2075 (NC_014365)	89	Uncultured bacterium clone cloC-08 (JX268325)	99	0.0	0.0	1.1	0.0
	22	<i>Desulfarculus baarsii</i> DSM 2075 (NC_014365)	83	Uncultured nitrogen-fixing bacterium clone SB-24 (EU090278)	94	5.0	5.2	26.7	25.6
<i>Desulfovibrionales</i>									
<i>Desulfovibrionaceae</i>	18	<i>Desulfovibrio magneticus</i> MBC34 (NZ_ALAO01000346)	88	Uncultured nitrogen-fixing bacterium clone SB-37 (EU090291)	96	0.0	0.0	4.4	1.3
<i>Syntrophobacterales</i>									
<i>Syntrophobacteraceae</i>	14	<i>Syntrophobacter fumaroxidans</i> MPOB (NC_008554)	93	Uncultured bacterium clone SN1P1047 (JX154753)	94	0.0	0.0	1.1	1.3
<i>Nitrospirae</i>									

Nitrospirales

Nitrospiraceae

1	<i>Thermodesulfovibrio yellowstonii</i> DSM 11347 (NC_011296)	93	' <i>Bradyrhizobium japonicum</i> ' clone nifH 1-60 (GQ289578)	95	0.0	0.0	1.1	3.8
10	<i>Thermodesulfovibrio yellowstonii</i> DSM 11347 (NC_011296)	87	' <i>Bradyrhizobium japonicum</i> ' clone nifH 1-60 (GQ289578)	94	0.0	0.0	8.9	1.3

Chlorobi

Chlorobia

Chlorobiales

Chlorobiaceae

15	<i>Chloroherpeton thalassium</i> ATCC 35110 (NC_011026)	91	<i>Chloroherpeton thalassium</i> ATCC 35110 (NC_011026)	91	0.0	0.0	1.1	1.3
16	<i>Chlorobium phaeovibrioides</i> DSM 265 (NC_009337)	89	Uncultured nitrogen-fixing bacterium clone SB-38 (EU090292)	98	0.0	0.0	1.1	2.6

Spirochaetes

Spirochaetales

Spirochaetaceae

21	<i>Treponema azotonutricium</i> ZAS-9 (NC_015577)	90	Uncultured bacterium clone pTKY2312_10 (JN217110)	92	0.0	0.0	1.1	0.0
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Verrucomicrobia

Opitutae

Puniceicoccales

Puniceicoccaceae

13	<i>Coraliomargarita akajimensis</i> DSM 45221 (CP001998)	90	Uncultured bacterium clone JSS4-10 (HM750471)	91	0.0	0.0	0.0	2.6
17	<i>Coraliomargarita akajimensis</i> DSM 45221 (CP001998)	94	<i>Opitutaceae</i> bacterium TAV5 (AGJF01000003)	98	5.0	0.0	2.2	3.8

* Identity of in silico translated nucleic acid sequences as determined by BLASTX analysis.