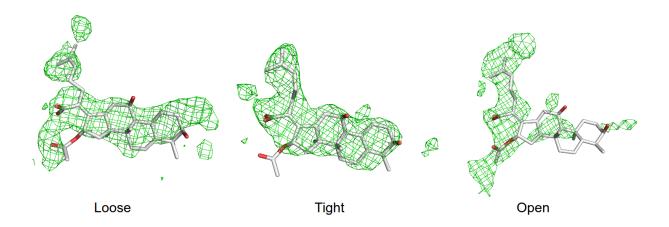
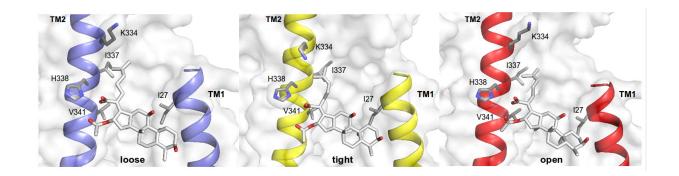


Supplementary Figure 1 - 24-Bromofusidic acid bound at TM1-TM2

A Chemical structure of 24-bromofusidic acid. **B, C** Anomalous density for the bromine atom at TM1/TM2 in **B** tight and **C** open monomer, contoured at 5 σ , 1.3098 e Å³ (red mesh). The anomalous signal of the bromine atom is located at position C24.

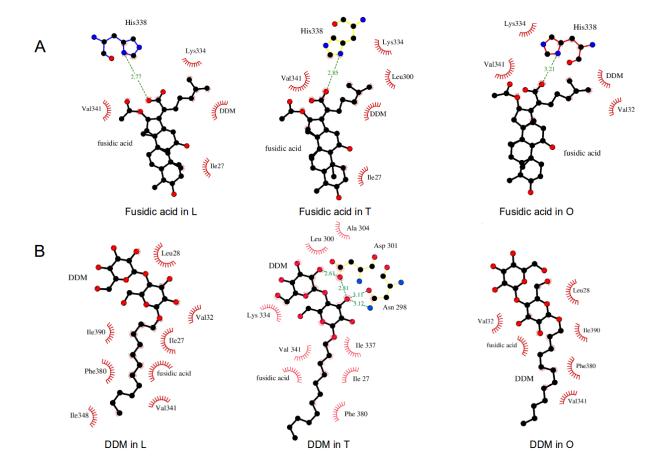


Supplementary Figure 2 - Polder and Fo-Fc electron density map for fusidic acid at TM2 groove Polder maps contoured at 3.5, 4.0 and 3.0 σ for fusidic acid bound at loose, tight and open conformer, respectively. Polder maps were calculated by phenix.polder²⁷. Fusidic acid is shown as sticks (carbon = white; oxygen = red).



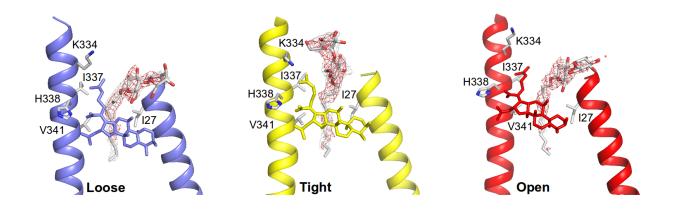
Supplementary Figure 3 - Localization of fusidic acid in each of the three protomers at the TM1/TM2 groove of asymmetric AcrB

Fusidic acid (white stick presentation with oxygen atoms in red) between TM1 (right helix) and TM2 (left helix) in the loose (on the left, blue), tight (middle, yellow), and open (right, red) conformer of asymmetric AcrB. On TM1, I27 is indicated as grey stick, on TM2, from top to bottom, K334, I337, H338 and V341 side chains are indicated in grey sticks (N atoms in blue).



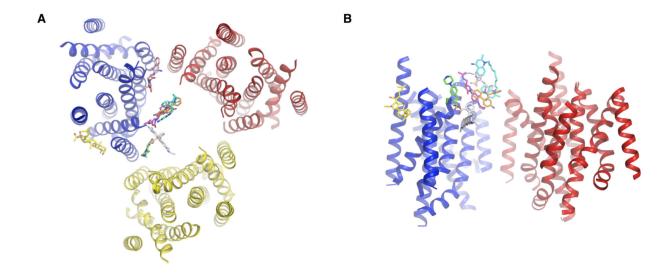
Supplementary Figure 4 - Ligplot representations of fusidic acid and dodecyl-β-D-maltoside binding in the TM1/TM2 area

- **A** Ligand interactions of fusidic acid in the loose (L), tight (T) and open (O) conformer.
- ${\bf B}$ Ligand interaction of dodecyl- β -D-maltoside in the loose (L), tight (T) and open (O) conformer.



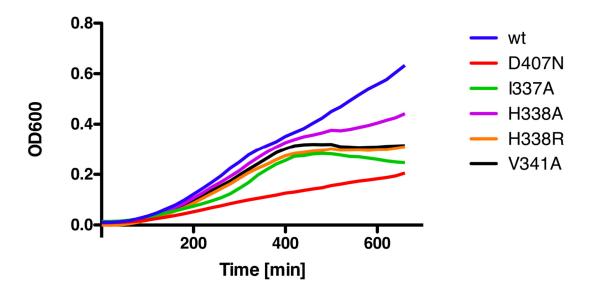
Supplementary Figure 5 - Densities and assignment to dodecyl-β-D-maltoside located at the TM1/TM2 groove

Observed Fo-Fc and 2Fo-Fc SA composite omit maps. Shown are the Fo-Fc densities (at 2.5, 2.5, and 2.0 sigma, in red mesh), 2Fo-Fc densities (at 1.0 sigma, in grey mesh), and DDM assignment at the TM1/TM2 groove of the Loose (colored in blue), Tight (yellow), and Open (red) protomers, respectively.



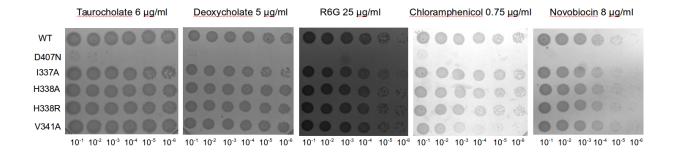
Supplementary Figure 6 - Superimposition of the L protomer of the symmetric AcrB structures with assigned drug molecules bound at the central cavity with the L protomer of the asymmetric fusidic acid AcrB complex structure

Symmetric structures of AcrB comprising one monomer in the asymmetric unit were superimposed on the L monomer (blue) of asymmetric AcrB structure with bound fusidic acid (this work, comprising a trimer in the asymmetric unit, L protomer, blue; T protomer, yellow, O protomer, red). The position of substrates from the symmetric structures are shown in colored stick representation: Rhodamine 6G (white, PDB: 10Y8), Ethidium (orange, PDB: 10Y9), Dequalinium (cyan. PDB: 10YD), Ciprofloxacin (green, PDB: 10YE), Linezolid (pink, PDB: 4K7Q), Ampicillin (salmon, PDB: 2RDD). Fusidic acid from this work is shown in yellow stick representation. A Top view of the transmembrane domain viewed from the periplasm. B View parallel to the membrane plane. Here, the T protomer (yellow in A) has been omitted for clarity.



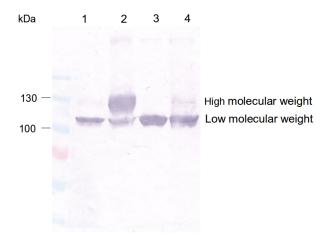
Supplementary Figure 7 - Phenotypic analysis of AcrB wildtype and TM2 mutants in liquid culture containing 60 μg ml $^{-1}$ fusidic acid

Representation of growth curves of *E. coli* BW25113 $\Delta acrB$ complemented with pET24acrB wild type and the indicated mutants on LB supplemented with 60 μg ml⁻¹ fusidic acid. The normalized OD₆₀₀ is plotted versus time of growth.

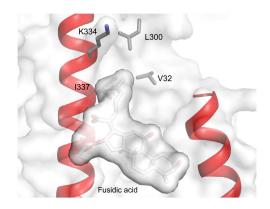


Supplementary Figure 8 - Drug LB-agar plate dilution test with the indicated AcrB variants

E. coli BW25113ΔacrB was complemented with *acrB* wildtype or mutant genes expressed from plasmid. For the assay, colonies of these cells were picked and grown over night in LB_{Kan50} medium at 37 °C, diluted to OD₆₀₀ 10^{-1} – 10^{-6} , and 4μl were spotted on an LB_{Kan50} agar plate supplemented with Taurocholate (6 μg/ml), Deoxycholate (5 μg/ml), Rhodamine-6G (25 μg/ml), Chloramphenicol (0.75 μg/ml), or Novobiocin (8 μg/ml). Comparison between wild type and the D407N mutant (deficient in proton translocation) shows in all variants no difference in resistance, with the exception of V341A in the presence of chloramphenicol. Cell dilutions are indicated at the bottom of each plate.

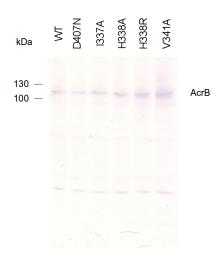


Supplementary Figure 9 - Determination of the extent of MTS-rhodamine cross-linking to AcrB-cl_C338 Membranes containing AcrB-cl_C338 were treated as described in Materials and Methods. Briefly, membranes (80 μ l) were washed with cross-linking buffer (CLB), and incubated with either MTS-rhodamine (20 μ M) or *N*-ethylmaleimide (NEM, 5 mM) in the absence of AcrB substrate for 15 min on ice. After addition of 420 μ l ice-cold CLB excess thiol-reactive reagent was removed by centrifugation. Subsequently, membranes were resuspended in CLB (80 μ l) containing 1.5% SDS and methoxypolyethylene glycol maleimide (average molecular weight = ~5000) (MAL-PEG, 3 mM) and incubated at room temperature for 2 h. As a positive control, membranes without prior treatment with NEM or MTS-rhodamine were treated with MAL-PEG in parallel. Membranes without addition of thiol-reactive reagents were used as a negative control. Samples were analyzed by SDS-PAGE and Western blot using α -His antibody as probe. Lane 1: Detection of AcrB-cl_C338 without NEM, MTS-rhodamine, or MAL-PEG treatment; 2: Detection of MAL-PEG only treated AcrB-cl_C338 sample; 3: NEM+MAL-PEG treated sample; 4: MTS-rhodamine + MAL-PEG treated sample.

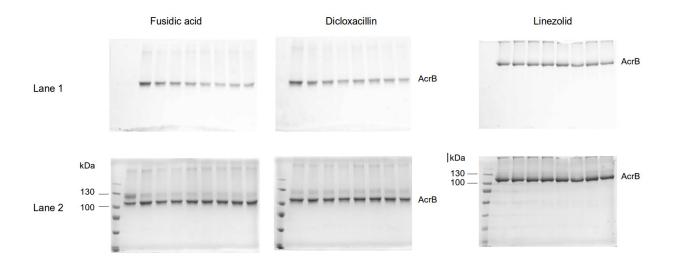


Supplementary Figure 10 - Fusidic acid localization proximal to a hydrophobic area in the AcrB O monomer

In the O conformer, fusidic acid is localized proximal to a hydrophobic pocket, comprising V32, L300, K334 and I337 (as indicated).

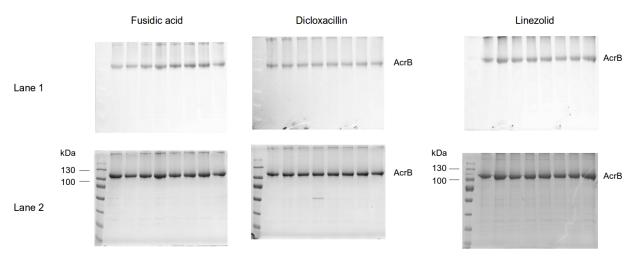


Supplementary Figure 11 – Uncropped image of Figure 4a

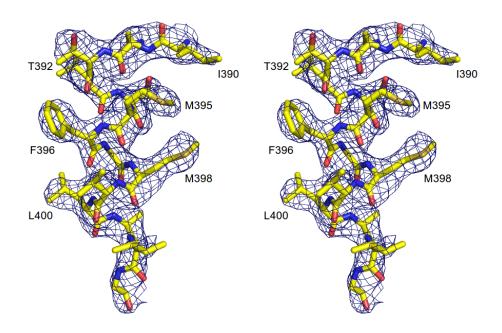


Supplementary Figure 12 – Uncropped images of Figure 5 for in-gel fluorescence (Lane 1) and Coomassie-stained SDS-PAGE analysis of MTS-rhodamine modified AcrB-cl_C338 (Lane 2)

In Lane 2, Row 2 of fusidic acid is AcrB-cl_C338 sample labeled with MAL-PEG. This is a control to show that C338 is able to react with thiol-reactive compounds.



Supplementary Figure 13 – Uncropped images of Figure 5 for in-gel fluorescence (Lane 1) and Coomassie-stained SDS-PAGE analysis of MTS-rhodamine modified AcrB-cl_C14 (Lane 2)



Supplementary Figure 14 – Stereo image of a representative portion of the electron density map of AcrB/DARPins in complex with fusidic acid. Residues 390-403 are shown in stick representation with the 2Fo-Fc electron density map of that region contoured at 1.2σ (blue mesh).

Supplementary Table 1 - Data collection and refinement statistics

	A aug /D A DDina in a complete	A sup /D A DDisas in a susual su
	AcrB/DARPins in complex with fusidic acid	AcrB/DARPins in complex with
	With fusible acid	24-bromofusidic acid
Data collection		2 : 5: 5: 1: 1: 1: 1: 1: 1: 1: 1: 1: 1: 1: 1: 1:
Space group	P2 ₁ 2 ₁ 2 ₁	P2 ₁ 2 ₁ 2 ₁
Cell dimensions		
a, b, c (Å)	145.65, 163.25, 246.04	145.49, 163.54, 245.10
α; β; γ (°)	90.0, 90.0, 90.0	90.0, 90.0, 90.0
Resolution (Å)	49.12- 2.50 (2.54-2.50)*	49.02- 2.60 (2.64-2.60)
R _{merge}	0.158 (2.290)	0.259 (2.228)
Ι/σΙ	9.9 (0.8)	10.6 (1.5)
Completeness (%)	100.0 (99.9)	100.0 (100.0)
Redundancy	6.7 (6.7)	13.5 (13.6)
Refinement		
Resolution (Å)	2.5	
No. reflections	192130	
R _{work} / R _{free}	22.49 / 26.15	
No. atoms		
Protein	25948	
Ligand/ion	874	
Water	815	
B-factors		
Protein	62.62	
Ligand/ion	108.47	
Water	45.65	
R.m.s. deviations		
Bond length (Å)	0.0069	
Bond angles (º)	1.1048	

Number of crystals for each structure =1. * Highest resolution shell is shown in parenthesis.

Supplementary Table 2 - Primer sequences (PHO: phosphorylated at 5' end)

Plasmid	Primer sequences
pET24acrB _{His}	AcrBfor (5'-GGATCCCATATGCCTAATTTCTTTATCGATC-3') acrBrev (5'-AAGCTTCTCGAGATGATCGACAGTATGGCTG-3')
pET24acrB-I337A _{His}	For_AcrB_l337A (5'-AAGCTTCTCGAGATGATCGACAGTATGGCTG-3') rev_AcrB337-338 (5'PHO-GAACGGCGTGGTGTCATATGGGTAAAC-3)
pET24acrB-H338A _{His}	For_AcrB_H338A (5'-GTGAAAATCTCTATTGCCGAAGTGGTTAAAACGC-3') rev_AcrB337-338 (5'PHO-GAACGGCGTGGTGTCATATGGGTAAAC-3)
pET24acrB-H338R _{His}	For_AcrB_H338R (5'-GTGAAAATCTCTATTAGAGAAGTGGTTAAAACGC-3') rev_AcrB337-338 (5'PHO-GAACGGCGTGGTGTCATATGGGTAAAC-3)
pET24acrB-V341A _{His}	For_AcrB_V341A (5'-CGAAGTGGCAAAAACGCTGGTCGAAGCGATCATCCTCGTGTTCCTGG-3') rev_AcrB341 (5'PHO-TGAATAGAGATCTTCACGAACGGCGTGGTGTCGTATGGG-3')

Supplementary Methods

Abbreviations:

ACN: Acetonitrile

Br₂: Bromine liquid

CCl₄: Carbon tetrachloride

DBU: 1,8-Diazabicyclo[5.4.0]undec-7-ene

DCM: Dichloromethane

DMF: N,N-Dimethylformamide

EA Ethyl acetate

eq.: Equivalent

EtOH: Ethanol

HPLC: High performance liquid chromatography

KH₂PO₄: Potassium dihydrogen phosphate

LC-MS: Liquid chromatography mass Spectroscopy

MeOH Methanol

NaOH: Sodium hydroxide

Na₂SO₄: Sodium sulfate

NMR: Nuclear magnetic resonance

PE Petroleum ether

R_f: Retention factor in chromatography

Rt: Retention time

R.T.: Room temperature

SCRC: Sinopharm Chemical Reagent Co., Ltd.

TEA: Triethylamine

TFA: Trifluoroacetic acid

TLC: Thin layer chromatography

Tol.: Toluene

1. Deliverable of BC1102

1.1 Original deliverables:

Basilea China shall synthesize **24-Bromo-fusidic acid sodium salt BC1111** and deliver 50-100 mg to Basilea Pharmaceutica International Ltd.:

2. Synthesis of BC1111

2.1 General comment

Reference:

Patent: WO2005/007669A1

Instrumentation: All reagents and solvents were of commercial quality and used without further purification unless otherwise noted. The NMR spectra were obtained with a Bruker AVANCE III 400 MHz spectrometer using CDCl₃ or DMSO-d₆ as solvent. The chemical shifts are expressed in ppm with CDCl₃ or DMSO-d₆ as internal standard. MS (ESI-MS) was recorded on a Waters UPLC H-Class with SQD2 MS spectrometer (Instrument No.03) or Waters 2695/2998 HPLC with SQD MS spectrometer (Instrument No.39). Purity of the products were checked using a Agilent 1200 series instrument (Instrument No.21) equipped with a SunFireTM C18 (3.5μm), 150×4.6 mm column. Peaks were detected at 254 nm or 230 nm or 210 nm absorption. Thin layer chromatography (TLC) was performed using glass plates pre-coated with silica gel GF₂₅₄ with a layer thickness of 0.25 mm purchased from Tsingdao Haiyang Co. Ltd. The conditions for ¹H NMR, LC-MS are listed below unless otherwise noted. The HPLC conditions are listed in the analytical data respectively.

NMR analysis

Varian: MercuryPlus-400M Bruker: AVANCE III 400

Solvent: CDCl₃ or DMSO-d₆

Method A:

LC-MS: Instrument No. 03

Instrument: LC/MS system (Instrument No. 33)

Agilent 1100 Series

Components: G1310A/G1315B/G1313A/G1946D

Agilent software (Agilent ChemStations Ver.A. 10.02[1757])

Pump A: Water (prepared using a Millipore ultra pure water system) with 0.1%

formic acid

Pump B: ACN (LC-MS grade) with 0.1% formic acid

Column: Agilent SB-C18 (3.5 µm), 4.6 mmx50mm

Column temp: room temperature

Detection: DAD

Flow: 1.5 mL/min

Gradient program:

Step	Time (min)	Phase A (%)	Comments	
1	0	10	Equilibration	
2	1 90		10	Linear gradient
3	4	5	95	
4	5.5	5	95	

Method B:

LC-MS:

Instrument No. 39

Instrument: LC/MS system (Instrument No. 39)

Waters 2695/2998 HPLC with SQD MS

Waters Empower2 software

Pump A: Water (prepared using a Millipore ultra pure water system, resistivity:

18.2 $M\Omega$.CM) with 0.1% formic acid

Pump B: ACN (LC-MS grade) with 0.1% formic acid

Column: Atlantis ® T3 (3 µm), 100×4.6 mm

Column temp: 30 °C

Detection: λ = 254 nm, 230 nm and 218 nm/PDA

Flow: 1.0 mL/min Gradient elution program:

Step	Time (min)	Phase A (%)	Phase B (%)	Comments
1	0	95	5	Equilibration
2	1	95	5	Linear gradient
3	13	5	95	
4	15	5	95	

Method C:

HPLC: Instrument No. 09

HPLC method description:

Instrument: Agilent 1100 Series (HPLC09)

Agilent software (Agilent ChemStations Rev.A. 10.02[1757])

Pump A: Water (prepared using a Millipore ultra pure water system,

resistivity: $18.2M\Omega$.CM) with $10mM K_2HPO_4$ (pH=7.2)

Pump B: Acetonitrile (Ourchem®, HPLC grade)

Column: Atlantis® T3 (3µm), 150×4.6 mm

Column temp: R. T.

Sample conc: 0.8mg/mL

Injection vol: 8µL

Detection: λ = 254nm, and 230nm, and 210nm/DAD

Flow: 1.0mL/min

Gradient program:

Step	Time (min)	Phase A (%)	Phase B (%)	Comments

1	0	45	55	Equilibration
2	1	45	55	Linear gradient
3	13	5	95	
4	16.5	5	95	
5	17	95	5	

Method D:

HPLC: Instrument No. 21

HPLC method description:

Instrument: Agilent 1200 Series (HPLC21)

Agilent software (ChemStation for LC 3D system B.04.02 SP1 [208])

Pump A: Water (prepared using a Millipore ultra pure water system, resistivity:

 $18.2 \text{ M}\Omega.\text{CM}$) with 0.1% TFA

Pump B: ACN (Ourchem®, HPLC grade) with 0.1% TFA

Column: Prontosil 120-3-C18-SH (3 μm) 150×4.6 mm

Column temp: room temperature

Detection: λ = 230 nm and 210 nm/DAD

Flow: 1.0 mL/min

Gradient program:

Step	Time (min)	Phase A (%)	Phase B (%)	Comments
1	0	98	2	Equilibration
2	5	98	2	Linear gradient
3	18	62	38	
4	25	5	95	
5	30	5	95	

2.2 Synthesis of BAL0117224 (BC1111)

2.2.1 Synthesis of BC1111a01

2.2.1.1 General comment

The product was obtained in 85.9% yield after purification.

Reference:

3.31

g

Patent: WO2005/007669A1

2.2.1.2 Experimental Procedure

Lab Journal No.: JIANL-180-1

Experiment No.: JIBC1111a01-001

Under nitrogen atmosphere, in a 100 mL round-bottomed flask, a mixture of

of Fusidic acid (note 1, 25.18 mmol, 1.0 eq.) and 13.01 g

of TEA (note 2, 32.74 mmol, 1.3 eq.) in 38 of DMF (note 3) was stirred at 25°C for 30 minutes. Then mL

5.31 of Chloromethyl pivalate (note 4, 35.26 mmol, 1.4 eq.) was added dropwise at 25°C over 25 minutes. After addition, the reaction mixture was stirred at 50°C for 18 hours. When TLC showed that the reaction was complete, the mixture was cooled to 25°C and then concentrated under vacuum. The residue was purified by silica gel column chromatography eluting with PE/EA=3/1 (TLC: PE/EA=3/1, R_f=0.25) to give

13.65 g of the product as a white foam.

Product: 13.65 g

Yield: 85.9%

Notes

- 1. Industrial grade
- 2. SCRC
- 3. SCRC
- 4. SCRC

Analytical Methods

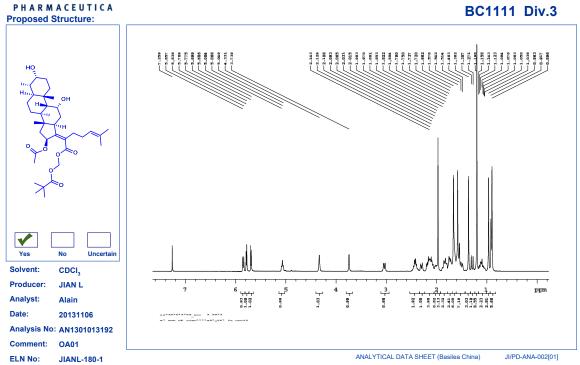
1H NMR (400 MHz, CDCl₃) δ ppm: 5.85 (d, J=8.4Hz, 1H, H-16), 5.69, 5.78 (AB, J=5.6Hz, 2H, H-32), 5.07 (t, J=7.2Hz, 1H, H-24), 4.33 (s, 1H, H-3), 3.74 (s, 1H, H-11), 3.04 (d, J=11.2Hz, 1H, H-13), 2.35-2.52 (m, 2H, H-2(1H), 12(1H)), 2.25-2.35 (m, 1H, H-4), 2.05-2.20 (m, 4H, H-22, 23), 1.95-2.05 (m, 1H, H-15(1H)), 1.97 (s, 3H, H-31), 1.46-1.90 (m, 8H, H-1(1H), 2(1H), 5, 6, 7(1H), 12(1H), 15(1H)), 1.66 (s, 3H, H-26), 1.58 (s, 3H, H-27), 1.36 (s, 3H, H-28), 1.29 (d, J=14.4Hz, 1H, H-9), 1.20 (s, 9H, H-35), 1.00-1.19 (m, 2H, H-1(1H), H-7(1H)), 0.96 (s, 3H, H-18), 0.85-0.95 (m, 6H, H-19, 29)

2.2.1.3 Analytical data

¹H NMR analysis: Varian: MercuryPlus-400M

Solvent: CDCl₃

basilea NMR of BC1111a01



2.2.2 Synthesis of BC1111a02

2.2.2.1 General comment

The experimental result indicated that the product was labile over silica gel. Thus the crude product was used directly in the next step without further purification after work up. The crude product was analyzed by LCMS but it coudn't be ionized.

Reference:

Patent: WO2005/007669A1

2.2.2.2 Experimental Procedure

Lab Journal No.: JIANL-189-1

Experiment No.: JIBC1111a02-003

Under nitrogen atmosphere, in a 250 mL round-bottomed flask,

7.15 g of <u>BC1111a01</u> (note 1, 11.33 mmol, 1.0 eq.) was dissolved in

40 mL of $\underline{CCl_4}$ (note 2) and the resulting mixture was cooled to -10°C. Then a

solution of

1.99 g of Br_2 (note 3, 12.47 mmol, 1.1 eq.) in

40 mL of $\underline{CCl_4}$ (note 2) was dropwise added in the course of 1 hour with

continuous stirring at -10°C. After addition, the resulting reaction mixture was further stirred at -10°C for 1 hour. When TLC showed that the

reaction was complete, the mixture was concentrated under vacuum to

give the crude product as a light yellow foam. The crude product was

used directly in the next step without further purification.

Product: 8.95 g

Yield: 100%

Notes

1. JIANL-180-1

2. SCRC, dried over 4 Å molecular sieve

3. SCRC

2.2.3 Synthesis of BC1111a03

2.2.3.1 General comment

The product was prepared in about 40% yield after purification by silica gel column chromatography.

Reference:

Patent: WO2005/007669A1

2.2.3.2 Experimental Procedure

Lab Journal No.: JIANL-185-1

Experiment No.: JIBC1111a03-002

Under Argon atmosphere, to a solution of

3.78 g of <u>BC1111a02</u> (note 1, 4.78 mmol, 1.0 eq.) in

50 mL of CCl₄ (note 2) was added

1.46 g of DBU (note 3, 9.56 mmol, 2.0 eq.). The mixture was refluxed at 77°C

for 16 hours. Then the reaction mixture was filtered through Celite. The

filter cake was washed with

100 mL of PE and

100 mL of EA successively. The combined filtrate and washings were

concentrated under reduced pressure. The residue was purified by silica

gel column chromatography eluting with PE/EA=5/1 (TLC: PE/EA=3/1,

 $R_f=0.25$) to give

1.29 g of the product as a light yellow foam.

Product: 1.29 g

Yield: 38.1%

Notes

1. JIANL-184-1

2. SCRC, anhydrous

3. SY-001471

Analytical Methods

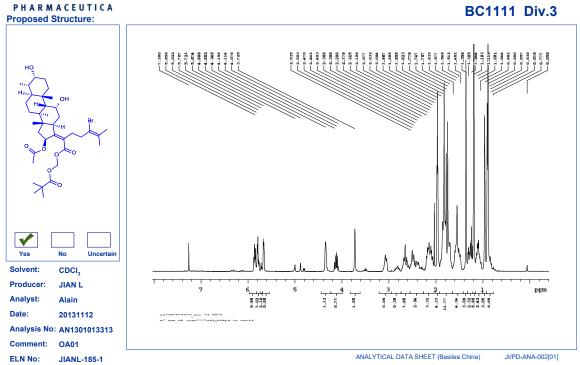
¹H NMR (400 MHz, CDCl₃) δ ppm: (the sample was not clean) 5.85 (m, 1H, H-16), 5.68, 5.82 (AB, 2H, H-32), 4.35 (s, 1H, H-3), 3.72 (s, 1H, H-11), 3.02-3.12 (m, 1H, H-13), 2.06-2.22, 2.35-2.58 and 2.58-2.75 (3m, 7H, H-2(1H), 4, 12(1H), 22, 23), 1.92-2.00 (m, 4H, H-15(1H), 31), 1.45-1.65 and 1.65-1.92 (m, 8H, H-1(1H), 2(1H), 5, 6, 7(1H), 12(1H), 15(1H)), 1.85 (s, 3H, H-26), 1.75 (s, 3H, H-27), 1.36 (s, 3H, H-28), 1.25-1.32 (m, 1H, H-9), 1.24 (s, 9H, H-35), 1.03-1.20 (m, 2H, H-1(1H), 7(1H)), 0.96 (s, 3H, H-18), 0.85-0.95 (m, 6H, H-19, 29)

2.2.3.3 Analytical data

¹H NMR analysis: Varian: MercuryPlus-400M

Solvent: CDCl₃

basilea NMR of BC1111a03



2.2.4 Synthesis of BC1111a04

2.2.4.1 General comment

A main impurity with similar polarity to the product couldn't be removed by silica gel column chromatography. Then the product was tried to be purified by recrystallization using EA/Toluene as solvent according to the patent but failed. Later, the product was treated with DCM/MeOH/PE and most of the impurity was precipitated and removed by filtration.

After then the filtrate was concentrated to give the product with better purity which was further purified by preparative HPLC.

Reference:

Patent: WO2005/007669A1

2.2.4.2 Experimental Procedure

Lab Journal No.: JIANL-191-001

Expe	riment No	o.: JIBC1111a04-003
		Under Argon atmosphere, to a solution of
3.26	g	of <u>BC111a03</u> (note 1, 4.59 mmol, 1.0 eq.) and
0.32	g	of <u>DBU</u> (note 2, 2.11 mmol, 0.46 eq.) in
40	mL	of MeOH (note 3) was added dropwise a mixed solution of
25	mL	of MeOH (note 3) and
25	mL	of $\underline{\text{H}_2\text{O}}$ over 1 hour. Then the mixture was stirred at 20°C for 5 hours.
		Then the mixture was acidified with 1.0 M aqueous solution of KH_2PO_4
		until pH=4-5. The resulting mixture was extracted with
100	mLX3	of EA. The combined organic phases were washed with
60	mL	of H ₂ O and
60	mLX2	of brine successively, dried over Na ₂ SO ₄ , and concentrated under
		vacuum. The residue was purified by silica gel column chromatography
		(Eluent: PE/EA/HCOOH=50/50/0.5) to give
2.30	g	of the product as white solid. The resulting solid was dissolved in
5	mL	of DCM and
1	mL	of MeOH followed by dropwise addition of
30	mL	of PE at 20°C. Most of the impurity was precipitated and then removed by
		filtration. Then the filtrate was concentrated to give
0.70	g	of the desired product with better purity which was further purified by
		preparative HPLC (note 4) to give
0.27	g	of the pure product as a white solid.

Product: 270 mg

Yield: 9.9%

Notes

1. JIANL-190-1

2. SY-001471

3. SCRC

4. Prep. HPLC condition:

Pump A: Water with 0.1% TFA

Pump B: Acetonitrile

Column: Atlantis[®] Prep T3 OBD™ 19×150mm 5µm

Column temp: rt

Detection: λ = 210nm

Flow: 14 mL/min

Gradient program:

Step	Time (min)	Phase A (%)	Phase B (%)	Comments		
1	1 0.0		1 0.0		20	Linear gradient
2	2 11.0 40 60		60			
3	11.1	40	60			
4	15.0	0	100			

Analytical Methods

¹H NMR (400 MHz, CDCl₃) δ ppm: (some EA was left in the sample) 8.02 (s, 1H, COO $\underline{\text{H}}$), 5.88 (d, J=8.4Hz, 1H, H-16), 4.38 (s, 1H, H-3), 3.76 (s, 1H, H-11), 3.08 (d,

J=11.2Hz, 1H, H-13), 2.48-2.80 (m, 4H, H-22, 23), 2.00-2.22 (m, 4H, H-4, 2(1H), 12(1H), 15(1H)), 1.97 (s, 3H, H-31), 1.83 (s, 3H, H-26), 1.76 (s, 3H, H-27), 1.78-1.93, 1.70-1.78 and 1.46-1.65 (3m, 8H, H-1(1H), 2(1H), 5, 6, 7(1H), 12(1H), 15(1H)), 1.37 (s, 3H, H-28), 1.26-1.35 (m, 1H, H-9), 1.03-1.18 (m, 2H, H-1(1H), 7(1H)), 0.98 (s, 3H, H-18), 0.87-0.95 (m, 6H, H-19, 29)

¹³C NMR (400 MHz, CDCl₃) δ ppm: 173.6 (C-21), 170.3 (C-30), 151.8 (C-17), 130.9 (C-25), 127.7 (C-20), 119.7 (C-24), 74.0 (C-16), 71.1 (C-3), 67.7 (C-11), 48.9 (C-9), 48.2 (C-14), 44.0 (C-15), 39.0, 38.4 and 37.4 (C-5, 8 13), 36.3, 36.1 and 35.2 (C-4, 12, 23), 31.3, 29.5 and 29.2 (C-1, 2, 7), 27.4 (C-10), 24.8, 23.2 and 22.8 (C-6, 19, 22), 20.6, 20.5 and 20.2 (C-18, 28, 31), 17.3 (C-26, 27), 15.5 (C-29)

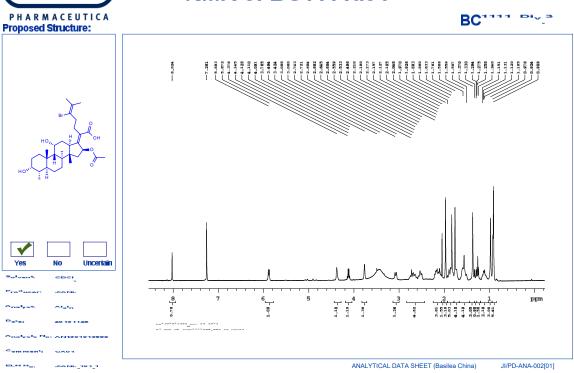
2.2.4.3 Analytical data

¹H NMR analysis: Varian: MercuryPlus-400M

Solvent: CDCl₃



NMR of BC1111a04

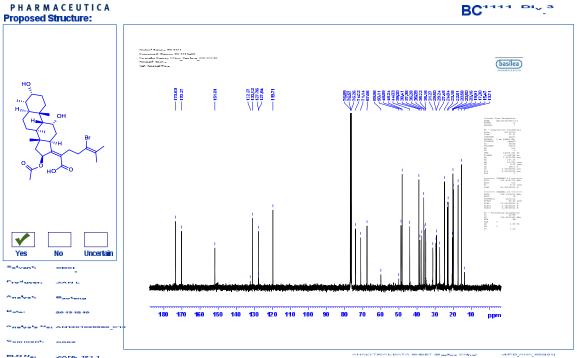


¹³C NMR analysis: Bruker: AVANCE III 400

Solvent: CDCl₃



13C-NMR of BC1111a04



2.2.5 Synthesis of BAL0117224 (BC1111)

2.2.5.1 General comment

Reference:

Patent: WO2005/007669A1

2.2.5.2 Experimental Procedure

Lab Journal No.: JIANL-199-1 Experiment No.: JIBC1111-001

To a solution of

80	mg	of <u>BC1111a04</u> (note 1, 0.13 mmol, 1.0 eq.) in
1.0	mL	of MeOH (note 2) was added a solution of
0.13	mL	of 1.0 M aqueous solution of NaOH (note 3, 0.13 mmol, 1.0 eq.) at
		20°C. The resulting solution was concentrated and the residue was
		dissolved in
5	mL	of EtOH (note 4) followed by dropwise addition of
5	mL	of EA (note 5) at 20°C, but no precipitate formed. Then the resulting
		mixture was concentrated under reduced pressure until about 2 mL of
		oily residue was left. Then
5	mL	of <u>EA</u> (note 5) was added at 20°C and a lot of white precipitate formed.
		The white solid was collected by filtration and then dissolved in
1	mL	of ACN and
5	mL	of H ₂ O. The resultant mixture was lyophilized to afford

Product: 67 mg

Yield: 80.7%

Purity: 92.3% (210 nm)

of the product as a white solid.

Notes

67

1. JIANL-191-1

mg

- 2. SCRC
- 3. SCRC
- 4. SCRC
- 5. SCRC

Analytical Methods

¹H NMR (400 MHz, DMSO-d₆+D₂O) δ ppm: 5.61 (d, J=8.0Hz, 1H, H-16), 4.11 (s, 1H, H-3), 3.50 (s, 1H, H-11), 2.80 (d, J=11.2Hz, 1H, H-13), 2.65-2.75 (m, 1H, H-4), 2.40-2.50, 2.13-2.38 and 2.00-2.10 (3m, 6H, H-22, 23, 2(1H), 12(1H)), 1.85 (s, 3H, H-31), 1.75 and 1.77 (2s, 6H, H-26, 27), 1.82-1.92 and 1.56-1.70 (2m, 4H, H-15, 2(1H), 12(1H)), 1.28-1.55 and 0.93-1.10 (2m, 8H, H-1, 5, 6, 7, 9), 1.24 (s, 3H, H-28), 0.87 (s, 3H, H-18), 0.80 (s, 3H, H-19), 0.78 (d, J=6.4Hz, 3H, H-29)

MS m/z (-ESI): 593.4/595.6 ([M-H]⁻, 68%/100%).

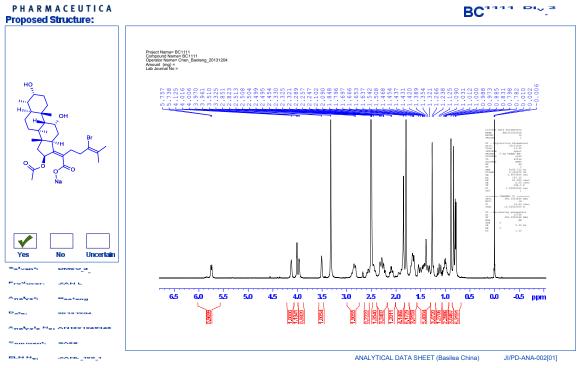
HPLC: Rt =22.86 min (Purity: 88.5% at 254 nm, 91.5% at 230 nm, and 92.3% at 210 nm).

2.2.5.3 Analytical data

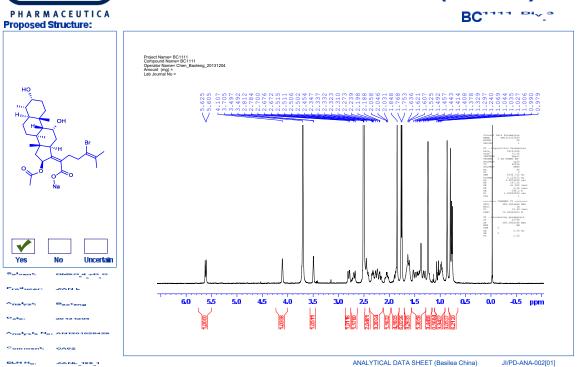
¹H NMR analysis: Bruker: AVANCE III 400

Solvent: d₆-DMSO

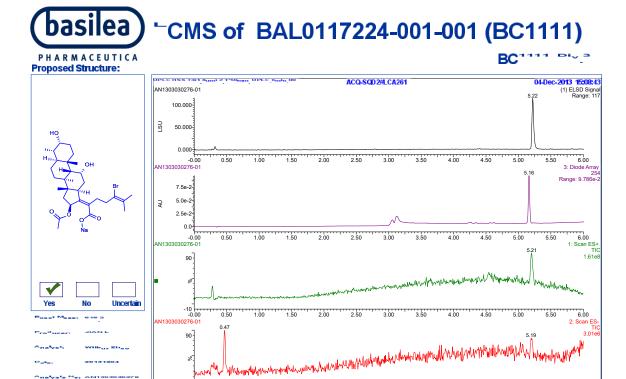
basilea NMR of BAL0117224-001-001 (BC1111)



basilea NMR of BAL0117224-001-001 (BC1111)



LC-MS analysis: Method A

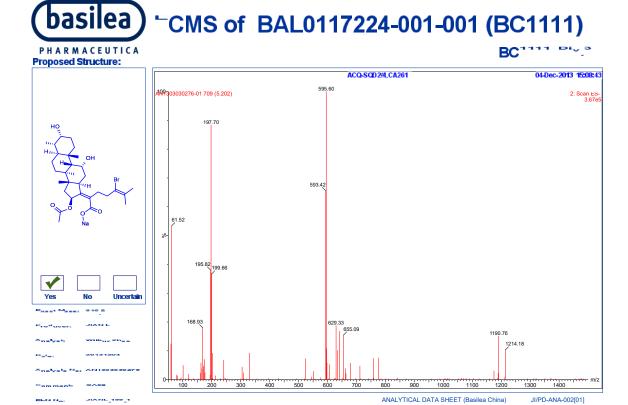


4.00

ANALYTICAL DATA SHEET (Basilea China)

4.50

JI/PD-ANA-002[01]



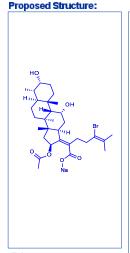
HPLC: Method D

basilea) HPLC of BAL0117224-001-001 (BC1111)

Data file : C:\CHEM32\1\DATA\20131204\AN1302212940.D Injection Date: 12/4/2013 3:42:48 PM







Sample Name: BC1111
Concentration: 0.400mg/ml
Inj. vol. : 8 µl 03:42:->
Acq. Method : C:\CHEMM2\\\METHODS\89-4TFA-2.M
Column : sun fire 4.6*150 3.5 Signal. HPLC-21 Thu, 5. Dec. 2013 11:04:43 am

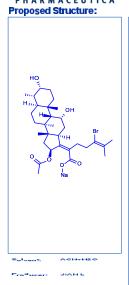
ANALYTICAL DATA SHEET (Basilea China)

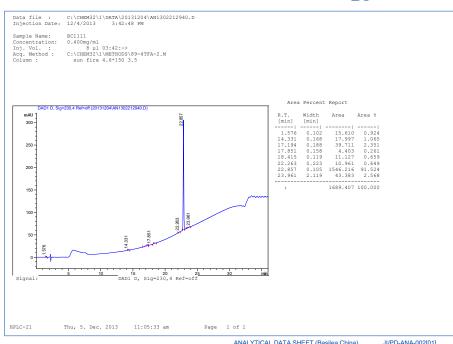
JI/PD-ANA-002[01]

HPLC of BAL0117224-001-001 (BC1111)

PHARMACEUTICA







ANALYTICAL DATA SHEET (Basilea China)

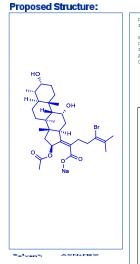
JI/PD-ANA-002[01]

basilea HPLC of BAL0117224-001-001 (BC1111)





JI/PD-ANA-002[01]

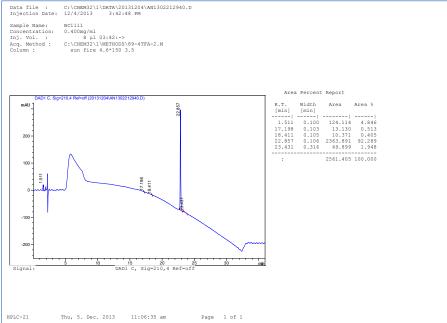


Froducer: JIANL

Pate: 20131204

Comment, OA02

ELN No: JIANE_199_1



ANALYTICAL DATA SHEET (Basilea China)

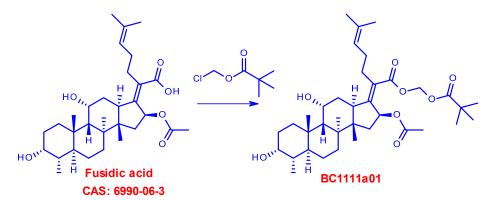
3. Conclusion

The delivered target compounds were listed in the following table.

Compounds	Batch No.	Analytical No.
BAL0117224-001-001 (BC1111)	JIANL-199-1	¹ HNMR: AN1301025428 ¹ HNMR: AN1301025429 HPLC: AN1302212940 LCMS: AN1303030276
BAL0117224-001-002 (BC1111)	JIANL-206-1	¹ HNMR: AN1301025575 ¹ HNMR: AN1301025576 HPLC: AN1302213052 LCMS: AN1303030304

4. Batch tracking

4.1 Synthesis of BC1111a01



Reaction conditions:

ſ	Francisco est No	D-4-	Fusid	ic acid		Chloromet	hyl pivalate	TE	Α	DMF	Temp.	Time
ı	Experiment No.	No. Date	Batch No.	g	mmol	g	mmol/eq.	g	mmol/eq.	mL	°C	hr
	JIBC1111a01-001	11/3/2013	industrial grage	13.01	25.18	5.31	35.26/1.4	3.31	32.74/1.3	38	50	18

Results:

					crude product				pure product				
	Experiment No.	Batch No.	Date	o. Date		Analytical date		Qty.	Yield	Analytical date		Qty.	Yield
				HPLC	LCMS	NMR	mg	%	HPLC	LCMS	NMR	g	%
	JIBC1111a01-001	JIANL-180-1	11/4/2013								AN13010 13192	13.65	85.9

4.2 Synthesis of BC1111a02

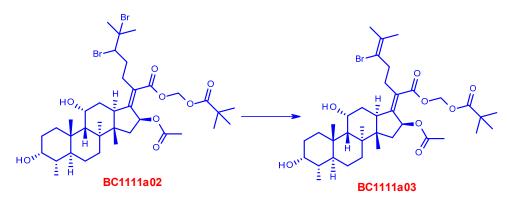
Reaction conditions:

Experiment No.	Date		BC1111a01		Br ₂	CCI ₄	Temp.	Time	
Experiment No.	Date	Batch No.	g	mmol	g	mmol/eq.	mL	°C	hr
JIBC1111a02-001	11/5/2013	JIANL-180-1	3.01	4.77	0.84	5.25/1.1	40	0	2
JIBC1111a02-002	11/11/2013	JIANL-180-1	3.02	4.78	0.84	5.26/1.1	40	-10	2
JIBC1111a02-003	11/21/2013	JIANL-180-1	7.15	11.30	2.00	12.5/1.1	80	-10	2

Results:

				crud	e product			pure product					
Experiment No.	Batch No.	Date	Aı	nalytical da	ite	Qty.	Qty. Yield Analytical date		Qty.	Yield			
			HPLC	LCMS	NMR	g	%	HPLC	LCMS	NMR	g	%	
JIBC1111a02-001	JIANL-181-1	11/5/2013								AN130101 3193	1.783	47.2	
JIBC1111a02-002	JIANL-184-1	11/11/2013				3.78	100.0						
JIBC1111a02-003	JIANL-189-1	11/21/2013				8.95	100.0						

4.3 Synthesis of BC1111a03



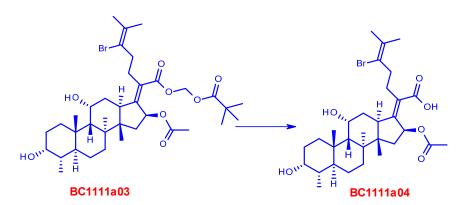
Reaction conditions:

Reaction conditions.	cuction continuous.											
Evmeriment No	Data	BC1111a02				DBU	CCI ₄	Temp.	Time			
Experiment No.	Date	Batch No.	g	mmol	g	mmol/eq.	mL	°C	hr			
JIBC1111a03-001	11/5/2013	JIANL-181-1	1.28	1.62	0.49	3.25/2.0	20	77	16			
JIBC1111a03-002	11/11/2013	JIANL-184-1	3.78	4.78	1.46	9.56/2.0	50	77	16			
JIBC1111a03-003	11/21/2013	JIANL-189-1	8.95	11.3	3.46	22.6/2.0	120	77	16			

Results:

				crud	e product			pure product					
Experiment No.	Batch No.	Date	Analytical date			Qty.	Yield	Α	nalytical da	te	Qty.	Yield	
			HPLC	LCMS	NMR	mg	%	HPLC	LCMS	NMR	g	%	
JIBC1111a03-001	JIANL-182-1	11/6/2013					failed						
JIBC1111a03-002	JIANL-185-1	11/12/2013								AN130101 3313	1.29	38.1	
JIBC1111a03-003	JIANL-190-1	11/22/2013									3.26	40.6	

4.4 Synthesis of BC1111a04



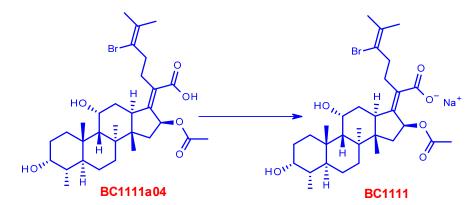
Reaction conditions:

Experiment No.	Date	В	BC1111a03 DBU MeOH		MeOH	H ₂ O	Temp.	Time		
Experiment No.	Date	Batch No.	g	mmol	g	mmol/eq.	mL	mL	°C	hr
JIBC1111a04-001	11/5/2013	JIANL-181-1	0.45	0.63	0.05	0.33/0.53	7.7	4	20	4
JIBC1111a04-002	11/13/2013	JIANL-185-1	1.23	1.74	0.12	0.80/0.46	22.5	12.5	20	4
JIBC1111a04-003	11/21/2013	JIANL-190-1	3.26	4.59	0.32	2.11/0.46	65.0	25.0	20	6

Results:

vesures.												
				crud	e product			pure	product			
Experiment No.	Batch No.	Date	Analytical date			Qty.	Yield	А	nalytical da	te	Qty.	Yield
			HPLC	LCMS	NMR	g	%	HPLC	LCMS	NMR	g	%
JIBC1111a04-001	JIANL-183-001	11/7/2013					failed					
JIBC1111a04-002	JIANL-186-001	11/14/2013		AN130339 6701	AN130101 3347	1	68.9		AN130339 7045		0.15	14.5
JIBC1111a04-003	JIANL-191-001	11/25/2013				2				AN130101 3596	0.27	9.9

4.5 Synthesis of BAL0117224 (BC1111)



Reaction conditions:

Experiment No.	Date	ВС	1111a04			NaOH	MeOH	H₂O	Temp.	Time
Experiment No.	Date	Batch No.	mg	mmol	mg	mmol/eq.	mL	mL	°င	hr
JIBC1111-001	12/4/2013	JIANL-191-1	80	0.13	5	0.13/1.0	1.0	0.134	20	0
JIBC1111-002	12/6/2013	JIANL-191-1	190	0.32	13	0.32/1.0	2.4	0.319	20	0

Results:

Results.				crude product					pure product				
Experiment No.	Batch No.	Date	Aı	nalytical da	te	Qty.	Yield	А	te	Qty.	Yield		
			HPLC	LCMS	NMR	mg	%	HPLC	LCMS	NMR	mg	%	
JIBC1111-001	JIANL-199-001	12/4/2013						AN130221 2940	AN130303 0276	AN130102 5428	67	80.7	
JIBC1111-002	JIANL-206-001	12/6/2013						AN130221 3052	AN130303 0304	AN130102 5575	152	77.2	