

Supplementary Methods

Animals and atrazine exposure. All animal procedures were performed according to the guidelines for animal models in research defined by the Ethics Committee and approved by the Ministry of France (reference project number is 01861.02), and by persons who qualified for laboratory animal care and use. Eight-week-old adult CD1 mice were treated by oral gavage from E6.5 to E15.5 with atrazine in oil at a concentration of 100 mg/kg/day. The control mice were treated with the same volume of oil. The 12 week old F1 male progeny were crossed with unrelated untreated 12-week-old female and progeny of F2 were crossed again to receive F3 male progeny. 12 week-old F1 and F3 ATZ-derived or control-derived male mice were euthanized, and reproductive organs, liver and hypothalamus were dissected. At least five independent experiments were performed with at least six-to-ten animals in each group.

Testosterone and FSH quantification. Serum was collected from ketamine/xylazine-anaesthetized adult animals by terminal cardiac exsanguination, and aliquots were stored at -20°C. Testosterone levels in the serum were assayed in duplicate using a commercial radioimmunoassay (RIA) based on competitive binding with ¹²⁵I-labeled testosterone (Immunotech, Beckman Coulter, Villepinte, France), according to the manufacturer's recommendations. FSH measurements were conducted according to a standard protocol provided by a FSH measurement kit (KA2330, Abnova, Walnut CA, USA). In each hormone measurement, the data from at least 10 treated and control animals were averaged and plotted, and the results were expressed in nanograms per milliliter.

Analysis of histology, apoptosis and germ cells/Sertoli cells numbers. For the histological analysis, testis samples were fixed in Bouin's solution and embedded in paraffin. Histological sections (5- μ m thick) were stained with hematoxylin and eosin (H&E). The staging of cells in seminiferous tubules was performed according to an established method ([Russell 1990](#)), with analysis of at least 600 tubule sections. For immunohistochemistry (IHC), the animals were perfused, and the testes were fixed for 24 hours in 4% (wt/vol) paraformaldehyde (PFA), dehydrated and embedded in paraffin. For IHC, 5- μ m-thick testis sections were incubated overnight at 4°C with goat anti-ZBTB16 (diluted at 1:500) and rat anti-GATA1 (diluted at 1:50). The sections were counterstained with 0.001% (vol/vol) 4,6-diamidino-2-phenylindole dihydrochloride (DAPI) and mounted in Vectashield (Vector Laboratories, UK).

Cells sorting by flow cytometry. Germ cells were isolated from control and ATZ-derived F3 male testes. The testes were subjected to collagenase/DNAse I treatment and purified by FACS, as described previously (Zhu, Shao et al. 2011). The dispersed cells were fixed in 1% (wt/vol) buffered PFA for 20 min at 4°C and permeabilized in a solution of 0.2% (wt/vol) saponin containing propidium iodide to label the DNA. Cells were sorted at concentration of 1X10⁶ cells/ml on FACSCalibur cell-sorting machine. The data were analyzed by CellQuestPro software, which was included with the sorter.

Spermatozoa count. Spermatozoa counts were conducted according to an established protocol ([Vallet-Erdtmann, Tavernier et al. 2004](#)). Briefly, mice were euthanized, and each epididymis was dissected, rapidly frozen in liquid nitrogen and stored at -80°C until the sperm heads were counted as follows. The organ was first cut with a scalpel into several fragments and homogenized in 50 ml 0.15 M NaCl containing 0.005% (vol/vol) Triton X-100 (Sigma). After homogenization with three rounds of sonication (12 kHz), an aliquot of the cell suspension was loaded onto a Malassez hemocytometer, and spermatozoa heads were counted. The data from at least ten controls or F3 ATZ-derived animals were averaged and plotted, and presented as averaged sperm counts normalized to control; a t-test was performed using Excel software.

Antibodies. The following commercial antibodies were used: rat anti-GATA1 (sc-265) and goat anti-PRM2 (sc-23104) antibodies from Santa Cruz, rabbit anti-H3K4me3 (07-473), mouse anti-H2B (05-1352) and rabbit anti-H4K5Ac (07-327) from Millipore, rabbit anti-Ddx4 (ab13840) from Abcam, mouse anti-ACTB (A1978) from Sigma Aldrich and goat anti-PLZF (AF2944) from R&D systems. Secondary HRP antibodies were purchased from Jackson Laboratories. Fluorescent secondary Alexa antibodies were purchased from Invitrogen.

Histone purification and Western blot analysis.

Protein samples from F3 mouse testes were prepared using the EpiSeeker Histone Extraction Kit (Abcam, 113476) according to the manufacturer's supplied protocol. Briefly, mice testes were homogenized and centrifuged at 900 g for 5 min. The pellets were resuspended in lysis buffer and left on ice for 30 min. After centrifugation, the supernatant fractions containing acid-soluble proteins were transferred to new tubes, and the Balance-DTT buffer was added. The protein concentrations were determined using the Pierce™ 660nm Protein Assay (ThermoScientific, France). Five ug (H4K5ac), 10 ug (H2B) or 20 µg (H3K4me3) of protein were run on a 4-20% gradient gel (BioRad, USA) for 1 h. Proteins were transferred onto ImmobilonPSQ membranes (Millipore, France) using an electro-blotter system (TE77X; Hoefer, USA) and modified Towbin buffer (48 mM Tris base, 40 mM glycine and 0.1% (wt/vol) SDS) and methanol (20% (vol/vol) anode; 5% (vol/vol) cathode) for 2 hours. Proteins were detected using an H2B mouse monoclonal antibody, a PRM2 goat polyclonal antibody, an ACTB mouse monoclonal antibody and a H3K4me3 or a H4K5Ac rabbit polyclonal antibody. The antibodies were incubated in hybridization buffer. The primary antibodies were detected using secondary antibodies conjugated to horseradish peroxidase for 1 h. The signals were developed using the ECL-Plus Chemiluminescence kit (GE Healthcare, USA) and the ImageQuant 350 system (GE Healthcare, USA).

Meiotic surface spreads

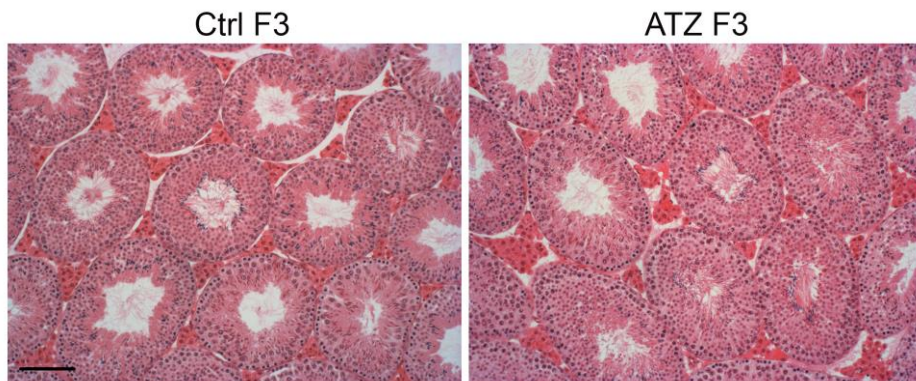
The surface spreads were prepared in 4 independent experiments from F3 control or ATZ progeny as described in a previous study ([Peters, Plug et al. 1997](#)). The analysis of the meiotic substages was performed according to previous studies ([Mahadevaiah, Turner et al. 2001](#), [Page, de la Fuente et al. 2012](#)). The images were taken from randomly chosen pachytene stages of prophase I of meiosis and reanalyzed independently by two researchers.

Fractionation of germ cells

Germ cells were separated according to ([Bellve, 1979](#)). Briefly, 8 C57Bl/6J twelve week-old male mice were perfused with PBS, and testes were dissected. Testes were digested with collagenase, trypsin and DNase. The cells were loaded into 2-4% gradient of BSA and allowed to sediment. The spermatocytes and spermatids fractions were collected. The purity of the fractions was confirmed by DAPI staining of the small cell sample. Cells were fixed with PFA and used for ChIP-seq. The library and data analysis were done similar to analysis of total testis extracts.

Preparation and Immunohistochemistry of embryonic testis histological sections

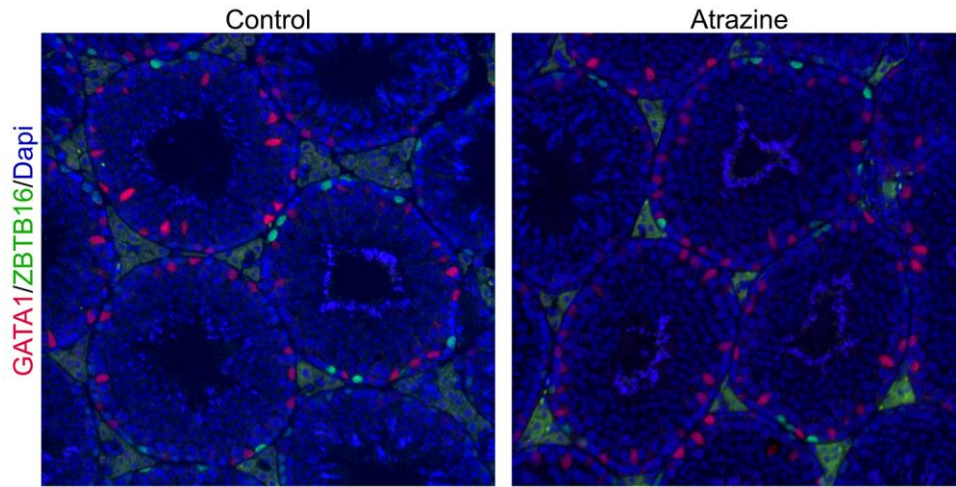
To perform immunohistochemistry in E15.5 and E18.5 embryo, the testis from control or ATZ-treated groups were fixed in PFA 4% (v/v) solution for 16 hours, then were dehydrated and embedded in paraffin. Organs were cut and every 5th sections (5 µm-thick) were taken for a slide. In total, testes from at least 3 animals of each group were used. The sections were deparaffined and rehydrated and slides were placed in 0.01M solution of citrate buffer, pH6 at 80°C for 45min to unmask the epitopes. The sections were washed three times for five minutes in 1X PBS Tween 0.05% (v/v), then incubated with primary antibody anti-Ddx4 diluted PBS-Tween 0.05% (v/v) overnight at 4C ° in a humid chamber. This antibody recognizes all germ cells. The following day the sections were washed with PBS-Tween 0.05% and then incubated with florescent secondary antibody for 1 hour in a humidified chamber. The sections were all counterstained with 0.001 % (vol/vol) 4,6-diamidino-2-phenylindole dihydrochloride (DAPI) and mounted in Vectashield (Vector Laboratories, UK). The images were taken using an AxioImager microscope equipped with an AxioCam MRc5 camera and AxioVision software version 4.8.2 (Zeiss, Le Pecq, France) with a 20X objective lens. To quantify the number of Ddx4-positive cells, we take at least 10 pictures in at least 3 different slides per replicate. We counted the number of Ddx4-positive cells in each tubule present on the picture and we report this number to the surface of seminiferous tubule measure using ImageJ software.



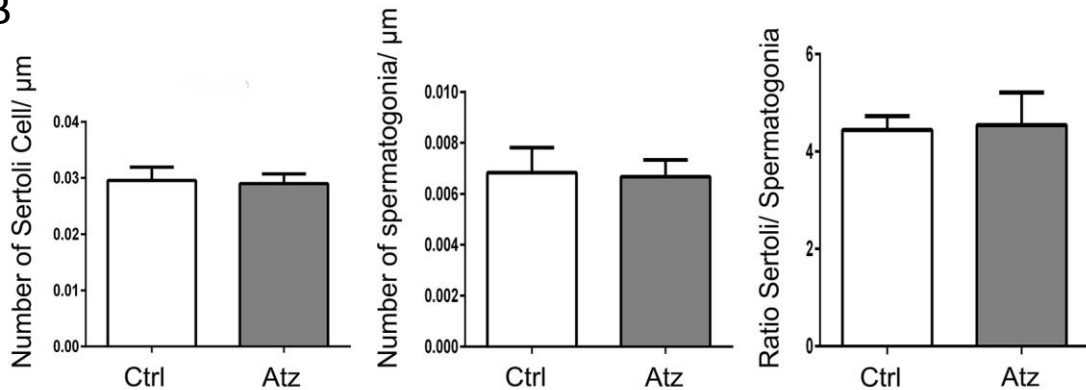
Supplementary_Figure_S1. Embryonic exposure to ATZ does not affect morphology in the third generation of males after treatment.

Representative image of H&E staining of the histological sections of F3 control (vehicle) and ATZ-derived mouse testes (12 weeks old). The sections from testis tissue were prepared as described in the Supplementary Methods section. We compared sections of seminiferous tubules at similar stages in at least 4 different samples of each group control (Ctrl F3) or ATZ (ATZ F3) derived, scale bar:150 μ M.

A

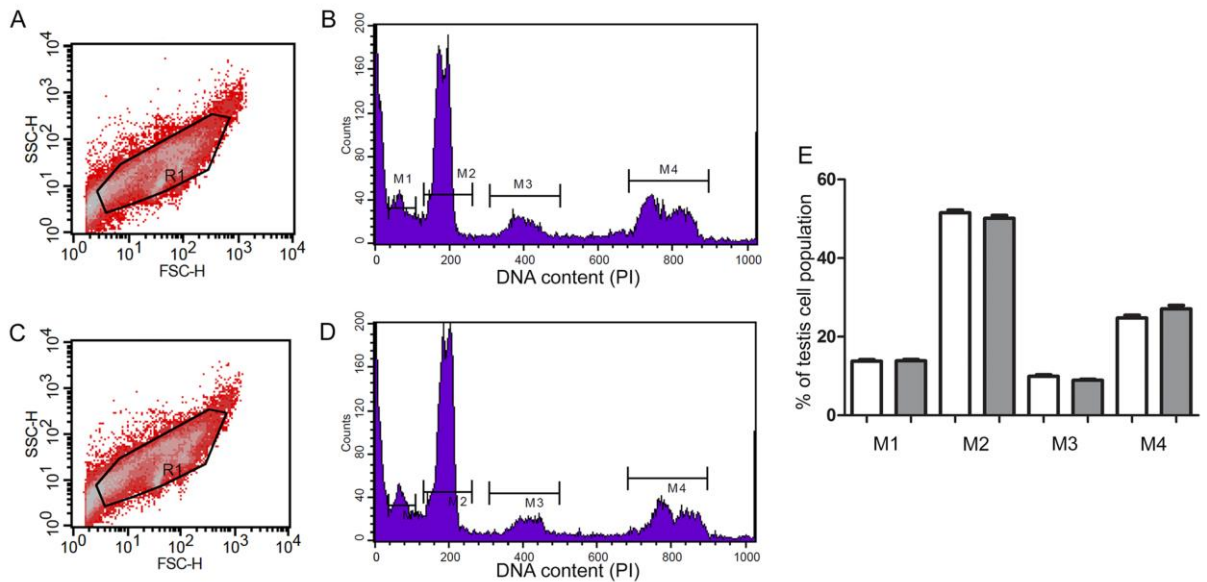


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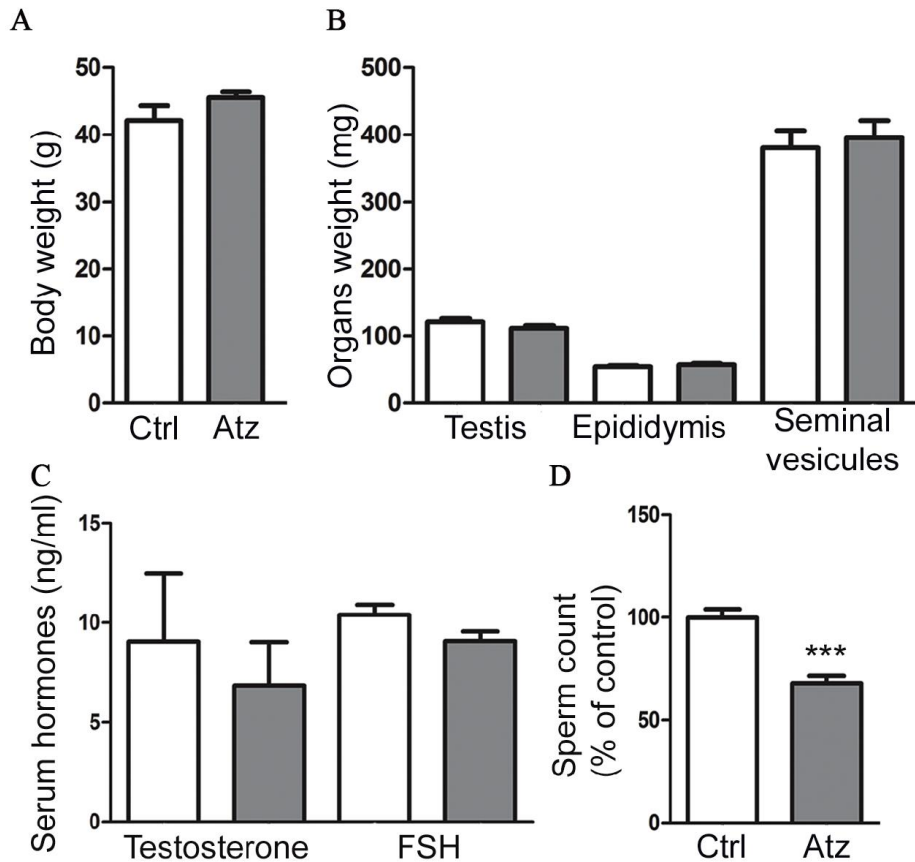
Supplementary_Figure_S2. Embryonic exposure to ATZ does not affect the number of undifferentiated spermatogonia and Sertoli cells in the F3 generations after treatment.

(A) Representative image of testis sections: Sertoli and spermatogonia cells are immunostained with anti- GATA1 or anti- ZBTB16 antibodies. (B) Quantitative analysis: cells, Sertoli cells and spermatogonia were counted manually at stage VII in F3 mouse testes. The contour of each tubule section was measured using the ImageJ software. The values indicate the cell counts per micrometer of tubule circumference. Ratio of the number of Sertoli cells per spermatogonia is also indicated. $n=4$, Scale bar: 50 μm . The immunostaining of testis sections was performed as described in the Supplementary Methods section.



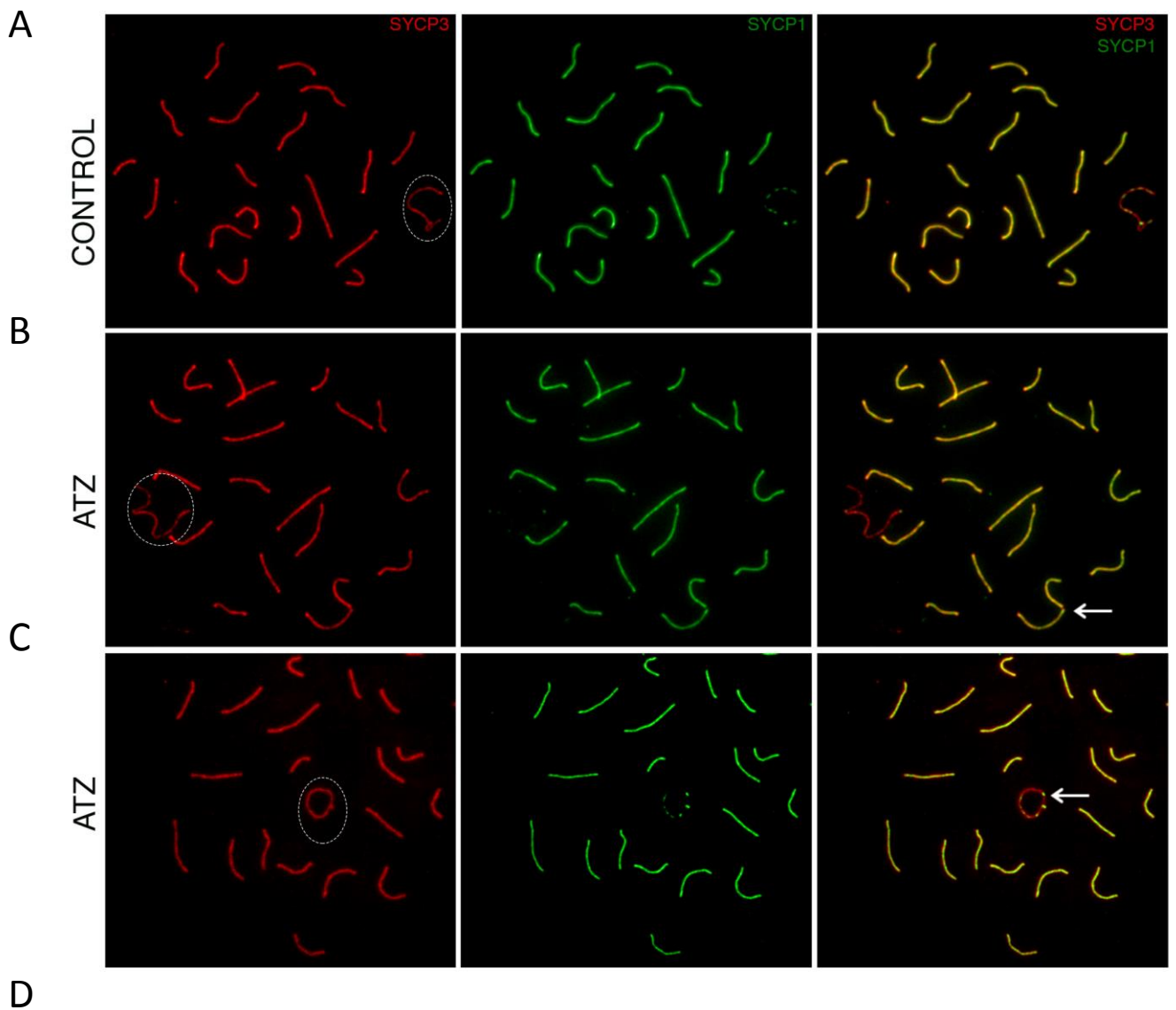
Supplementary_Figure_S3. Embryonic exposure to ATZ does not affect cell populations in seminiferous tubules in the F3 generation of males after treatment.

(A-D) Representative image of a flow cytometric analysis of germ cells from (A-B) control and (C-D) ATZ-derived testes. DNA content of cells was evaluated by propidium iodide incorporation (PI), which discriminates among the four populations: M1, spermatozoa; M2, haploid cells; M3, diploid cells; and M4, tetraploid cells (meiotic cells). (E) Quantitation of the percentage of cell type in each population from F3 control (white bars) and ATZ-derived mice (grey bars); n=8. The details of the flow cytometric analysis of germ cells are described in the Supplementary Methods section.



Supplementary_Figure_S4. Embryonic exposure to ATZ does not affect the weight of reproductive organs but decreases spermatozoa number in the epididymis in the F3 generation of males after treatment.

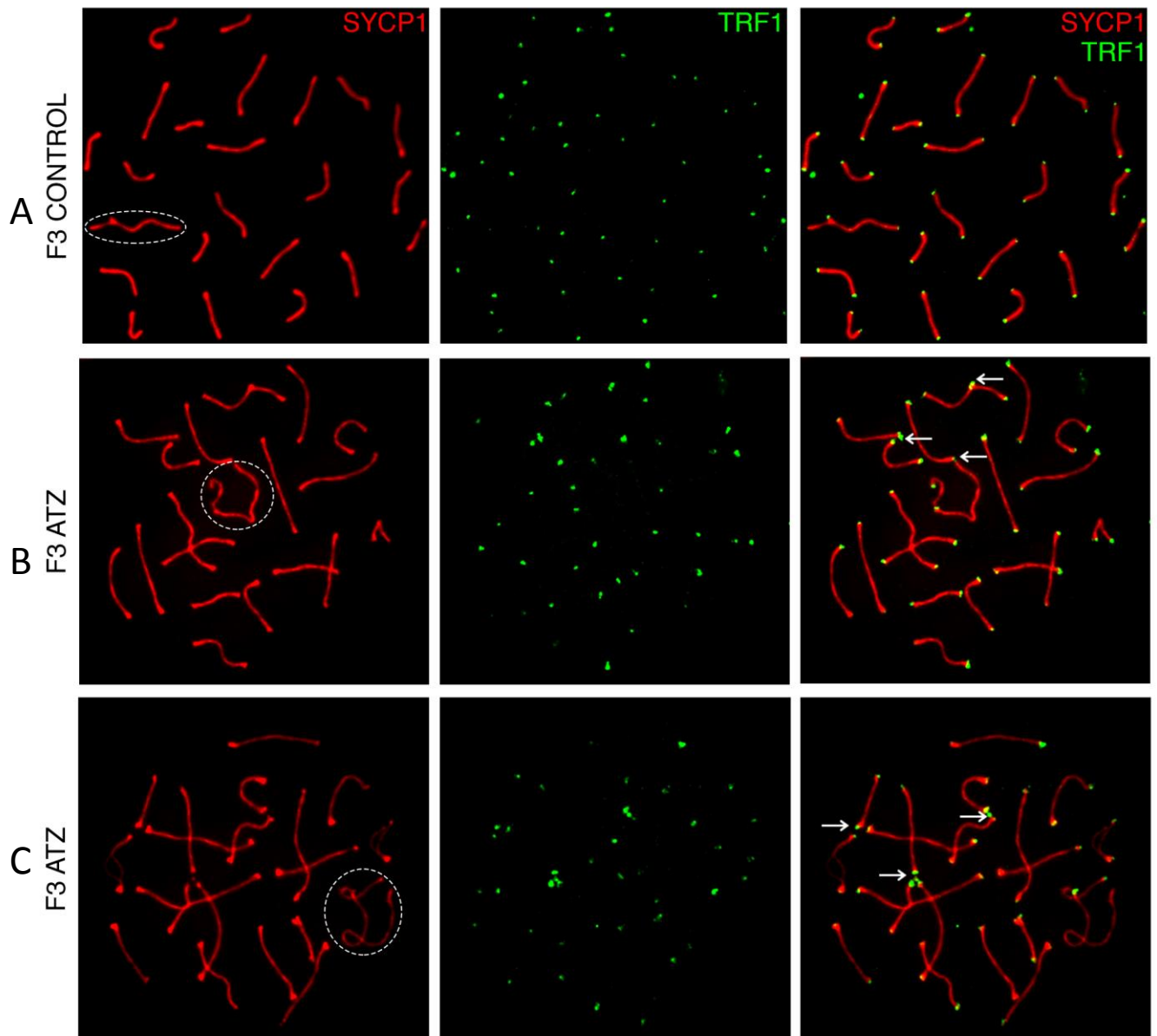
Weight of (A) body and (B) reproductive organs in F3 control (with bar) and ATZ (grey bar) mice ($n \geq 14$). (C), Testosterone and FSH level are not significantly decreased in serum of F3 control and ATZ-derived mice ($n \geq 8$). (D) The number of spermatozoa in the epididymis decreased in F3 ATZ-derived males compared to the control ($n \geq 10$). Significant differences ($P^{***} < 0.001$, t-test) shown by asterisks. The measurements of hormones and spermatozoa counts are described in the Supplementary Methods section.



	Normal SCs	Telomere defects	"ring" sex body	Incomplete synapse
control, n=199	185	10	0	4
ATZ, n=195	156	25	10	4

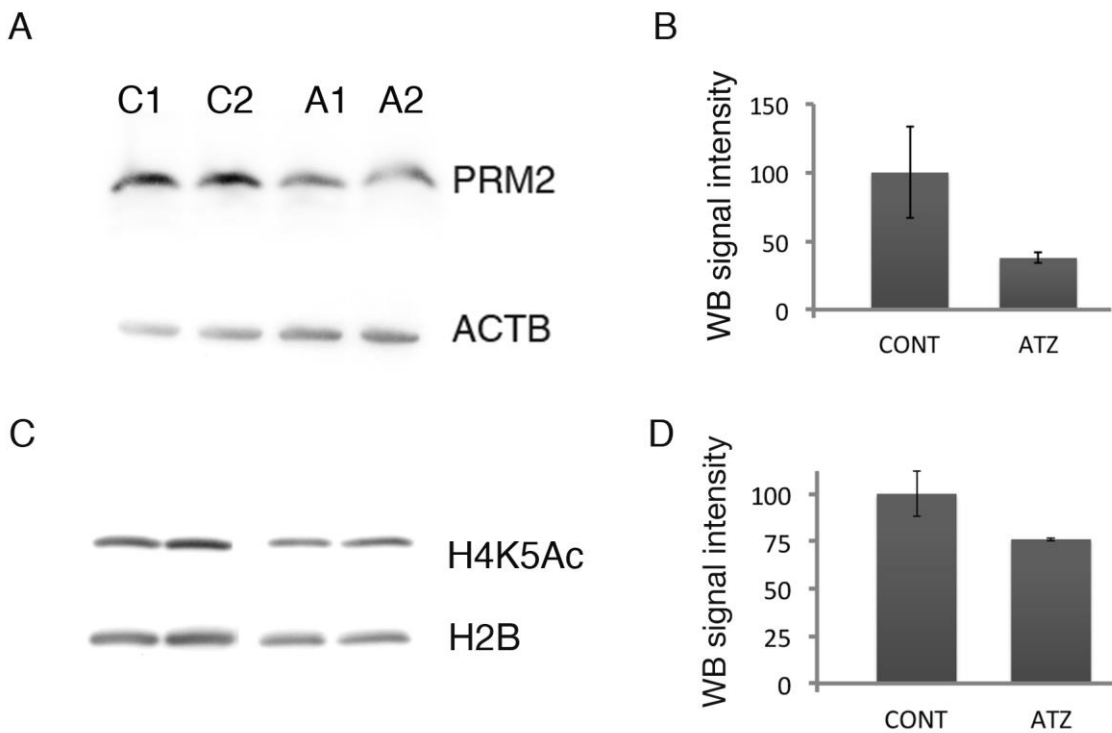
Supplementary_Figure_S5. Embryonic exposure to ATZ affects meiosis in F3 ATZ-derived males.

Surface spreads were prepared from F3 generation testes of the control or ATZ-derived males from 4 independent experiments as described in Supplementary Methods section. The spreads were immunostained against the major protein of lateral element of synaptonemal complex (SC), SYCP3 (in red), and against protein of the SC central element, SYCP1 (in green). (A) Representative images of control testis spread, normally SYCP3 is detected along the length of every chromosomes, SYCP1 is present in all autosomal chromosomes and is visible as punctuated or absent immunostaining pattern in sex chromosomes, (B) In ATZ-derived males the cells with anomalies of the telomere connections are increased (C) the formation in sex chromosomes of the "ring" like structure. X and Y sex chromosomes (sex body) is circled, the arrow shows the chromosomes with abnormal phenotypes. N=199 (control), n=195 (ATZ), $p < 0.01$, Pearson's chi-squared test.



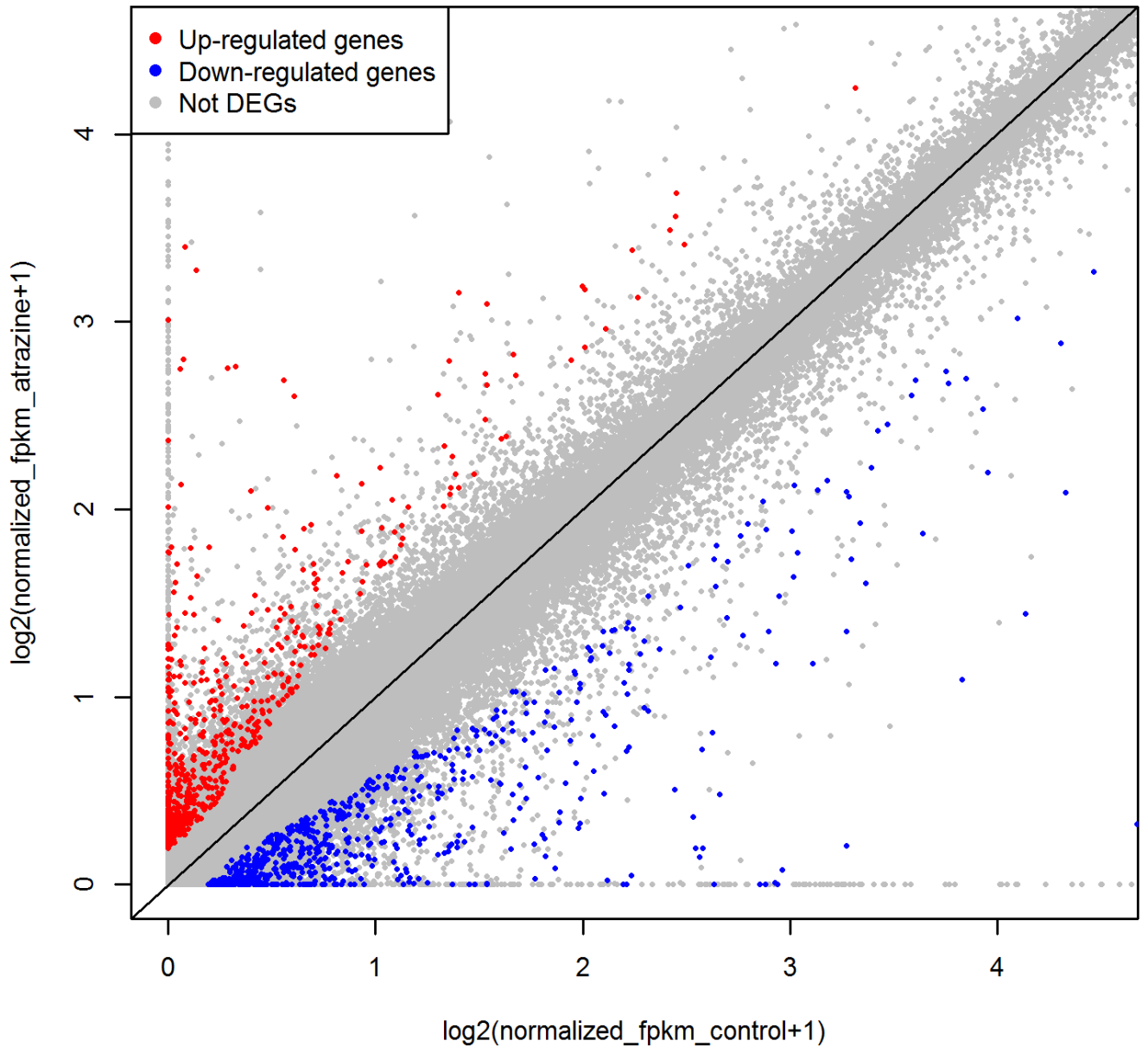
Supplementary_Figure_S6. Embryonic exposure to ATZ causes defects in SC in F3 ATZ-derived males.

Surface spreads were prepared from the testes of the F3 generation of control or ATZ-derived males as described in Supplementary Methods section. The spreads were immunostained against the major protein of synaptonemal complex, SYCP3 (in red), and TERF1 (in green), a telomere –binding protein. (A) In control cells the TERF1 signal is detected at the end of each chromosomes. (B) The telomeres of different chromosomes are connected in cells of ATZ-derived males. (C), The telomeres of different chromosomes were connected in ATZ-derived sample, X and Y sex chromosomes (sex body) are circled, the arrows show the chromosomes with abnormal phenotypes.



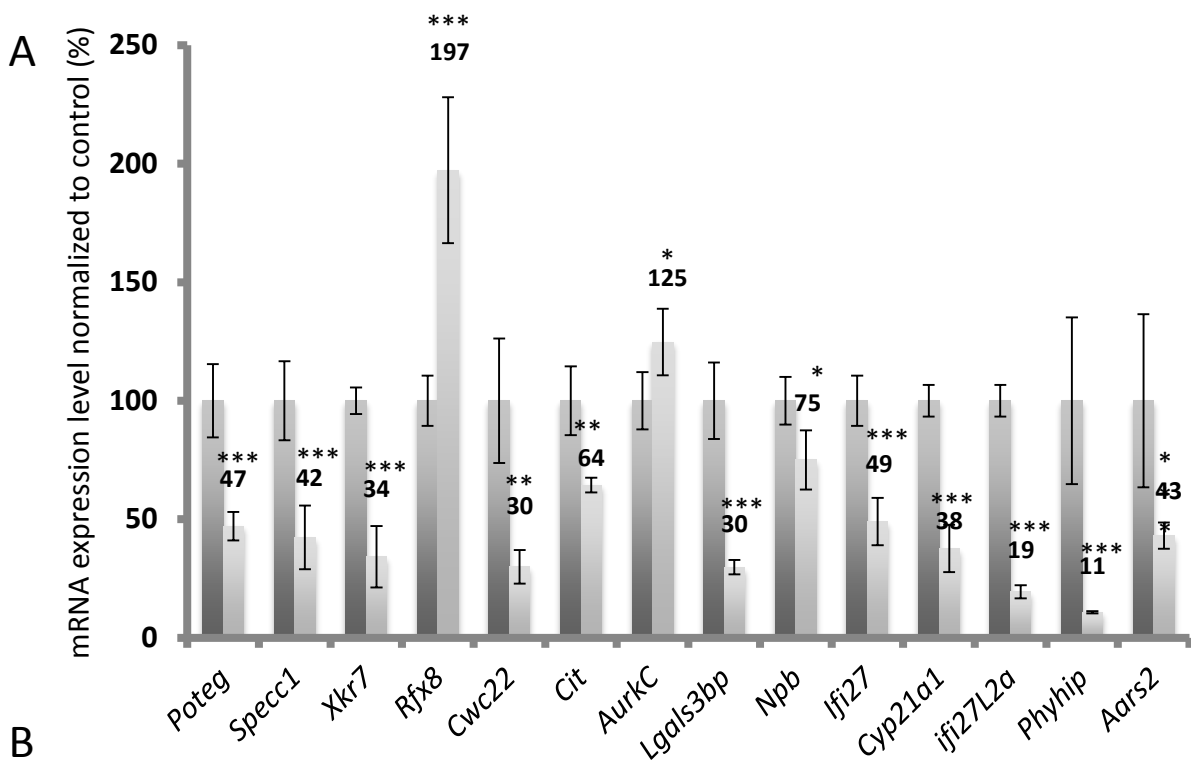
Supplementary_Figure_S7. The PRM2 and H4K5Ac proteins are decreased in F3 ATZ progeny males.

The total protein extracts or purified histone fractions from F3 control or ATZ progeny testes were prepared as described in Supplementary Methods. Two biological replicates of protein extracts were used. (A) The signal of PRM2 intensity was normalized to ACTB, (B) quantitative analysis of the blot in A. (C) The signal of H4K5Ac intensity was normalized to intensity of unmodified histone H2B, (D) quantitative analysis of the Western blot in C. The peak intensities was calculated using Image J software.



Supplementary_Figure_S8: Embryonic exposure to ATZ globally affects gene expression in the testis tissue in third generation of males

The scatter plot of differentially expressed genes. Each dot is the \log_2 mean normalized FPKM (fragments per kilobase of exon per million mapped fragments) value from three biological replicates. Up-regulated transcripts are depicted in red, down-regulated ones in blue and not differentially expressed transcripts are in grey, (619 transcripts are upregulated and 703 downregulated).

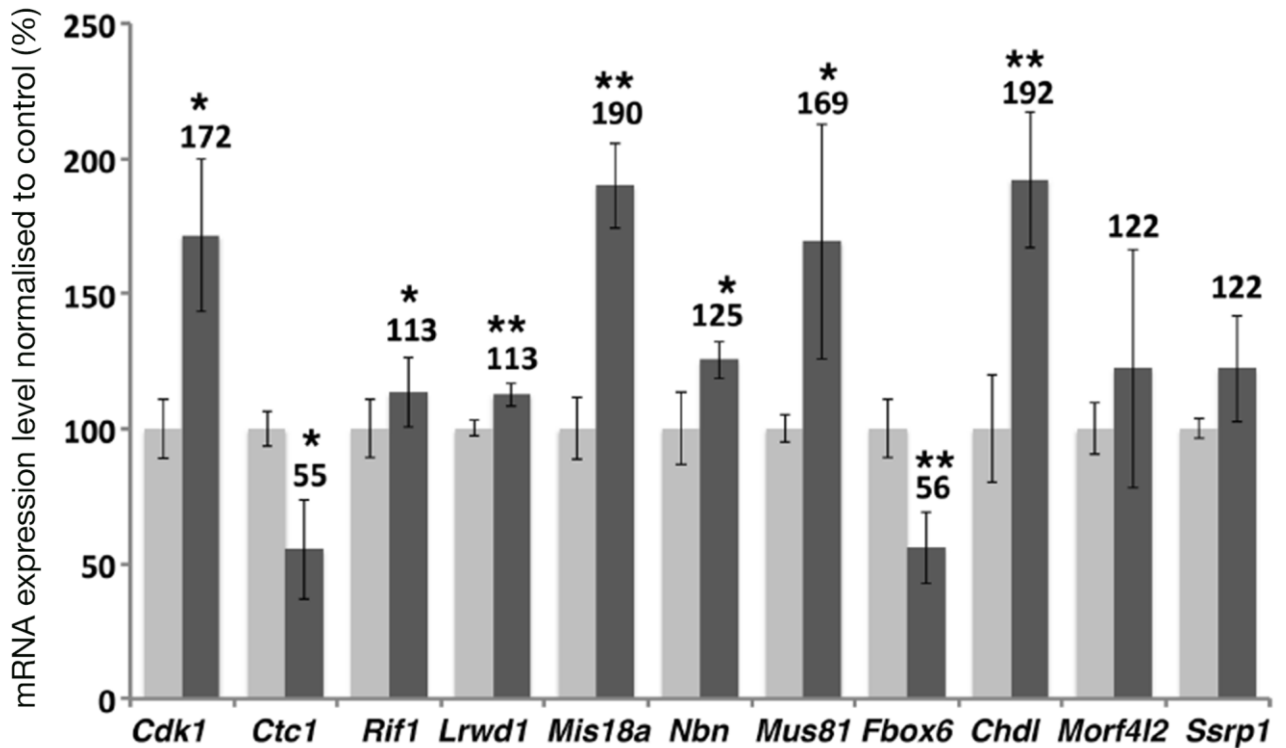


B

gene	RNA-seq FC	FDR RNA-seq	qPCR FC	p-value qPCR	process
<i>Poteg</i>	0.49	0.013	0.47	0.00346	Spermatogenesis
<i>Specc1</i>	0.46	0.017	0.42	0.00196	Spermatogenesis, role in cancer
<i>Xkr7</i>	0.48	0.050	0.34	0.00066	ND
<i>Rfx8</i>	0.42	0.001	0.42	0.00503	Transcription
<i>Cwc22</i>	0.27	0.019	0.30	0.01010	Splicing
<i>Cit</i>	0.00	0.025	0.64	0.01406	Spermatogenesis
<i>AurkC</i>	3.48	0.023	1.25	0.03762	Spermatogenesis
<i>Lgals3bp</i>	0.22	0.008	0.30	0.00256	Cell adhesion
<i>Npb</i>	0.47	0.044	0.75	0.02218	G-protein coupled receptor b
<i>Ifi27</i>	0.33	0.019	0.49	0.00044	Regulation of transcription
<i>Cyp21a1</i>	0.23	0.034	0.38	0.00011	C21-steroid hormone biosynthetic
<i>Ifi27L2a</i>	0.10	0.049	0.19	0.00503	Immune response
<i>Phyhip</i>	0.01	0.032	0.11	0.00002	Mitophagy
<i>Aars2</i>	0.45	0.040	0.43	0.01473	Mitochondrial respiratory chain complex assembly
<i>Spin2d</i>	7.59	0.007	10.69	0.05092	ND
<i>Crtam</i>	>10	0.042	4.71	0.00878	Regulation of immune response

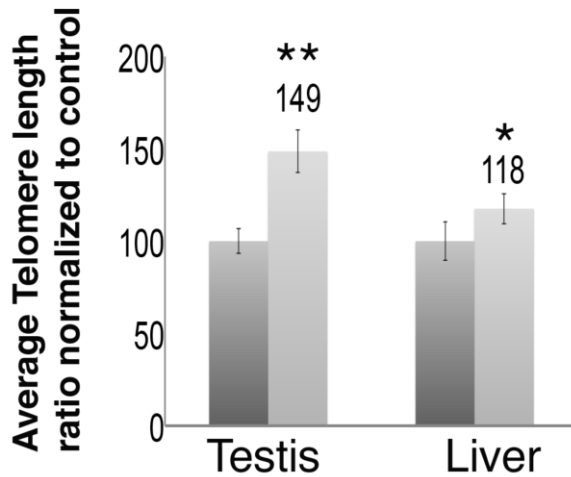
Supplementary_Figure_S9. Comparison of RNA-seq data and qPCR gene expression data for mRNA analysis.

(A) Differentially expressed genes identified by RNA-seq and confirmed by qPCR. Total RNA was extracted, cDNA isolated and qPCR performed as described in the Methods section. Primer sequences are indicated in Supplemental_Table_S12. At least 4 independent experiments were performed and averaged (* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, t-test). Control samples are in dark grey, ATZ-derived in light grey. (B) Comparison of gene expression determined by RNA-seq and qPCR data.



Supplementary_Figure_S10. Embryonic exposure to ATZ affects the expression of several genes belonging to the DNA damage pathway.

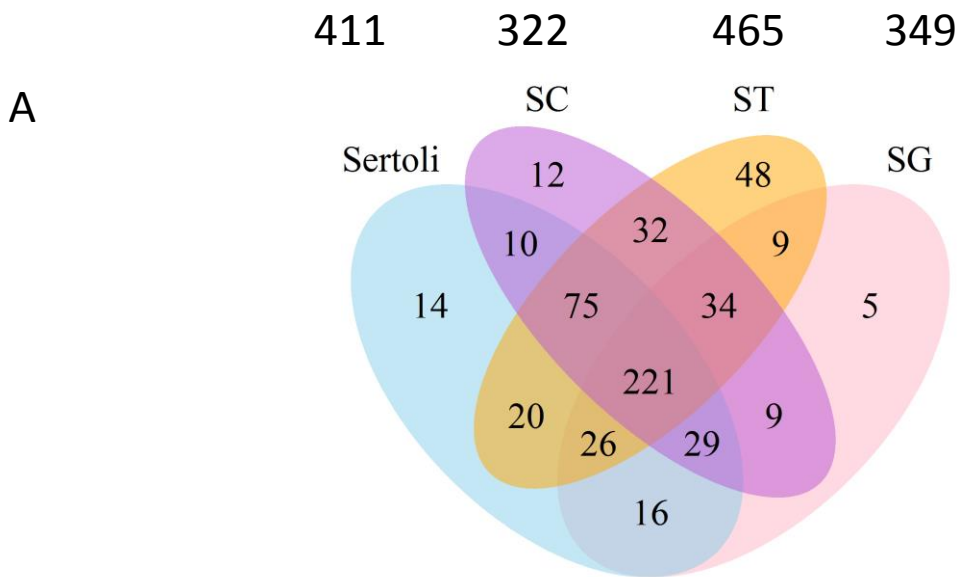
QPCR confirmation of differentially expressed genes of DNA damage response. The details of the experiments are described in the Methods section. The data represent the average of at least 4 independent experiments and are expressed as mRNA expression level in the experimental sample compared to the control (%) ($p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, t-test). The primer sequences are indicated in Supplemental_Table_S12.



Supplementary_Figure_S11. Embryonic exposure to ATZ affects the telomeres length.

Genomic DNA was isolated from testis or liver tissues using Proteinase K and phenol: chloroform extraction. Concentration of DNA was estimated by fluorescent method. Equal amounts of DNA were used for qPCR. The primers sequences are indicated in Supplementary_Table_S12.

Data from at least 6 independent experiments were averaged and expressed as the average telomere length (ATLR) normalized to control, (* $p < 0.05$, ** $p < 0.01$, t-test).

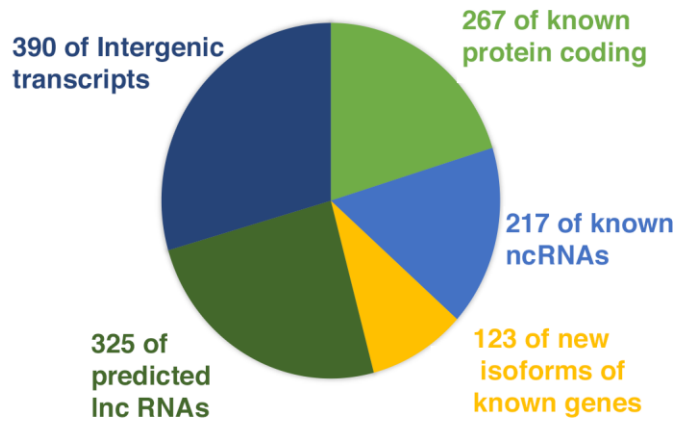


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Gene symbol	Gene Name
<i>2810474O19Rik</i>	RIKEN cDNA 2810474O19 gene
<i>Acp6</i>	acid phosphatase 6, lysophosphatidic
<i>Aox3</i>	aldehyde oxidase 3
<i>Esrrb</i>	estrogen related receptor, beta
<i>Flnb</i>	filamin, beta
<i>Gatad2b</i>	GATA zinc finger domain containing 2B
<i>Gtf2i</i>	general transcription factor II I
<i>Hsp90aa1</i>	heat shock protein 90, alpha
<i>Islr2</i>	immunoglobulin superfamily containing leucine-rich repeat 2
<i>Manba</i>	mannosidase, beta A, lysosomal
<i>Mcf2</i>	mcf.2 transforming sequence
<i>Ncor1</i>	nuclear receptor co-repressor 1
<i>Prrc2b</i>	proline-rich coiled-coil 2B
<i>Rfc1</i>	replication factor C (activator 1) 1
<i>Rfc5</i>	replication factor C (activator 1) 5
<i>Rif1</i>	replication timing regulatory factor 1
<i>Ssrp1</i>	structure specific recognition protein 1
<i>Sumo3</i>	small ubiquitin-like modifier 3
<i>Esrrb</i>	estrogen related receptor, beta

Supplementary_Figure_S12: The origin of differentially expressed testis genes in the third generation of males after treatment.

A). The differentially expressed genes were compared with gene expression in different cell types from previously published dataset (Gan, Wen et al. 2013). Each oval represents a fraction, Sertoli (Sertoli cells), SC (spermatocytes), ST (spermatids), SG (Spermatogonia). A large number of genes (221) are shared between all cell fractions. The highest number (48) of unique differentially expressed genes are found in spermatid fraction. B). DEGs from spermatogonia fraction are known to be interacting with POU5F1.



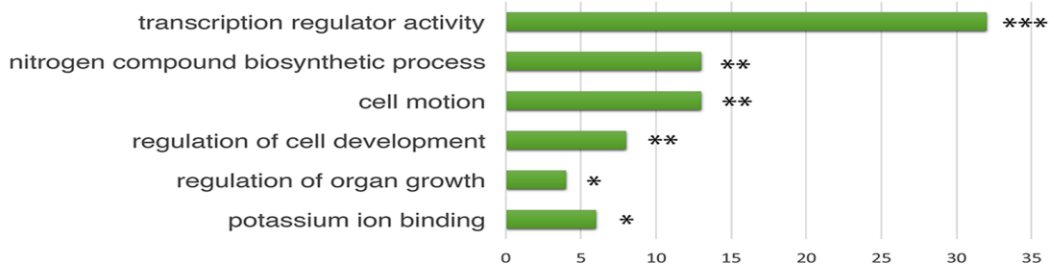
Supplementary_Figure_S13. Group of transcripts that are differentially expressed in the F3 generation of the ATZ -derived males.

The pie chart shows the distribution of the differentially expressed RNA types in the testis tissue of F3 ATZ-derived males. The RNA-Seq data were analyzed as described in the Methods section.

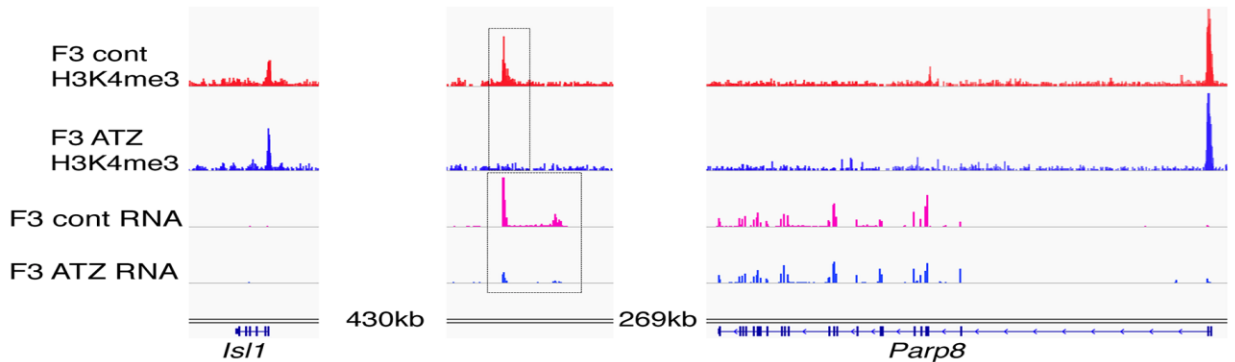
A

Term Name	Number of genes	P-Value	Genes
transcription regulator activity	32	1.77E-04	<i>Sox2, Sox5, Sox14, Meis1, Max, Lbh, Olig3, Gata3, Gata4, Hey2, Foxb2, Atoh8, Foxd1, Tlx1, Klf5, Klf12, Tbx3, , Nr4a2, Rybp, Tead1, Ldb2, Arntl, Isl1, Med13l, Ppargc1a, Med10, Mef2d, Hhex, Ets1, Vgll3, Prdm2, Rbpj</i>
nitrogen compound biosynthetic process	13	1.07E-03	<i>Bcat1, Dctd, Adss, Nampt, Adcy1, Atp11b, Nr4a2, Atp5g3, Cmpk2, Gm4953, Gata3, Mat2b, Tph2</i>
cell motion	13	5.32E-03	<i>Fut8, 4921509c19rik, Plxna2, Gm14147, Nr4a2, Tnp1, Isl1, Kitl, Alcam, Tns1, Bmp7, Foxd1, Ch11</i>
regulation of cell development	8	6.82E-03	<i>Tbx3, Ntrk2, Sox2, Sox5, Isl1, Bmp7, Meis1, Ngf</i>
regulation of organ growth	4	1.06E-02	<i>Gata4, Hey2, Cxadr, Bcl2l1l</i>
potassium ion binding	6	1.90E-02	<i>Kcnma1, Slc24a3, Kcnh7, Kcnj3, Kcnip4, Kcnv1</i>

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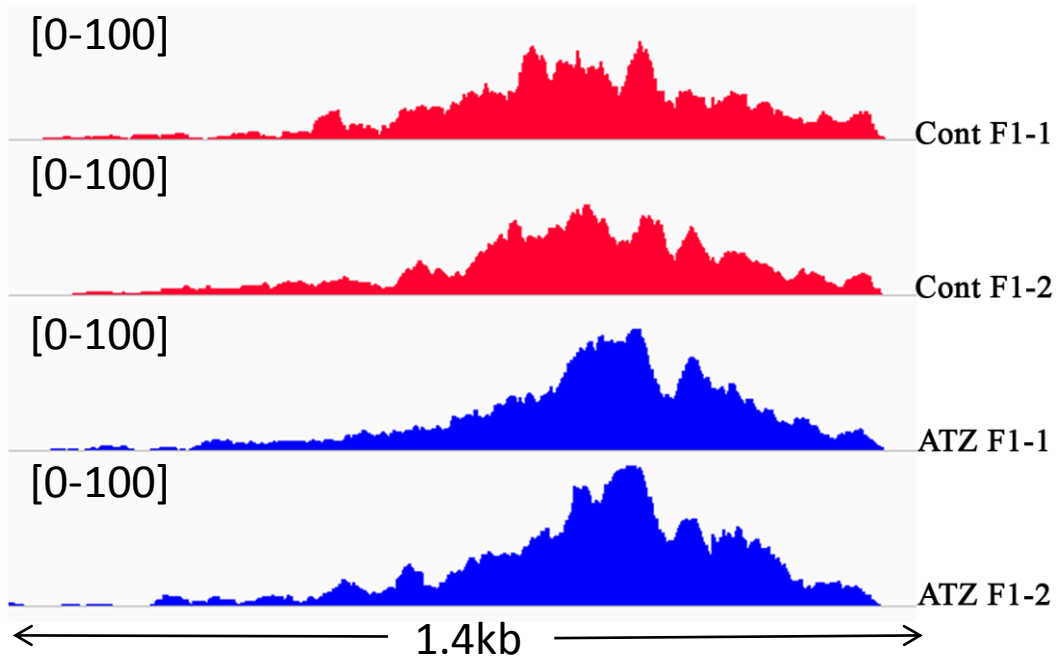
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Supplementary_Figure_S14: The expression of the new long noncoding RNA is globally affected in the F3 generation.

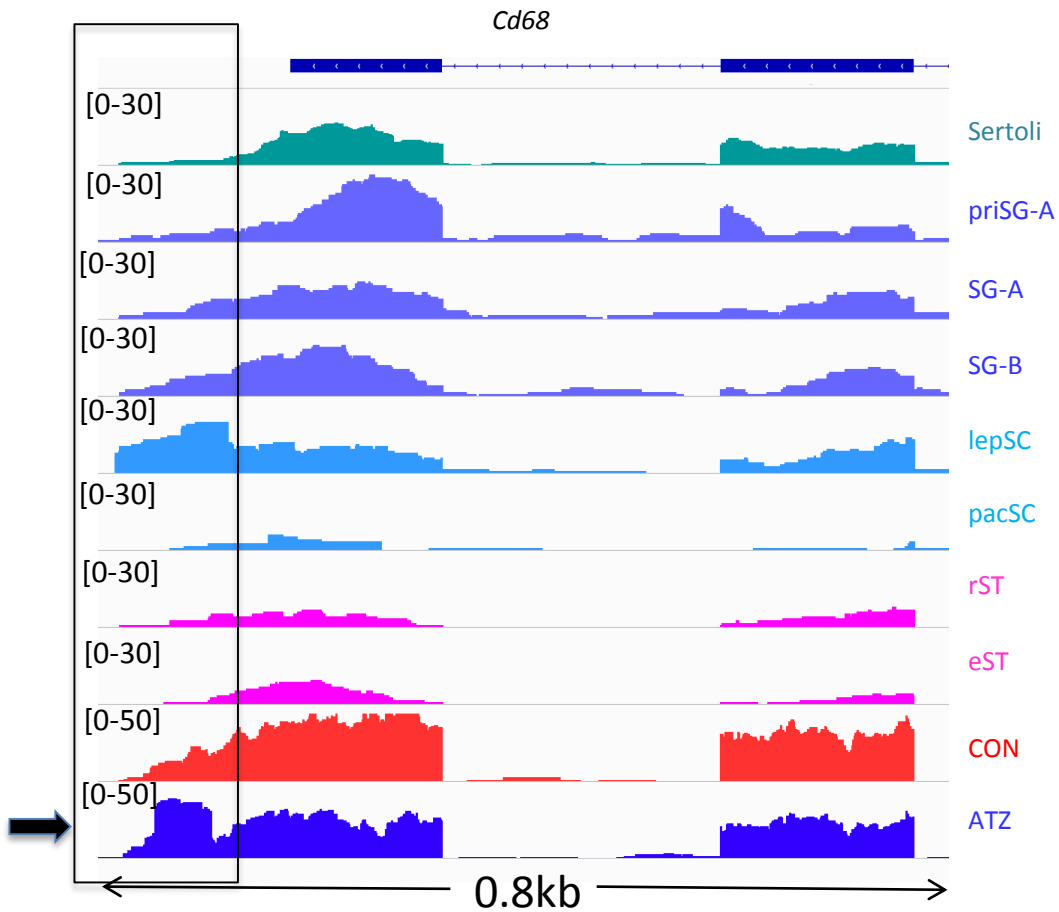
(A) Gene Ontology (GO) term enrichment of genes located upstream and downstream of LincRNA. GO terms were sorted based on p-values (* $P < 0.05$; ** $P < 0.01$, *** $P < 0.001$, Fisher exact test). (B) The plot represents the number of genes in each GO category. (C) A new predicted long noncoding RNA is located distally from the *Isl1* and *Parp8* genes. Both the expression of noncoding RNA and the H3K4me3 occupancy in the promoter are dramatically decreased. H3K4me3 peaks and RNA expression were determined as described in the Methods section and represent the averaged and normalized values of two (ChIP-seq) or three (RNA-seq) biological replicates for the F3 control (in red) or ATZ-derived (in blue) males testes.

Adat1 H3K4me3



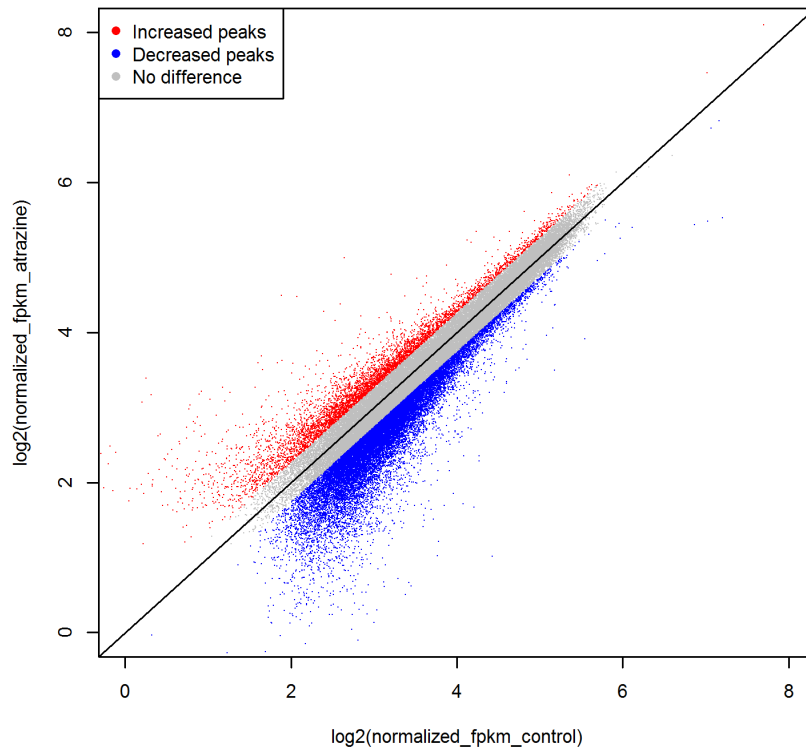
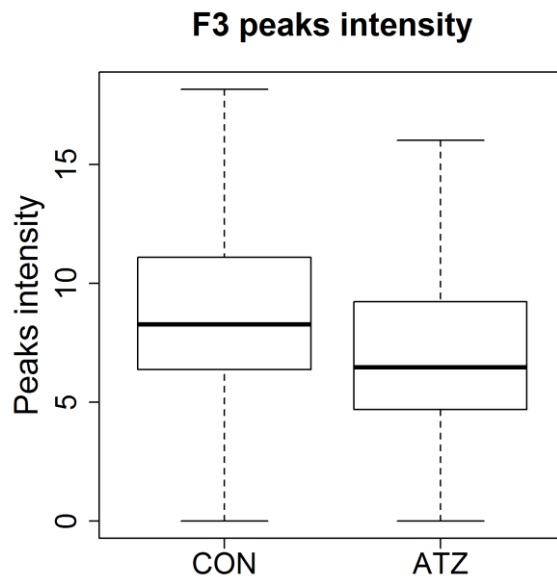
Supplementary_Figure_S15. Embryonic exposure to ATZ affects the H3K4me3 regions in promoters of *Adat1* gene in F1 ATZ-derived males.

Histone H3K4me3 occupancy is changed in the promoter of the *Adat1* gene in the testes of the F1 generation of ATZ-derived males. Each plot represents the normalized number of tags in the control (red) and ATZ-derived samples (blue). Two biological replicates are shown for the control and the ATZ-derived animals. The IGV genome viewer v 2.3.36 was used to visualize the ChIP-seq data for the *Adat1* gene.



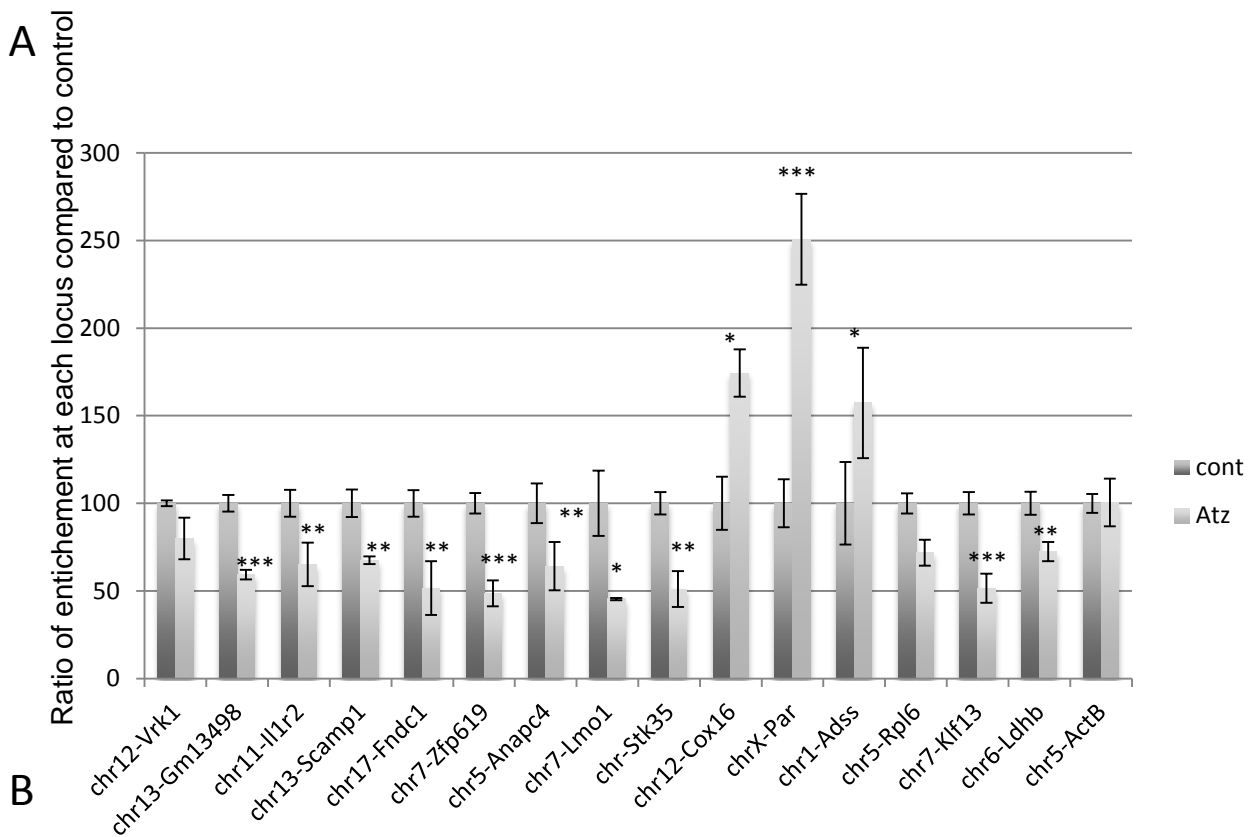
Supplementary_Figure_S16. The altered APA transcript of the *Cd68* gene was identified in F3 ATZ progeny males.

The RNA profiles from different cell types based on previous work were reanalyzed (Gan, Wen et al. 2013). The *CD68* gene has only one transcript. The altered polyadenylated site is shown by an arrow. Three biological replicates of RNA-seq data in *Cd68* transcripts were merged for simplicity in the control (red) and ATZ-derived (blue) transcripts and are represented as averaged and normalized tag numbers. The direction of transcription is on the opposite strand. priSG-A: primitive type A spermatogonia; SG-A: type A spermatogonia; SG-B: type B spermatogonia; lepSC: leptotene spermatocytes; pacSC: pachytene spermatocytes; rST: round spermatids; eST: elongative spermatids; CON: F3 testis, control; ATZ: F3 testis, atrazine. The IGV genome viewer v 2.3.36 was used to visualize the RNA-seq data.

A**B**

Supplementary_Figure_S17. The scattered plot and boxplot of H3K4me3 peaks with FC above 1.2.

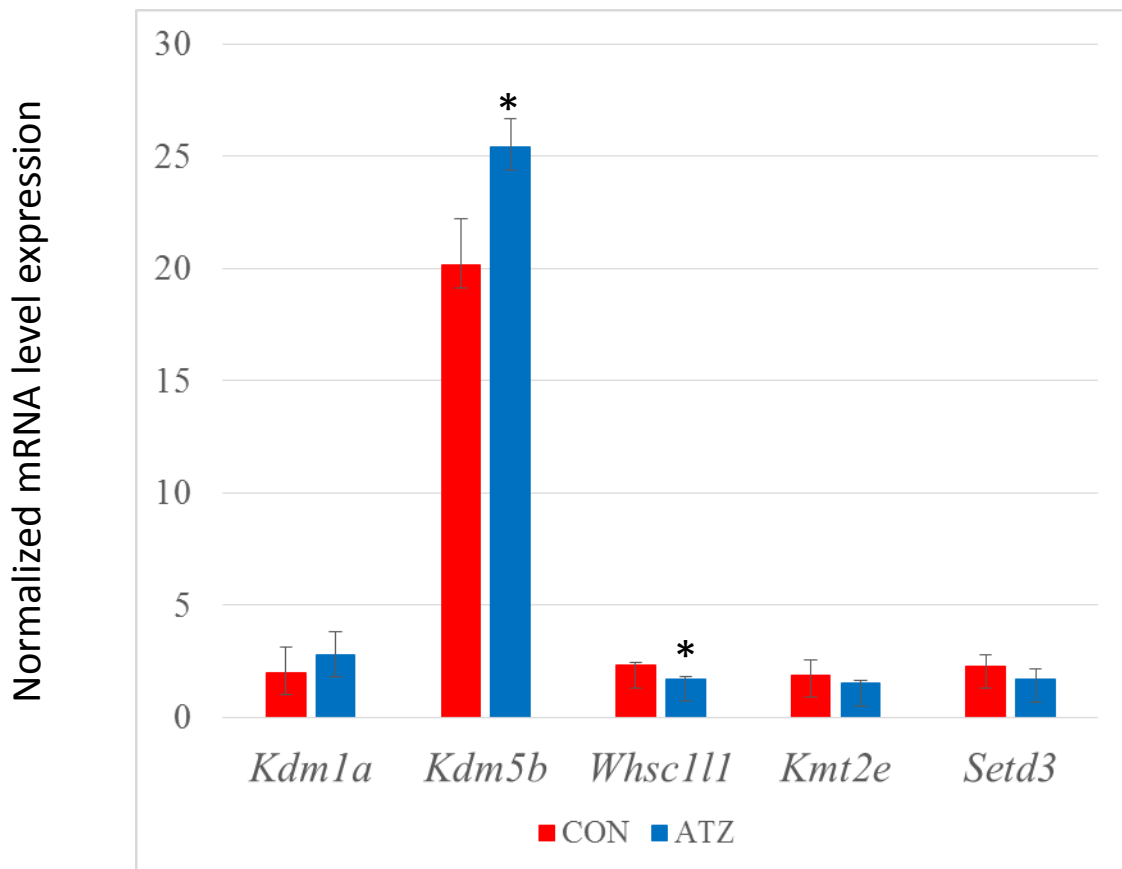
(A) The scattered plot of differential peaks. With fold change above 1.2, we found 5010 peaks have increased and 19479 peaks have decreased H3K4me3 occupancy. Each dots represents the log2 of mean normalized FPKM value from two biological replicates. The increased occupancy peaks are depicted in red, decreased occupancy peaks are in blue, peaks with no changes in H3K4me3 occupancy are shown in grey color. (B) Boxed plot of H3K4me3 peaks intensity



Supplementary_Figure_S18. Comparison of ChIP-seq and ChIP-qPCR data.

(A) ChIP-qPCR was performed as described in the Methods section. The primer sequences are indicated in Supplemental_Table_S12. The average of at least 4 independent experiments is plotted and expressed as the ratio of enrichment of ChIP product compared to Input and normalized to the control, * $p < 0.05$, ** $p < 0.01$, ***, $p < 0.001$, t-test. The control samples are in dark grey, the ATZ-derived in light grey.

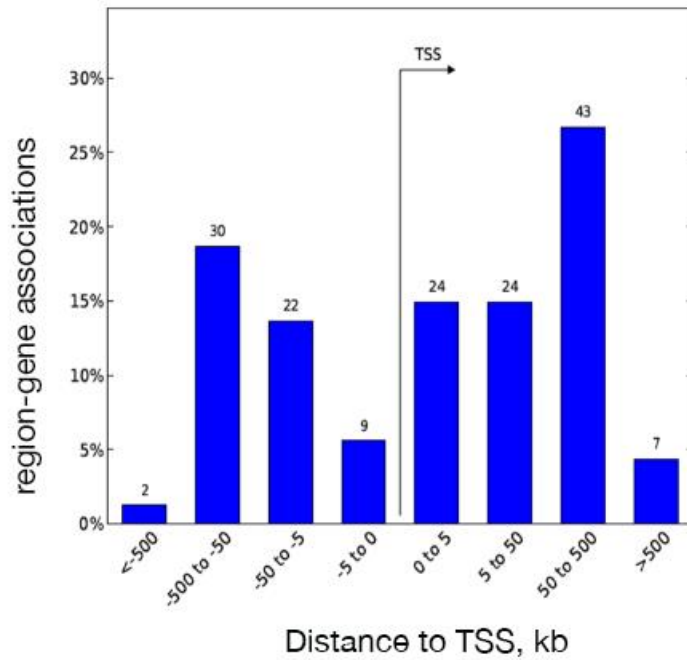
(B) Comparison of ChIP-seq and ChIP-qPCR data



Supplementary_Figure_S19. The expression level of histone H3 lysine 4 methyltransferase mRNA (*Whsc1l1*) is decreased and the expression levels of histone H3 lysine demethylases (*Kdm5b*) is increased in the testes of F3 ATZ-lineage males.

The expression levels were determined using RNA-seq data. Each plot are represented as the averaged FPKM value of three biological replicates. FPKMs were calculated using the tag number. The control samples are in red, ATZ-derived samples in blue, * $p < 0.05$, t-test.

A



B

GO Biological Process

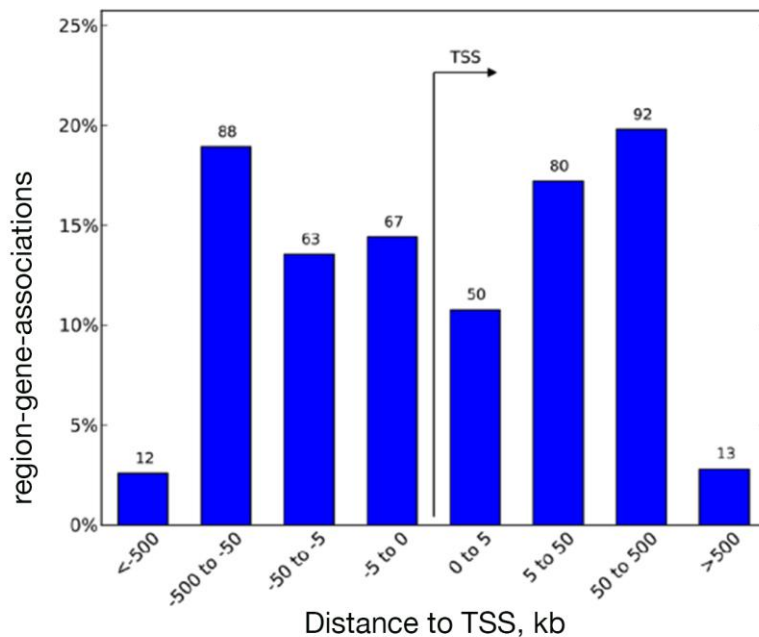
Binomial Region Hits

GO Biological Process	0	1	2	3	4	5	6	7	8	9	10	11	Genes
epithelial cell development													<i>Slc26a3, Dicer1, Trip13, Mei1, Rapgef3, Sod1, Dmrt1, Sox18, Shroom3, Foxc2, Bbs4</i>
anatomical structure homeostasis													<i>Mapt, Dicer1, Fgfr1op, Spry1, Crocc, Bbs4</i>
microtubule cytoskeleton organization													<i>Mapt, Dicer1, Fgfr1op, Spry1, Crocc, Bbs4</i>
negative regulation of transferase activity													<i>Fgfr1op, Spry1, Hexim2, Pif1, Zfyve28</i>
multicellular organismal homeostasis													<i>Bbs4, Abca4, Arrdc3, Gpr98, Il20rb, Sod1</i>
cellular component disassembly													<i>Apaf1, Arid2, Mtlf3, Mtrf1</i>
inorganic anion transport													<i>Ano6, Gabrb3, Slc26a3, Slc34a2</i>
nucleic acid phosphodiester bond hydrolysis													<i>Apaf1, Dicer1, Dnase2a, Endou, Wm</i>
inner ear receptor cell differentiation													<i>Cux1, Dicer1, Gabrb3, Gpr98, Sod1</i>
tissue homeostasis													<i>Gpr98, Abca4, Bbs4, Sod1</i>
mechanoreceptor differentiation													<i>Gpr98, Cix1, Dicer1, Sod1, Gabrb3</i>
response to insulin stimulus													<i>Foxc2, Ggh, Ipin1, Ptpn1, Sh2b2</i>
meiotic cell cycle													<i>Spin1, Mei1, Spdy1, Trip13</i>
neural tube closure													<i>Apaf1, Bbs4, Shroom3, Zic5</i>
ATP catabolic process													<i>Abca4, Abca6, Abcc4, Wm</i>
primary neural tube formation													<i>Apaf1, Bbs4, Shroom3, Zic5</i>
tube closure													<i>Apaf1, Bbs4, Shroom3, Zic5</i>
purine ribonucleoside monophosphate catabolic process													<i>Abca4, Abca6, Abcc4, Wm</i>
purine nucleoside monophosphate catabolic process													<i>Abca4, Abca6, Abcc4, Wm</i>
nucleoside monophosphate catabolic process													<i>Abca4, Abca6, Abcc4, Wm</i>

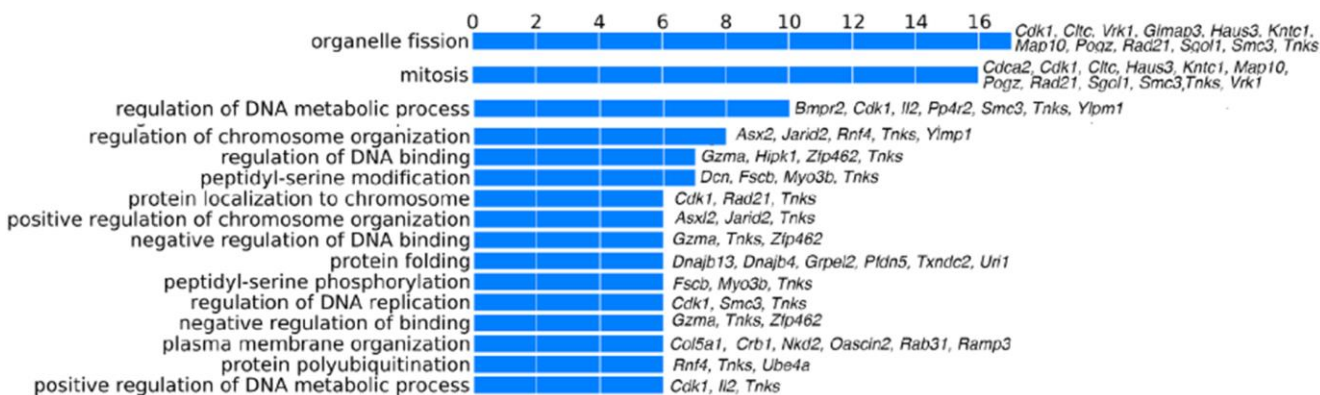
Supplementary_Figure_S20. Differential H3K4me3 peaks in spermatocyte fraction in testis of F3 ATZ-derived males are enriched in regions of genes involved in epithelial cell development and cytoskeleton organization

Differential H3K4me3 peaks were compared with normalized peaks intensity from purified spermatocytes or spermatids. (A) The distance of differential peaks to TSS of the genes, (B) The subset of differential H3K4me3 peaks with high signal in spermatocytes were annotated with GREAT. The GO term of biological function is shown on the left, the corresponding genes are on the right.

A



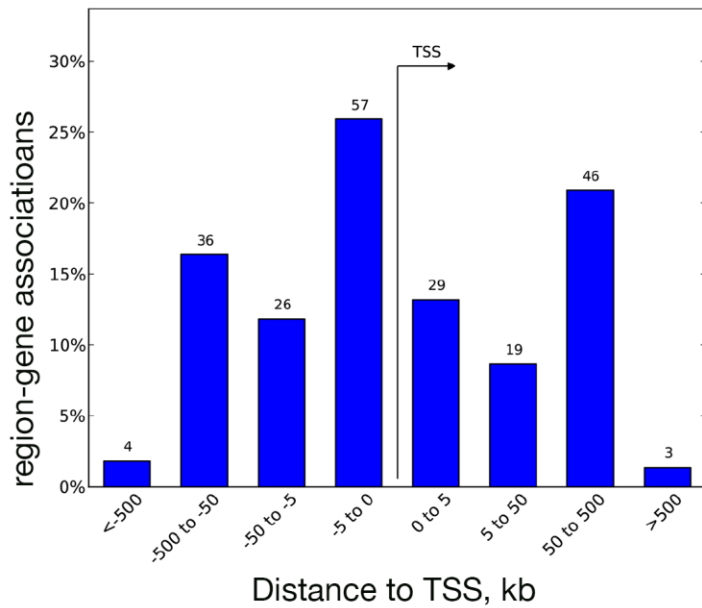
B



Supplemental_Figure_S21. Differential H3K4me3 peaks in spermatocyte fraction in F3 ATZ-derived males are enriched in regions of genes involved in mitosis.

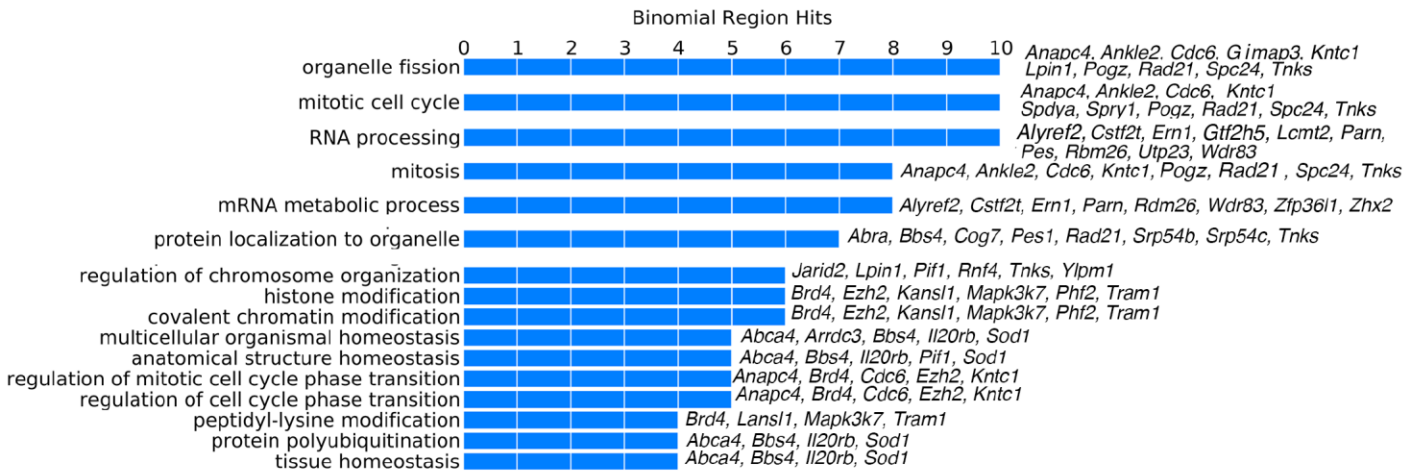
Differential H3K4me3 peaks were compared with normalized peaks intensity from purified spermatocytes or spermatids. (A) The distance of differential peaks to TSS of the gene. (B) The subset of h3K4me3 peaks with high signal in spermatids were annotated with GREAT. The GO term of biological function is shown on the left, the corresponding genes are on the right.

A



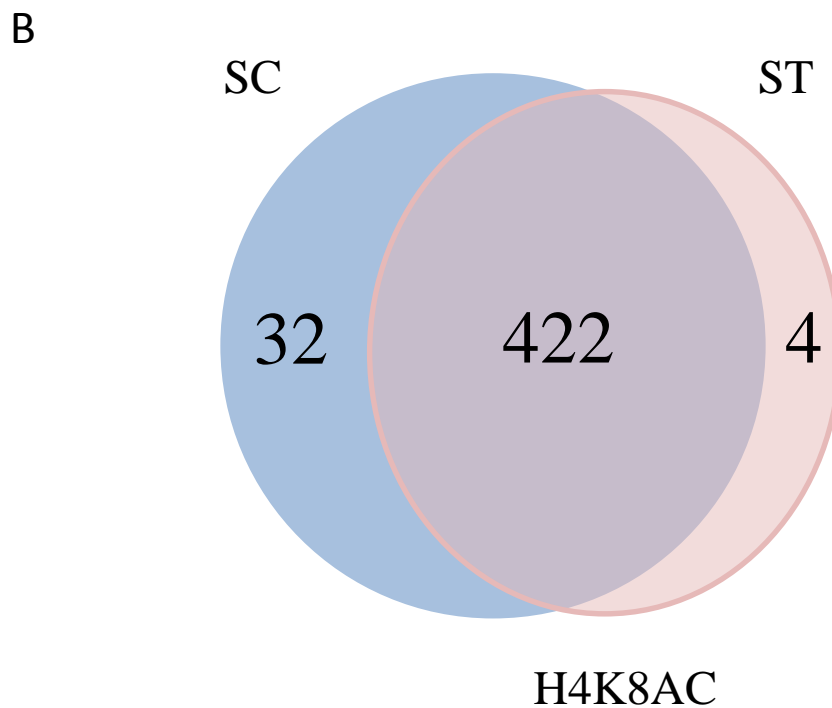
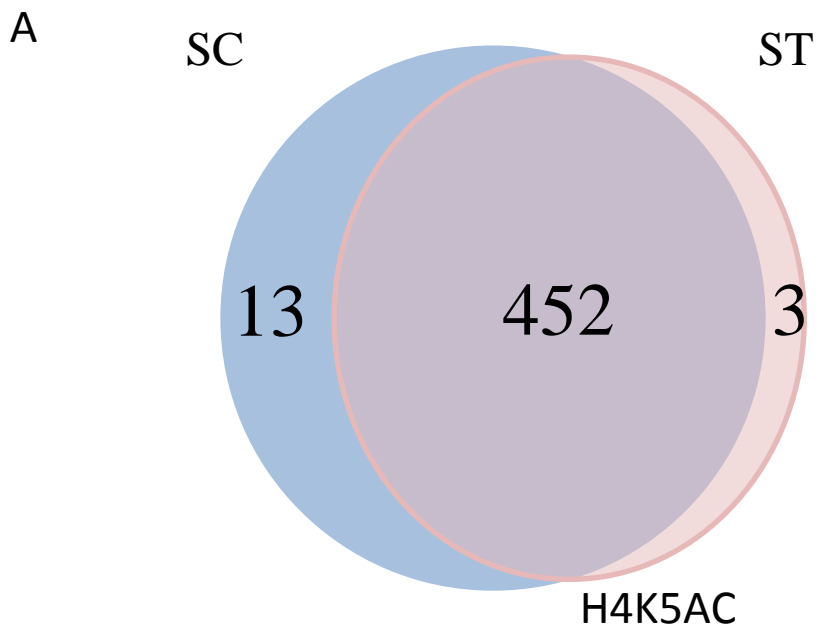
B

GO Biological Process



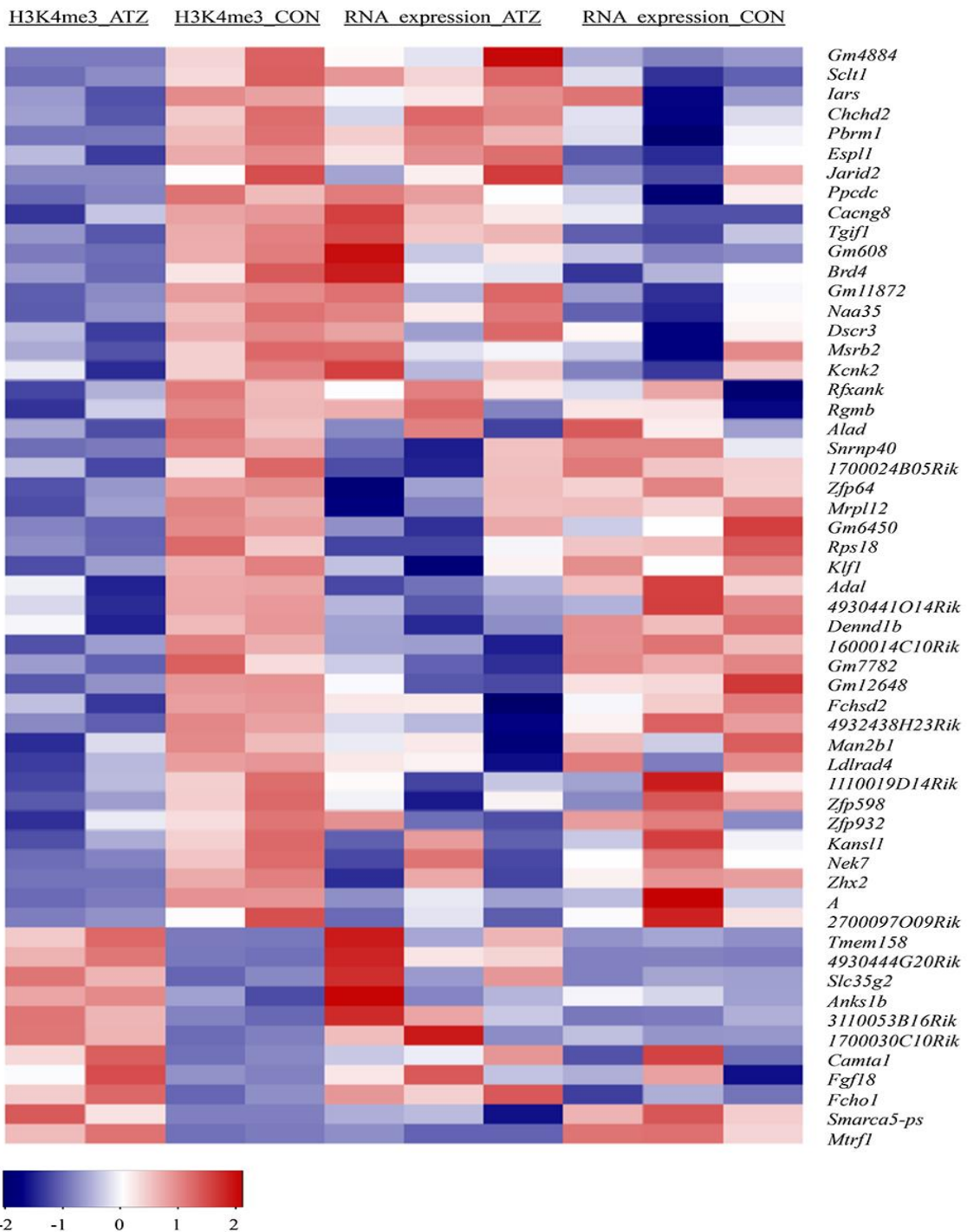
Supplementary_Figure_S22. Differential H3K4me3 peaks in spermatogonia fraction in F3 ATZ-derived males are enriched in regions for the genes associated with cell cycle, mitosis and RNA processing functions

Differential H3K4me3 peaks were compared with published datasets. (A) The distance of differential peaks to TSS of the genes, B) The subset of peaks with high signal in spermatogonia were annotated with GREAT. The GO term of biological function is shown on the left, the corresponding genes are on the right.



Supplementary_Figure_S23. The association of differential H3K4me3 peaks in F3 ATZ-derived males with H4K5Ac and H8K8ac marks from spermatocyte and spermatid fractions .

Differential H3K4me3 peaks from testis of F3 ATZ progeny ChIP-seq data were compared with published H4K5Ac and H4K8ac datasets. (A) The venn diagrams of differential peaks of testis fractions with H4K5Ac and (B) The venn diagrams of differential peaks of testis fractions with H4K8ac. SC(Spermatocytes), ST(Spermatids).

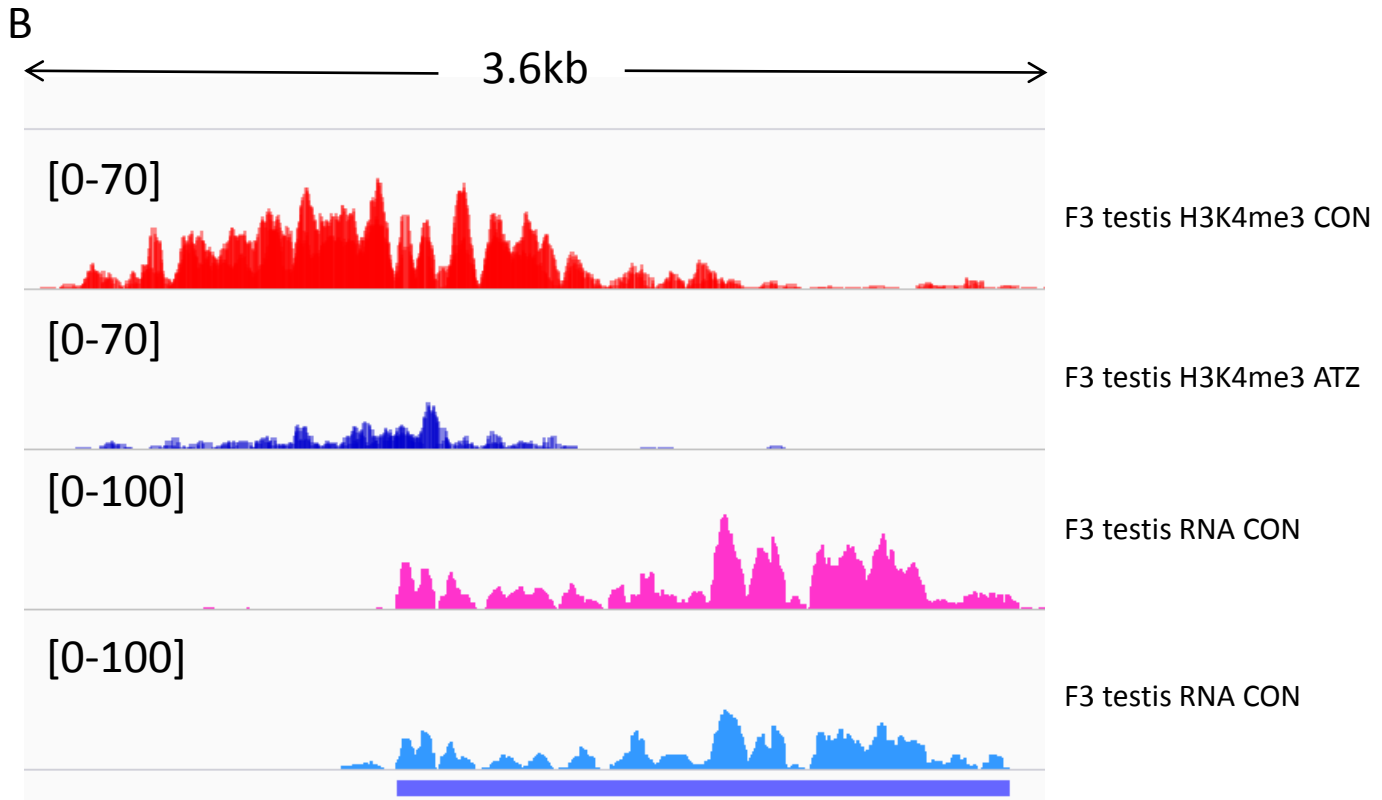


Supplementary_Figure_S24: The association of intensity of differential H3K4me3 at promoters with altered gene expression level (FC>1.2) .

H3K4me3 differential peaks associated with promoters were identified using published datasets based on signal ratio of H3K4me1 to H3K4me3. From all differential peaks, we found only 26% of them are associated with gene promoters. We presented here the signal for ChIP and corresponding RNA expression. Each column represents a biological replicate. Up-regulated genes and peaks with increased H3K4me3 occupancy are depicted in red, down-regulated RNA and peaks with decreased H3K4me3 occupancy in blue.

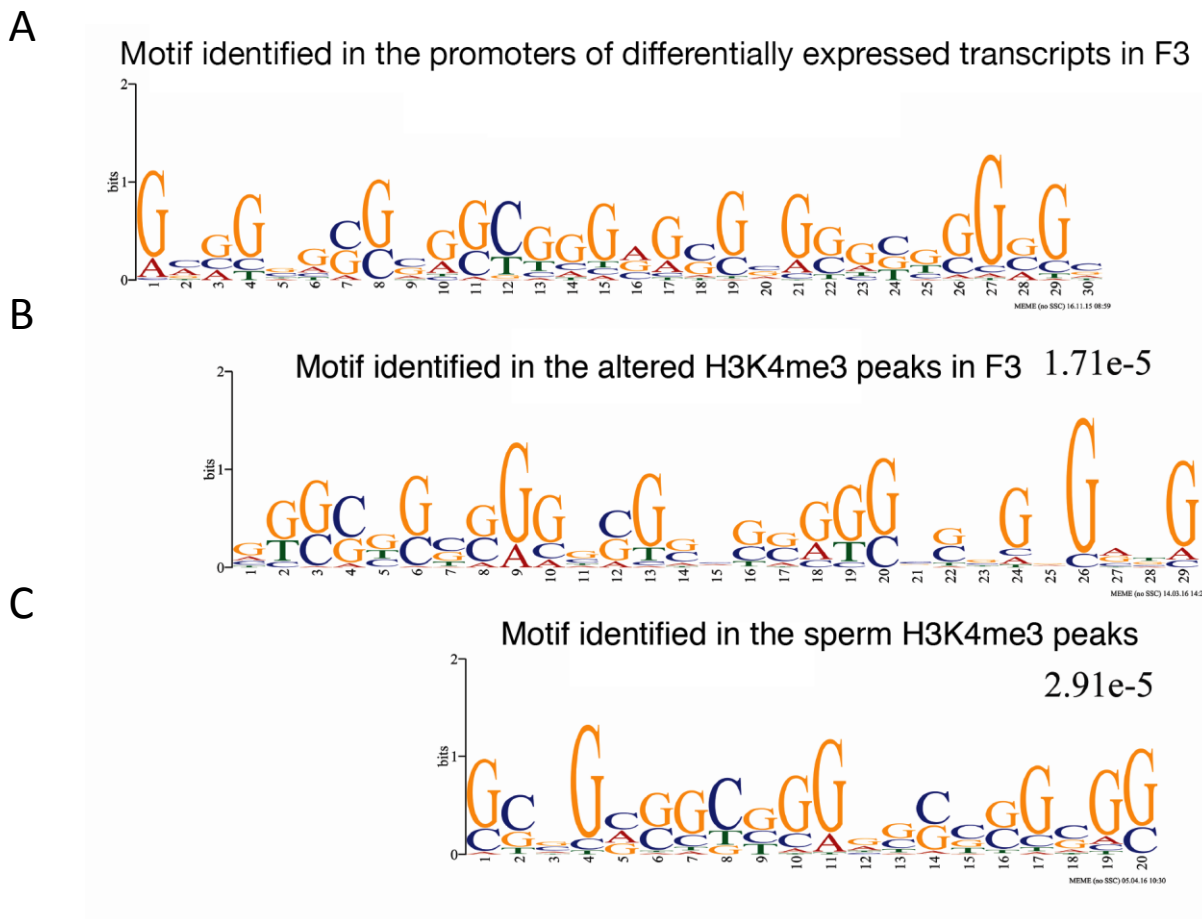
A

Chr	Start	End	P-val	FDR	peak presence in ATZ	Nearest Differentially expressed RNA	Fold Change RNA	type RNA
chr11	110041351	110042160	0,002	0,078	no	<i>Abca9</i>	1.2	coding, novel isoform
chr4	140820871	140821290	0,007	0,078	no	<i>Arhgef19</i>	0,7	coding, novel isoform
chr1	169171501	169171621	0,001	0,043	no	<i>Uck2</i>	0,6	coding, novel isoform
Chr8*	35142601	35144570	0,011	0,078	no	Intergenic RNA	0,6	novel isoform
chr10	21778021	21779480	0,006	0,065	yes	Intergenic RNA	21,0	novel isoform
Chr8	36322681	36324820	0,003	0,078	no	Intergenic RNA	0,8	novel isoform
chr10	21787571	21787970	0,002	0,043	yes	4930444G20Rik	3,0	protein coding
chr13	66815341	66815900	0,012	0,078	no	Intergenic RNA	0,8	novel isoform
chr7	46435861	46436150	0,000	0,073	no	Intergenic RNA	0,3	novel isoform



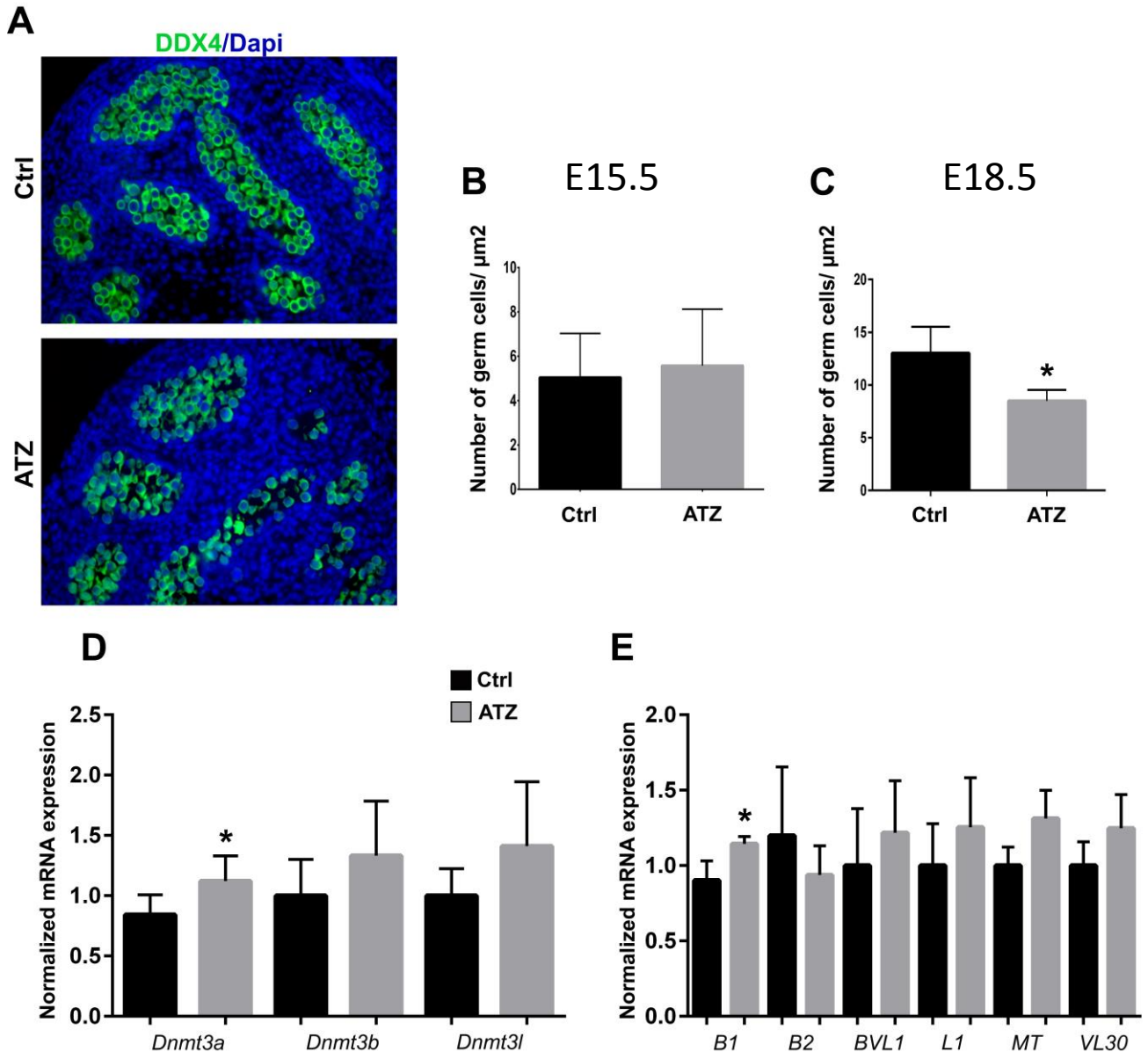
Supplementary_Figure_S25. Differential H3K4me3 peaks lost or gained in F3 ATZ-derived males

(A) Differential H3K4me3 peaks from testis of F3 ATZ progeny ChIP-seq data (chr8:35142601-35144570). (B) An example of a very low peak intensity in **F3 ATZ-derived males** near intergenic RNA expression



Supplementary_Figure_S26. The enriched motifs identified in the promoters of new RNA transcripts, the differential H3K4me3 peaks in F3 ATZ-derived males and the strongest peaks in sperm are similar.

(A) The motif enrichment was identified 500 bp upstream of the TSSs of transcripts with alternative TSSs in F3 ATZ-derived males. (B) Motif found in differential H3K4me3 peaks in F3 ATZ-derived males. (C) Motif found in the strongest H3K4me3 peaks identified during sperm histone retention analysis (Erkek et al. 2013). The MEME application was used as described in the Methods section, and the *q-value* of similarity between top and down motifs was estimated by TomTom.



Supplementary_Figure_S27. Germ cell analysis in F1 ATZ exposed male embryos.

The testes from F1 control or ATZ exposed males were fixed with PFA and paraffin slides were prepared as described in Supplementary Methods. A) The sections were immunostained against anti- DDX4 antibody, a marker of germ cells. (B) The number of germ cells were counted in three biological replicates, B) at E15,5 and C) at E18.5. D) QPCR analysis of *de novo* DNA methyltransferases from E18.5 male gonads, total RNA was extracted as described in Material and Methods and RT PCR was performed using n=5, *, p<0.05, t-test, E) qPCR analysis of retroelements, n=5, *, p<0.05, t-test.

Pearson correlation coefficients for RNA-seq and H3K4me3 ChIP-seq samples

Pearson correlation coefficients of RNA-seq samples

A

	ATZ_1	ATZ_2	ATZ_3	CON_1	CON_2	CON_3
ATZ_1	1.00	0.98	0.96	0.94	0.85	0.97
ATZ_2	0.98	1.00	0.93	0.96	0.89	0.98
ATZ_3	0.96	0.93	1.00	0.91	0.80	0.94
CON_1	0.94	0.96	0.91	1.00	0.94	0.96
CON_2	0.85	0.89	0.80	0.94	1.00	0.85
CON_3	0.97	0.98	0.94	0.96	0.85	1.00

Pearson correlation coefficients of ChIP-seq samples

B

	ATZ_1	ATZ_2	CON_1	CON_2
ATZ_1	1.00	0.94	0.96	0.96
ATZ_2	0.94	1.00	0.93	0.92
CON_1	0.96	0.93	1.00	0.97
CON_2	0.96	0.92	0.97	1.00

Differentially expressed genes in F3 ATZ-derived testes, functionally annotated by DAVID.

Term Name	Number of genes	P Value	Genes
response to DNA damage stimulus	17	0.001	up regulated in ATZ : <i>Cdk1, Pgap2, Ssrp1, Nbn, Usp1, Morf4l2, Xab2, Ercc8, Pnkp, Chd1l, Rif1, Bre, Fbxo6, Mus81, Bcl3, Rev3l</i> ; down regulated in ATZ: <i>Mms19, Ercc8, Bre, Mus81</i>
nucleotide binding	71	0.006	up regulated in ATZ: <i>Kif23, Rnasel, Gtpbp10, Aurkc, Slfn5, Mcm7, Clp1, Ndufs2, Rdm1, Cdk1, Nadsyn1, Arl9, Cdkl3, Stk4, Dapk3, Mark2, Pnkp, Glul, Hhat, NrK, Srxn1, Rev3l, Enox1, Prps1, Abca2, Chd1l, Ddx19a, Rab11a, Nat10, Prkaa1, Mat2a, Safb2, Epha7, Tubal3, Pprc1</i> ; down regulated in ATZ: <i>Acvrl1, Rps6kb2, Lace1, Dhx34, Nlrp4e, Pdxk, Hsp90aa1, Pik3cd, Ptbp1, Ifi47, Pkdcc, Rfc5, Rfc1, Ube2m, Aars2, Clcn3, Chkb, Hk2, Irak4, Igtp, Naip2, Ptk2b, Hnrnpd, Guf1, Rbm26, Pdk2, Chdh, Axl, Idh3b, Psm8, Acsm3, Cirbp, Aox3, Tnk2, Cit, Katnal1</i>
ion binding	115	0.011	up regulated in ATZ: <i>Rnasel, Zfp94, Plod1, Clp1, Rnf219, Luc7l2, Rps27a, Stk4, Mark2, Srxn1, Prps1, Zfp619, Nr2c2, Pxn, Wt1, Trim68, Mbnl3, B4galt4, Trpc1, Klf12, Zbtb42, Cblb, Dohh, Klf2, Cyb5r4, Kcnab1, Npnt, Zfp644, Kcnq3, P4ha3, Ndufs2, Zswim2, Dnajc24, Rev3l, Enox1, Inpp1, Ing3, Repl1, Eri2, Gatad2b, Slc39a7, Prkaa1, Cybasc3, Dtna, Mat2a, Sun1, Racgap1, Anxa4, Itpr2, Zc3h11a, Gca</i> ; down regulated in ATZ : <i>Gm8935, Rabgef1, Dhx34, Zfp688, Pdxk, Polg, Nudt14, Zfp78, Nbr1, Fxyd1, Clcn3, Cacnb4, Ttc3, Irak4, Sgip1, Esrrb, Idh3b, Myl12b, Trim63, Zfp579, Frem1, Tnk2, Cryzl1, Acvrl1, Lyar, Phf23, Pcgf3, Trim6, Zfp1, Gatad1, Kdm5b, Zcchc7, Zcchc8, Gmpr, Gpr98, Cyp27a1, Adam17, Zfpm2, Haghl, Ppm1m, Mecr, Lims2, Ecel1, Glis1, Zbtb16, Dchs1, Rnf123, Naip2, Mus81, Eno3, Scnm1, Rbm26, Adarb1, Cyp21a1, Trim28, Mid1, Capn3, Manba, Acsm3, Plscr2, Rassf1, Aox3, Cit, Scn4a</i>
multicellular organism reproduction	20	0.004	up regulated in ATZ : <i>Tdrd7, Htt, Il11ra1, Racgap1, Hmga1, Wt1, Nr2c2, Calca, Gm2854, Gm2790, Prmt7, Gm14332, Gm3763, Chrna7, Spata6, Gm2964</i> ; down regulated in ATZ : <i>Cav1, Tmbim6, Hk2, Zbtb16, Sbf1, Creb3l4, Cit</i>
amine biosynthetic process	7	0.007	up regulated in ATZ : <i>Bcat1, Chdh, Glul, Mat2a, Chrna7</i> ; down regulated in ATZ : <i>Chdh, Asl, Prodh</i>
mitochondrion	47	0.011	up regulated in ATZ : <i>Bcat1, Mrps34, Rnasel, Cyb5r2, Tdrd7, Acp6, Gtpbp10, 1700021f05rik, Ndufs5, Stard4, Rab11a, Atp5h, Adck4, Ndufs2, Atpaf2, Park7, Pex11b, Glul, Mtrf1, Phb2</i> ; down regulated in ATZ : <i>Cav1, Gcat, Hk2, Chchd3, Bnip3, Stoml2, Ccdc58, Lace1, Mrpl55, Guf1, Cox18, Gtf3c2, Sardh, Cox16, Chdh, Pdk2, Mmadhc, Polg, Fdxr, Idh3b, Acsm3, Cyp27a1, Mrpl48, Aars2, Surf1, Mecr, Prodh</i>
intracellular organelle lumen	40	0.022	up regulated in ATZ : <i>Midn, Ing3, Tdrd7, Gtpbp10, Sox6, Senp5, Wt1, Blzf1, Grwd1, Gm3763, P4ha3, Nat10, Dcaf17, Rdm1, Tead2, Sun1, Carf, Med6, Trpc4ap, Pop4, Gm2964</i> ; down regulated in ATZ : <i>Lyar, Bnip3, Zbtb16, Tcf7l2, Ercc8, Mus81, Mrpl55, Sardh, Olfm1, Pdk2, Polg, Trim28, Fdxr, Smad2, Acsm3, Cirbp, Pbx1, Hdac9, Aars2, Prodh</i>
lymphocyte activation	11	0.015	up regulated in ATZ : <i>Cblb, Nbn, Il11ra1, Bcl3, Chrna7</i> ; down regulated in ATZ : <i>Pik3cd, Adam17, Cacnb4, Fas, Hdac9, Ncor1</i>
vesicle-mediated transport	19	0.028	up regulated in ATZ : <i>Stx3, Caly, Htt, Hook2, Blzf1, Gapvd1, Stxbp5, Exoc4, Chrna7</i> ; down regulated in ATZ : <i>Cav1, Dennd1a, Ap4m1, Vps33b, Cd36, Stx16, Zfp1, Rabgef1, Snap23, Cox16</i>
G2/M transition DNA damage checkpoint	3	0.027	up regulated in ATZ : <i>Cdk1, Nbn</i> ; down regulated in ATZ : <i>Bre</i>

Differentially expressed genes in F3 ATZ-derived Liver, functionally annotated by DAVID.

Term Name	Number of genes	P -value	Genes
oxidation reduction	8	0.001	<i>Me1, Cyp2c44, Akr1b3, Glud1, Pgd, Cat, Dus1l, Hadha</i>
mitochondrion	8	0.040	<i>Me1, Mtrf1, Glud1, Slc25a22, Mtcp1, Cat, Timm23, Hadha</i>

Differentially expressed genes in F3 ATZ-derived brain

ID	Gene Name	GOTERM_BP_FAT
<i>Pdk1</i>	3-phosphoinositide dependent protein kinase-1	May play a general role in signaling processes and in development
<i>Arl16</i>	ADP-ribosylation factor-like 16	Nucleotide binding, GTP binding, purine nucleotide binding, guanyl nucleotide binding, ribonucleotide binding
<i>B9d1</i>	B9 protein domain 1	Specifically or prominently expressed in mouse blastocysts compared to 4-cell stage embryos
<i>Thumpd1</i>	THUMP domain containing 1	acetylation, phosphoprotein,
<i>Wwp2</i>	WW domain containing E3 ubiquitin protein ligase 2	E3 ubiquitin-protein ligase which accepts ubiquitin from an E2 ubiquitin-conjugating enzyme in the form of a thioester and then directly transfers the ubiquitin to targeted substrates
<i>Nnat</i>	neuronatin	May participate in the maintenance of segment identity in the hindbrain and pituitary development
<i>Nvl</i>	nuclear VCP-like	Nucleotide binding, nucleoside binding, purine nucleoside binding, ATP binding
<i>4933434E20Rik</i>	RIKEN cDNA 4933434E20 gene	ND
<i>Gm8399</i>	predicted gene 8399	ND
<i>Gm6189</i>	predicted gene 6189	ND
<i>Psmc3ip</i>	proteasome (prosome, macropain) 26S subunit, ATPase 3, interacting protein	Plays an important role in meiotic recombination
<i>Slc1a5</i>	solute carrier family 1 (neutral amino acid transporter), member 5	Insulin-dependent transporter for L-serine and, to a lesser extent, uptake of L-alanine and L-glutamate
<i>Tns1</i>	tensin 1	Cell motion, cell-substrate junction assembly, cell migration, cell junction assembly, cell junction organization, cell motility, localization of cell,
<i>Txn2</i>	thioredoxin 2	Has an anti-apoptotic function and plays an important role in the regulation of mitochondrial membrane potential
<i>Trim26</i>	tripartite motif-containing 26	Zinc ion binding, ion binding, cation binding, metal ion binding, transition metal ion binding
<i>Zbtb25</i>	zinc finger and BTB domain containing 25	DNA binding, zinc ion binding, ion binding, cation binding, metal ion binding, transition metal ion binding,
<i>Gm25736</i>	predicted gene 25736	ND
<i>Gm25475</i>	predicted gene 25475	ND
<i>Gm22771</i>	predicted gene 22771	ND
<i>Rps2-ps10</i>	ribosomal protein S2, pseudogene 10	ND

Differentially expressed new mRNA isoforms in F3 ATZ-derived testis

extended TSS	<p>exon gained: <i>Klf12, Gm13318, Htt, Specc1, Gatad2b, Xkr7, 9130204L05Rik, Scn4a, Gm1043, Zscan30, Zfp78, A630077J23Rik, Gm7120, 4930432E11Rik, Elp5, Slfn5, Nxn12, Mark2, Ccdc28a, Gm13794, 4930443O20Rik, Itfg2, 1700128F08Rik, Ptk2b, Psmb8, Gm21982, Poteg, Gm13853, Gm16347, 1700026J12Rik, Hbb-bs</i> ; exon number no difference: <i>Scgb2b22-ps, Stard8, Prrc2b, Scgb2b23-ps, Zfp619, Pglyrp4, Lca5, 1700021L23Rik, Naip2, Plod1, Arl9, Slc19a1, Katnal1, Gm13794, Kihl15, Tex101, Fam186a, Safb2, Pkdcc, Tspan8, Mmadhc, 1110004E09Rik, Nlrp4e, Ifi27, Anxa4, 4930444M15Rik, Dennd1a, Supt20, Blzf1</i>; exon loss: <i>Bre, Zfp951, Igf2, Gm17359, Tubal3, Luc7l2, Prodh, Stx16, Kazald1, Zcchc8, Stag3, Nup35, Gpaal1, Clp1, Prep, 2410002F23Rik, Gm8540, If7, Gm21982, Gm10125, Igtp, Nadsyn1, Nudt14, Racgap1, Rgs11, Trpc1</i></p>
shortened TSS	<p>exon gained: <i>Cyb5r2, Zc3h11a, Creb3l4, Tmem8, Srxn1, Tmem123, Gm21982, Gtf3c2, Gm7205</i>; exon number no difference: <i>Usp25, Nbn, Gm12973, Slc35a5, Rnf219, Trim6, Prkaa1, Hhipl2, Chd1l, Gm14012, Phf23, Pcgf3, Slc6a7, Tprn, Cyp27a1, Veph1, Epm2a, 1700057H15Rik, Sox6, Adarb1, Esrrb, Kcnq3</i>; exon loss: <i>Ap4m1, Ssrp1, Cers5, Chmp3, Rgcc, Ifit2, Gm14012, Maged2, Myom2, Klf2, Rnasel, Cul5, Serinc2, Calca, Tmem123, Gtpbp10, Apoc2, Gm21982, Rbm48, Phyhip, Hook2, Gm8540, Bcl3, Sun1, Dcun1d4, Epb4.1l1, Vangl2, Paqr3, 9230104L09Rik, Kihl25, Ptprf, Klrb1, P4ha3, Nmrk1, Pdxk, Mecr, Vmn2r19, Hif3a, Cblb, Fam105b, Pik3cd, Unc45b, Fas, Wdr78, Serac1, Tdrd5, Slc39a12, Pxn, Akap9, Fndc1, Rftn1, Cox16, Gzmk, Col6a6, Gm13490, A530072M11Rik, Rasgrf1, Hapln1, Serinc5, Flnb, Cit, Btrc, Col4a4, Mid1, Lhfpl2, Kcnab1, Ppp1r12b, Frem1, Glis1, Hhat, Hdac9, Dtna, Gabbr2, Maml3, Nrg3</i></p>
non altered TSS	<p>exon gained: <i>Zfp644, Serpine2, Atpaf2, Tanc1, Hsp90aa1</i>; exon number no difference: <i>Cwc22, Prss38, Cops3, Hap1, Inpp1, Rfc1, Gm14012</i>; exon loss: <i>Gm8935, E330020D12Rik, Cwc22, Rbm26, BC029214, Serp2, Dennd6b, Gm14012, Surf1, Idh3b, Flii, Trim68, Lrwd1</i></p>

Differentially expressed new mRNA isoforms in F3 ATZ-derived testis

transcripts with additional exons at 3' UTR	1700128F08Rik, Poteg, Gm13794, Gm13318, Gm8935, Zfp644, Gm7205, lqcj, Gm1043, E330020D12Rik, Klhl15, Gm16347, Gtf3c2, Cwc22, lgtf, Ifnar2, Kcnq3, Rbm26, 2610020H08Rik, 4930432E11Rik, 1700128F08Rik, Calca, 1700057H15Rik, Gm13853, Gatad2b, Klf12, Ifit2, Dcun1d4, Cit, Cblb, Cblb, Col4a4, Katnal1, 9230104L09Rik, Zfp78, Gm8540, Gm8540, Gm8540, Gm14012, Safb2, Creb3l4, Zc3h11a, Trpc1, Nrg3, Mark2, Col6a6, 2410002F23Rik, A630077J23Rik, Arl9, Prss38, Pxn, Zfp951, Luc7l2, Specc1, Serinc2, Ceacam3, Klrb1, Nudt14, Srxn1, Trim6, Gzmk, Mid1, Bre, Serpine2, Scgb2b23-ps, Scgb2b23-ps, Scgb2b22ps, Slc35a5, Slc35a5, Cox16, Scn4a, Slc6a7, Hif3a, BC029214, Epm2a, Naip2, Slc19a1, Hook2, Atpaf2, Kcnab1, Gpaa1, Gm21982, Anxa4, Metap1d, Metap1d, Tex22, Tmem123, Tmem123, Frem1, Prrc2b, Stx16, Pkdcc, Ssrp1, Apoc2
shortened 3'UTR	Maged2, Gm14012, Chmp3, Surf1, Gm7120, Irf7, Gm12973, Idh3b, Lca5, Rgs11, Nlrp4e, Flii, Trim68, Plscr2, Vmn2r19, Klhl25, Zswim2, Lrwd1, Gm21982, Ppp1r12b, Serinc5, Nup35, Gm10125, Gm17359, Ppp1r12b, Myom2
non altered 3'UTR	Htt, Stard8, Zfp619, Xkr7, Pglyrp4, Tubal3, Elp5, Slfn5, Kazald1, Nxn12, Stag3, Tspan8, Prep, Psmb8, Ifi27, Gm13794, Supt20, Serp2, Tanc1, Wdr78, lgf2, Dennd1a, Esrrb, 9130204L05Rik, Zscan30, 1700021L23Rik, Plod1, Prodh, Ccdc28a, Zcchc8, 4930443O20Rik, Itfg2, Tex101, Fam186a, Clp1, Mmadhc, 1110004E09Rik, Ptk2b, Nadsyn1, 1700026J12Rik, 4930444M15Rik, Hbb-bs, Racgap1, Blzf1, 1700057G04Rik, Gmpr, Lrrd1, Lztr1, Mtrf1, Soat2, Stoml1, Sun1, Tsr3, Wdr67, Zbtb42, Ap4m1, Usp25, Nbn, Tmem8, Prkaa1, Hhip12, Phf23, Klf2, Pcgf3, Rnasel, Phyhip, Tprn, Cyp27a1, Sun1, Epb4.111, P4ha3, Nmrk1, Mecr, Pik3cd, Unc45b, Fas, Slc39a12, Akap9, Adarb1, A530072M11Rik, Rasgrf1, Hapln1, Flnb, Btrc, Lhfpl2, Rasgrf1, Glis1, Dtna, Gm13490, Cops3, Cwc22, Dennd6b, Hap1, Hsp90aa1, Inpp1, Rfc1, Cyb5r2, Cyb5r2, Rnf219, Cers5, Rgcc, Chd11, Cul5, Gtpbp10, Rbm48, Bcl3, Vangl2, Paqr3, Ptprf, Pdxk, Fam105b, Serac1, Tdrd5, Veph1, Sox6, Fndc1, Rftn1, Hhat, Hdac9, Gabbr2, Maml3

Differential transcripts with altered APA sites in F3 ATZ-derived testis

Gene	strand	Coordinates
<i>Spin2-ps2</i>	-	chrX:71922607-71923384
<i>Gm11499</i>	-	chr11:91990925-91991246
<i>Gm6177</i>	+	chr1:162823251-162823715
<i>Mid1</i>	+	chrX:166427518-166428501
<i>Mpdz</i>	-	chr4:80925242-80925673
<i>Spin2-ps4</i>	-	chrX:72135002-72135772
<i>Spin2-ps3</i>	+	chrX:72050138-72050899
<i>Spin2-ps4</i>	-	chrX:72135002-72135763
<i>Htatsf1</i>	+	chrX:54310068-54310383
<i>Mid1</i>	+	chrX:166427579-166428727
<i>Spin2-ps3</i>	+	chrX:72050129-72050899
<i>Gm2500</i>	-	chr4:42336528-42337018
<i>Gm9969</i>	+	chr4:42681536-42682026
<i>Htatsf1</i>	+	chrX:54310068-54310381
<i>Slfn5</i>	+	chr11:82774402-82776443
<i>4933404K08Rik</i>	+	chr13:23373238-23374136
<i>Rpl21-ps8</i>	-	chr18:82692936-82693413
<i>Cwc22</i>	-	chr2:77754484-77755220
<i>Gm5687</i>	+	chr18:13056936-13057575
<i>Cd68</i>	-	chr11:69477715-69478006
<i>1700045I11Rik</i>	+	chr4:42041568-42042058

Differential transcripts with altered APA sites in F3 ATZ-derived liver

Gene	strand	Coordinates
<i>Gm12651</i>	-	chr4:93570715-93571161
<i>Gm14414</i>	+	chr2:177244980-177245552
<i>Gm7887</i>	-	chr6:48474553-48474973
<i>Gm13701</i>	-	chr2:80817995-80818295
<i>Rps8-ps2</i>	-	chr2:175804078-175804765
<i>Pop4</i>	-	chr7:39047839-39048328
<i>4632419I22Rik</i>	+	chr11:86020455-86024183
<i>Gm14438</i>	+	chr2:175096057-175096680
<i>Htatsf1</i>	+	chrX:54310068-54310383
<i>I830012O16Rik</i>	+	chr19:34685921-34687891
<i>Rps8-ps2</i>	-	chr2:175804115-175804738
<i>Htatsf1</i>	+	chrX:54310068-54310381
<i>Ube2j2</i>	+	chr4:155331190-155331675
<i>Rpl21-ps8</i>	-	chr18:82692936-82693413
<i>Dcaf17</i>	+	chr2:70926404-70926701
<i>9430015G10Rik</i>	+	chr4:155499526-155500328
<i>Gm14438</i>	+	chr2:175096030-175096717
<i>Gm14414</i>	+	chr2:177244980-177245589

Differential expressed transcripts with altered APA sites in F3 ATZ-derived brain

Gene	strand	Coordinates
<i>Gm14734</i>	-	chrX:69760043-69760489
<i>Gm11499</i>	-	chr11:91990925-91991246
<i>Mid1</i>	+	chrX:166427518-166428501
<i>Gm10320</i>	-	chr13:99259303-99259783
<i>Gm5528</i>	+	chr1:72050862-72051344
<i>Mid1</i>	+	chrX:166427579-166428727
<i>Gm11886</i>	+	chr4:23349660-23349962
<i>Tcte2</i>	-	chr17:13682139-13683705
<i>Gm14702</i>	+	chrX:66223680-66224125
<i>Gm13468</i>	+	chr2:47472605-47473049
<i>Gm14414</i>	+	chr2:177244980-177245589

Functional annotation of altered F3 ATZ-derived testis H3K4me3 peaks

ID	Desc	Hyper P-value	Obs Genes	Genes
GO:0031323	regulation of cellular metabolic process	2,42E-04	188	2410141K09Rik,2610044O15Rik8,5730507C01Rik,AW822073,Abra,Adcy1,Ankle2,App,Arfgap3,Arhgap24,Arhgap29,Arhgef19,Arid2,Asap2,Asxl2,Aym1,Bach2,Barhl2,Basp1,Bmpr2,Brd4,Camta1,Cc2d1a,Ccne1,Cdk1,Cdk5rap3,Cebpb,Cep290,Cers6,Cux1,Cux2,D10Bwg1379e,Dicer1,Dmrt1,EU599041,Edn1,Ednra,Eef2k,Efna5,Epc2,Epha2,Ern1,Etv6,Evi5,Ezh2,Fgfr1op,Fnip1,Foxc2,Foxh1,Foxk1,Galr1,Gdi2,Gm10323,Gm13157,Gm13212,Gm13225,Gm13235,Gm6712,Gm7148,Gm9376,Gm9791,Gmeb1,Gtf2e1,Gtf2e2,Gtf2h5,Gzma,Hexim2,Hipk1,Hnrnpk,lars,lkzf5,Il2,Il21,Ino80c,Jarid2,Jmjd1c,Klf1,Klf13,Klf2,Lamtor1,Leo1,Lhx4,Lmo1,Lpin1,Lrrtm3,Lzts1,Maf,Magi3,Map2k6,Map3k7,Mapkbp1,Med10,Mkl2,Mrpl12,Mtif3,Mtnr1a,Nfatc2ip,Nfe2l3,Nfya,Nkd2,Nr3c2,Pfdn5,Pgam5,Phf2,Phf1,Pif1,Ppp1r3b,Ppp4r2,Prex2,Prg4,Ptpn3,Rad21,Rad23b,Ramp3,Rapgef3,Rbm26,Rbpms,Rfx8,Rgma,Rgmb,Rnf4,Rpl38,Runx1,Rxra,Sall3,Sap30l,Shox2,Siah2,Sipa1l2,Smarmarcal1,Smc3,Sod1,Sox14,Sox18,Sox6,Sp5,Spata24,Spdya,Sprtn,Spry1,Srsf4,Srsf7,Taf12,Tbc1d15,Tcf15,Tgfb3,Tgif1,Tiam1,Tnfsf8,Tnks,Tollip,Tram1,Trim24,Trim27,Tsc22d2,Tshz1,Tshz3,Uri1,Usp2,Wapal,Wnt11,Ylpm1,Ywhaq,Zbtb44,Zcchc9,Zfat,Zfhx3,Zfp191,Zfp369,Zfp36l1,Zfp462,Zfp516,Zfp521,Zfp534,Zfp536,Zfp64,Zfp640,Zfp71rs1,Zfp715,Zfp85rs1,Zfp930,Zfp932,Zfp960,Zfp97,Zfpm2,Zfyve28,Zhx2,Zmym2

Functional annotation of altered F3 ATZ-derived testis H3K4me3 peaks

GO:0006355	regulation of transcription, DNA-dependent	2,43E-04	133	2410141K09Rik,2610044015Rik,5730507C01Rik, AW822073, Abra, App, Arid2, Asxl2, Aym1, Bach2, Barhl2, Basp1, Bmpr2, Brd4, Camta1, Cc2d1a, Cdk5rap3, Cebpb, Cep290, Cers6, Cux1, Cux2, Dicer1, Dmrt1, EU599041, Edn1, Epc2, Ern1, Etv6, Ezh2, Fnip1, Foxc2, Foxh1, Foxk1, Gm10323, Gm13157, Gm13212, Gm13225, Gm13235, Gm6712, Gm7148, Gm9376, Gm9791, Gmeh1, Gtf2e1, Gtf2e2, Gtf2h5, Hexim2, Hipk1, Hnrnpk, Ikzf5, Il2, Ino80c, Jarid2, Jmjd1c, Klf1, Klf13, Klf2, Leo1, Lhx4, Lmo1, Lpin1, Maf, Map2k6, Map3k7, Mapkbp1, Med10, Mkl2, Mrpl12, Nfatc2ip, Nfe2l3, Nfya, Nr3c2, Pfdn5, Phf2, Phf1, Rad21, Ramp3, Rbpms, Rfx8, Rgma, Rgmb, Rnf4, Runx1, Rxra, Sall3, Sap30l, Shox2, Smarcal1, Sox14, Sox18, Sox6, Sp5, Spata24, Taf12, Tcf15, Tgfbr3, Tgif1, Tnfsf8, Tnks, Tram1, Trim24, Trim27, Tsc22d2, Tshz1, Tshz3, Uri1, Usp2, Wnt11, Ylpm1, Ywhaq, Zbtb44, Zfat, Zfhx3, Zfp191, Zfp369, Zfp462, Zfp516, Zfp521, Zfp534, Zfp536, Zfp64, Zfp640, Zfp71rs1, Zfp715, Zfp85rs1, Zfp930, Zfp932, Zfp960, Zfp97, Zfpm2, Zhx2, Zmym2
GO:0048387	negative regulation of retinoic acid receptor signaling pathway	7,31E-04	3	Ezh2, Tgif1, Zfp536
GO:0032318	regulation of Ras GTPase activity	9,29E-04	16	Arfgap3, Arhgap24, Arhgef19, Asap2, Efna5, Epha2, Evi5, Ezh2, Gdi2, Prex2, Rapgef3, Sod1, Spry1, Tbc1d15, Tiam1, Wnt11
GO:0048318	axial mesoderm development	2,84E-03	3	Epha2, Foxh1, Rpl38
GO:0060119	inner ear receptor cell development	3,19E-03	5	Cecr2, Dicer1, Gabrb3, Gpr98, Sod1

Functional annotation of altered F3 ATZ-derived testis H3K4me3 peaks

GO:0033002	muscle cell proliferation	3,45E-03	4	<i>Foxc2,Naa35,Rxra,Tgfb3</i>
GO:0048385	regulation of retinoic acid receptor signaling pathway	3,96E-03	3	<i>Ezh2,Tgif1,Zfp536</i>
GO:0048285	organelle fission	4,36E-03	20	<i>Anapc4,Ankle2,Cdc6,Cdca2,Cdk1,Cltc,Gimap3,Haus3,Kntc1,Lpin1,Map10,Nsun2,Pogz,Rad21,Sgol1,Smc3,Spc24,Tnks,Vrk1,Wapal</i>
GO:0031124	mRNA 3'-end processing	4,88E-03	5	<i>App,Cpsf6,Cstf2t,Leo1,Parn</i>
GO:0030004	cellular monovalent inorganic cation homeostasis	4,88E-03	5	<i>Cftr,Nr3c2,Rab20,Rab7,Slc26a3</i>
GO:0032204	regulation of telomere maintenance	5,31E-03	3	<i>Pif1,Tnks,Ylpm1</i>
GO:0001946	lymphangiogenesis	5,31E-03	3	<i>Bmpr2,Foxc2,Sox18</i>
GO:0034502	protein localization to chromosome	6,01E-03	4	<i>Cdk1,Rad21,Tnks,Wapal</i>
GO:0060491	regulation of cell projection assembly	6,52E-03	7	<i>Arhgap24,Bbs4,Cdc42ep4,Epha2,Kank1,Myo10,Myo3b</i>
GO:0016070	RNA metabolic process	6,81E-03	105	<i>Abra,Alyref2,App,Asxl2,Asz1,Bach2,Barhl2,Bmpr2,Brd4,Camta1,Cc2d1a,Cebpb,Cep290,Cpsf6,Cstf2t,Cux1,Cux2,Dicer1,Dido1,Dmrt1,Epc2,Ern1,Etv6,Ezh2,Fdxacb1,Foxc2,Foxh1,Foxk1,Gmeb1,Gtf2e1,Gtf2e2,Gtf2h5,Hexim2,Hipk1,Hnrnpk,lars,lkzf5,Ino80c,Jarid2,Jmjd1c,Klf1,Klf13,Klf2,Lcmt2,Leo1,Lhx4,Lpin1,Maf,Map2k6,Map3k7,Med10,Mkl2,Mrpl12,Nfe2l3,Nfya,Nr3c2,Nsun2,Parn,Pes1,Phf2,Phf1,Polr3a,Pop4,Ppp4r2,Rbm26,Rbpms,Rfx8,Rnf4,Runx1,Rxra,Sall3,Sap30l,Sf3a1,Snrnp40,Sox14,Sox18,Sox6,Sp5,Spata24,Srsf4,Srsf7,Taf12,Tgif1,Tram1,Trim24,Trim27,Tshz1,Tshz3,Uri1,Utp23,Wdr83,Ylpm1,Zbtb44,Zfat,Zfhx3,Zfp191,Zfp369,Zfp36l1,Zfp516,Zfp521,Zfp536,Zfp64,Zfpm2,Zhx2,Zmym2</i>
GO:0036303	lymph vessel morphogenesis	6,90E-03	3	<i>Bmpr2,Foxc2,Sox18</i>
GO:0060113	inner ear receptor cell differentiation	7,52E-03	6	<i>Cecr2,Cux1,Dicer1,Gabrb3,Gpr98,Sod1</i>
GO:0050880	regulation of blood vessel size	7,69E-03	7	<i>Cftr,Edn1,Ednra,Foxc2,Kcnj8,Sod1,Tgfb3</i>
GO:0006378	mRNA polyadenylation	8,27E-03	4	<i>App,Cpsf6,Cstf2t,Leo1</i>

Functional annotation of altered F3 ATZ-derived testis H3K4me3 peaks

GO:0035150	regulation of tube size	8,33E-03	7	<i>Cftr,Edn1,Ednra,Foxc2,Kcnj8,Sod1,Tgfr3</i>
GO:0007067	mitosis	8,67E-03	18	<i>Anapc4,Ankle2,Cdc6,Cdca2,Cdk1,Cltc,Haus3,Kntc1,Map10,Nsun2,Pogz,Rad21,Sgol1,Smc3,Spc24,Tnks,Vrk1,Wapal</i>
GO:0090382	phagosome maturation	8,74E-03	3	<i>Rab20,Rab31,Rab7</i>
GO:0006662	glycerol ether metabolic process	8,74E-03	3	<i>Pdia4,Pdia5,Txndc2</i>
GO:0043631	RNA polyadenylation	9,58E-03	4	<i>App,Cpsf6,Cstf2t,Leo1</i>
GO:0090304	nucleic acid metabolic process	1,10E-02	124	<i>Abra,Alyref2,Apaf1,App,Asxl2,Asz1,Bach2,Barhl2,Bmpr2,Brd4,Camta1,Cc2d1a,Ccne1,Cdc6,Cdk1,Cebpb,Cep290,Cpsf6,Cstf2t,Cux1,Cux2,Dicer1,Dido1,Dmrt1,Dnase2a,Endou,Epc2,Ern1,Etv6,Ezh2,Fdxac1,Foxc2,Foxh1,Foxk1,Gmeb1,Gtf2e1,Gtf2e2,Gtf2h5,Haus3,Hexim2,Hipk1,Hist1h2al,Hnrnpk,lars,lkzf5,Ino80c,Jarid2,Jmjd1c,Klf1,Klf13,Klf2,Lcmt2,Leo1,Lhx4,Lpin1,Maf,Map2k6,Map3k7,Med10,Mkl2,Mrpl12,Nfe2l3,Nfya,Nr3c2,Nsun2,Orc5,Parn,Pes1,Phf2,Phtf1,Pif1,Polr3a,Pop4,Ppp4r2,Rad21,Rad23b,Rbm26,Rbpms,Rdm1,Rfx8,Rnf4,Runx1,Rxra,Sall3,Sap30l,Sf3a1,Smarcal1,Smc3,Snrnp40,Sox14,Sox18,Sox6,Sp5,Spata24,Sprtn,Srsf4,Srsf7,Taf12,Tgif1,Top3a,Tram1,Trim24,Trim27,Trip13,Tshz1,Tshz3,Uri1,Utp23,Wdr83,Wrn,Ylpm1,Zbtb44,Zfat,Zfx3,Zfp191,Zfp369,Zfp36l1,Zfp516,Zfp521,Zfp536,Zfp64,Zfpm2,Zhx2,Zmym2</i>
GO:0051453	regulation of intracellular pH	1,10E-02	4	<i>Cftr,Rab20,Rab7,Slc26a3</i>
GO:0030641	regulation of cellular pH	1,26E-02	4	<i>Cftr,Rab20,Rab7,Slc26a3</i>
GO:0051301	cell division	1,36E-02	23	<i>Anapc4,Ankle2,Bbs4,Ccne1,Cdc6,Cdc7,Cdca2,Cdk1,Dicer1,Evi5,Haus3,Kntc1,Map10,Nsun2,Pogz,Rab11fip3,Rad21,Sgol1,Smc3,Spc24,Tnks,Vrk1,Wapal</i>

Functional annotation of altered F3 ATZ-derived testis H3K4me3 peaks

GO:0001895	retina homeostasis	1,43E-02	4	<i>Abca4, Bbs4, Gpr98, Sod1</i>
GO:0007205	protein kinase C-activating G-protein coupled receptor signaling pathway	1,43E-02	4	<i>Agk, Dgkh, Edn1, Il2</i>
GO:0055010	ventricular cardiac muscle tissue morphogenesis	1,65E-02	5	<i>Foxc2, Foxh1, Rxra, Tgfbr3, Zfpm2</i>
GO:0055067	monovalent inorganic cation homeostasis	1,74E-02	6	<i>Cftr, Edn1, Nr3c2, Rab20, Rab7, Slc26a3</i>
GO:0015991	ATP hydrolysis coupled proton transport	1,82E-02	4	<i>Atp6v0e, Atp6v0e2, Atp6v1e1, Atp6v1g3</i>
GO:1902115	regulation of organelle assembly	1,82E-02	4	<i>Bbs4, Edn1, Myh4, Rnf4</i>
GO:0045931	positive regulation of mitotic cell cycle	1,82E-02	4	<i>App, Brd4, Cdk1, Usp2</i>
GO:0007009	plasma membrane organization	1,88E-02	8	<i>Ano6, Col5a1, Crb1, Nkd2, Pacsin2, Rab31, Ramp3, Sod1</i>
GO:0006200	ATP catabolic process	1,90E-02	10	<i>Abca4, Abca6, Abca9, Abcc4, Atp5j, Cftr, Dnahc7b, Mdn1, Vwa8, Wrn</i>
GO:0006996	organelle organization	2,00E-02	84	<i>4933400A11Rik, Anapc4, Ankle2, Arhgef10l, Arid2, Arl6, Bbs4, Brd4, Cdc6, Cdca2, Cdk1, Cep290, Cltc, Coro2b, Crocc, Dctn6, Dicer1, Dtnbp1, Epc2, Ezh2, Fam101b, Fam109a, Fat1, Fcho1, Fgfr1op, Gimap3, Golph3l, Gzma, Haus3, Hist1h2al, Jarid2, Jmjd1c, Kansl1, Klf1, Kntc1, Lamtor1, Leo1, Lpin1, Map10, Map3k7, Mapt, Mrpl12, Msrb2, Myh4, Nkd2, Nsun2, Nup153, Pes1, Pfn2, Phactr1, Phf2, Pif1, Pogz, Ptpn1, Rab20, Rab31, Rab7, Rad21, Rad23b, Rnf40, Rpl38, Rpsaps10, Sclt1, Sgol1, Sh2b2, Shroom1, Shroom3, Slc25a4, Smarcal1, Smc3, Sod1, Spc24, Spry1, Taf12, Tmed5, Tnks, Tomm34, Tram1, Trip13, Ttl6, Vrk1, Wapal, Wrn, Zfp462</i>

Functional annotation of altered F3 ATZ-derived testis H3K4me3 peaks

GO:1901360	organic cyclic compound metabolic process	2,08E-02	157	<p><i>Abca4, Abca6, Abca9, Abcc4, Abra, Adcy1, Adss, Alad, Alyref2, Amd2, Apaf1, App, Asxl2, Asz1, Atp5j, Bach2, Barhl2, Bmpr2, Brd4, Camta1, Cc2d1a, Ccne1, Cdc6, Cdk1, Cebpb, Cep290, Cftr, Cmah, Cpsf6, Cstf2t, Cux1, Cux2, Cyp51, Dctd, Dicer1, Dido1, Dio3, Dmrt1, Dnahc7b, Dnase2a, Endou, Epc2, Ern1, Etv6, Ezh2, Fdxacb1, Foxc2, Foxh1, Foxk1, Gmeb1, Gtf2e1, Gtf2e2, Gtf2h5, Haus3, Hexim2, Hipk1, Hist1h2al, Hnrnpk, Iars, Ikzf5, Ino80c, Jarid2, Jmjd1c, Klf1, Klf13, Klf2, Lcmt2, Ldhb, Leo1, Lhx4, Lpin1, Maf, Map2k6, Map3k7, Mdn1, Med10, Mkl2, Mmadhc, Mme, Mrpl12, Nfe2l3, Nfya, Nmnat3, Nr3c2, Nsun2, Nuggc, Oard1, Orc5, Parn, Pde4a, Pes1, Phf2, Phf1, Pif1, Polr3a, Pop4, Ppcdc, Ppp4r2, Rab7, Rad21, Rad23b, Rbm26, Rbpms, Rdm1, Rfx8, Rnf4, Runx1, Rxra, Sall3, Sap30l, Sf3a1, Smarcal1, Smc3, Snrnp40, Sox14, Sox18, Sox6, Sp5, Spata24, Sprtn, Srd1, Srp54b, Srp54c, Srsf4, Srsf7, Taf12, Tgif1, Tgm3, Top3a, Tph2, Tram1, Trim24, Trim27, Trip13, Tshz1, Tshz3, Uck2, Uck2, Uri1, Utp23, Vwa8, Wdr83, Wrn, Ylpm1, Zbtb44, Zfat, Zfhx3, Zfp191, Zfp369, Zfp36l1, Zfp516, Zfp521, Zfp536, Zfp64, Zfpm2, Zhx2, Zmym2</i></p>
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Functional annotation of altered F3 ATZ-derived testis H3K4me3 peaks

GO:0031623	receptor internalization	2,26E-02	4	<i>Cltc,Cxcr7,Rab31,Ramp3</i>
GO:0009169	purine ribonucleoside monophosphate catabolic process	2,28E-02	10	<i>Abca4,Abca6,Abca9,Abcc4,Atp5j,Cftr,Dnahc7b,Mdn1,Vwa8,Wrn</i>
GO:0031123	RNA 3'-end processing	2,32E-02	5	<i>App,Cpsf6,Cstf2t,Leo1,Parn</i>
GO:0009128	purine nucleoside monophosphate catabolic process	2,38E-02	10	<i>Abca4,Abca6,Abca9,Abcc4,Atp5j,Cftr,Dnahc7b,Mdn1,Vwa8,Wrn</i>
GO:0048469	cell maturation	2,39E-02	9	<i>App,Cftr,Ii21,Klf1,Klf2,Sc1t1,Slc26a3,Sox18,Trip13</i>
GO:0008088	axon cargo transport	2,50E-02	4	<i>App,Dtnbp1,Mapt,Sod1</i>
GO:0009125	nucleoside monophosphate catabolic process	2,59E-02	10	<i>Abca4,Abca6,Abca9,Abcc4,Atp5j,Cftr,Dnahc7b,Mdn1,Vwa8,Wrn</i>
GO:0006898	receptor-mediated endocytosis	2,65E-02	6	<i>Cltc,Cxcr7,Fcho1,Pacsin2,Rab31,Ramp3</i>
GO:0006885	regulation of pH	2,72E-02	5	<i>Cftr,Edn1,Rab20,Rab7,Slc26a3</i>
GO:0044802	single-organism membrane organization	3,47E-02	18	<i>Abca4,Ankle2,Ano6,Arl6,Col5a1,Crb1,Fcho1,Nkd2,Pacsin2,Rab20,Rab31,Rab7,Ramp3,Sc1t1,Sod1,Spam1,Srp54b,Srp54c</i>
GO:0007017	microtubule-based process	3,51E-02	21	<i>App,Bbs4,Ccdc39,Cltc,Crocc,Dicer1,Dnahc14,Dnahc7b,Dtnbp1,Fgfr1op,Haus3,Ift43,Kif21b,Kif5c,Map10,Mapt,Sgol1,Smc3,Sod1,Spry1,Tll6</i>

Functional annotation of altered F3 ATZ-derived testis H3K4me3 peaks

ID	Desc	HyperP	Obs Genes	Genes
GO:0009161	ribonucleoside monophosphate metabolic process	3,67E-02	13	<i>Abca4,Abca6,Abca9,Abcc4,Adss,Atp5j,Cftr,Dnahc7b,Mdn1,Uck2,Uck2,Vwa8,Wrn</i>
GO:0000278	mitotic cell cycle	3,82E-02	20	<i>Anapc4,Ankle2,Cdc6,Cdca2,Cdk1,Cltc,Haus3,Kntc1,Map10,Nsun2,Pogz,Rad21,Sgol1,Smc3,Spc24,Spdya,Spry1,Tnks,Vrk1,Wapal</i>
GO:0007140	male meiosis	3,96E-02	4	<i>Asz1,Mei1,Spdya,Trip13</i>
GO:0051225	spindle assembly	3,96E-02	4	<i>Cltc,Dicer1,Haus3,Map10</i>
GO:0022411	cellular component disassembly	3,96E-02	7	<i>Apaf1,Arid2,Gzma,Mtif3,Mtrf1,Noxo1,Vrk1</i>
GO:0006873	cellular ion homeostasis	3,99E-02	16	<i>App,Cftr,Edn1,Galr1,Hrh4,Ii2,Nr3c2,Nucb2,Rab20,Rab7,Scara5,Slc24a3,Slc26a3,Slc34a2,Sod1,Trpm1</i>
GO:0006821	chloride transport	4,51E-02	6	<i>Ano6,Cftr,Clcn3,Clc6,Gabrb3,Slc26a3</i>
GO:0009123	nucleoside monophosphate metabolic process	4,54E-02	13	<i>Abca4,Abca6,Abca9,Abcc4,Adss,Atp5j,Cftr,Dnahc7b,Mdn1,Uck2,Uck2,Vwa8,Wrn</i>
GO:0006457	protein folding	4,69E-02	9	<i>Dnajb13,Dnajb4,Fkbp14,Grpel2,Pdia5,Pfdn5,Qsox1,Txndc2,Uri1</i>
GO:0033044	regulation of chromosome organization	4,69E-02	8	<i>Asxl2,Jarid2,Lpin1,Pif1,Rnf4,Tnks,Wapal,Ylpm1</i>

Functional annotation of altered F1 ATZ-derived testis H3K4me3 peaks

ID	Desc	HyperP	Obs Genes	Genes
GO:0072132	mesenchyme morphogenesis	3,48E-05	5	<i>Hey1, Isl1, Osr1, Sox9, Twist1</i>
GO:0048863	stem cell differentiation	3,74E-05	15	<i>Cyp26a1, Eomes, Hey1, Isl1, Jarid2, Med10, Msi2, Nanos2, Nog, Osr1, Pou5f1, Sox2, Sox9, Twist1, Zhx2</i>
GO:0048864	stem cell development	4,26E-05	13	<i>Cyp26a1, Eomes, Hey1, Isl1, Med10, Msi2, Nanos2, Nog, Pou5f1, Sox2, Sox9, Twist1, Zhx2</i>
GO:0001706	endoderm formation	9,93E-05	5	<i>Eomes, Lhx1, Nog, Pou5f1, Sox2</i>
GO:0001714	endodermal cell fate specification	1,06E-04	3	<i>Eomes, Pou5f1, Sox2</i>
GO:0002495	antigen processing and presentation of peptide antigen via MHC class II	1,92E-04	4	<i>H2-Aa, H2-Ab1, H2-DMb1, March1</i>
GO:0003203	endocardial cushion morphogenesis	2,43E-04	4	<i>Hey1, Isl1, Sox9, Twist1</i>
GO:2000543	positive regulation of gastrulation	2,89E-04	3	<i>Lhx1, Osr1, Otx2</i>
GO:0002504	antigen processing and presentation of peptide or polysaccharide antigen via MHC class II	3,04E-04	4	<i>H2-Aa, H2-Ab1, H2-DMb1, March1</i>
GO:0072028	nephron morphogenesis	3,09E-04	5	<i>Hey1, Irx1, Lhx1, Osr1, Sox9</i>
GO:0007492	endoderm development	4,56E-04	6	<i>Eomes, Lhx1, Nog, Otx2, Pou5f1, Sox2</i>
GO:0019827	stem cell maintenance	5,27E-04	8	<i>Eomes, Med10, Nanos2, Nog, Pou5f1, Sox2, Sox9, Zhx2</i>

Functional annotation of altered F1 ATZ-derived testis H3K4me3 peaks

ID	Desc	HyperP	Obs Genes	Genes
GO:0033993	response to lipid	5,31E-04	21	<i>Abl2, Aicda, Alpl, Cftr, Cnr1, Cyp26a1, Il15, Isl1, Lbp, Lox, Mapk3, Nr3c2, Osr1, Pou5f1, Ptgfr, Rbp1, Slc34a2, Sox2, Sox9, Tat, Tgfbr2</i>
GO:0002478	antigen processing and presentation of exogenous peptide antigen	7,77E-04	4	<i>H2-Aa, H2-Ab1, H2-DMb1, H2-Q2</i>
GO:0072006	nephron development	9,58E-04	7	<i>Acta2, Hey1, Irx1, Lhx1, Magi2, Osr1, Sox9</i>
GO:0043583	ear development	1,03E-03	11	<i>Chd7, Grhl3, H2-DMb1, H2-Q2, Maf, Nog, Osr1, Otx2, Rpl38, Sox2, Sox9</i>
GO:0072073	kidney epithelium development	1,10E-03	6	<i>Hey1, Irx1, Lhx1, Magi2, Osr1, Sox9</i>
GO:0045621	positive regulation of lymphocyte differentiation	1,29E-03	6	<i>Cd83, H2-Aa, H2-DMb1, Ikzf1, Il15, Tgfbr2</i>
GO:0001708	cell fate specification	1,29E-03	6	<i>Eomes, Isl1, Otx2, Pou5f1, Sox2, Sox9</i>
GO:0042471	ear morphogenesis	1,35E-03	8	<i>Chd7, Grhl3, Nog, Osr1, Otx2, Rpl38, Sox2, Sox9</i>
GO:0001711	endodermal cell fate commitment	1,38E-03	3	<i>Eomes, Pou5f1, Sox2</i>

Functional annotation of altered F1 ATZ-derived testis H3K4me3 peaks

ID	Desc	HyperP	Obs Genes	Genes
GO:0072009	nephron epithelium development	1,41E-03	5	<i>Hey1,Irx1,Lhx1,Osr1,Sox9</i>
GO:0003197	endocardial cushion development	1,42E-03	4	<i>Hey1,Isl1,Sox9,Twist1</i>
GO:0050870	positive regulation of T cell activation	1,42E-03	8	<i>Cd83,Coro1a,H2-Aa,H2-Ab1,H2-DMb1,Ikzf1,Ill15,Tgfbr2</i>
GO:0019884	antigen processing and presentation of exogenous antigen	1,62E-03	4	<i>H2-Aa,H2-Ab1,H2-DMb1,H2-Q2</i>
GO:0060009	Sertoli cell development	1,73E-03	3	<i>Fndc3a,Nup210l,Sox9</i>
GO:0072087	renal vesicle development	1,73E-03	3	<i>Hey1,Lhx1,Osr1</i>
GO:0072079	nephron tubule formation	1,73E-03	3	<i>Irx1,Osr1,Sox9</i>
GO:0072088	nephron epithelium morphogenesis	1,84E-03	4	<i>Irx1,Lhx1,Osr1,Sox9</i>
GO:0060993	kidney morphogenesis	1,87E-03	5	<i>Hey1,Irx1,Lhx1,Osr1,Sox9</i>
GO:0001649	osteoblast differentiation	2,14E-03	6	<i>Figl1,Hey1,Nog,Sox2, Twist1,Wwox</i>
GO:0019886	antigen processing and presentation of exogenous peptide antigen via MHC class II	2,14E-03	3	<i>H2-Aa,H2-Ab1,H2-DMb1</i>
GO:0007398	ectoderm development	2,14E-03	3	<i>Grhl3,Lhx1,Pou5f1</i>
GO:0035987	endodermal cell differentiation	2,60E-03	3	<i>Eomes,Pou5f1,Sox2</i>
GO:0072210	metanephric nephron development	2,95E-03	4	<i>Irx1,Lhx1,Osr1,Sox9</i>
GO:0032526	response to retinoic acid	3,15E-03	6	<i>Abi2,Cyp26a1,Osr1,Pou5f1, Sox2,Sox9</i>

Functional annotation of altered F1 ATZ-derived testis H3K4me3 peaks

ID	Desc	HyperP	Obs Gene s	Genes
GO:0071300	cellular response to retinoic acid	3,28E-03	4	<i>Abl2, Cyp26a1, Osr1, Sox9</i>
GO:0045582	positive regulation of T cell differentiation	3,60E-03	5	<i>Cd83, H2-Aa, H2-DMb1, Ikzf1, Tgfbr2</i>
GO:0070849	response to epidermal growth factor stimulus	3,69E-03	3	<i>Garem, Mapk3, Sox9</i>
GO:0045619	regulation of lymphocyte differentiation	3,82E-03	7	<i>Cd83, H2-Aa, H2-DMb1, Ikzf1, Il15, Ndfip1, Tgfbr2</i>
GO:0010033	response to organic substance	4,02E-03	43	<i>Abl2, Aicda, Alpl, Cd83, Cfr, Cnr1, Coro1a, Cyp26a1, Fut8, Garem, Gm13305, H2-Aa, H2Ab1, Idi1, Ifnlr1, Il11ra2, Il15, Isl1, Jak1, Klf6, Lbp, Lhx1, Lox, Magi2, Mapk3, Nog, Nos1, Nr3c2, Nub1, Osr1, Pdgfd, Pou5f1, Ptgfr, Rad51, Rbp1, Slc34a2, Snrpn, Sox2, Sox9, Tat, Tgfbr2, Tmbim6, Wwox</i>
GO:0048002	antigen processing and presentation of peptide antigen	4,17E-03	5	<i>H2-Aa, H2-Ab1, H2-DMb1, H2-Q2, March1</i>
GO:0045580	regulation of T cell differentiation	4,49E-03	6	<i>Cd83, H2-Aa, H2-DMb1, Ikzf1, Il15, Tgfbr2</i>
GO:0060008	Sertoli cell differentiation	5,02E-03	3	<i>Fndc3a, Nup210l, Sox9</i>
GO:0001503	ossification	5,28E-03	9	<i>Alpl, Figl1, Hey1, Nog, Rpl38, Sox2, Sox9, Twist1, Wwox</i>

Functional annotation of altered F1 ATZ-derived testis H3K4me3 peaks

ID	Desc	HyperP	Obs Genes	Genes
GO:0046320	regulation of fatty acid oxidation	5,78E-03	3	<i>Cnr1,Ppargc1a, Twist1</i>
GO:0090190	positive regulation of branching involved in ureteric bud morphogenesis	5,78E-03	3	<i>Lhx1,Nog, Sox9</i>
GO:0072078	nephron tubule morphogenesis	5,78E-03	3	<i>Irx1,Osr1, Sox9</i>
GO:0050863	regulation of T cell activation	6,27E-03	9	<i>Cd83, Coro1a, H2-Aa, H2-Ab1, H2-DMb1, Ikzf1, Il15, Ndfip1, Tgfbr2</i>
GO:0060042	retina morphogenesis in camera-type eye	6,37E-03	4	<i>Lhx1, Pvr13, Sox2, Sox9</i>
GO:0034502	protein localization to chromosome	6,61E-03	3	<i>Casc5, Lrwd1, Tnks</i>
GO:2000826	regulation of heart morphogenesis	6,61E-03	3	<i>Isl1, Sox9, Twist1</i>
GO:0007369	gastrulation	6,63E-03	7	<i>Eomes, Lhx1, Nog, Otx2, Pou5f1, Sox2, Tgfbr2</i>
GO:0002696	positive regulation of leukocyte activation	6,94E-03	9	<i>Cd83, Coro1a, H2-Aa, H2-Ab1, H2DMb1, Ikzf1, Il15, Lbp, Tgfbr2</i>
GO:2000779	regulation of double-strand break repair	7,50E-03	3	<i>Figl1, Rad51, Twist1</i>
GO:0009072	aromatic amino acid family metabolic process	7,50E-03	3	<i>Dct, Tat, Tph1</i>
GO:0001837	epithelial to mesenchymal transition	7,52E-03	4	<i>Eomes, Hey1, Nog, Sox9</i>
GO:0035019	somatic stem cell maintenance	7,52E-03	4	<i>Nog, Sox2, Sox9, Zhx2</i>
GO:0042474	middle ear morphogenesis	8,46E-03	3	<i>Nog, Osr1, Rpl38</i>
GO:0060795	cell fate commitment involved in formation of primary germ layer	8,46E-03	3	<i>Eomes, Pou5f1, Sox2</i>

Functional annotation of altered F1 ATZ-derived testis H3K4me3 peaks

ID	Desc	HyperP	Obs Gene s	Genes
GO:0090189	regulation of branching involved in ureteric bud morphogenesis	8,46E-03	3	<i>Lhx1,Nog,Sox9</i>
GO:0046635	positive regulation of alpha-beta T cell activation	8,79E-03	4	<i>Cd83,H2-Ab1,Ikzf1,Tgfbr2</i>
GO:0090183	regulation of kidney development	9,48E-03	4	<i>Lhx1,Nog,Osr1,Sox9</i>
GO:0002066	columnar/cuboidal epithelial cell development	9,49E-03	3	<i>Insm1,Klf5,Spdef</i>
GO:0061333	renal tubule morphogenesis	9,49E-03	3	<i>Irx1,Osr1,Sox9</i>
GO:0002064	epithelial cell development	9,94E-03	10	<i>Acta2,Capza3,Cftr,Fndc3a,Insm1,Klf5,Magi2,Nup210l,Sox9,Spdef</i>
GO:0006282	regulation of DNA repair	1,10E-02	4	<i>Bre,Figl1,Rad51,Twist1</i>
GO:2001252	positive regulation of chromosome organization	1,17E-02	4	<i>Jarid2,Mapk3,Nos1,Tnks</i>
GO:0035065	regulation of histone acetylation	1,30E-02	3	<i>Mapk3,Nos1,Twist1</i>

Differentially expressed genes in F3 ATZ-derived testis compared to control, containing the binding sites of Sp1/Sp3/Sp4 and Wt1 sites with more than >70% conservative score in mammalian species

SP4	SP3	SP1	WT1
<i>Zfp644</i>	<i>Cul5</i>	<i>Cul5</i>	<i>Cul5</i>
<i>Cul5</i>	<i>Zfp644</i>	<i>Zfp644</i>	<i>Zfp644</i>
<i>Ppp1r12b</i>	<i>Maged2</i>	<i>Ppp1r12b</i>	<i>Safb2</i>
<i>Hsp90aa1</i>	<i>Ppp1r12b</i>	<i>Hsp90aa1</i>	<i>Dennd1a</i>
<i>Dennd1a</i>	<i>Dennd1a</i>	<i>Maged2</i>	<i>Maged2</i>
<i>Surf1</i>	<i>Hsp90aa1</i>	<i>Dennd1a</i>	<i>Hsp90aa1</i>
<i>Pik3cd</i>	<i>Pik3cd</i>	<i>Mark2</i>	<i>Flii</i>
<i>Maged2</i>	<i>Rbm26</i>	<i>Flii</i>	<i>Mark2</i>
<i>Cops3</i>	<i>Flii</i>	<i>Pik3cd</i>	<i>Gm10125</i>
<i>Rbm26</i>	<i>Atpaf2</i>	<i>Atpaf2</i>	<i>Lrwd1</i>
<i>Flii</i>	<i>Mark2</i>	<i>Rbm26</i>	<i>Atpaf2</i>
<i>Irf7</i>	<i>Safb2</i>	<i>Safb2</i>	<i>Clp1</i>
<i>Atpaf2</i>	<i>Cops3</i>	<i>Cops3</i>	<i>Slc35a5</i>
<i>Serpine2</i>	<i>Itfg2</i>	<i>Anxa4</i>	<i>Itfg2</i>
<i>BC029214</i>	<i>BC029214</i>	<i>Tex101</i>	<i>Anxa4</i>
<i>Gpaa1</i>	<i>Anxa4</i>	<i>Ssrp1</i>	<i>Stard8</i>
<i>Ssrp1</i>	<i>Ssrp1</i>	<i>Tanc1</i>	<i>Usp25</i>
<i>Usp25</i>	<i>Usp25</i>	<i>Pkdcc</i>	<i>Tanc1</i>
<i>Phf23</i>	<i>Pkdcc</i>	<i>Phf23</i>	<i>Pkdcc</i>
<i>Pkdcc</i>	<i>Stag3</i>	<i>Prrc2b</i>	<i>Phf23</i>
<i>Tanc1</i>	<i>Tanc1</i>	<i>Stx16</i>	<i>Stx16</i>
<i>Luc7l2</i>	<i>Phf23</i>	<i>Klhl15</i>	<i>Stag3</i>
<i>Stx16</i>	<i>Klhl15</i>	<i>Stag3</i>	<i>Arl9</i>

Differentially expressed genes in F3 ATZ-derived testis, containing the binding sites of Sp1/Sp3/Sp4 and Wt1 sites with more than >70% conservative score in mammalian species

SP4	SP3	SP1	WT1
<i>Stag3</i>	<i>Prrc2b</i>	<i>Ap4m1</i>	<i>Metap1d</i>
<i>Klhl15</i>	<i>Stx16</i>	<i>Sun1</i>	<i>Prep</i>
<i>Prrc2b</i>	<i>Sun1</i>	<i>Ptprf</i>	<i>Chmp3</i>
<i>Sun1</i>	<i>Luc7l2</i>	<i>Stoml1</i>	<i>Zbtb42</i>
<i>Dcun1d4</i>	<i>Prep</i>	<i>Hif3a</i>	<i>Klhl15</i>
<i>Prep</i>	<i>Ptprf</i>	<i>Tmem8</i>	<i>Stoml1</i>
<i>Hook2</i>	<i>Stoml1</i>	<i>Zc3h11a</i>	<i>Trpc1</i>
<i>Stoml1</i>	<i>Hif3a</i>	<i>Trpc1</i>	<i>Zc3h11a</i>
<i>Ptprf</i>	<i>Tmem8</i>	<i>Serpine2</i>	<i>Tmem8</i>
<i>Zc3h11a</i>	<i>Trpc1</i>	<i>Cers5</i>	<i>Tspan8</i>
<i>Tmem8</i>	<i>Zc3h11a</i>	<i>Tsr3</i>	<i>Nudt14</i>
<i>Flnb</i>	<i>Serpine2</i>	<i>Zswim2</i>	<i>Serpine2</i>
<i>Trpc1</i>	<i>Rgcc</i>		<i>Ptprf</i>
<i>Nbn</i>	<i>Zswim2</i>		
<i>Wdr67</i>	<i>Prkaa1</i>		
<i>Prkaa1</i>			
<i>Cers5</i>			
<i>Tsr3</i>			
<i>Rgcc</i>			
<i>Gm21982</i>			

The altered H3K4me3 peaks near the genes, containing Sp1/Sp3/Sp4 and Wt1 binding sites sites with more than >70% conservative score in mammalian species

SP4	SP3	SP1	WT1
<i>Dhx40</i>	<i>Fgf18</i>	<i>Fgf18</i>	<i>Fgf18</i>
<i>Fgf18</i>	<i>Pabpc1l</i>	<i>C030017K20Rik</i>	<i>Trim27</i>
<i>C030017K20Rik</i>	<i>C030017K20Rik</i>	<i>Dnajib4</i>	<i>Scube2</i>
<i>Irak2</i>	<i>Dnajib4</i>	<i>Tmem158</i>	<i>Irak2</i>
<i>Scube2</i>	<i>Scube2</i>	<i>Scube2</i>	<i>Rapgef3</i>
<i>1700030C10Rik</i>	<i>Tmem158</i>	<i>3110053B16Rik</i>	<i>Gm16257</i>
<i>Tmem158</i>	<i>Rapgef3</i>	<i>Dhx40</i>	<i>3110053B16Rik</i>
<i>Trim27</i>	<i>Gm16257</i>	<i>Zfat</i>	<i>Zfp36l1</i>
<i>3110053B16Rik</i>	<i>3110053B16Rik</i>	<i>Gm14290</i>	<i>Gm17591</i>
<i>Zfp36l1</i>	<i>Zfat</i>	<i>Etv6</i>	<i>Gm14290</i>
<i>Gm17591</i>	<i>Dhx40</i>	<i>Rapgef3</i>	<i>Tmem158</i>
<i>Zfat</i>	<i>4933411K20Rik</i>	<i>Gm16257</i>	<i>Alad</i>
<i>Ogfr</i>	<i>Gabrb3</i>	<i>4933411K20Rik</i>	<i>Gabrb3</i>
<i>Rapgef3</i>	<i>Zhx2</i>	<i>Pabpc1l</i>	<i>Zhx2</i>
<i>Gm16257</i>	<i>Cux1</i>	<i>Gabrb3</i>	<i>Cux1</i>
<i>Gabrb3</i>	<i>Gm16599</i>	<i>Zhx2</i>	<i>Gm16599</i>
<i>Zhx2</i>	<i>Gpr27</i>	<i>Cux1</i>	<i>Zbtb44</i>
<i>Cux1</i>	<i>Tmem52</i>	<i>Gm16599</i>	<i>Pde4a</i>
<i>Gm16599</i>	<i>Kansl1</i>	<i>Kansl1</i>	<i>Arrdc3</i>
<i>Kif21b</i>	<i>1700081L11Rik</i>	<i>1700081L11Rik</i>	<i>1700023H06Rik</i>
<i>Col9a3</i>	<i>Sox18</i>	<i>Tmem52</i>	<i>Pole3</i>
<i>Arrdc3</i>	<i>1700030C10Rik</i>	<i>Sox18</i>	<i>Gpr27</i>
<i>1700023H06Rik</i>	<i>Gm16256</i>	<i>1700030C10Rik</i>	<i>Sox18</i>
<i>Gpr27</i>	<i>Tram1</i>	<i>Anks1b</i>	<i>Tatdn2</i>
<i>Tmem52</i>	<i>Anks1b</i>	<i>Tram1</i>	<i>Anks1b</i>
<i>Tatdn2</i>	<i>Siva1</i>	<i>Kcnb1</i>	<i>Gm16256</i>
<i>Sox18</i>	<i>8030451A03Rik</i>	<i>Gm16256</i>	<i>1700030C10Rik</i>
<i>Kansl1</i>	<i>Tnc</i>	<i>8030451A03Rik</i>	<i>Tram1</i>
<i>1700081L11Rik</i>	<i>Ldlrad4</i>	<i>Tnc</i>	<i>Kcnb1</i>
<i>Mei1</i>	<i>Pdia5</i>	<i>Pdia5</i>	<i>Sdk2</i>
<i>Tram1</i>	<i>Kcnb1</i>	<i>Sdk2</i>	<i>Pdia5</i>
<i>Ldlrad4</i>	<i>Sdk2</i>	<i>Crocc</i>	<i>Crocc</i>
<i>Cntnap2</i>	<i>1600014C10Rik</i>		
<i>Anks1b</i>			
<i>Kcnb1</i>			
<i>Pdia5</i>			
<i>Gm16256</i>			
<i>1600014C10Rik</i>			
<i>Crocc</i>			
<i>Akr1e1</i>			

Supplementary Table S11

Genes located near F3 testis differential peaks and retained in human sperm nucleosomes

<i>Official_gene_symbol</i>	Name
<i>Kcnj16</i>	potassium inwardly-rectifying channel, subfamily J, member 16
<i>Prg4</i>	proteoglycan 4 (megakaryocyte stimulating factor, articular superficial zone protein)
<i>Slc16a1</i>	solute carrier family 16 (monocarboxylic acid transporters), member 1
<i>Fgf18</i>	fibroblast growth factor 18
<i>Mtnr1a</i>	melatonin receptor 1A
<i>Ms4a5</i>	membrane-spanning 4-domains, subfamily A, member 5
<i>Zcchc7</i>	zinc finger, CCHC domain containing 7
<i>Dmbp1</i>	dystrobrevin binding protein 1
<i>Acss3</i>	acyl-CoA synthetase short-chain family member 3
<i>Rasgef1b</i>	RasGEF domain family, member 1B; hypothetical protein LOC100044232
<i>Cdkn3</i>	cyclin-dependent kinase inhibitor 3
<i>A2m</i>	alpha-2-macroglobulin
<i>Tmed5</i>	transmembrane emp24 protein transport domain containing 5
<i>Rab31</i>	RAB31, member RAS oncogene family
<i>Nkain3</i>	Na ⁺ /K ⁺ transporting ATPase interacting 3
<i>Tram111</i>	translocation associated membrane protein 1-like 1
<i>Ppcdc</i>	phosphopantothenoylcysteine decarboxylase
<i>Arrdc4</i>	arrestin domain containing 4
<i>Cog7</i>	component of oligomeric golgi complex 7
<i>Arl6</i>	ADP-ribosylation factor-like 6
<i>Tph2</i>	tryptophan hydroxylase 2
<i>Rgmb</i>	RGM domain family, member B
<i>Prss48</i>	predicted gene 1019
<i>Itga11</i>	integrin alpha 11
<i>Cpm</i>	carboxypeptidase M
<i>Gpx8</i>	glutathione peroxidase 8 (putative)
<i>Xylt1</i>	xylosyltransferase 1
<i>Rfx8</i>	RIKEN cDNA 4933400N17 gene
<i>Cux2</i>	cut-like homeobox 2
<i>Cftr</i>	cystic fibrosis transmembrane conductance regulator homolog
<i>Qsox1</i>	quiescin Q6 sulfhydryl oxidase 1
<i>Grhpr</i>	glyoxylate reductase/hydroxypyruvate reductase
<i>Arl4d</i>	ADP-ribosylation factor-like 4D; hypothetical protein LOC100044157

Supplementary Table S11

<i>Nr3c2</i>	nuclear receptor subfamily 3, group C, member 2
<i>Dkk2</i>	dickkopf homolog 2 (<i>Xenopus laevis</i>)
<i>Fchs2</i>	FCH and double SH3 domains 2
<i>Med10</i>	mediator of RNA polymerase II transcription, subunit 10 homolog (<i>NUT2</i> , <i>S. cerevisiae</i>)
<i>Tmem202</i>	transmembrane protein 202
<i>Hebp2</i>	heme binding protein 2
<i>Tmco1</i>	transmembrane and coiled-coil domains 1
<i>Evi5</i>	ecotropic viral integration site 5
<i>Slc25a4</i>	solute carrier family 25 (mitochondrial carrier, adenine nucleotide translocator), member 4
<i>Stk33</i>	serine/threonine kinase 33
<i>Gzma</i>	granzyme A
<i>Nkain1</i>	Na ⁺ /K ⁺ transporting ATPase interacting 1
<i>Kank1</i>	KN motif and ankyrin repeat domains 1
<i>Fgl2</i>	fibrinogen-like protein 2
<i>Il1r2</i>	interleukin 1 receptor, type II
<i>Phf1</i>	putative homeodomain transcription factor 1
<i>Oxr1</i>	oxidation resistance 1
<i>Tmem200b</i>	transmembrane protein 200B
<i>Abcc4</i>	ATP-binding cassette, sub-family C (CFTR/MRP), member 4
<i>Cgnl1</i>	cingulin-like 1
<i>Ldhb</i>	lactate dehydrogenase B; predicted gene 5514
<i>Epc2</i>	enhancer of polycomb homolog 2
<i>Ywhaq</i>	tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation protein, theta polypeptide
<i>Arhgap29</i>	Rho GTPase activating protein 29
<i>Pla2g4a</i>	phospholipase A2, group IVA (cytosolic, calcium-dependent)
<i>Gpx5</i>	glutathione peroxidase 5
<i>Zfp361l1</i>	zinc finger protein 36, C3H type-like 1
<i>Als2cr12</i>	amyotrophic lateral sclerosis 2 (juvenile) chromosome region, candidate 12 (human)
<i>Veph1</i>	ventricular zone expressed PH domain homolog 1 (zebrafish)
<i>Cdc6</i>	cell division cycle 6 homolog
<i>Tnc</i>	tenascin C
<i>Hmcn1</i>	hemicentin 1
<i>Sox6</i>	SRY-box containing gene 6

<i>Slc24a3</i>	solute carrier family 24 (sodium/potassium/calcium exchanger), member 3
<i>Myo3b</i>	myosin IIIB
<i>Gtf2e1</i>	general transcription factor II E, polypeptide 1 (alpha subunit)
<i>Mdn1</i>	midasin homolog (yeast)
<i>Slc38a9</i>	solute carrier family 38, member 9
<i>Sgol1</i>	shugoshin-like 1 (S. pombe)
<i>Mapkbp1</i>	mitogen-activated protein kinase binding protein 1
<i>Pdzr3</i>	PDZ domain containing RING finger 3
<i>Col4a1</i>	collagen, type IV, alpha 1
<i>Tnks</i>	tankyrase, TRF1-interacting ankyrin-related ADP-ribose polymerase
<i>Lmo1</i>	LIM domain only 1
<i>Fndc3a</i>	fibronectin type III domain containing 3A
<i>Cpsf6</i>	cleavage and polyadenylation specific factor 6
<i>Wnt11</i>	wingless-related MMTV integration site 11
<i>Frmd4b</i>	FERM domain containing 4B
<i>Sgk1</i>	serum/glucocorticoid regulated kinase 1
<i>Fam109a</i>	family with sequence similarity 109, member A
<i>Abca9</i>	ATP-binding cassette, sub-family A (ABC1), member 9
<i>Fscb</i>	fibrous sheath CABYR binding protein
<i>Mme</i>	membrane metallo endopeptidase
<i>Tnfai3</i>	tumor necrosis factor, alpha-induced protein 8
<i>Nfe2l3</i>	nuclear factor, erythroid derived 2, like 3
<i>Fam65b</i>	family with sequence similarity 65, member B
<i>Zfat</i>	zinc finger and AT hook domain containing
<i>Actr6</i>	ARP6 actin-related protein 6 homolog (yeast)
<i>Mast4</i>	microtubule associated serine/threonine kinase family member 4
<i>Dynlrb2</i>	dynein light chain roadblock-type 2
<i>Maf</i>	avian musculoaponeurotic fibrosarcoma oncogene homolog
<i>Ergic1</i>	endoplasmic reticulum-golgi intermediate compartment (ERGIC) 1
<i>Nat8l</i>	N-acetyltransferase 8-like
<i>Lhfp12</i>	lipoma HMGIC fusion partner-like 2
<i>Phactr1</i>	phosphatase and actin regulator 1
<i>Afap111</i>	actin filament associated protein 1-like 1
<i>Dicer1</i>	Dicer1, Dcr-1 homolog (Drosophila)
<i>Isca1</i>	iron-sulfur cluster assembly 1 homolog (S. cerevisiae)
<i>Psm7</i>	proteasome (prosome, macropain) 26S subunit, non-ATPase, 7
<i>Tmem158</i>	transmembrane protein 158

<i>Rasef</i>	RAS and EF hand domain containing
<i>Tiam1</i>	T-cell lymphoma invasion and metastasis 1
<i>Zfp64</i>	zinc finger protein 64
<i>Klkb1</i>	kallikrein B, plasma 1
<i>Tgif1</i>	TGFB-induced factor homeobox 1
<i>Abra</i>	actin-binding Rho activating protein
<i>Tulp4</i>	tubby like protein 4
<i>Txndc2</i>	thioredoxin domain containing 2 (spermatzoa)
<i>Dir2</i>	disrupted in renal carcinoma 2 (human)
<i>Alcam</i>	activated leukocyte cell adhesion molecule
<i>Pdzrn4</i>	PDZ domain containing RING finger 4
<i>Galm</i>	galactose mutarotase
<i>Sema5b</i>	sema domain, seven thrombospondin repeats (type 1 and type 1-like), transmembrane domain (TM) and short cytoplasmic domain, (semaphorin) 5B
<i>Zc3h13</i>	zinc finger CCCH type containing 13
<i>Tmem144</i>	transmembrane protein 144
<i>Dlgap1</i>	discs, large (Drosophila) homolog-associated protein 1
<i>Bcl2l14</i>	BCL2-like 14 (apoptosis facilitator)
<i>Lphn3</i>	latrophilin 3
<i>Mtif3</i>	mitochondrial translational initiation factor 3
<i>Cd180</i>	CD180 antigen
<i>Abca4</i>	ATP-binding cassette, sub-family A (ABC1), member 4
<i>Cabin1</i>	calcineurin binding protein 1
<i>Fnip1</i>	folliculin interacting protein 1
<i>Spin1</i>	spindlin 1
<i>Slc26a3</i>	solute carrier family 26, member 3
<i>Edn1</i>	endothelin 1
<i>Mmadhc</i>	methylmalonic aciduria (cobalamin deficiency) cblD type, with homocystinuria
<i>Mkl2</i>	MKL/myocardin-like 2
<i>Siah3</i>	seven in absentia homolog 3 (Drosophila)
<i>Pdia4</i>	protein disulfide isomerase associated 4
<i>Barhl2</i>	BarH-like 2 (Drosophila)
<i>Atp10a</i>	ATPase, class V, type 10A
<i>Foxk1</i>	forkhead box K1
<i>Cops8</i>	COP9 (constitutive photomorphogenic) homolog, subunit 8 (Arabidopsis thaliana)
<i>Parn</i>	poly(A)-specific ribonuclease (deadenylation nuclease)
<i>Kif21b</i>	kinesin family member 21B
<i>Hrh4</i>	histamine receptor H4
<i>Rnd3</i>	Rho family GTPase 3
<i>Arrdc3</i>	arrestin domain containing 3

<i>Lmbr1</i>	limb region 1
<i>Lpin1</i>	lipin 1
<i>Galnt10</i>	UDP-N-acetyl-alpha-D-galactosamine:polypeptide N-acetylgalactosaminyltransferase 10
<i>Mak16</i>	MAK16 homolog (<i>S. cerevisiae</i>)
<i>App</i>	amyloid beta (A4) precursor protein
<i>Vrk1</i>	vaccinia related kinase 1
<i>Kcnj8</i>	potassium inwardly-rectifying channel, subfamily J, member 8
<i>Utp23</i>	UTP23, small subunit (SSU) processome component, homolog (yeast)
<i>Basp1</i>	brain abundant, membrane attached signal protein 1
<i>Tram1</i>	translocating chain-associating membrane protein 1
<i>Zhx2</i>	zinc fingers and homeoboxes 2
<i>Fam120a</i>	family with sequence similarity 120, member A
<i>Wdfy3</i>	WD repeat and FYVE domain containing 3
<i>Arid2</i>	AT rich interactive domain 2 (ARID, RFX-like); RIKEN cDNA 1700124K17 gene
<i>N6amt1</i>	N-6 adenine-specific DNA methyltransferase 1 (putative)
<i>Ramp3</i>	receptor (calcitonin) activity modifying protein 3
<i>Map3k7</i>	mitogen-activated protein kinase kinase kinase 7; predicted gene 8188
<i>Ylpm1</i>	YLP motif containing 1
<i>Grid1</i>	glutamate receptor, ionotropic, delta 1
<i>Dcn</i>	decorin
<i>Sap30l</i>	SAP30-like
<i>Mapt</i>	microtubule-associated protein tau
<i>Myh4</i>	myosin, heavy polypeptide 4, skeletal muscle
<i>Traf7d1</i>	TRAF type zinc finger domain containing 1
<i>Cul1</i>	cullin 1
<i>Ptplad2</i>	protein tyrosine phosphatase-like A domain containing 2
<i>Ndst3</i>	N-deacetylase/N-sulfotransferase (heparan glucosaminyl) 3
<i>Cstf2t</i>	cleavage stimulation factor, 3' pre-RNA subunit 2, tau
<i>Pak6</i>	p21 protein (Cdc42/Rac)-activated kinase 6
<i>Dennd1b</i>	DENN/MADD domain containing 1B
<i>Prkrir</i>	protein-kinase, interferon-inducible double stranded RNA dependent inhibitor, repressor of (P58 repressor)
<i>Dmrt1</i>	doublesex and mab-3 related transcription factor 1

<i>Spam1</i>	sperm adhesion molecule 1
<i>Crb1</i>	crumbs homolog 1 (<i>Drosophila</i>)
<i>Gxylt1</i>	glycosyltransferase 8 domain containing 3
<i>Runx1</i>	runt related transcription factor 1
<i>Wapal</i>	wings apart-like homolog (<i>Drosophila</i>)
<i>Gpr98</i>	G protein-coupled receptor 98
<i>Adcy1</i>	adenylate cyclase 1
<i>Cdca2</i>	cell division cycle associated 2
<i>Asap2</i>	development and differentiation enhancing factor 2
<i>Ano6</i>	anoctamin 6
<i>Nek11</i>	NIMA (never in mitosis gene a)-related expressed kinase 11
<i>Cdc7</i>	cell division cycle 7 (<i>S. cerevisiae</i>)
<i>Shroom3</i>	shroom family member 3
<i>Spry1</i>	sprouty homolog 1 (<i>Drosophila</i>)
<i>Scamp1</i>	secretory carrier membrane protein 1
<i>Ednra</i>	endothelin receptor type A
<i>Ccdc39</i>	coiled-coil domain containing 39
<i>Trhde</i>	TRH-degrading enzyme
<i>Phf2</i>	PHD finger protein 2
<i>Anks1b</i>	ankyrin repeat and sterile alpha motif domain containing 1B
<i>Efna5</i>	ephrin A5
<i>Stxbp5l</i>	syntaxin binding protein 5-like
<i>Kif5c</i>	kinesin family member 5C
<i>Sall3</i>	sal-like 3 (<i>Drosophila</i>)
<i>Ptpn3</i>	protein tyrosine phosphatase, non-receptor type 3
<i>Bmpr2</i>	bone morphogenic protein receptor, type II (serine/threonine kinase)
<i>Lzts1</i>	leucine zipper, putative tumor suppressor 1
<i>Rnf38</i>	ring finger protein 38
<i>Otud6b</i>	OTU domain containing 6B; predicted gene 12444
<i>F11</i>	coagulation factor XI
<i>Atp6v1g3</i>	ATPase, H ⁺ transporting, lysosomal V1 subunit G3
<i>Dgkh</i>	diacylglycerol kinase, eta
<i>Lhx4</i>	LIM homeobox protein 4
<i>Rpl38</i>	ribosomal protein L38
<i>Ino80c</i>	INO80 complex subunit C
<i>Atp5j</i>	ATP synthase, H ⁺ transporting, mitochondrial F0 complex, subunit F pseudogene
<i>Arid1b</i>	AT rich interactive domain 1B (SWI-like)
<i>Kcnk2</i>	potassium channel, subfamily K, member 2
<i>Galr1</i>	galanin receptor 1

<i>Trpc7</i>	transient receptor potential cation channel, subfamily C, member 7
<i>Pdia5</i>	protein disulfide isomerase associated 5
<i>Mctp2</i>	multiple C2 domains, transmembrane 2
<i>Dio3</i>	deiodinase, iodothyronine type III
<i>Exoc8</i>	exocyst complex component 8
<i>Rps29</i>	ribosomal protein S29
<i>Sdk2</i>	sidekick homolog 2 (chicken)
<i>Zbtb40</i>	zinc finger and BTB domain containing 40
<i>Ppp4r2</i>	protein phosphatase 4, regulatory subunit 2
<i>Nek7</i>	NIMA (never in mitosis gene a)-related expressed kinase 7
<i>Bach2</i>	BTB and CNC homology 2
<i>Sclt1</i>	sodium channel and clathrin linker 1
<i>Zfp2</i>	zinc finger protein, multitype 2
<i>Rad23b</i>	RAD23b homolog (S. cerevisiae)
<i>Anapc4</i>	anaphase promoting complex subunit 4
<i>Chchd2</i>	coiled-coil-helix-coiled-coil-helix domain containing 2
<i>Chsy3</i>	chondroitin sulfate synthase 3
<i>Abca6</i>	ATP-binding cassette, sub-family A (ABC1), member 6
<i>Zfx3</i>	zinc finger homeobox 3
<i>Apaf1</i>	apoptotic peptidase activating factor 1
<i>Trim24</i>	tripartite motif-containing 24
<i>Znfm2</i>	zinc finger, MYM-type 2
<i>Cep170</i>	centrosomal protein 170
<i>Siah2</i>	seven in absentia 2
<i>Sox14</i>	SRY-box containing gene 14
<i>Fam198b</i>	RIKEN cDNA 1110032E23 gene
<i>Cdh10</i>	cadherin 10
<i>Tnfrsf8</i>	tumor necrosis factor (ligand) superfamily, member 8
<i>Cbl1</i>	Casitas B-lineage lymphoma-like 1
<i>Kctd3</i>	predicted gene 7553; potassium channel tetramerisation domain containing 3
<i>Pgk2</i>	phosphoglycerate kinase 2
<i>Magi3</i>	membrane associated guanylate kinase, WW and PDZ domain containing 3
<i>Scrn1</i>	secernin 1
<i>Dctd</i>	dCMP deaminase
<i>Crocc</i>	ciliary rootlet coiled-coil, rootletin
<i>Clic6</i>	chloride intracellular channel 6
<i>Sp5</i>	trans-acting transcription factor 5
<i>Camta1</i>	calmodulin binding transcription activator 1
<i>Muc5b</i>	mucin 5, subtype B, tracheobronchial
<i>Il20rb</i>	interleukin 20 receptor beta

<i>Gtf2h5</i>	general transcription factor IIIH, polypeptide 5
<i>Msrb2</i>	methionine sulfoxide reductase B2
<i>Smpd3a</i>	sphingomyelin phosphodiesterase, acid-like 3A
<i>Nmnat3</i>	nicotinamide nucleotide adenylyltransferase 3
<i>Cdk1</i>	cell division cycle 2 homolog A (S. pombe)
<i>Stk35</i>	serine/threonine kinase 35
<i>Asxl2</i>	additional sex combs like 2 (Drosophila)
<i>Tollip</i>	toll interacting protein
<i>Tmem66</i>	transmembrane protein 66
<i>Arglu1</i>	arginine and glutamate rich 1
<i>Akap9</i>	A kinase (PRKA) anchor protein (yotiao) 9
<i>Rnf32</i>	ring finger protein 32
<i>Gpr149</i>	G protein-coupled receptor 149
<i>Dscr3</i>	Down syndrome critical region gene 3
<i>Zbtb44</i>	zinc finger and BTB domain containing 44
<i>Adss</i>	adenylosuccinate synthetase, non muscle
<i>Rad21</i>	RAD21 homolog (S. pombe)
<i>Rgma</i>	RGM domain family, member A
<i>Agk</i>	predicted gene 8546; acylglycerol kinase
<i>Klf13</i>	Kruppel-like factor 13
<i>Gdi2</i>	predicted gene 4973; guanosine diphosphate (GDP) dissociation inhibitor 2
<i>Nucb2</i>	nucleobindin 2
<i>Edil3</i>	EGF-like repeats and discoidin I-like domains 3
<i>Nup153</i>	nucleoporin 153
<i>Tshz3</i>	teashirt zinc finger family member 3
<i>Mtrf1</i>	mitochondrial translational release factor 1
<i>Atg16l2</i>	autophagy related 16 like 2 (S. cerevisiae)
<i>Prex2</i>	phosphatidylinositol-3,4,5-trisphosphate-dependent Rac exchange factor 2
<i>Map2k6</i>	mitogen-activated protein kinase kinase 6
<i>Col6a3</i>	collagen, type VI, alpha 3
<i>Trpm1</i>	transient receptor potential cation channel, subfamily M, member 1
<i>Uck2</i>	uridine-cytidine kinase 2
<i>Vps52</i>	vacuolar protein sorting 52 (yeast)
<i>Clybl</i>	citrate lyase beta like
<i>Sipa112</i>	hypothetical protein LOC676463; signal-induced proliferation-associated 1 like 2
<i>Sod1</i>	superoxide dismutase 1, soluble
<i>Ezh2</i>	enhancer of zeste homolog 2 (Drosophila)
<i>Ppm1l</i>	protein phosphatase 1 (formerly 2C)-like
<i>Cep290</i>	centrosomal protein 290
<i>Tsc22d2</i>	TSC22 domain family, member 2
<i>Gpc6</i>	glypican 6
<i>Rbm26</i>	RNA binding motif protein 26

<i>Spock1</i>	sparc/osteonectin, cwcv and kazal-like domains proteoglycan 1
<i>Il2</i>	interleukin 2
<i>Pld5</i>	phospholipase D family, member 5
<i>Jmjd1c</i>	jumonji domain containing 1C
<i>Clcn3</i>	chloride channel 3
<i>Shroom1</i>	shroom family member 1
<i>Btg1</i>	B-cell translocation gene 1, anti-proliferative
<i>Gabbr3</i>	gamma-aminobutyric acid (GABA) A receptor, subunit beta 3
<i>Arih1</i>	ariadne ubiquitin-conjugating enzyme E2 binding protein homolog 1 (Drosophila)
<i>Sema3c</i>	sema domain, immunoglobulin domain (Ig), short basic domain, secreted, (semaphorin) 3C
<i>Shox2</i>	short stature homeobox 2
<i>Lrrtm3</i>	leucine rich repeat transmembrane neuronal 3
<i>Coro2b</i>	coronin, actin binding protein, 2B
<i>Wrn</i>	Werner syndrome homolog
<i>Ivns1abp</i>	influenza virus NS1A binding protein
<i>Tgfbr3</i>	transforming growth factor, beta receptor III
<i>Asz1</i>	ankyrin repeat, SAM and basic leucine zipper domain containing 1
<i>Ccdc62</i>	coiled-coil domain containing 62
<i>Serp2</i>	stress-associated endoplasmic reticulum protein family member 2
<i>Kpna4</i>	karyopherin (importin) alpha 4
<i>Hs3st5</i>	heparan sulfate (glucosamine) 3-O-sulfotransferase 5
<i>Arhgap24</i>	Rho GTPase activating protein 24
<i>Fat1</i>	FAT tumor suppressor homolog 1 (Drosophila)
<i>Ggh</i>	gamma-glutamyl hydrolase
<i>Ppfi2</i>	protein tyrosine phosphatase, receptor type, f polypeptide (PTPRF), interacting protein (liprin), alpha 2
<i>Ube2e2</i>	ubiquitin-conjugating enzyme E2E 2 (UBC4/5 homolog, yeast)
<i>Srbd1</i>	S1 RNA binding domain 1
<i>Pfn2</i>	profilin 2
<i>Zcchc9</i>	zinc finger, CCHC domain containing 9
<i>Sdc3</i>	syndecan 3
<i>Lmod3</i>	leiomodin 3 (fetal)
<i>Kansl1</i>	KAT8 regulatory NSL complex subunit 1
<i>Map10</i>	microtubule-associated protein 10
<i>Asic2</i>	acid-sensing (proton-gated) ion channel 2
<i>Cers6</i>	ceramide synthase 6
<i>Vwa8</i>	von Willebrand factor A domain containing 8

<i>Tmem178b</i>	transmembrane protein 178
<i>Caap1</i>	caspase activity and apoptosis inhibitor 1
<i>Tenn3</i>	teneurin transmembrane protein 3
<i>Focad</i>	focadhesin
<i>Saysd1</i>	SAYSVFN motif domain containing 1
<i>Scp2d1</i>	SCP2 sterol-binding domain containing 1
<i>Tmem242</i>	transmembrane protein 242
<i>Gsap</i>	gamma-secretase activating protein
<i>Ldlrad4</i>	low density lipoprotein receptor class A domain containing 4
<i>Uri1</i>	URI1, prefoldin-like chaperone

Common regions affected in vinclozolin transgenerational and this studies

Official symbol	Official Full Name	Function of the protein	FC, in our data
<i>Cwc22</i>	CWC22 spliceosome-associated protein	Plays role in mRNA splicing	3.21
<i>Slmo2</i>	slowmo homolog 2 (Drosophila)	The protein has the phosphatidic acid transporter activity	2.34
<i>Lrrc61</i>	leucine rich repeat containing 61	ND	2.16
<i>Etv1</i>	Ets variant 1	The ETS proteins regulate genes that modulate the cell growth, angiogenesis, migration, proliferation and differentiation	1.68
<i>1700123k08Rik</i>	RIKEN cDNA 1700123K08 gene	ND	1.51
<i>Dcxr</i>	dicarbonyl L-xylulose reductase	Protein plays role oxidation-reduction process	1.47
<i>Sepw1</i>	selenoprotein W, muscle 1	Protein is highly expressed in proliferating myoblasts and suggested to be involved in muscle growth and differentiation by protecting the developing myoblasts from oxidative stress.	1.38
<i>Pibf1</i>	progesterone immunomodulatory binding factor 1	Mediator of progesterone that by acting on the phospholipase A2 enzyme interferes with arachidonic acid metabolism, induces a Th2 biased immune response, and by controlling NK activity exerts an anti-abortive effect	1.37
<i>Skap1</i>	src family associated phosphoprotein 1	The encoded protein plays a critical role in inside-out signaling by coupling T-cell antigen receptor stimulation to the activation of integrins	1.33
<i>Sepn1</i>	selenoprotein N, 1	The function of this gene in mouse is not known, however, mutations in the homologous gene in human are associated with multiminicore disease and rigid spine muscular dystrophy	1.31
<i>Bop1</i>	block of proliferation 1	Block of proliferation 1 (BOP1) protein plays an oncogenic role in hepatocellular carcinoma by promoting epithelial-to-mesenchymal transition.	1.28
<i>Itgb3</i>	integrin beta 3	Integrins are known to participate in cell adhesion as well as cell-surface mediated signalling	1.26
<i>Arl6ip4</i>	ADP-ribosylation factor-like 6 interacting protein 4	Involved in modulating alternative pre-mRNA splicing with either 5 distal site activation or preferential use of 3 proximal site. In case of infection by Herpes simplex virus (HSV1), may act as a splicing inhibitor of HSV1 pre-mRNA.	1.22

Supplementary Table S13
The primers used for QPCR analysis

Gene	5'→3', forward	5'→3', reverse
<u>Poteg</u>	CACTTATCCACCACCCAAACC	GTCGTCCCAAAGTCTAOCCTCAG
<u>Specc1</u>	ATGCTOCATCAACGACTAACC	TAGTCCAGAACCTOOGTTCTC
<u>Xkr7</u>	CCACTTAOCACAGAAACAGAAACC	ATAGTOGOCCTTCTTTCAACC
<u>Rfx8</u>	T TAAATCCTOCACCTTCTOCAGTTCTTG	GGCTCAOCTGTGTCTOCAT
<u>Cwc22</u>	GAGAAOCTOCAOCCAAGTGAA	TCAAGGAACAGTAOCGACACCAT
<u>Cit</u>	AAACTTCTAOCCTGTCCAAAACAACCTC	GCCTOCATCGTTGCTTCT
<u>AurkC</u>	CAGCAOCTCAOCCAGAAGCATT	T CCTTGAGOOAGACCAAGTA
<u>Lgals3bp</u>	AAAGGTTCCAGAGGTTGTCCAC	CCAGGGACTCAAGTACAGAAGA
<u>Npb</u>	GTAAGGAGOCCTTGTGOCAG	T CAGGTGCTCTGGCAATCAGT
<u>Ifi27</u>	GCTTCAOCCOCTCAGGAAT	GTCOAGCAGCTCAGCAGACT
<u>Cyp21a1</u>	CAGTTCOCCAAOCCATCTCATOC	TCAOCCOCTGAATOCAGAAG
<u>Ifi272a</u>	GAGTCCAAGGAOCCCTGCTGATT	ACTOCCTAAGTTCTTCTATGCTCT
<u>Phytip</u>	AGCATGOCAGAOCCACTGT	GATGGCATAGTGGTAGGCTGTGT
<u>Aars2</u>	CCAOCCAGOOCTACTAOCOA	CCCAATGATCAOCCAAGTCTOCC
<u>Spir2d</u>	CTOCAAGGCTTTCTGGTGT	CAGGAAOCCATCAOCCGGTCCAC
<u>Crtam</u>	GAAAGGAGTCATAACACGGTCA	ACCAGCTTCTTCAOCCATTCCAG
<u>Cdk1</u>	CCAGAGCTTGAAGATOCCTGAAGA	GTACCGATCTOCAGAAGTGTG
<u>Ctc1</u>	CTGCTCAGATACAGAAAGAAATOC	GGGCTAATTCAOCCAAGTGTC
<u>Rif1</u>	AGCAACAGTGAATCTOCTGG	GOGAACCTTCTTTCATCOGT
<u>Irwd1</u>	CAACACAGATOCCTGAAGTGCC	AGTGAGGTAGGTGAAGGCTG
<u>Mis18</u>	CGTTGAAAGTTACACCTTAGGGTC	GGCTGTCAGGACTTCTTCCA
<u>Nbn</u>	GCCAAGCAGACAAGAACTCAG	GTTATTOCTACATCAACAAGCAG
<u>Mus81</u>	GCATCATAAGCAGOCAGGA	CTGTCAAGAACAAGOOCCAGT
<u>F box6</u>	GTTGTCCAGGAAAGCTTGTGOCA	GAACCCGTGTGCTGAAAGAAOC
<u>Chd1</u>	ACCAAGATAGOCCTGTTTCTG	GCATOCCAACTTACAATACTACTTTC
<u>Morf42</u>	GCTOCTCTCATCTTACTTCTCTG	GAAAGAAGGAACATTGTGGTTGG
<u>Ssrp1</u>	CAGCATTGAAAGGAGGAGTATGG	TCTTCATCAGAGTCCAGCATAATOGT
<u>Dnmt3a</u>	GCAGGAGAGGGCAAAGAACAG	CAGCCACTCATCOOGTTTTC
<u>Dnmt3b</u>	T TTAAGTAACCCGTGCTOCA	GGAAATGCTACAAGAOCACAG
<u>Dnmt3l</u>	GCCCTTCTTCTGGATATTCATGGAC	ACTCTTCTTCTTGGGGTCCAG
<u>Retro- B1</u>	TGGTGGTGCATGOCCTTAAT	CCTGGTGTGCTGGAACCTACT ^(a)
<u>Retro- B2</u>	GGCTGGTGGAGATGGCTCAGT	TACACTGTAGCTGTCTTCAGACA ^(a)
<u>Retro- BM1</u>	CTAGAGCATTGGAAGGAGGTGC	ACCACTATAGAATCACAGTACCAC ^(b)
<u>Retro- L1</u>	GTGGGCGAAGAGGACGATGG	CTTGGCAGTCTAGGGGGC ^(c)
<u>Retro-MT</u>	ATGTCTTGGGGAGGACTGTG	AGCCCCAGCTAACAGAACT ^(d)
<u>Retro-V30</u>	CTTCATGAOCCAAOCCCTTCA	GTATGAGTTTCTTCTGCOA ^(b)

(a) A Allen T, et al 2004

(b) Willard J. Costain et al 2006

(c) Fransen E et al, 1998

(d) Peaston A. et al, 2004

Supplementary Table S13
The primers used for QPCR analysis

chr regions	forward 5'-->3'	reverse 5'-->3'
chr1-Adss	GTTAGTGGTGCTCAACGTCTACGA	TGAGAACTAAGCTGGAGGCGG
chr2-Stk35	TGAAGCAGACAACCACATCCA	TGTTAGTAGATGTAGGAAGACCCA
chr5-Rpl6	GTCAGCCATATCATCACCGAG	ACTACATGCCTTCTCCTTCCA
chr5-Anapc4	AAGTTCCCAAGACGTAATCCTTCC	GTGTTCTATGTTCTGATCCAGTC
chr6-Ldhd	ATACTGCATTAATCCATCCCATCC	CTCAGAATCCGAATCCGAATCAC
chr7-Klf13	GACTTCCATATGCTGACCCAC	CTTCCTTGCCTCATCTTCTGTC
chr7-Zfp619	TCACCAGCATTACAGAAACCA	GGGACAAAGTTAGCAAAGAAAGG
chr7-Lmo1	CCTGACTCTGCCATTTCTTCC	ATCCTGCTTTGCCCTCTGAC
chr11-Ilf1r2	TTTGCTTTGGGACTTCGGTG	TGCTGAATATGAATGAGAGGCGG
chr12-Vrk1	GAACCCACCTCTGTCTCCTTCC	GTCCACATAGAGCAACATCCGT
chr12-Cox16	ACCAATACGCCACTCAGCCAA	GCCAATCGTTTCCGTTTGCCT
chr13-Gm13498	TAGACTGCAAAGGACCCACTG	AAACGATCATCAGGAACGGG
chr13-Scamp1	AGAGTTTCCATCCCAAGCCA	TCTTAACCATTCTGCCTTCTTCC
chr17-Fndc1	ATCTTCTTCAATGTCCTCATCCTC	CTGCTGGAATGGAGTTGTATAGG
chrX-Par	GCGCTCTTCATTTATTGATTCGT	ACACAGGGACGGACAGACAG
Tel_length	CGGTTTGTTTGGGTTTGGGTTTGGGT TTGGGTTTGGGTT	GGCTTGCCTTACCCTTACCCTTACC CTTACCCTTACCCT
36B4_F	ACTGGTCTAGGACCCGAGAAG	TCAATGGTGCCTCTGGAGATT

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