

Usually Multiple Acids Move In and out Transporters 14 is an *Arabidopsis* amino acid exporter involved in phloem unloading of amino acids in roots

Supplemental tables S1-7

Table S1. List of primers used. Primers sequences are given from 5' to 3'. Underlined lower case represents bases added for the creation of *attB1* and *attB2* in forward and reverse primers, respectively.

Name	Type	Sequence
UMAMIT14 promoter	Forward	<u>acaagttgtacaaaaaaagcaggctc</u> CACTAAAATATTACATTGACC
	Reverse	accacttgcataagaagctgggtCTCATGTTTAAAGCCATATCTCAACGTAG
UMAMIT14 promoter (for modifying pPWYTk)	Forward	GAGAAGATCTCTCGAGGTACAAAAAAGCAGGCTGGCACTAAAATATTACATTGACC
	Reverse	TATAGAGCTCTGTTTAAAGCCATATCTCAACGTAG
UMAMIT14 genomic	Forward	<u>acaagttgtacaaaaaaagcaggctc</u> GATATGGCTTAAAACATGGAAG
	Reverse	accacttgcataagaagctgggtCTCAGACTGATTGTTAGGCCT
UMAMIT14 CDS	Forward	<u>gacaagttgtacaaaaaaagcaggctc</u> AGATATGGCTTAAAACATGGAAG
	Reverse	gaccacttgcataagaagctgggtACYAGACTGATTGTTAGGCCT
UMAMIT14 CDS (venus fusion)	Forward	GAGAGTCGACACCATGGCTTAAAACATGGAAG
	Reverse	ATATAGCGCTTGCAGTGATTGTTAGG
UMAMIT18 CDS	Forward	<u>gacaagttgtacaaaaaaagcaggctc</u> AATAAGATGAAAGTGGAAAGCATG
	Reverse	gaccacttgcataagaagctgggtACYAGGTACTGGTAACCACACCGTTAGT
UMAMIT14 T-DNA detection	Forward	CTATCAAATTGCATTACTTAGTGG
UMAMIT14 T-DNA detection	Reverse	GGCGATACGACGCTAGAACGTGAG
Actin2 qPCR	Forward	GGTAACATTGTGCTCAGTGGTGG
	Reverse	AACGACCTTAATCTTCATGCTGC
UMAMIT14 qPCR	Forward	ATGGTCATTGTTGCGATCTT
	Reverse	TGGTCGTCTTGCTTTTC

Table S2. Relative abundance of amino acids secreted by yeast expressing UMAMIT14 and UMAMIT18. Amino acid contents represented in Figure 2 were used. The average of content obtained for the empty plasmid was subtracted to the content of each biological replicates of UMAMIT14 and UMAMIT18. Each value, which represents amino acid secretion above the background level, was divided by the sum of these normalized values, and expressed as a percentage. Stars indicate significant difference between UMAMIT14 and UMAMIT18 results (t-test; p-value<0.5). GABA: gamma-amino-butyric acid.

	Gln+	Ala*	Glu*	Ser*	Gly*	Asn	Asp*	Pro*	Thr	Val*	Cys	His	Lys	Ile*	Leu*	Met	GABA	Phe	
	Arg*																		
UMAMIT18		77.8	5.8	3.7	1.8	1.8	1.7	-0.4	1.1	2.3	2.0	0.5	0.7	0.0	0.5	0.2	0.0	0.0	0.06
UMAMIT14		43.3	27.6	7.7	5.9	4.6	2.0	1.1	2.5	2.5	1.7	0.0	0.4	0.0	0.4	0.3	0.0	0.0	0.06

Table S3. Physiological traits of *umamit* mutants. Characteristics of nine-week-old *Arabidopsis* plants grown in soil in long day conditions. Biomass represents all plant tissue collected from the aerial parts, minus the seeds. Significant differences ($p<0.05$) are indicated by different letters according to one-way ANOVA in conjunction with Tukey's test (n=4 biological replicates).

Table S4. Shoot and root amino acid content of *umamit* mutants. Amino acids were extracted from five-week-old *Arabidopsis* plants growing in hydroponic conditions. Values are expressed in nmol.mg⁻¹ DW. Significant differences ($p<0.05$) are indicated by different letters according to ANOVA in conjunction with Tukey's test (n=4 biological replicates). n.d.: not detected.

Table S5. Amino acid secreted by *umamit* mutants. The data are the ones used in Figure 6. Plants were grown for two weeks in liquid J medium supplemented with 20 mM NH₄NO₃; the medium was replaced with fresh J medium and collected after three days for analysis. Values are expressed in nmol.mg⁻¹ DW. Significant differences (p<0.05) are indicated by different letters according to one-way ANOVA in conjunction with Tukey's test (n=6 biological replicates).

	Ala	Asn	Asp	Gaba	Gln	Glu	Gly	His	Ile	Leu	Phe	Ser	Thr	Val	Total
Wild Type	1.26 (a)	7.74 (a)	1.03 (a)	1.64 (a)	15.3 (a)	3.16 (a)	3.23 (a)	0.28 (a)	0.12 (a)	0.14 (a)	0.15 (a)	0.80 (a)	0.46 (a)	0.50 (a)	35.43 (a)
<i>umamit14-1</i>	1.09 (a)	5.83 (b)	0.86 (a)	0.98 (b)	10.4 (b)	2.57 (b)	2.45 (b)	0.21 (b)	0.08 (b)	0.09 (b)	0.10 (a)	0.56 (b)	0.32 (b)	0.11 (a)	26.80 (b)
<i>umamit18-1</i>	0.79 (b)	6.49 (a)	0.85 (a)	1.11 (b)	8.11 (b)	2.48 (b)	2.67 (b)	0.29 (a)	0.07 (b)	0.08 (b)	0.16 (a)	0.57 (b)	0.30 (b)	0.10 (b)	24.04 (b)
<i>umamit14-1</i>	0.34	3.07	0.71	0.47	1.90	1.74	2.32	0.29	0.02	0.02	0.13	0.32	0.12	0.03	11.47
<i>umamit18-1</i>	(c)	(d)	(b)	(b)	(c)	(c)	(b)	(a)	(c)	(c)	(a)	(c)	(c)	(c)	(c)

Table S6. CPM and dry weight of samples used in the shoot-to-root transfer assay using [³H]Gln.

	Sample	CPM					Dry weight (mg)			
		Fed leaf	Shoot	Root	Medium	Total	Fed leaf	Shoot	Root	Total
		1	6908	716	24	88	7648	0.6	2.71	0.58
WT	2	11967	2680	259	185	14907	0.4	2.88	0.72	4
	3	11000	1702	88	156	12791	0.35	3.36	0.44	4.15
	1	9648	1884	81	36	11613	0.81	2.75	0.89	4.45
<i>umamit14-1</i>	2	8101	1588	61	41	9750	0.47	2.88	1.1	4.45
	3	13739	2803	138	51	16681	0.31	2.45	1.45	4.21
	1	9374	3378	54	41	12807	0.71	2.72	1.19	4.62
<i>umamit14-1</i> <i>umamit18-1</i>	2	6877	3620	114	41.8	10611	0.29	1.9	1.34	3.53
	3	11742	3726	126	40.8	15595	0.33	2.97	0.96	4.26
	4	7623	2361	43	31	10027	0.34	2.24	0.97	3.55
<i>umamit14-1</i> <i>UMAMIT14</i>	1	5729	1480	174	126	7383	0.62	2.44	1.01	4.07
	2	7836	1098	144	112	9078	0.34	3.45	1.14	4.93
	3	9343	953	77	89	10373	0.77	2.81	0.84	4.42

Table S7. CPM and dry weight of samples used in the shoot-to-root transfer assay using [¹⁴C]sucrose. n.d. means the radioactivity found in the sample was similar to the background.

	Sample	CPM					Dry weight (mg)			
		Fed leaf	Shoot	Roots	Medium	Total	Fed leaf	Shoots	Roots	Total
WT	1	5618	8982	3355	n.d.	17955	0.35	2.85	1.07	4.27
	2	8205	11955	1958	n.d.	22118	0.42	3.16	0.98	4.56
	3	8496	12928	2915	n.d.	24339	0.27	3.52	0.62	4.41
<i>umamit14-1</i>	1	6806	4677	1574	n.d.	13057	0.37	3.4	0.7	4.47
	2	5552	8440	2281	n.d.	16273	0.51	2.72	0.84	4.07
	3	11804	6834	1626	n.d.	20264	0.44	2.67	0.65	3.76
<i>umamit14-1</i> <i>umamit18-1</i>	1	6707	9296	1625	n.d.	17628	0.48	2.56	0.83	3.87
	2	4679	4364	1805	n.d.	10848	0.41	2.27	0.79	3.47
	3	5687	9341	1926	n.d.	16954	0.36	2.21	0.9	3.47
<i>umamit14-1</i> <i>UMAMIT14-1</i>	1	5743	8070	2193	n.d.	16006	0.43	2.97	0.85	4.25
	2	4589	10473	2482	n.d.	17544	0.37	2.7	0.67	3.74
	3	9563	11386	3097	n.d.	24046	0.33	2.33	0.41	3.07