

Supporting Information

Bioprinting 3D Microfibrous Scaffolds for Engineering Endothelialized Myocardium and Heart-on-a-Chip

Yu Shrike Zhang^{1,2,3,*}, Andrea Arneri^{1,2,4,†}, Simone Bersini^{1,2,5,†}, Su-Ryon Shin^{1,2,3}, Kai Zhu^{1,2,6}, Zahra Goli Malekabadi^{1,2,7}, Julio Aleman^{1,2}, Cristina Colosi^{1,2,8}, Fabio Busignani^{1,2,9}, Valeria Dell'Erba^{1,2,10}, Colin Bishop¹¹, Thomas Shupe¹¹, Danilo Demarchi⁹, Matteo Moretti⁵, Marco Rasponi⁴, Mehmet Remzi Dokmeci^{1,2,3}, Anthony Atala¹¹, Ali Khademhosseini^{1,2,3,12,13,*}

¹Biomaterials Innovation Research Center, Division of Biomedical Engineering, Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, Cambridge, MA 02139, USA

²Harvard-MIT Division of Health Sciences and Technology, Massachusetts Institute of Technology, Cambridge, MA 02139, USA

³Wyss Institute for Biologically Inspired Engineering, Harvard University, Cambridge, MA 02139, USA

⁴Bioengineering Department, Politecnico di Milano, Milan 20133, Italy

⁵Cell and Tissue Engineering Lab, IRCCS Istituto Ortopedico Galeazzi, Milan 20161, Italy

⁶Department of Cardiac Surgery, Zhongshan Hospital, Fudan University, Shanghai 210032, China

⁷Department of Biomechanical Engineering, Amirkabir University of Technology, Tehran 64540, Iran

⁸Department of Chemistry, Sapienza Università di Roma, Rome, 00185, Italy

⁹Department of Electronics and Telecommunications, Politecnico di Torino, Torino 10129, Italy

¹⁰Department of Biomedical Engineering, Politecnico di Torino, Torino 10129, Italy

¹¹Wake Forest Institute for Regenerative Medicine, Winston-Salem, NC 27101, USA

¹²Department of Bioindustrial Technologies, College of Animal Bioscience and Technology, Konkuk University, Seoul 143-701, Republic of Korea

¹³Department of Physics, King Abdulaziz University, Jeddah 21569, Saudi Arabia

E-mails: alik@bwh.harvard.edu, yszhang@research.bwh.harvard.edu

[†] A.A. and S.B. contributed equally to this work.

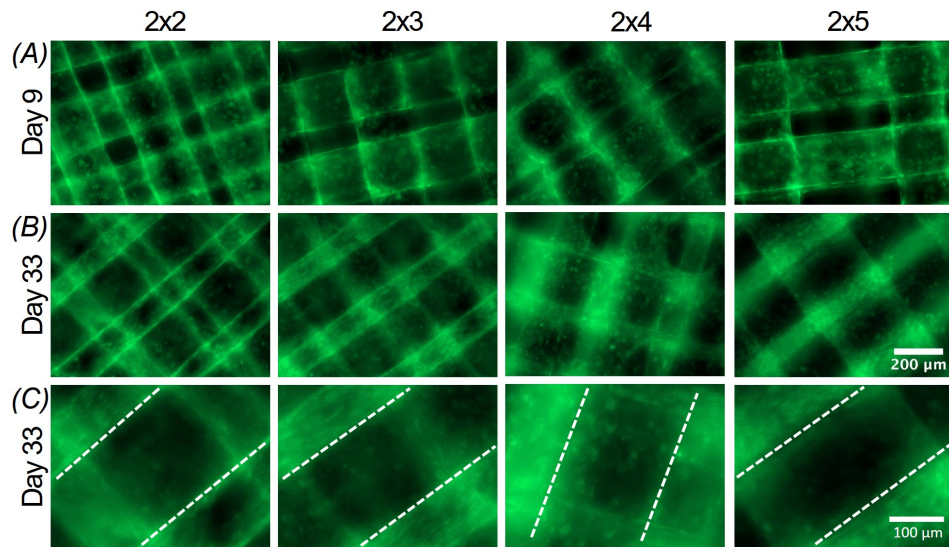


Figure S1. (A, B) Fluorescence micrographs showing the spreading of GFP-HUVECs in the bioprinted microfibrinous scaffolds with different aspect ratios of unit grids at Day 9 and Day 33. (C) High-magnification fluorescence micrographs showing the spreading of GFP-HUVECs in the bioprinted microfibrinous scaffolds with different aspect ratios of unit grids at Day 33. The dotted lines in each panel represent a microfiber in between, showing the outward migrated HUVECs at the bottom of the culture well.

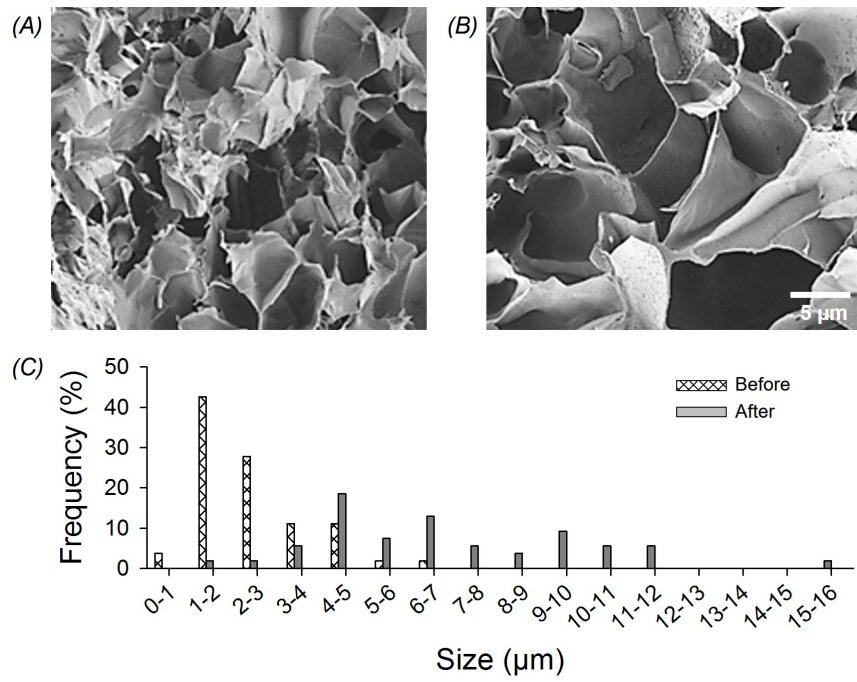
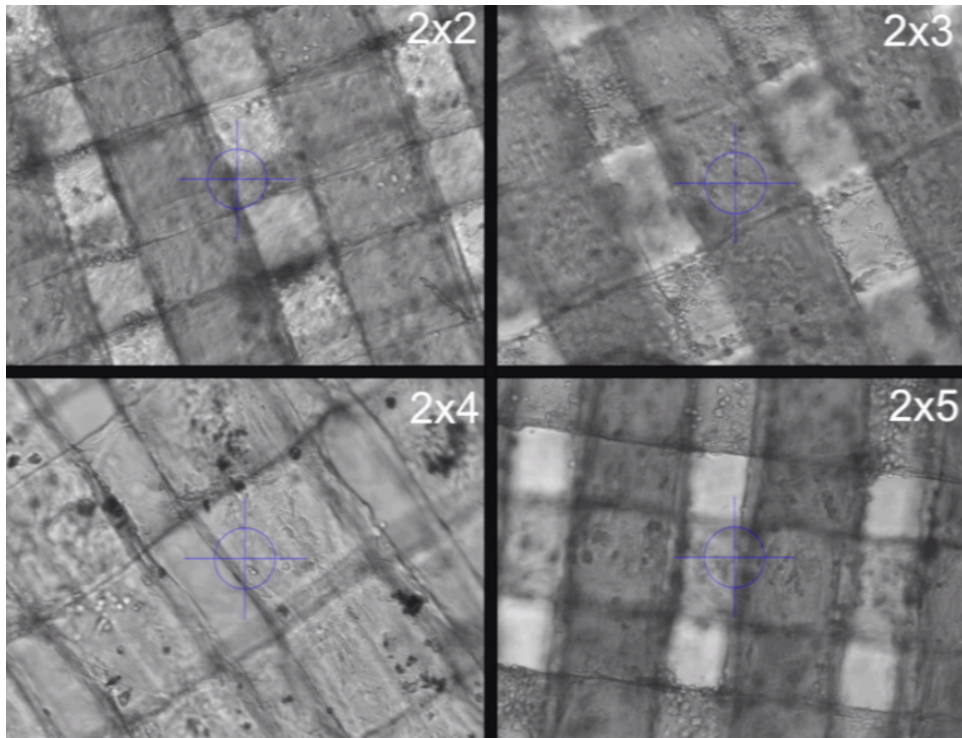
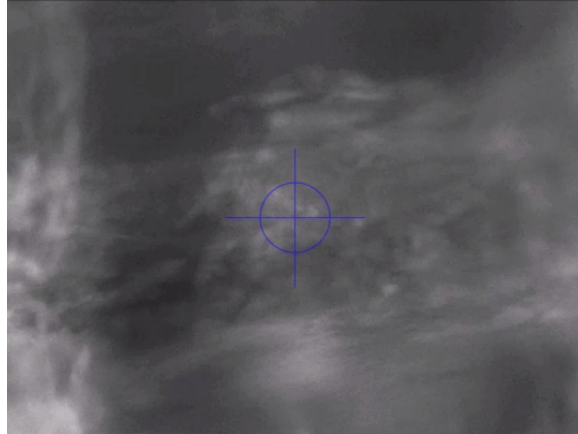


Figure S2. (A, B) SEM images showing the change in pore size of the bioprinted microfibers in regions close to the peripheries, (A) before and (B) after alginate removal. (C) Pore size distribution for the microfibers before and after alginate removal.



Movie S1. Videos showing the beating of the myocardial tissues on bioprinted microfibrous scaffolds with different aspect ratios of unit cells.



Movie S2. Video taken in fluorescence mode showing the beating of the endothelialized myocardial tissue construct. The HUVECs forming the endothelium inside the microfibers were GFP-fluorescent.