

Analytical and Bioanalytical Chemistry

Electronic Supplementary Material

**It is all about the solvent: on the importance of the mobile phase for
ZIC-HILIC glycopeptide enrichment**

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Table_S6A - 216_2016_51_MOESM2_ESM.pdf

MATERIALS AND METHODS

Table S1 Ion trap settings applied in this study, following the MIRAGE guidelines (www.beilstein-mirage.org)

MS Settings - General	
ESI probe	CaptiveSpray™
Capillary voltage	1.3 kV
SPS	<i>m/z</i> 900
Compound stability	100%
Trap Drive Level	100%
Spectra averaging	5
Dry gas temperature	150°C
Dry Gas flow	3 L/min
Maximum accumulation time	200 ms
Ion mode	positive
MS-Scan	
MS Scan mode	Enhanced scan
ICC target	200000
Mass detection range	<i>m/z</i> 350-1800
MS2	
MS scan mode	ultrascan
SPS MS(n)	automatic
MS(n) spectra averages	5
MS(n) ICC target	40000
Preferred charge state	≥Doubly,
Active exclusion	Off
Mass detection range	<i>m/z</i> 100-2000
Isolation width	3 Da
Exclude singly charged ions	on
No. of precursor ions	3
SmartFrag	Enhanced
SmartFrag Start amplitude	30%
SmartFrag End amplitude	120%
Fragmentation width	5 <i>m/z</i>
Resolution	
Ultra Scan	32,000 u/s @ 0.5 u FWHM
Enhanced Scan	8,100 u/s @ 0.3 u FWHM

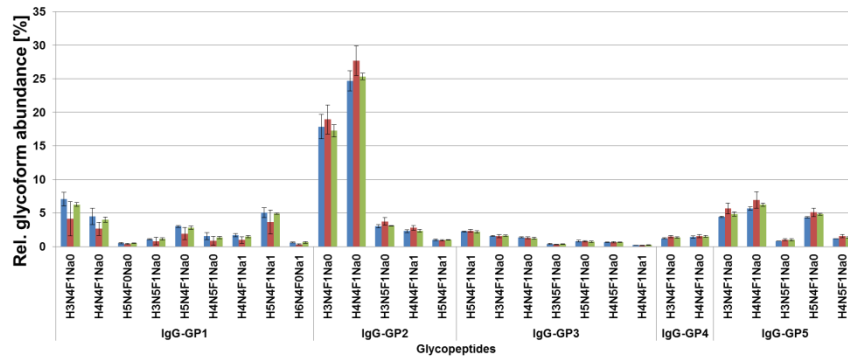
Glycopeptide synthesis

Standard amino acids for SPPS were purchased from Iris Biotech GmbH (Marktredwitz, Germany), and polystyrene and NovaPEG resin were from Novabiochem (Darmstadt, Germany). Fmoc-labelled asparagine (Asn) carrying a disialylated biantennary N-glycan was initially prepared from egg yolks as described previously [1,2]. Sialic acid residues were selectively protected by esterification with benzyl bromide prior their use in SPPS [3]. The coupling of the glycosylated Asn building blocks was performed as described by Unverzagt and co-workers [4]. Synthesised glycopeptides were purified using preparative C18-RP HPLC on an Agilent 1200 series ELSD HPLC system (Agilent, Santa Clara, CA). The lyophilisate was dissolved in water and subjected to C18-RP chromatography (XBridge BEH C18 column, 130 Å, 3.5 µm, 4.6 mm x 150 mm, Waters, Milford, MA) using a linear gradient (1 mL/min) from 1% to 50% acetonitrile containing 0.1% TFA within 35 min, followed by an increase to 100% over 10 min. Fractions of interest from several runs were combined and analysed by MS to confirm their composition.

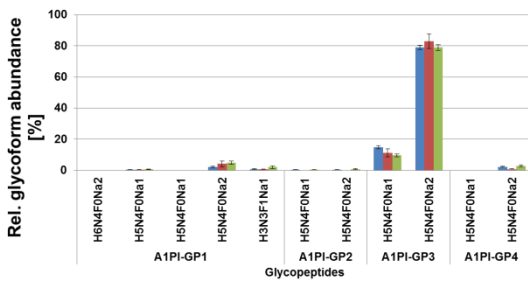
RESULTS AND DISCUSSION

Incubation Time Does Not Influence Drop-HILIC Enrichment Efficiency

A



B



Legend

H – Hex
N – HexNAc
F – Fuc
Na – Neu5Ac

■ ACN Drop 1 min
■ ACN Drop 3 min
■ ACN Drop 5 min

IgG2-GP1: ¹⁶⁸TKPREEQFNSTFR¹⁸⁰
IgG2-GP2: ¹⁷²EEQFNSTFR¹⁸⁰
IgG1-GP3: ¹⁷²TKPREEQYNSTYR¹⁸⁴
IgG4-GP4: ¹⁷³EEQFNSTYR¹⁸¹
IgG1-GP5: ¹⁷⁶EEQYNSTYR¹⁸⁴

A1PI-GP1: ⁶⁴QLAHQSNSTNIFFSPVSIATAFAMLSLGTK⁹³
A1PI-GP2: ⁹⁴ADTHDEILEGLNFNLTPEAQIHEGFQELLR¹²⁵
A1PI-GP3: ²⁶⁸YLGNATAIFFLPDEGK²⁸³
A1PI-GP4: ²⁶⁸YLGNATAIFFLPDEGKQLQHLENLTHDIITK²⁹⁸

Fig. S1 Effect of incubation time on Drop HILIC efficiency

Table S2 List of IgG glycopeptides identified after HILIC enrichment

Table S2A Raw Data after quantification of identified glycopeptides

SNO	m/z	RT	Z	Calculate d mass	Glycan	Peptide	Hex	HexNAc	H	N	F	Na	Glycan Name	ACN spin E1	ACN spin E2	ACN spin E3	ACN drop 1 min E1	ACN drop 1 min E2	ACN drop 1 min E3	ACN Drop 3 min E1	ACN Drop 3 min E2	ACN Drop 3 min E3	ACN Drop 5 min E1	ACN Drop 5 min E2	ACN Drop 5 min E3	EtOH drop E1	EtOH drop E2	EtOH drop E3	IPA drop E1	IPA drop E2	IPA drop E3	
1	1028.89	17.1	3	3083.72	HexNAc2 (Deoxyhexose)1 + (Man)3(GlcNAc)2	168-180 TKPREQFNSTR	0	2	3	4	1	0	H3N4F1NA0	2.66E+08	5.00E+08	2.12E+08	6.21E+08	4.79E+08	6.61E+08	1.17E+08	2.90E+08	5.78E+08	5.12E+08	4.69E+08	4.75E+08	1.37E+08	2.12E+08	4.73E+08	5.95E+08	5.95E+08		
2	1082.89	17.3	3	3245.84	Hex1(HexNAc)2(Deoxyhexose)1 + (Man)3(GlcNAc)2	168-180 TKPREQFNSTR	1	2	4	4	1	0	H4N4F1NA0	1.97E+08	2.94E+08	1.53E+08	4.89E+08	3.08E+08	3.16E+08	1.02E+08	1.79E+08	3.05E+08	3.12E+08	3.15E+08	2.96E+08	1.53E+08	1.07E+08	1.62E+08	3.19E+08	3.40E+08	3.65E+08	
3	1088.21	16.2	3	3261.59	Hex1(HexNAc)2 + (Man)3(GlcNAc)2	168-180 TKPREQFNSTR	2	2	5	4	0	0	H5N4F0NA0	3.00E+07	3.68E+07	1.67E+07	4.53E+07	2.79E+07	4.70E+07	1.75E+07	2.95E+07	3.80E+07	4.49E+07	3.18E+07	3.90E+07	2.05E+07	1.71E+07	1.55E+07	3.16E+07	3.03E+07	4.36E+07	
4	1096.52	17.3	3	3286.53	HexNAc3(Deoxyhexose)1 + (Man)3(GlcNAc)2	168-180 TKPREQFNSTR	0	3	5	5	1	0	H5N5F1NA0	4.69E+07	6.27E+07	3.61E+07	8.82E+07	8.28E+07	9.80E+07	2.10E+07	4.42E+07	1.22E+08	1.11E+08	6.94E+07	9.09E+07	2.76E+07	3.24E+07	3.34E+07	1.12E+08	1.10E+08	1.32E+08	
5	1136.84	17.1	3	3407.51	Hex1(HexNAc)2(Deoxyhexose)1 + (Man)3(GlcNAc)2	168-180 TKPREQFNSTR	2	2	5	4	1	0	H5N4F1NA0	1.29E+08	2.08E+08	9.97E+07	2.55E+08	2.39E+08	2.59E+08	6.14E+07	1.31E+08	2.37E+08	2.45E+08	1.81E+08	2.32E+08	1.81E+08	2.32E+08	1.81E+08	2.32E+08	2.11E+08	2.42E+08	
6	1150.52	17.4	3	3448.54	Hex1(HexNAc)3(Deoxyhexose)1 + (Man)3(GlcNAc)2	168-180 TKPREQFNSTR	1	3	4	5	1	0	H5N5F1NA0	3.98E+07	1.22E+08	3.55E+07	1.25E+08	8.60E+07	1.75E+08	1.78E+07	4.37E+07	1.34E+08	1.21E+08	7.89E+07	1.10E+08	3.04E+07	3.25E+07	4.93E+07	8.12E+07	1.02E+08	1.31E+08	
7	1185.15	18.1	4	3536.58	Hex1(HexNAc)2(Deoxyhexose)1 + (Man)3(GlcNAc)2	168-180 TKPREQFNSTR	1	2	4	4	1	0	H4N4F1NA1	7.49E+07	9.57E+07	5.93E+07	1.37E+08	1.20E+08	1.66E+08	3.36E+07	5.99E+07	1.27E+08	1.16E+08	1.01E+08	1.32E+08	4.39E+07	4.90E+07	4.90E+07	1.29E+08	1.41E+08		
8	1234.18	18.2	3	3698.67	Hex1(HexNAc)2(Deoxyhexose)1 (NeuAc)1 + (Man)3(GlcNAc)2	168-180 TKPREQFNSTR	2	2	5	4	1	1	H5N4F1NA1	2.38E+08	3.14E+08	1.69E+08	4.13E+08	3.44E+08	4.88E+08	1.20E+08	2.32E+08	4.64E+08	4.05E+08	3.52E+08	3.88E+08	1.68E+08	1.49E+08	1.63E+08	3.12E+08	3.43E+08	4.32E+08	
9	1239.21	16.8	3	3714.6	Hex1(HexNAc)2 (NeuAc)1 + (Man)3(GlcNAc)2	168-180 TKPREQFNSTR	3	2	6	4	0	0	H6N4F0NA1	1.69E+07	2.50E+07	1.49E+07	4.98E+07	4.56E+07	1.25E+07	2.54E+07	2.85E+07	6.36E+07	3.11E+07	4.84E+07	2.81E+07	2.11E+07	2.11E+07	2.11E+07	4.15E+07	4.86E+07	5.58E+07	
10	1301.74	19.6	2	2601.15	HexNAc2(Deoxyhexose)1 + (Man)3(GlcNAc)2	172-180 EQGNFSTR	0	2	3	4	1	0	H3N4F1NA0	1.37E+09	1.31E+09	1.38E+09	1.32E+09	1.58E+09	1.53E+09	1.25E+09	1.34E+09	1.40E+09	1.55E+09	1.17E+09	1.32E+09	1.15E+09	1.06E+09	1.17E+09	1.37E+09	1.59E+09	1.68E+09	
11	1301.74	19.6	2	2601.15	HexNAc2(Deoxyhexose)1 + (Man)3(GlcNAc)2	172-180 EQGNFSTR	0	2	3	4	1	0	H3N4F1NA0	1.37E+09	1.31E+09	1.38E+09	1.32E+09	1.58E+09	1.53E+09	1.25E+09	1.34E+09	1.40E+09	1.55E+09	1.17E+09	1.32E+09	1.15E+09	1.06E+09	1.17E+09	1.37E+09	1.59E+09	1.68E+09	
12	1403.08	19.6	2	2804.15	HexNAc3(Deoxyhexose)1 + (Man)3(GlcNAc)2	172-180 EQGNFSTR	0	3	5	5	1	0	H3N5F1NA0	2.44E+08	2.52E+08	2.30E+08	2.40E+08	2.64E+08	2.44E+08	2.51E+08	2.71E+08	2.61E+08	2.62E+08	2.17E+08	2.36E+08	2.15E+08	1.97E+08	2.06E+08	2.15E+08	2.38E+08	2.47E+08	
13	1403.08	19.6	2	2804.15	HexNAc3(Deoxyhexose)1 + (Man)3(GlcNAc)2	172-180 EQGNFSTR	1	2	4	4	1	1	H4N4F1NA1	1.97E+08	2.08E+08	1.72E+08	1.95E+08	1.95E+08	1.77E+08	1.80E+08	2.06E+08	2.00E+08	1.87E+08	1.82E+08	1.76E+08	1.58E+08	1.43E+08	1.52E+08	1.69E+08	1.85E+08	1.93E+08	
14	1403.08	19.6	2	2804.15	HexNAc3(Deoxyhexose)1 (NeuAc)1 + (Man)3(GlcNAc)2	172-180 EQGNFSTR	2	2	5	4	1	1	H5N4F1NA1	7.81E+07	7.75E+07	5.78E+07	8.19E+07	7.27E+07	9.21E+07	6.00E+07	6.54E+07	6.67E+07	8.31E+07	7.10E+07	7.72E+07	5.75E+07	8.53E+07	1.31E+08	1.44E+08	1.48E+08		
15	1424.84	16	3	1244.08	Hex1(HexNAc)2(Deoxyhexose)1 (NeuAc)1 + (Man)3(GlcNAc)2	172-184 TKPREQFNSTR	2	2	5	4	1	1	H5N4F1NA1	1.62E+08	1.70E+08	1.39E+08	1.84E+08	1.86E+08	1.85E+08	1.24E+08	1.77E+08	2.00E+08	1.81E+08	1.42E+08	1.86E+08	1.34E+08	1.51E+08	1.55E+08	1.60E+08	1.66E+08	1.85E+08	
16	1039.52	15.4	3	3115.54	HexNAc2(Deoxyhexose)1 + (Man)3(GlcNAc)2	172-184 TKPREQFNSTR	0	2	3	4	1	0	H3N4F1NA0	1.18E+08	1.43E+08	1.09E+08	1.29E+08	1.20E+08	1.38E+08	7.51E+07	1.15E+08	1.44E+08	1.28E+08	1.14E+08	1.35E+08	9.23E+07	8.82E+07	9.84E+07	1.20E+08	1.20E+08	1.61E+08	
17	1093.54	15.3	3	3278.76	Hex1(HexNAc)2(Deoxyhexose)1 + (Man)3(GlcNAc)2	172-184 TKPREQFNSTR	1	2	4	4	1	0	H4N4F1NA0	1.10E+08	1.10E+08	8.36E+07	1.07E+08	1.12E+08	1.19E+08	6.62E+07	9.88E+07	1.09E+08	1.15E+08	7.57E+07	9.84E+07	9.22E+07	9.29E+07	8.76E+07	1.05E+08	1.57E+08	1.95E+08	
18	1107.19	15.6	3	3318.54	HexNAc3(Deoxyhexose)1 + (Man)3(GlcNAc)2	172-184 TKPREQFNSTR	0	3	5	5	1	0	H3N5F1NA0	1.88E+07	3.13E+07	1.80E+07	3.92E+07	2.61E+07	2.97E+07	1.48E+07	1.88E+07	2.76E+07	2.87E+07	2.81E+07	2.94E+07	1.81E+07	1.92E+07	2.11E+07	3.75E+07	3.73E+07	3.89E+07	
19	1147.51	15.2	3	3439.49	Hex1(HexNAc)2(Deoxyhexose)1 + (Man)3(GlcNAc)2	172-184 TKPREQFNSTR	2	2	5	4	1	0	H5N4F1NA0	6.75E+07	6.78E+07	5.57E+07	7.44E+07	5.52E+07	7.24E+07	3.95E+07	6.40E+07	6.40E+07	5.97E+07	6.24E+07	4.47E+07	6.24E+07	7.33E+07	5.40E+07	7.42E+07	9.15E+07	7.68E+07	
20	1161.19	15.4	3	3480.56	Hex1(HexNAc)3(Deoxyhexose)1 + (Man)3(GlcNAc)2	172-184 TKPREQFNSTR	1	3	4	5	1	0	H4N5F1NA0	4.62E+07	5.68E+07	4.06E+07	5.82E+07	5.04E+07	4.92E+07	3.29E+07	4.95E+07	5.62E+07	5.21E+07	4.93E+07	4.90E+07	3.44E+07	3.04E+07	3.51E+07	4.85E+07	4.50E+07	7.29E+07	
21	1190.52	16	3	3568.54	Hex1(HexNAc)2(Deoxyhexose)1 (NeuAc)1 + (Man)3(GlcNAc)2	172-184 TKPREQFNSTR	1	2	4	4	1	1	H5N4F1NA1	1.53E+07	1.41E+07	1.20E+07	1.69E+07	1.41E+07	1.65E+07	1.24E+07	1.97E+07	1.57E+07	1.89E+07	1.19E+07	1.20E+07	9.54E+06	1.09E+07	1.05E+07	1.48E+07	1.92E+07	2.01E+07	
22	1309.56	18	2	2617.1	HexNAc2(Deoxyhexose)1 + (Man)3(GlcNAc)2	173-181 EQGNFSTR	0	2	3	4	1	0	H3N4F1NA0	9.72E+07	9.39E+07	8.34E+07	1.05E+08	9.03E+07	1.05E+08	9.70E+07	1.03E+08	1.09E+08	1.05E+08	1.03E+08	9.93E+07	8.01E+07	5.94E+07	6.45E+07	6.74E+07	8.63E+07	9.62E+07	
23	1390.57	17.7	2	2779.14	Hex1(HexNAc)2(Deoxyhexose)1 + (Man)3(GlcNAc)2	173-181 EQGNFSTR	0	2	4	4	1	0	H3N4F1NA0	1.01E+08	1.05E+08	8.54E+07	1.29E+08	1.00E+08	1.22E+08	1.10E+08	1.01E+08	1.18E+08	1.30E+08	1.16E+08	1.04E+08	9.81E+07	7.72E+07	8.88E+07	1.19E+08	1.21E+08		
24	1317.65	16.4	2	2634.06	HexNAc2(Deoxyhexose)1 + (Man)3(GlcNAc)2	176-184 EQGNFSTR	0	2	3	4	1	0	H3N4F1NA0	3.61E+08	3.81E+08	3.37E+08	3.57E+08	3.61E+08	3.76E+08	3.84E+08	4.05E+08	4.03E+08	3.80E+08	3.68E+08	3.75E+08	3.23E+08	2.82E+08	3.26E+08	3.25E+08	3.48E+08	3.62E+08	
25	932.73	16.3	3	2795.15	HexNAc3(Deoxyhexose)1 + (Man)3(GlcNAc)2	176-184 EQGNFSTR	1	2	4	4	1	0	H3N4F1NA0	5.18E+08	5.11E+08	4.60E+08	4.75E+08	4.72E+08	4.57E+08	4.80E+08	4.98E+08	4.70E+08	5.44E+08	4.26E+08	4.89E+08	4.76E+08	4.46E+08	4.43E+08	4.91E+08	5.36E+08	5.31E+08	
26	946.41	16.7	3	2836.2	HexNAc3(Deoxyhexose)1 + (Man)3(GlcNAc)2	176-184 EQGNFSTR	0	3	5	5	1	0	H3N5F1NA0	7.25E+07	7.04E+07	6.12E+07	6.61E+07	6.76E+07	6.74E+07	6.65E+07	7.44E+07	7.37E+07	8.55E+07	7.95E+07	6.36E+07	6.34E+07	5.55E+07	7.34E+07	6.60E+07	6.27E+07		
27	1479.59	16.5	2	2957.16	Hex1(HexNAc)2(Deoxyhexose)1 + (Man)3(GlcNAc)2	176-184 EQGNFSTR	2	2	5	4	1	0	H5N4F1NA0	3.60E+08	3.75E+08	3.20E+08	3.55E+08	3.48E+08	3.80E+08	3.38E+08	3.62E+08	3.99E+08	3.36E+08	3.85E+08	3.49E+08	2.85E+08	3.35E+08	3.35E+08	3.67E+08	3.51E+08		
28	1000.4	16.6	3	2998.19	Hex1(HexNAc)3(Deoxyhexose)1 + (Man)3(GlcNAc)2	176-184 EQGNFSTR	1	3	4	5	1	0	H4N5F1NA0	9.77E+07	1.07E+08	8.98E+07	9.61E+07	9.51E+07	9.76E+07	1.06E+08	1.11E+08	1.07E+08	1.12E+08	1.05E+08	1.00E+08	1.03E+08	7.72E+07	9.50E+07	9.09E+07	9.11E+07	9.21E+07	

Table S2B Normalized Data after quantification of the identified glycopeptides

SNO	m/z	RT	Z	Calculate d mass	Glycan	Peptide	Hex	HexNAc	H	N	F	Na	Glycan Name	ACN spin E1	ACN spin E2	ACN spin E3	ACN drop 1 min E1	
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Table S3 List of A1PI glycopeptides identified after HILIC enrichment

Table S3A Raw Data after quantification of identified glycopeptides

SNO	m/z	RT	Calculate d mass	Glycan	PEP	Hex	HexNAc	H	N	F	Na	Glycan Name	ACN spin E1	ACN spin E2	ACN spin E3	ACN drop 1 min E1	ACN drop 1 min E2	ACN drop 1 min E3	ACN Drop 3 min E1	ACN Drop 3 min E2	ACN Drop 3 min E3	ACN Drop 5 min E1	ACN Drop 5 min E2	ACN Drop 5 min E3	EtOH drop E1	EtOH drop E2	EtOH drop E3	IPA drop E1	IPA drop E2	IPA drop E3		
1	1388.55	40.8	4	5547.36	(Hex)3(HexNAc)2(NeuAc)2+(Man)3(GlcNAc)2	64-93QLAHQSNSTNIFFSPVSIATAFAMLSLGTK	3	2	5	4	0	2	HSN4F0N2	2.97E+07	1.47E+07	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00
2	1352.08	40.9	4	5401.4	(Hex)2(HexNAc)2(NeuAc)2+(Man)3(GlcNAc)2	64-93QLAHQSNSTNIFFSPVSIATAFAMLSLGTK	2	2	5	4	0	1	HSN4F0N1	6.53E+08	3.90E+08	3.14E+08	2.11E+07	3.22E+07	1.11E+08	3.57E+07	4.21E+07	2.52E+07	4.08E+08	8.96E+07	8.98E+07	5.74E+07	1.43E+08	1.36E+08	2.94E+08	2.24E+09	1.93E+09	
3	1279.38	38.4	4	5110.52	(Hex)2(HexNAc)2(NeuAc)1+(Man)3(GlcNAc)2	64-93QLAHQSNSTNIFFSPVSIATAFAMLSLGTK	2	2	5	4	0	1	HSN4F0N1	5.97E+07	6.28E+07	5.91E+07	2.88E+06	8.83E+06	1.40E+07	3.48E+06	6.64E+06	2.45E+06	4.51E+07	9.30E+06	1.18E+07	7.59E+06	1.53E+07	1.68E+07	2.80E+08	1.97E+08	1.97E+08	
4	1275.36	39.7	4	5094.56	(Hex)2(HexNAc)2(NeuAc)1+(Man)3(GlcNAc)2	64-93QLAHQSNSTNIFFSPVSIATAFAMLSLGTK	2	2	5	4	0	2	HSN4F0N2	1.45E+07	2.01E+07	1.79E+07	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00
5	1475.63	38.1	3	4422.39	(HexNAc)1(Deoxyhexose)1+(Man)3(GlcNAc)2	64-93QLAHQSNSTNIFFSPVSIATAFAMLSLGTK	0	1	3	3	1	1	HSN3F1N1	2.99E+08	2.24E+08	1.68E+08	4.40E+06	1.24E+07	4.09E+07	4.33E+06	3.65E+06	3.79E+06	2.14E+08	3.04E+07	3.31E+07	3.67E+07	7.75E+07	8.38E+07	1.76E+09	1.06E+09	9.47E+08	
6	1402.85	36	4	5604.36	(Hex)2(HexNAc)2(NeuAc)1+(Man)3(GlcNAc)2	94-125ADTHDELEGLNFLNLTPEAQIHGEGFQELLR	2	2	5	4	0	1	HSN4F0N1	2.66E+07	3.18E+07	4.12E+07	3.32E+06	6.90E+06	7.84E+06	0.00E+00	0.00E+00	0.00E+00	0.00E+00	3.43E+07	5.16E+06	5.89E+06	6.26E+06	1.32E+07	1.21E+07	1.52E+08	9.19E+07	1.21E+08
7	1180.83	38	5	5896.05	(Hex)2(HexNAc)2(NeuAc)2+(Man)3(GlcNAc)2	94-125ADTHDELEGLNFLNLTPEAQIHGEGFQELLR	2	2	5	4	0	2	HSN4F0N2	9.37E+07	6.59E+07	6.22E+07	1.15E+08	6.55E+08	1.61E+07	0.00E+00	0.00E+00	0.00E+00	7.09E+07	1.29E+07	1.20E+07	1.29E+07	2.23E+07	2.63E+07	4.40E+08	2.83E+08	2.30E+08	
8	1224.6	39	3	3668.7	(Hex)2(HexNAc)2(NeuAc)1+(Man)3(GlcNAc)2	268-298YLGNAITAIFFLPDEGK	2	2	5	4	0	1	HSN4F0N1	2.77E+08	4.02E+08	3.72E+08	1.68E+08	2.44E+08	6.08E+08	1.21E+08	1.45E+08	4.73E+07	5.92E+08	1.63E+08	2.47E+08	1.16E+08	2.22E+08	2.85E+08	7.13E+08	6.85E+08	6.88E+08	
9	1321.29	34.7	3	3959.67	(Hex)2(HexNAc)2(NeuAc)2+(Man)3(GlcNAc)2	268-298YLGNAITAIFFLPDEGK	2	2	5	4	0	2	HSN4F0N2	3.56E+08	2.92E+09	2.36E+09	9.21E+08	1.19E+09	3.44E+09	7.22E+08	1.55E+09	5.24E+08	1.41E+09	1.83E+09	1.35E+09	2.28E+09	2.77E+09	8.12E+09	8.20E+09	7.02E+09		
10	1092.17	34.6	5	5453.8	(Hex)2(HexNAc)2(NeuAc)1+(Man)3(GlcNAc)2	268-298YLGNAITAIFFLPDEGKLOHLENLTHDITK	2	2	5	4	0	1	HSN4F0N1	1.64E+07	6.15E+07	4.29E+07	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00	0.00E+00
11	1150.36	35.9	5	5744.95	(Hex)2(HexNAc)2(NeuAc)2+(Man)3(GlcNAc)2	268-298YLGNAITAIFFLPDEGKLOHLENLTHDITK	2	2	5	4	0	2	HSN4F0N2	2.89E+08	5.49E+08	3.75E+08	1.88E+07	2.69E+07	9.62E+07	6.43E+06	4.73E+06	4.32E+06	1.66E+08	3.58E+07	6.92E+07	4.10E+07	8.86E+07	7.69E+07	6.33E+08	3.33E+08	6.00E+08	
12	1438.8	36	4	5744.24	(Hex)2(HexNAc)2(NeuAc)2+(Man)3(GlcNAc)2	268-298YLGNAITAIFFLPDEGKLOHLENLTHDITK	2	2	5	4	0	2	HSN4F0N2	5.88E+09	8.11E+07	5.53E+07	5.47E+06	4.18E+06	1.87E+07	0.00E+00	0.00E+00	0.00E+00	3.03E+07	2.74E+06	1.05E+07	7.15E+06	1.84E+07	1.48E+07	1.36E+08	6.96E+07	1.27E+08	
													5.38E+09	4.83E+09	3.89E+09	1.15E+09	1.53E+09	4.36E+09	8.93E+08	1.75E+09	4.14E+08	6.81E+09	1.75E+09	2.31E+09	1.64E+09	2.89E+09	3.43E+09	1.54E+10	1.33E+10	1.20E+10		

Table S3B Normalized Data after quantification of the identified glycopeptides

	m/z	RT	Calculate d mass	Glycan	PEP	Hex	HexNAc	H	N	F	Na	Glycan Name	ACN spin E1	ACN spin E2	ACN spin E3	ACN drop 1 min E1	ACN drop 1 min E2	ACN drop 1 min E3	ACN Drop 3 min E1	ACN Drop 3 min E2	ACN Drop 3 min E3	ACN Drop 5 min E1	ACN Drop 5 min E2	ACN Drop 5 min E3	EtOH drop E1	EtOH drop E2	EtOH drop E3	IPA drop E1	IPA drop E2	IPA drop E3		
1	1388.55	40.8	4	5547.36	(Hex)3(HexNAc)2(NeuAc)2+(Man)3(GlcNAc)2	64-93QLAHQSNSTNIFFSPVSIATAFAMLSLGTK	3	2	5	4	0	2	HSN4F0N2	0.55	0.30	0.41	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.92	0.63	0.65	
2	1352.08	40.9	4	5401.4	(Hex)2(HexNAc)2(NeuAc)2+(Man)3(GlcNAc)2	64-93QLAHQSNSTNIFFSPVSIATAFAMLSLGTK	2	2	5	4	0	1	HSN4F0N1	12.14	8.09	8.08	1.84	2.10	2.54	3.99	2.41	6.10	5.99	4.77	3.89	3.50	4.93	3.95	19.09	16.84	16.12	
3	1279.38	38.4	4	5110.52	(Hex)2(HexNAc)2(NeuAc)1+(Man)3(GlcNAc)2	64-93QLAHQSNSTNIFFSPVSIATAFAMLSLGTK	2	2	5	4	0	1	HSN4F0N1	1.11	1.30	1.52	0.25	0.58	0.32	0.39	0.38	0.59	0.66	0.53	0.51	0.46	0.53	0.49	1.82	1.48	1.64	
4	1275.36	39.7	4	5094.56	(Hex)2(HexNAc)2(NeuAc)1+(Man)3(GlcNAc)2	64-93QLAHQSNSTNIFFSPVSIATAFAMLSLGTK	2	2	5	4	0	2	HSN4F0N2	0.27	0.42	0.46	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.19	0.23	0.22	0.21	0.22	0.26
5	1475.63	38.1	3	4422.39	(HexNAc)1(Deoxyhexose)1+(Man)3(GlcNAc)2	64-93QLAHQSNSTNIFFSPVSIATAFAMLSLGTK	0	1	3	3	1	1	HSN3F1N1	5.56	4.63	4.32	0.38	0.81	0.94	0.49	0.21	0.92	3.15	1.73	1.43	2.24	2.68	2.44	11.46	8.00	7.89	
6	1402.85	36	4	5604.36	(Hex)2(HexNAc)2(NeuAc)1+(Man)3(GlcNAc)2	94-125ADTHDELEGLNFLNLTPEAQIHGEGFQELLR	2	2	5	4	0	1	HSN4F0N1	0.49	0.66	1.06	0.29	0.45	0.18	0.00	0.00	0.00	0.50	0.29	0.25	0.38	0.46	0.35	0.99	0.69	1.01	
7	1180.83	38	5	5896.05	(Hex)2(HexNAc)2(NeuAc)2+(Man)3(GlcNAc)2	94-125ADTHDELEGLNFLNLTPEAQIHGEGFQELLR	2	2	5	4	0	2	HSN4F0N2	1.74	1.37	1.60	0.10	0.43	0.37	0.00	0.00	0.00	1.04	0.73	0.52	0.79	0.77	0.77	2.86	2.12	1.91	
8	1224.6	39	3	3668.7	(Hex)2(HexNAc)2(NeuAc)1+(Man)3(GlcNAc)2	268-298YLGNAITAIFFLPDEGK	2	2	5	4	0	1	HSN4F0N1	5.14	8.34	9.58	14.63	15.94	13.56	8.29	11.57	8.70	9.30	10.69	7.09	7.68	8.30	4.64	5.15	5.73		
9	1321.29	34.7	3	3959.67	(Hex)2(HexNAc)2(NeuAc)2+(Man)3(GlcNAc)2	268-298YLGNAITAIFFLPDEGK	2	2	5	4	0	2	HSN4F0N2	66.21	60.55	60.80	80.39	77.67	79.05	80.85	88.44	79.78	77.06	80.45	79.26	82.42	79.01	88.30	52.79	61.65	58.46	
10	1092.17	34.6	5	5453.8	(Hex)2(HexNAc)2(NeuAc)1+(Man)3(GlcNAc)2	268-298YLGNAITAIFFLPDEGKLOHLENLTHDITK	2	2	5	4	0	1	HSN4F0N1	0.30	1.27	1.10	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.24	0.20	0.27	
11	1150.36	35.9	5	5744.95	(Hex)2(HexNAc)2(NeuAc)2+(Man)3(GlcNAc)2	268-298YLGNAITAIFFLPDEGKLOHLENLTHDITK	2	2	5	4	0	2	HSN4F0N2	5.38	11.38	9.64	1.64	1.75	2.21	0.72	0.27	1.04	2.45	2.04	3.00	2.50	3.07	2.24	4.11	2.50	5.00	
12	1438.8	36	4	5744.24	(Hex)2(HexNAc)2(NeuAc)2+(Man)3(GlcNAc)2	268-298YLGNAITAIFFLPDEGKLOHLENLTHDITK	2	2	5	4	0	2	HSN4F0N2	1.09	1.68	1.42	0.48	0.27	0.43	0.00	0.00	0.00	0.45	0.16	0.45	0.64	0.64	0.43	0.88	0.52	1.05	

Glycopeptide synthesis

Target glycopeptides were synthesised using the 9-fluorenylmethyl chloroformate (Fmoc) based solid-phase peptide synthesis. Major steps involved in the glycopeptides synthesis are outlined in the figure 6.1, which includes (i) Isolation of glycoamino acid building block (ii) F-moc protection of the N α amino group of the glycosylated asparagine (iii) Selective Benzyl Esterification of Sialic acid and (iv) Solid phase glycopeptide synthesis (Fig. S2A).

Fmoc protection of glycosylated amino acids

The N α amino group of the glycosylated asparagine (50 mg) was protected using 9-fluorenylmethyl-succinimidyl-carbonate (FmocOsu) and trimethyl amine in a mixture of dioxane/water (1:1). The reaction was completed within two hours at pH 9. In order to prevent any possible cleavage of the Fmoc group in later stages, the reaction mixture was neutralized using acetic acid and the solvents were removed by rotovap evaporation and subsequent lyophilisation. The Fmoc protected N-glycan-Asn building blocks were purified by solid phase extraction (SPE) using 5 g C18 Sep-Pack Cartridges. The highly hydrophilic nature of the unprotected glycosylated molecule prevents its binding to the C18 materials. In contrary, the hydrophobic nature of the Fmoc group can also act as an affinity tag during this separation, since it imparts a unique hydrophobic group to the highly hydrophilic glycosylated amino acid. The compounds were eluted using 5 and 10% Acetonitrile and lyophilized yielding the Fmoc protected biantennary disialyated asparagine building block (80% yield). The presence of the desired compound was further verified by ESI-MS/MS analysis (Fig. S2-B).

Esterification of sialic acid

The sialic acid residues present on the purified glycosylated amino acid building block from egg yolk was selectively protected using benzyl esterification as described by Kajihara *et al* [2]. The amino acid was dissolved in water, titrated with a solution of cesium carbonate and then lyophilized. The dry cesium salt obtained was converted to the corresponding allyl ester by reaction with allyl bromide in DMF at room temperature under argon atmosphere. After the completion of the reaction (24 h), the glycosyl amino acid ester was precipitated using cold diethyl ether from the reaction mixture (61 % yield), purified and analyzed via ESI-MS/MS analysis (Fig. S2C). Target synthetic

glycopeptides were there further purified and analyzed by LC-ESI-MS/MS (Fig. S3A-S3C).

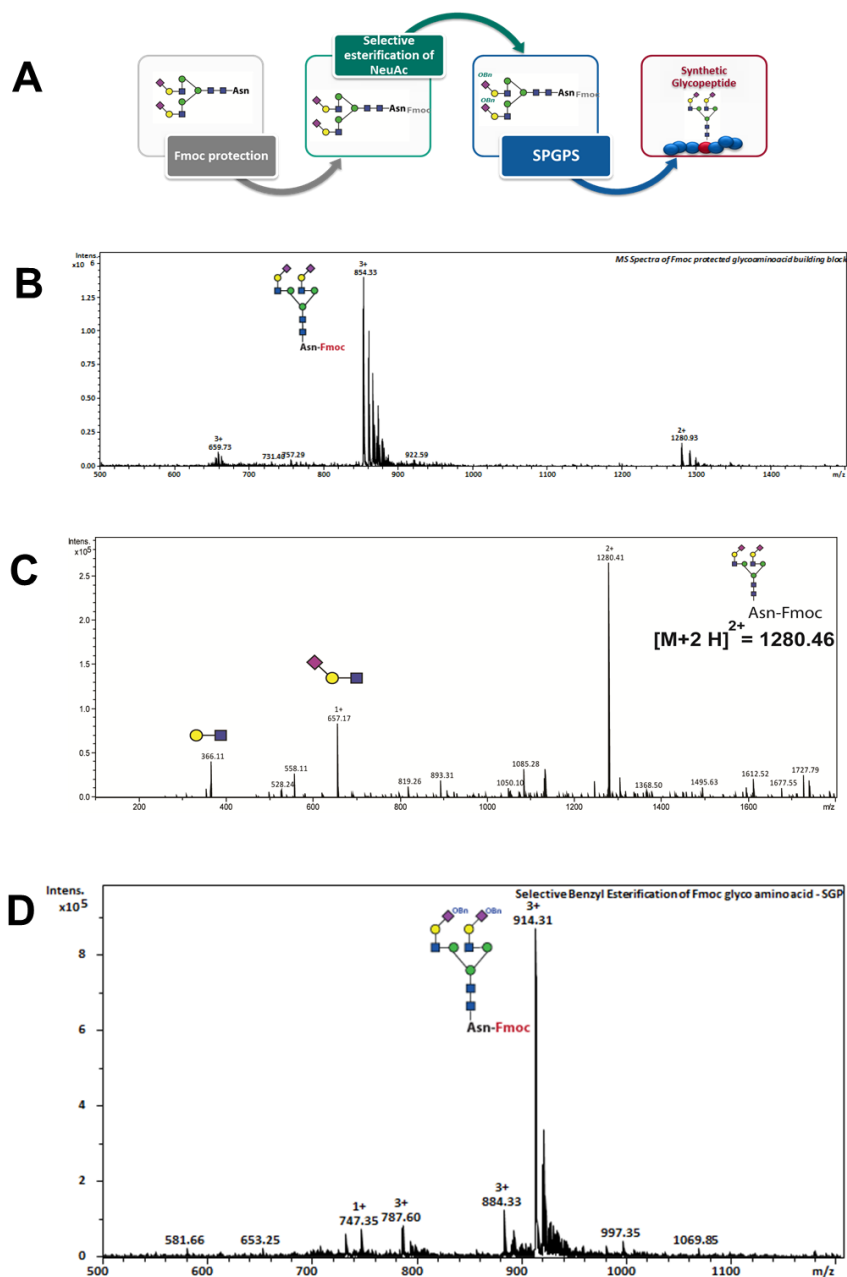


Fig. S2 (A) Schematic representation of major steps involved in solid phase glycopeptide synthesis. (B) MS spectra of Fmoc protected Asn carrying a disialylated, biantennary N-glycan. The signals at m/z 854.02 [M+3H]³⁺ and 1280.46 [M+2H]²⁺ correspond to the Fmoc protected glycosylated Asn. Other peaks observed correspond to sodium and potassium adducts of the target compound. (C) CID MSMS spectra of m/z 1280.46 [M+2H]²⁺ corresponding to the Fmoc protected glycosylated Asn (D) MS spectra of selective benzyl esterified Fmoc protected Asn carrying a disialylated, biantennary N-glycan (m/z 914.06 [M+3H]³⁺)

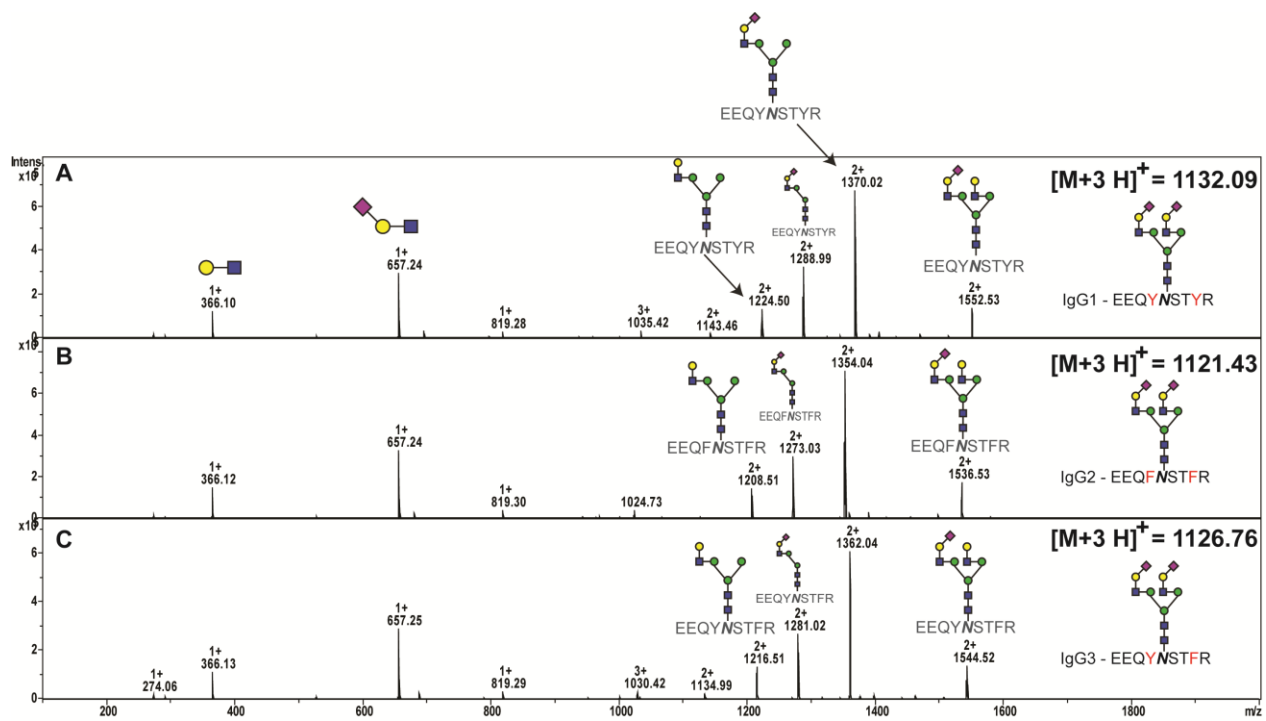


Fig. S3 MS/MS spectrum of a synthetic glycopeptide corresponding to tryptic peptide sequence derived from human Immunoglobulin G (A) IgG-1 (B) IgG-2 (C) IgG-3 subclass. In each case the triply charged precursor ion was isolated and fragmented by collision-induced dissociation

Table S4 GRAVY scores of non-glycosylated peptides identified by RP-nLC

Protein	Sequence NO	Sequence	Gravy Score
A1AT	64-93	QLAHQSNSTNIFFSPVSIATAFAMLSLGTK	0.43
A1AT	94-125	ADTHDEILEGLNFNLTEIPEAQIHEGFQELLR	-0.47
A1AT	268-283	YLGNATAIFFLPDEGK	0.16
A1AT	268-298	YLGNATAIFFLPDEGKLQHLENELTHDIITK	-0.20
IgG2	168-180	TKPREEQFNSTFR	-1.93
IgG2	172-180	EEQFNSTFR	-1.60
IgG1	172-184	TKPREEQYNSTYR	-2.56
IgG1	176-184	EEQYNSTYR	-2.51
IgG4	173-181	EEQFNSTYR	-2.06

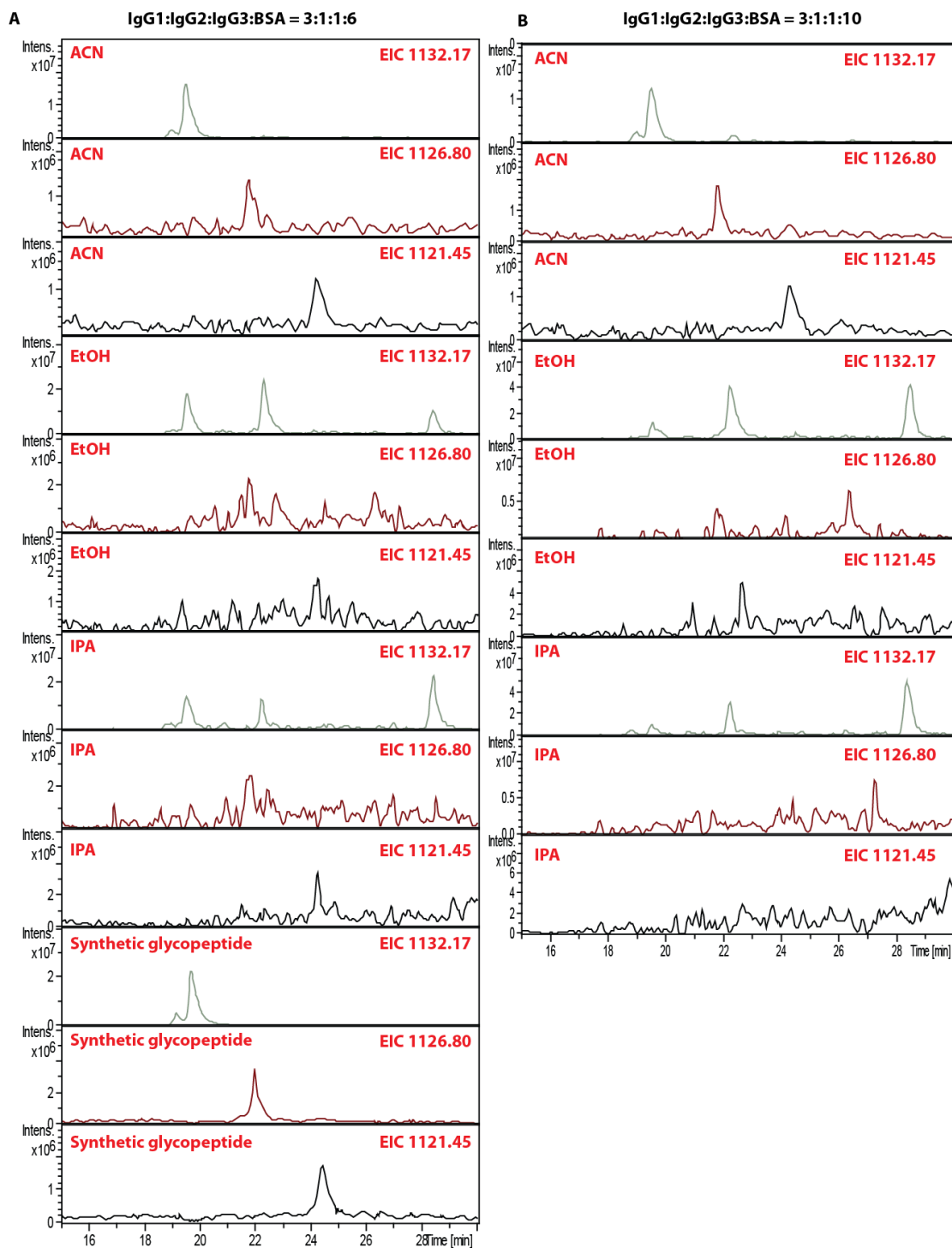


Fig. S4 (A-B) Glycopeptide enrichment efficiency evaluation using the synthetic glycopeptide. When Excess molar ratio of BSA was applied, the enrichment efficiency was significantly compromised in the case of EtOH and IPA as evident from the EIC of the synthetic glycopeptides.

Table S5 List of BSA peptides co-enriched after HILIC enrichment as reported by MASCOT/ProteinScape search for IgG and A1PI

Project Info

Name: Kathir **Date:** December 21, 2011

Sample Info & Protocols

Date: August 20, 2016

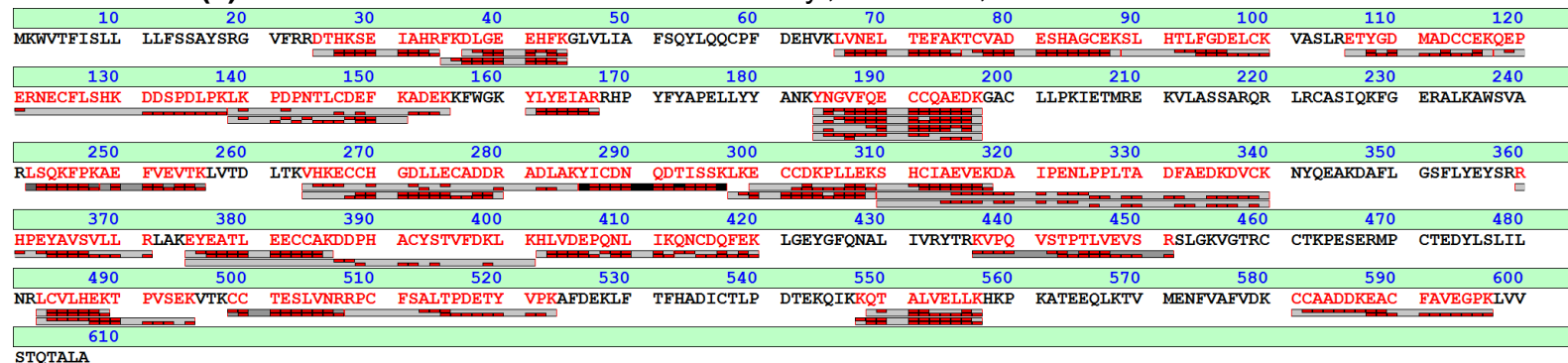
Name: abc efficiency check

Search Result Info

Search Result	Location	Search Engine	Database	Ident. Compounds
Amazon Mammalia_SPJuly2013_2016- 08-21 00:28:58	/Kathir/abc efficiency check/KA BSA IgG SGP 3ug ACN_RA2_01_8625.d	Mascot, 2.3.02	SwissProt_July2013, uniprotsprotJuly2013.fasta	31/250 (FDR:9.09%)
Amazon Mammalia_SPJuly2013_2016- 08-21 00:28:58	/Kathir/abc efficiency check/KA BSA IgG SGP 3ug ACN_RA2_01_8628.d	Mascot, 2.3.02	SwissProt_July2013, uniprotsprotJuly2013.fasta	16/250 (FDR:0.00%)
Amazon Mammalia_SPJuly2013_2016- 08-21 00:28:58	/Kathir/abc efficiency check/KA BSA IgG SGP 3ug ACN T2_RA2_01_8621.d	Mascot, 2.3.02	SwissProt_July2013, uniprotsprotJuly2013.fasta	34/250 (FDR:5.26%)

Protein 1: Serum albumin OS=Bos taurus GN=ALB PE=1 SV=4
Accession: ALBU_BOVIN **Score:** 2277.02
Database: SwissProt_July2013 **MW [kDa]:** 69.20
Seq. Coverage [%]: 58.60 % **pl:** 5.78
No. of Peptides: 38

Modification(s): Carbamidomethyl, Oxidation, Deamidated



Cmpd.	No. of Cmpds.	m/z meas.	Δ m/z [ppm]	z	Rt [min]	Score	P	Range	Sequence	Modification
3	3	597.3400	59.07	2	14.5	63.3	1	25-34	R.DTHKSEIAHR.F	
42	3	625.4220	172.36	2	17.8	71.0	1	35-44	R.FKDLGEEHFK.G	
41	3	487.8410	222.39	2	17.5	49.3	0	37-44	K.DLGEEHFK.G	
151	2	582.4180	170.06	2	25.0	79.3	0	66-75	K.LVNELTEFAK.T	
7	7	732.4000	139.13	2	14.8	98.2	0	76-88	K.TCVADESHAGCEK.S	Carbamidomethyl: 2, 11
140	1	473.9820	167.24	3	24.0	48.5	0	89-100	K.SLHTLFGDELCK.V	Carbamidomethyl: 11
23	4	747.8430	107.36	2	16.6	81.2	0	106-117	R.ETYGDMDADCCEK.Q	Carbamidomethyl: 9, 10; Oxidation: 6
62	1	636.1120	101.69	4	19.5	21.8	2	118-138	K.QEPERNECFLSHKDDSPDLPK.L	Carbamidomethyl: 8
128	1	789.0130	159.18	2	22.9	38.4	0	139-151	K.LKDPNTLCDEFK.A	Carbamidomethyl: 9

115	2	674.0740	117.84	3	22.3	23.8	1	139-155	K.LKPDPTLCDEFKADK.K	Carbamidomethyl: 9
108	3	464.3410	195.24	2	22.1	48.0	0	161-167	K.YLYEIAR.R	
104	3	874.9410	106.11	2	21.7	52.6	0	184-197	K.YNGVFQECCQAEDK.G	Carbamidomethyl: 8, 9; Deamidated: 6
100	3	874.9400	104.97	2	21.7	89.5	0	184-197	K.YNGVFQECCQAEDK.G	Carbamidomethyl: 8, 9; Deamidated: 2
78	3	874.9630	131.26	2	20.5	32.1	0	184-197	K.YNGVFQECCQAEDK.G	Carbamidomethyl: 8, 9; Deamidated: 10
75	5	874.4390	94.74	2	20.5	83.5	0	184-197	K.YNGVFQECCQAEDK.G	Carbamidomethyl: 8, 9
20	4	424.3490	220.51	2	16.0	41.5	1	242-248	R.LSQKFPK.A	
53	3	461.8320	182.67	2	19.3	46.3	0	249-256	K.AEFVEVTK.L	
34	2	1057.5490	97.36	2	17.0	93.4	1	264-280	K.VHKECCHGDLLECADDR.A	Carbamidomethyl: 5, 6, 13
57	1	871.4810	100.77	3	18.9	55.7	2	264-285	K.VHKECCHGDLLECADDRADLAK. Y	Carbamidomethyl: 5, 6, 13
49	6	722.4250	138.92	2	18.2	90.8	0	286-297	K.YICDNQDTISSK.L	Carbamidomethyl: 3
30	3	767.0040	143.18	2	16.9	65.4	1	298-309	K.LKECCDKP LLEK.S	Carbamidomethyl: 4, 5
28	2	646.3910	133.56	2	16.8	57.1	0	300-309	K.ECCDKP LLEK.S	Carbamidomethyl: 2, 3
19	3	536.8760	219.42	2	15.8	47.9	0	310-318	K.SHCIAEVEK.D	Carbamidomethyl: 3
186	4	1171.2790	42.83	3	27.3	102.6	2	310-340	K.SHCIAEVEKDAIPENLPPLTADF AEDKDVCK.N	Carbamidomethyl: 3, 30
190	3	1171.4960	-51.93	3	27.4	53.0	2	310-340	K.SHCIAEVEKDAIPENLPPLTADF AEDKDVCK.N	Carbamidomethyl: 3, 30; Deamidated: 15
96	2	480.7060	202.30	3	21.6	70.1	1	360-371	R.RHPEYAVSVLLR.L	
70	2	751.9260	153.60	2	19.8	77.9	0	375-386	K.EYEATLEECCA.K.D	Carbamidomethyl: 9, 10
156	1	1094.2250	375.61	3	24.7	54.5	2	375-401	K.EYEATLEECCA.KDDPHACYSTVF DKLK.H	Carbamidomethyl: 9, 10, 18
98	1	653.4550	142.80	2	21.4	59.9	0	402-412	K.HLVDEPQNLIK.Q	
17	2	534.8180	175.08	2	15.8	52.7	0	413-420	K.QNCDQFEK.L	Carbamidomethyl: 3
120	2	547.3940	139.89	3	22.7	73.2	1	437-451	R.KVPQVSTPTLVEVSR.S	
26	2	449.8320	194.80	2	16.8	43.2	0	483-489	R.LCVLHEK.T	Carbamidomethyl: 2
40	2	514.0300	165.72	3	17.8	51.0	1	483-495	R.LCVLHEKTPVSEK.V	Carbamidomethyl: 2
36	3	569.8390	151.62	2	17.5	67.8	0	499-507	K.CCTESLVNR.R	Carbamidomethyl: 1, 2
143	1	628.0330	617.83	3	24.1	59.5	0	508-523	R.RPCFSALTPDETYVPK.A	Carbamidomethyl: 3
113	2	571.9780	204.93	2	22.3	77.4	1	548-557	K.KQTALVELLK.H	

173	1	507.9300	229.76	2	26.2	52.2	0	549-557	K.QTALVELLK.H	
67	1	643.3430	112.01	3	19.9	84.0	1	581-597	K.CCAADDKEACFAVEGPK.L	Carbamidomethyl: 1, 2, 10

Table S6A List of peptides identified in the glycopeptide enriched fraction as reported by MASCOT/ProteinScape search for IgG and A1PI

MASCOT results provided as separate File.

Table S6B Number of identified peptides in the glycopeptide enriched fraction as reported by MASCOT/ProteinScape search for IgG and A1PI

	IgG		
	EtOH	ISO	ACN
NO of Peptides	35	40	41
	25	45	36
	31	47	40
Mean	30.33	44	39
SD	5.03	3.61	2.65
	A1PI		
	EtOH	ISO	ACN
NO of Peptides	63	72	57
	58	69	49
	54	68	58
Mean	58.33	69.67	54.67
SD	4.51	2.08	4.93

Table S6C P value score from statistical t-test performed using sigmaPlot

	p-value	
Solvent	IgG	A1AT
Acetonitrile and Ethanol	0.058 (No)	0.008 (Yes)
Acetonitrile and isopropanol	0.125 (No)	0.396 (NO)
Isopropanol and Ethanol	0.019 (Yes)	0.017 (Yes)

Table S6D Description of search parameters used in ProteinScape for protein identification

Peptide Identification

Search Method:	Amazon Human_SPJuly2013	Version:	1.2
Submitted Spectra:	ALL		
Precursor from Classification:	OFF		
Mascot			
Database:	SwissProt_July2013	Enzyme:	Trypsin
Fixed Modifications:	Carbamidomethyl (C)	Var. Modifications:	Deamidated (NQ) Oxidation (M)
Missed Cleavages:	2		
Instrument Type CID:	ESI-TRAP	Taxonomy: Homo sapiens (human)
Peptide Tolerance:	0.3 Da	Instrument Type ETD:	ETD-TRAP
Peptide Charge:	2+, 3+ and 4+	MS/MS Tolerance:	0.5 Da
Mass:	Monoisotopic	#¹³C:	1
Peptide Decoy:	ON	Percolator:	OFF
Significance Threshold:	0.05		

Protein List Compilation by Protein Extractor

Protein Filter

Min. No. of Significant Peptide IDs:	1
Max. MW [KDA]:	
Mascot	
Ions Score Cut-off:	15
Ions Score Threshold for Significant Peptide IDs:	60
Identity Threshold (only Mascot):	ON
Peptide Rank Cut-off:	10
Min. Peptide Length:	5

Assessment

Accept proteins if: Mascot > 40.0

Accept peptides if: Mascot > 20.0

For each peptide, accept: top hit compound only

In case of MS/MS spectra matching peptides from more than one identified protein, accept: peptide assigned to highest scoring protein

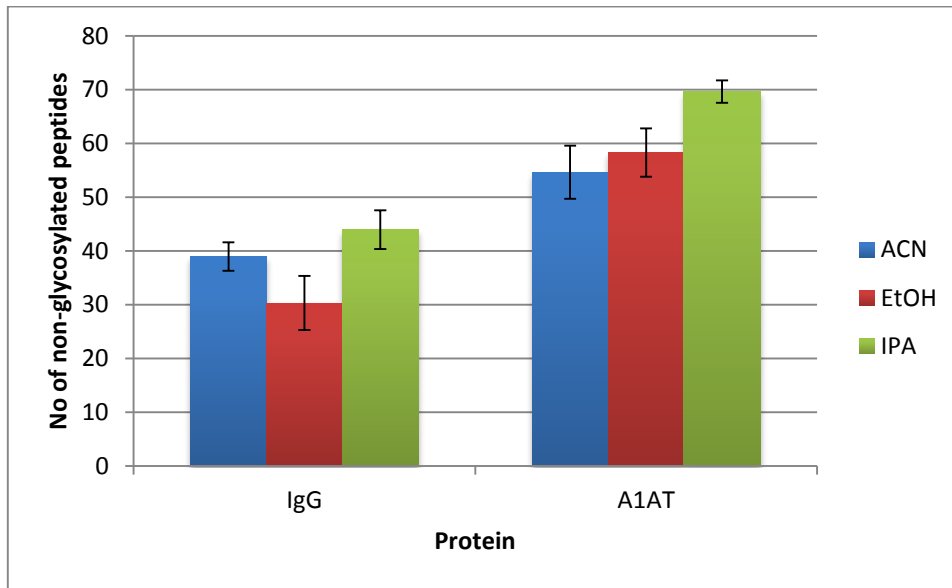


Fig. S5 Number of identified peptides co-enriched along with the glycopeptides using various solvents. As evident from the results, the number of non-glycosylated peptides co-enriched largely depends upon the mobile phase used

Table S7 Number of peptides peptides and glycopeptides identified in the human serum

High Abundance protein									
ACN	Trial 1	Trial 2	Trial 3		Trial 1	Trial 2	Trial 3	Mean	SD
No of GP	47.00	54.00	59.00		45.19	54.00	52.68	50.62	4.75
No of pep	57.00	46.00	53.00		54.81	46.00	47.32	49.38	4.75
	104.00	100.00	112.00						
EtOH									
No of GP	46.00	47.00	49.00		43.81	43.52	44.55	43.96	0.53
No of pep	59.00	61.00	61.00		56.19	56.48	55.45	56.04	0.53
	105.00	108.00	110.00						
ISO									
No of GP	23.00	23.00	22.00		22.33	24.21	23.66	23.40	0.97
No of pep	80.00	72.00	71.00		77.67	75.79	76.34	76.60	0.97
	103.00	95.00	93.00						
	Mean								
Relative percentage	ACN	EtOH	IPA	SD	SD	SD			
Glycopeptides	50.62	43.96	23.40	4.75	0.53	0.97			
Peptides	49.38	56.04	76.60	4.75	0.53	0.97			
serum depleted									
ACN	Trial 1	Trial 2	Trial 3		Trial 1	Trial 2	Trial 3	Mean	SD
No of GP	88.00	105.00	80.00		71.54	80.15	76.92	76.21	4.35
No of pep	35.00	26.00	24.00		28.46	19.85	23.08	23.79	4.35
	123.00	131.00	104.00						
EtOH									
No of GP	104.00	99.00	98.00		64.20	68.28	62.82	65.10	2.84
No of pep	58.00	46.00	58.00		35.80	31.72	37.18	34.90	2.84
	162.00	145.00	156.00						
ISO									
No of Glycopeptides	81.00	96.00	87.00		56.64	61.15	61.70	59.83	2.77
No of peptides	62.00	61.00	54.00		43.36	38.85	38.30	40.17	2.77
	143.00	157.00	141.00						
	Mean								
Relative percentage	ACN	EtOH	IPA	SD	SD	SD			
Glycopeptides	76.21	65.10	59.83	4.35	2.84	2.77			
Peptides	23.79	34.90	40.17	4.35	2.84	2.77			
serum non-depleted									
ACN	Trial 1	Trial 2	Trial 3		Trial 1	Trial 2	Trial 3	Mean	SD
No of Glycopeptides	80.00	74.00	74.00		74.77	68.52	73.27	72.18	3.26
No of peptides	27.00	34.00	27.00		25.23	31.48	26.73	27.82	3.26
	107.00	108.00	101.00		100.00	100.00	100.00	100.00	0.00
ETOH									
No of Glycopeptides	72.00	46.00	75.00		61.02	52.27	60.98	58.09	5.04
No of peptides	46.00	42.00	48.00		38.98	47.73	39.02	41.91	5.04
	118.00	88.00	123.00						
IPA									
No of Glycopeptides	66.00	25.00	35.00		49.25	23.15	26.92	33.11	14.11
No of peptides	68.00	83.00	95.00		50.75	76.85	73.08	66.89	14.11
	134.00	108.00	130.00						
	Mean								
Relative percentage	ACN	EtOH	IPA	SD	SD	SD			
Glycopeptides	72.18	58.09	33.11	3.26	5.04	14.11			
Peptides	27.82	41.91	66.89	3.26	5.04	14.11			

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