SUPPLEMENTAL FIGURE LEGENDS

Figure S1. Tumor infiltrating T cells lose mitochondrial mass. Refers to Figure 1. (**A**) MitoTracker Deep Red FM, MitoTracker Green FM, and TMRE staining of CD8⁺ T cells from spleen and LNs. The indicated histogram represents T cells pre-incubated with 100-111 μ M CCCP, which collapses membrane potential. Results from CCCP titration are tabulated below. (**B**) MitoTracker Green FM and intracellular VDAC staining of LN and TIL from d14 B16 tumors as in Figure 1A. (**C**) MitoTracker Deep Red FM staining of naïve (CD62L^{hi}CD44^{lo}), effector memory (CD62L^{lo}CD44^{hi}), and central memory (CD62L^{hi}CD44^{hi}) CD8⁺ T cells from LN and tumor-infiltrating compartments. (**D**) 2NBDG uptake and MitoTracker FM staining measurements from CD8⁺ and CD4⁺ T cells infiltrating day 18 MC38 tumors or (**E**) LLC tumors or from the nondraining or draining LN. Results represent 3 (A, B, D, E), or 6 (C) independent experiments. ** p < 0.01, **** p < 0.001 by unpaired (B) or paired (C) t test. Error bars indicate s.e.m.

Figure S2. Tumor infiltrating T cells lose mitochondrial mass and do not efficiently elaborate cytokines. Refers to Figure 2. (A) VDAC staining of OT-I T cells cells injected as in Figure 2. (B) Cytokine production of OT-I T cells injected as in Figure 2. Some were rechallenged with cognate peptide overnight and others were stimulated with PMA/ionomycin, as tabulated below from TIL or spleen of VV^{OVA} infected miceError bars indicate s.e.m.

Figure S3. **OT-I T cells do not lose mitochondrial activity when activated by tumor cells** *in vitro*. **Refers to Figure 3**. (**A**) Flow cytogram and tabulated data of CTV-labeled OT-I splenocytes cocultured with either 25 ng/mL SIINFEKL peptide or in a 1:4 ratio with B16^{OVA} cells, seeded 8 h prior to coculture, in the presence of 10 U/mL IL-2. (**B**) As in **A**, but using previously activated, purified effector OT-I T cells. Results represent the mean of 3 (of 8) independent experiments. Error bars indicate s.e.m.

Figure S4. Mitochondrial mass loss is a characteristic of exhausted T cells. Refers to

Figure 4. (**A**) VDAC staining of LN or TIL CD8⁺ T cells from B16-bearing mice expressing coinhibitory molecules PD-1 and LAG-3. (**B**) Co-inhibitory molecule expression on CD8⁺ and CD4⁺ T cells that are LN-resident or infiltrating MC38 or (**C**) LLC tumors on day 18 post inoculation (7-10 mm diameter tumors). (**D**) CD8⁺T cells were sorted flow cytometrically from B16-bearing animals based on MitoFM staining and washed extensively. Cells were then stimulated with anti-CD3/anti-CD28 for 16 h (final 4 h in the presence of a protein transport inhibitor), and then stained intracellularly for cytokines. Tabulated results for IFNγ staining are shown below. Results represent the mean of four independent experiments. Error bars indicate s.e.m. * < p 0.05, ** p < 0.01 by unpaired t test.

Figure S5. Regulatory T cells do not mediate mitochondrial dysfunction in tumor-

infiltrating CD8⁺ T cells. Refers to Figure 5. (**A**), Flow cytogram of CD4⁺ T cells from day 14 B16-bearing $Foxp3^{GFP.Cre.ERT2}$ or $Foxp3^{DTR.GFP}$ mice treated for 3 days with diphtheria toxin. (**B**), Representative flow cytogram and tabulated data of MitoTracker FM staining in CD8⁺ T cells from mice in **A**. (**C**), Flow cytogram and tabulated data of CellTrace Violet labeled CD8⁺ T cells stimulated with anti-CD3 and antigen presenting cells (CD4⁻CD8⁻ splenocytes) in the presence or absence of flow-cytometrically purified Treg cells (CD4⁺GFP⁺ cells from a *Foxp3^{GRP}* mouse). Results are representative of (**A**), or represent the mean of (**B**, **C**) two of three independent experiments. Error bars indicate s.e.m.

Figure S6. PGC1a is repressed progressively upon entry into the tumor

microenvironment. Refers to Figure 6. (**A**) qPCR analysis of *Ppargc1a* (encoding PGC1 α) from CD8⁺ T cells sorted from the indicated compartments based on co-inhibitory molecule expression from day 14 B16 tumors. Results are normalized to cyclophilin B expression and scaled to LN-resident CD4⁺ T cells. (**B**) PGC1 α MFI (per division) of congenically mismatched,

CTV-labeled OT-I T cells transferred into B16^{OVA} bearing mice for 72 h under the cover of anti-PD1 treatment or its isotype control as in Figure 4. (**C**) Representative flow cytogram of MitoFM staining of CD8⁺ T cells isolated from *Ppargc1athCd4^{Cre}* mice or *Cd4^{Cre}* controls after *in vitro* expansion. (**D**) Metabolic flux measurements (Seahorse) from the cells in **C**. (**E**) Representative flow cytogram and tabulated results of PGC1 α versus T-bet and (**F**) Ki67 staining. (**G**) Representative flow cytogram depicting phospho-Akt (S473) activation as a function of PD-1 status. Results are representative of three (**A**, **C**, **E**, **F**), five (**B**, **G**), or two (**D**) independent experiments. * p <0.05, **** p <0.0001 by unpaired (**C**) or paired (**E**,**F**) t test. Error bars indicate s.e.m.

Figure S7. Enforced PGC1 α expression results in general increases in mitochondrial function. Refers to Figure 7. (A) Fuel usage test of PGC1 α - or EV-expressing OT-I T cells. Cells were uncoupled with FCCP and then subjected to sequential inhibition of pyruvate oxidation (UK5099), fatty acid oxidation (etomoxir), and glutaminolysis (BPTES). Results are displayed as % of FCCP-uncoupled OCR sensitive to the appropriate inhibitor. (B) IFN γ and TNF α production of EV or PGC1 α expressing T cells prior to adoptive transfer. (C) Expression of PD-1 and LAG-3 on LN or TIL-resident reprogrammed T cells. Results represent the mean of three (A, B) or are representative of five (C) independent experiments. Error bars indicate s.e.m.



Figure S2 – related to Figure 2



Figure S3 – related to Figure 3



Figure S4 – related to Figure 4







Figure S6 - related to Figure 6

Figure S7 – related to Figure 7



SUPPLEMENTAL EXPERIMENTAL PROCEDURES

B16^{OVA} in vitro stimulation

Freshly isolated OT-I splenocytes or purified, previously activated OT-I CD8⁺ T cells were CellTrace Violet labeled and plated at various ratios on B16 or B16^{OVA} cells in 10% RPMI for 72 h. Proliferation and mitochondrial mass of the T cells was examined flow cytometrically.

Treg cell suppression assay

Lymph nodes and spleens from *Foxp3* reporter mice (*Foxp3*^{DTR.GFP}) were sorted based on expression of CD4 and GFP, then cocultured with CellTrace Violet-labeled CD8⁺ T cells from WT mice at a 1:4 ratio (Treg:Teff cell) in the presence of antigen presenting cells (CD4⁻CD8⁻ splenic cells at a 2:1 APC:T cell ratio) and 1 μ g/mL anti-CD3. After 72 h, proliferation and mitochondrial mass of the CD8⁺ T cells were analyzed by flow cytometry.

Fuel usage test

100,000 previously activated, transduced T cells (Generated in Figure 7) were plated on CellTak-coated Seahorse plates in minimal, unbuffered Seahorse media containing glucose, and glutamine. Basal measurements were taken and then cells were uncoupled with FCCP and subjected to inhibition by UK5099, etomoxir, and BPTES. S7A depicts the percentage of the total FCCP uncoupled OCR inhibited by these agents.