

Expanded View Figures

Figure EV1. Myeloid cells are the main source of IL-1 β upon MOG/CFA/PTx immunization.

A–C Analysis of IL-1 β expression by cells isolated from the dLN and stimulated with GM-CSF (A), LPS (B), and PMA/ionomycin (C). Data are representative FACS plots gated on VD⁻ cells with mean frequencies per group.

Data information: Cells (A–C) were isolated at day 7 after immunization and stimulated in the presence of monensin with indicated stimuli for 4 h. Data consist of n = 4 WT PBS-, n = 3 IL-1R1^{-/-} PBS-, n = 4 WT PTx-, n = 3 IL-1R1^{-/-} PTX MOG/CFA-immunized mice. Experiments were performed twice with similar results.



Figure EV2. Myeloid cells are the main source of IL-1a upon MOG/CFA/PTx immunization.

A Analysis of IL-1 α expression by cells isolated from the spleen and stimulated with GM-CSF. Data are representative FACS plots, gated on VD⁻ cells with mean frequencies per group.

B Frequencies (mean + SEM) of IL-1 α expression by CD11b⁺ cells shown in (A).

Data information: Cells (A, B) were isolated at day 7 after immunization and were stimulated with 20 ng/ml GM-CSF for 4 h in the presence of monensin. Data consist of n = 4 WT PBS-, n = 3 IL-1R1^{-/-} PBS-, n = 4 WT PTx-, and n = 3 IL-1R1^{-/-} PTx MOG/CFA-immunized mice. *P < 0.05, N.S., not significant; two-tailed unpaired *t*-test. Experiments were performed twice with similar results.





Figure EV3. IL-1R1 deletion impairs MOGspecific Th17 cell expansion.

- A Analysis of IL-17A expression by CD4 T cells isolated from the spleen and restimulated with MOG or OVA for 6 h.
- B Total cell numbers of antigen-specific Th17 cells isolated from the spleen shown in (A).

Data information: Cells (A, B) were isolated at day 9 after MOG/CFA/PTx immunization of n = 4 mice of each genotype. Data are (A) representative FACS plots, gated on VD⁻TCRβ⁺CD4⁺CD44⁺ cells with mean frequencies among CD4 T cells per group \pm SEM and (B) bar diagram (mean + SEM). *P < 0.05, N.S., not significant; two-tailed unpaired *t*-test.

Figure EV4. IL-17A-related cytokine expression by CD4 T cells is dependent on IL-1R1 signaling.

A–G Analysis of cytokine expression by CD4 T cells isolated from the (A, B, D, F) spleen and (C, E, G) dLN (depicted in Fig 3). Frequencies and total numbers of MOG-specific (B, C) GM-CSF⁺IFNγ⁺ cells, (D, E) GM-CSF⁺ cells, and (F, G) IFNγ⁺ cells. Data (A) are representative FACS plots, gated on VD⁻TCRβ⁺CD4⁺CD44⁺CD40L⁺ cells with mean frequencies among CD44⁺CD40L⁺ cells per group, and (B–G) bar diagram (mean + SEM).

Data information: Cells (A–G) were isolated at day 9 after immunization and restimulated in the presence of MOG for 6 h. Data consist of n = 5 WT PBS-, n = 4 WT PTx-, n = 4 IL-1R1^{-/-} PBS-, n = 4 IL-1R1^{-/-} PTx MOG/CFA-immunized mice. *P < 0.05, **P < 0.01, ***P < 0.001, N.S., not significant; two-tailed unpaired t-test. Experiments were performed at least twice with similar results.



Figure EV5. IL-23 promotes EAE severity in IL-1R1-deficient mice.

A EAE clinical scores (mean + SEM) of mice treated with IL-23 or PBS.

B Maximal EAE clinical scores (individual plots with mean) of mice shown in (A).

Data information: Mice were actively immunized with MOG/CFA/PTx and treated i.p. with either PBS or IL-23 (1 μ g/mouse) at day 3, 5, 7, 9, 11, and 13. Data consist of n = 7 WT PBS-, n = 4 IL-1R1^{AT} PBS-, n = 4 WT IL-23-, and n = 5 IL-1R1^{AT} IL-23-treated mice. **P < 0.01, N.S., not significant; two-tailed unpaired *t*-test. Experiments were performed twice with similar results.