1SUPPLEMENTAL INFORMATION2Chemistry

3 **Experiment Procedures**

4 General: Unless otherwise noted, all reactants or reagents including dry solvents were obtained from commercial suppliers and used as received. Unless otherwise $\mathbf{5}$ 6 noted, all reactions were performed with dry solvents under an atmosphere of 7 nitrogen in dried glassware using standard vacuum-line techniques. All work-up and 8 purification procedures were carried out with reagent-grade solvents in air. 9 Analytical thin-layer chromatography (TLC) was performed using E. Merck silica gel 10 60 F₂₅₄ precoated plates (0.25 mm). The developed chromatogram was analyzed by UV lamp (254 nm). Flash column chromatography was performed with E. Merck 11 silica gel 60 (230-400 mesh). Preparative thin-layer chromatography (PTLC) was 1213performed using Wako gel B5-F silica coated plates (0.75 mm) prepared in our 14laboratory. A reverse phase thin-layer chromatography (TLC) was performed using E. Merck silica gel 60 RP-18 F₂₅₄S precoated plates (0.25 mm). The high-resolution 1516 mass (HRMS) spectra were conducted on Thermo Fisher Scientific Exactive. 17Nuclear magnetic resonance (NMR) spectra were recorded on a JEOL 18JNM-ECA-400 (¹H 400 MHz, ¹³C 100 MHz) and JEOL JMN-ECA-600II with Ultra COOL[™] probe (¹H 600 MHz, ¹³C 150 MHz) spectrometer. Chemical shifts for ¹H 1920NMR are expressed in parts per million (ppm) relative to tetramethylsilane (δ 0.00 ppm). Chemical shift for ¹³C NMR are expressed in ppm relative to CDCl₃ (δ 77.0 2122ppm). Data are reported as follows: chemical shift, multiplicity (s = singlet, d = 23doublet, dd = doublet of doublets, t = triplet, m = multiplet, br = broad signal), 24coupling constant (Hz), and integration.



 $\begin{array}{c} 26 \\ 27 \end{array}$

Fig S1. Synthesis of glycosyl donors GD1 and GD2

29Reagents and reaction conditions: I) acetyl chloride, allyl alcohol, 70°C, 10 h; II) 30 TsOH⊕H₂O, benzaldehyde dimethyl acetal, 50°C, 3 h; III) NaH, benzyl bromide 31(BnBr), DMF, 0°C then room temperature (RT), overnight; IV) TsOH⊕H₂O, MeOH, 32CH₂Cl₂, RT; V) TrCl, DMAP, pyridine, 50°C, overnight; VI) NaH, R-X (methyl iodide 33 or BnBr), DMF, 0°C then RT, overnight; VII) TsOH⊕H₂O, MeOH, CH₂Cl₂, RT; VIII) 34AZADO, (diacetoxyiodo)benzene, buffer, CH₂Cl₂, 0°C, 3 h; IX) KHCO₃, BnBr, tetrabutylammonium iodide (TBAI), DMF, RT, 4 h; X) PdCl₂, sodium acetate, H₂O, 35acetic acid, RT, 48 h; XI) Ac₂O, Et₃N, CH₂Cl₂, RT, 4 h; XII) 4-MeC₆H₄-SH (tol-SH), 36 BF₃⊕Et₂O, CH₂Cl₂, 0°C to RT, 3 h. 37

38 Ts = tosyl, DMF = N,N-dimethylformamide, Tr = triphenylmethyl, DMAP =
 39 N,N-dimethyl-4-aminopyridine, AZADO = 2-aza-adamantane N-oxyl.

40



41

42 **Fig S2.** Synthesis of glycosyl donor **GD3**

43 Reagents and reaction conditions: I) NaH, BnBr, DMF, 0°C then RT, overnight; II)

44 PdCl₂, sodium acetate, H₂O, acetic acid, RT, 48 h; III) Ac₂O, Et₃N, CH₂Cl₂, RT, 4 h;

45 IV) 4-MeC₆H₄-SH (tol-SH), BF₃ \oplus Et₂O, CH₂Cl₂, 0°C to RT, 3 h.

46



47

48 **Fig S3.** Synthesis of glycosyl acceptor **GA1** and **GA2**

49 Reagent and reaction conditions: I) TrCl, pyridine, 50°C, overnight; II) BnBr, NaH,

50 DMF, 0°C then RT, 8 h; III) TsOH \oplus H₂O, MeOH, CH₂Cl₂, RT.



53 Fig S4. Synthesis of glycosyl acceptor GA3

Reagents and reaction conditions: I) BnBr, NaH, DMF, 0°C then RT, 3 h; II) (±)-CSA,
acetone dimethylacetal, RT, overnight; III) MeOH, H₂O, reflux, 48 h; IV) BnBr, NaH,
DMF, 0°C then RT, overnight; V) HOAc, H₂O, 65°C, 2 h; VI) Bu₂SnO, MeOH, reflux,
overnight; VII) PMB-CI, TBAB, MS4Å, benzene, reflux, 6 h; VIII) BnBr, NaH, DMF,
0°C then RT, overnight; IX) DDQ, CH₂Cl₂, H₂O, RT, 1 h.

59 CSA = camphorsulfonic acid, TBAB = tetrabutylammonium bromide, PMB-Cl = 60 *para*-methoxy benzyl chloride, MS = molecular sieves, DDQ = 61 2,3-dichloro-5,6-dicyano-1,4-benzoquinone.

62



- 64 Fig S5. Synthesis of glycosyl acceptor GA4
- Reagents and reaction conditions: I) BnBr, NaH, DMF, 0°C then RT, 3 h; II) 66 2,3-butanedione, trimethyl orthoformate, (±)-CSA, MeOH, reflux, 20 h; III) allyl

- bromide, NaH, DMF, 0°C then RT, 3 h; IV) TFA, CH₂Cl₂, RT, 15 min; V) BnBr, NaH,
- DMF, 0°C then RT, overnight; VI) PdCI₂, sodium acetate, H₂O, acetic acid, RT, 48 h.
- TFA = trifluoroacetic acid





- glycosylation product **G**' and the deprotection product **G** (the total yield of two
- anomers and ratio of β : α is shown in parenthesis)
- Reagent and reaction conditions: I) NIS, TfOH (cat.), CH₂Cl₂, -40 °C then RT, 4 h;
- 17 II) Pd(OH)₂/C, H₂ atm., EtOAc, MeOH, RT, overnight.
- 78
- 79
- 80
- 81
- 82



- Fig S7. Synthesis of 4,6-Me-GlcA- β (1,6)-Gal (G9)
- 86 Reagents and reaction conditions: I) TMSCHN₂, MeOH, toluene, RT, 30 min
- 87 TMSCHN₂ = trimethylsilyldiazomethane



90 **Fig S8.** Synthesis of 4-Me-GlcA- β -OMe (**G11**)

91 Reagents and reaction conditions: I) TsOH \oplus H₂O, benzaldehyde dimethyl acetal,

- 92 50°C, 3 h; II) NaH, benzyl bromide (BnBr), DMF, 0°C then room temperature (RT),
- 93 overnight; III) TsOH⊕H₂O, MeOH, CH₂Cl₂, RT; IV) TrCl, DMAP, pyridine, 50°C,
- 94 overnight; V) NaH, Me-I, DMF, 0°C then RT, overnight; VI) TsOH⊕H₂O, MeOH,

95 CH₂Cl₂, RT; VII) AZADO, (diacetoxyiodo)benzene, buffer, CH₂Cl₂, 0°C, 3 h; VIII)

96 Pd(OH)₂/C, H₂ atm., EtOAc, MeOH, RT, overnight.

97 Preparation of glycosyl donor GD1 (Fig. S1)

98 Benzyl(2S,3S,4S,5R)-4,5-bis(benzyloxy)-3-methoxy-6-(*p*-tolylthio)tetrahydro-2*H*-py

99 ran-2-carboxylate



100 GD1

89

101 The treatment of **7a** (ref. 1S) (239 mg, 0.50 mmol) with acetic anhydride (2 mL) 102 and triethylamine (2 mL) under ambient condition smoothly afforded the 103 corresponding acetyl glycoside **8a** within 4 h (the completion of the reaction was monitored by TLC). The reaction mixture was concentrated *in vacuo*, yielding a
 viscous residue, which was further purified by flash column chromatography
 (hexane/EtOAc = 5:1) to give **8a** as an oil liquid.

107 A dry CH₂Cl₂ was added to a Schlenk tube containing **8a** (ca. 0.50 mmol) and 108 Tol-SH (74.0 mg, 0.60 mmol, 1.2 equiv.) under a N₂ atmosphere, and the reaction 109 mixture was cooled down to 0 °C before catalytic amount of BF₃⊕Et₂O 1M in Et₂O 110 (10 μ L, 0.01 mmol, 0.02 equiv.) was added dropwise to the mixture. The reaction mixture was stirred for 3 h at RT and then cooled to 0 °C before 100 μ L of 111 112triethylamine was added slowly to quench the reaction. After that, the mixture was 113 allowed to increase to RT and concentrated in vacuo. The resultant viscous residue 114 was subjected to silica gel column chromatography (hexane/EtOAc = 5:1) to afford 115**GD1** as a colorless viscous liquid (mixture of two anomers) in 195 mg (0.34 mmol, 11667% yield in 2 steps).

¹H NMR (CDCl₃, 400 MHz) δ 2.30 (s, 3H), 2.32 (s, 1H, these two singlets are –Me in -tolyl group of two anomers), 3.37 (s, 1H), 3,41 (s, 3H, these two singlets are 4-O-Me group of two anomers), 3.48–3.51 (m, 1H), 3.57–3.59 (m, 0.3H), 3.76–3.84 (m, 2H) 4.56–4.92 (m, 6H), 5.21 (q, *J* = 12.4 Hz, 2.5H), 5.48 (d, *J* = 5.2 Hz, 0.7H), 7.01 (d, *J* = 7.9 Hz, 1.9H), 7.05 (d, *J* = 7.9 Hz, 0.7H), 7.29–7.37 (m, 23H).

¹³C NMR (CDCl₃, 100 MHz) δ 21.1, 60.5, 60.6, 67.1, 67.2, 70.8, 72.6, 75.4, 75.6,
75.7, 77.9, 79.0, 80.0, 80.7, 81.1, 81.5, 85.8, 87.9, 88.2, 127.6, 127.8(5), 127.9,
128.1, 128.2, 128.3, 128.3(8), 128.4(2), 128.5, 129.7, 132.8, 133.1, 135.2, 137.5,
137.7,138.2, 138.5, 98.7, 127.7, 127.9, 128.0, 128.2, 128.4, 137.3, 137.4, 137.7,
138.2, 138.5, 169.1.

- 127
- 128 **Preparation of glycosyl donor GD2** (Fig. S1)
- 129 ((2*S*,3*R*,4*S*,5*R*)-6-(Allyloxy)-3-benzyl-4,5-bis(benzyloxy)tetrahydro-2*H*-pyran-2-yl)m
- 130 ethanol



5b

131

132To a stirred solution of trityl ether 4 [ref. S1] (2.57 g, 4.0 mmol) in DMF (60 mL), a 60% dispersion of NaH in mineral oil (240 mg, 6.0 mmol, 1.5 equiv.) was added at 133 134 0°C. After stirring the mixture for 30 min, benzyl bromide (BnBr: 710 μ L, 6.0 mmol) 135was added, and the resultant mixture was stirred for a further 3 h at RT. After 136 adding MeOH (1.5 equiv.) to stop the reaction, the reaction mixture was extracted 137 three times with H₂O/EtOAc/hexane (2:1:1, 15 mL) to remove DMF, and dried over 138 Na₂SO₄. After filtration of the organic layer, the mixture was concentrated in *vacuo*. 139 The resultant crude product was dissolved in MeOH/CH₂Cl₂ (1:1, 30 mL) to obtain a 140 yellow solution, which was treated with TsOH \cdot H₂O (10 mol%) and stirred at RT for 6 141 h. NaHCO₃ was added to neutralize the solution and the resultant yellow solution 142was concentrated in *vacuo*, and then subjected to flash column chromatography 143 (hexane/EtOAc = 3:1) to yield alcohol **5b** as a yellow syrup (1.67 g, 3.4 mmol, 85%) 144yield).

- ¹H NMR (CDCl₃, 400 MHz) δ 3.50–3.56 (m, 2H), 3.65–3.78 (m, 3H), 3.97–4.06 (m,
- 2H), 4.14 (dd, J = 13.2, 5.2 Hz, 1H), 4.65 (dd, J = 12.0, 5.2 Hz, 2H), 4.74–4.91 (m,
 4H) 4.93–5.02 (m, 1H), 5.23 (d, J = 10.4 Hz, 1H), 5.33 (d, J = 17.2 Hz, 1H), 5.88–
 5.97 (m, 1H), 7.29–7.37 (m, 15H).
- ¹³C NMR (CDCl₃, 100 MHz) δ 61.8, 68.2, 70.8, 73.2, 75.0, 75.7, 79.9, 81.9, 95.6,
 118.2, 127.8, 127.9, 128.0, 128.3, 128.3(8), 128.4(2), 133.6, 138.1, 138.7.
- 151
- Benzyl(2S,3S,4S,5R)-6-(allyloxy)-3,4,5-tris(benzyloxy)tetrahydro-2*H*-pyran-2-carbo
 xylate



6b

154

155To a solution of **5b** (980 mg, 2.0 mmol,) in CH₂Cl₂ (4 mL) and buffer $(Na_2HPO_4/NaH_2PO_4, pH = 7)$, iodobenzene diacetate (1.29 g, 4.0 mmol) was added 156157at 0°C, followed by addition of 2-azaadamantane N-oxyl (AZADO: 15.0 mg, 0.1 mmol, 0.1 equiv.). The mixture was stirred at the same temperature (the completion 158159of the reaction was monitored by TLC). After 2 h, 5b was consumed and then the reaction mixture was extracted three times with CH₂Cl₂ and water. The organic 160 161 layer was dried over Na₂SO₄ and the solvent was removed *in vacuo*. A yellow liquid was obtained, to which was added dry DMF (5 mL), followed by KHCO₃ (1.0 g, 10.0 162mmol, 5.0 equiv.), BnBr (710 μ L, 6.0 mmol, 3.0 equiv.), and tetrabutylammonium 163 164 iodide (TBAI: 74.0 mg, 0.20 mmol, 0.1 equiv.). After stirring at RT for 3 h, the 165reaction mixture was extracted three times with EtOAc/H₂O (1:1, 10 mL). The 166 organic layer was concentrated *in vacuo*, and the residue was purified by silica gel 167 column chromatography (hexane/EtOAc = 5:1) to yield 6b as a colorless oil liquid 168 (713 mg, 1.2 mmol, 60% yield).

169 ¹H NMR (CDCl₃, 400 MHz) δ 3.57–3.63 (m, 1H), 3.74 (t, J = 10.0 Hz, 1H), 3.99– 170 4.06 (m, 2H), 4.18 (dd, J = 13.2, 5.2 Hz, 1H), 4.26–4.32 (m, 1H), 4.42–4.47 (m, 1H), 171 4.62–4.67 (m, 1H), 4.71–4.85 (m, 4H), 4.92–4.99 (m, 1H), 5.11–5.23 (m, 3H), 5.28– 172 5.35 (m, 1H), 5.86–5.96 (m, 1H), 7.10–7.15 (m, 2H), 7.21–7.25 (m, 3H), 7.29–7.34 173 (m, 15H)

¹³C NMR (CDCl₃, 100 MHz) δ 67.2, 68.6, 70.5, 73.4, 75.0, 75.8, 79.2, 79.6, 81.3,
96.1, 118.7, 127.6, 127.7, 127.8(8), 127.9(2), 128.1, 128.2, 128.3, 128.3(7),
128.4(1), 128.5, 133.2, 135.0, 137.8, 137.9, 138.5, 169.5

177

178 Benzyl(2S,3S,4S,5R)-3,4,5-tris(benzyloxy)-6-hydroxytetrahydro-2*H*-pyran-2-carbox

179 ylate



A mixture of **6b** (594 mg, 1.0 mmol), sodium acetate (205 mg, 2.5 mmol, 2.5 equiv.), $PdCl_2$ (212 mg, 1.2 mmol, 1.2 equiv.), and aqueous acetic acid (acetic acid/H₂O = 11:1, 12 mL) was stirred at RT for 48 h. After removing acetic acid and water in *vacuo*, the residue was subjected to silica gel column chromatography (hexane/EtOAc = 3:1) to afford **7b** as a white amorphous solid (421 mg, 0.76 mmol, 76% yield).

¹H NMR (CDCl₃, 400 MHz) δ 3.11 (s, 1H), 3.60 (dd, J = 9.2, 3.2 Hz, 1H), 3.76 (t, J = 9.6 Hz, 1H), 3.95–4.03 (m, 1H), 4.49 (dd, J = 16.0, 9.6 Hz, 2H), 4.65–4.80 (m, 4H), 4.86–4.93 (m, 1H), 5.09–5.23 (m, 3H), 7.12–7.15 (m, 2H), 7.25–4.26 (m, 3H), 7.29– 7.32 (m, 15H)

¹³C NMR (CDCl₃, 100 MHz) δ 67.2, 70.5, 73.3, 74.8, 75.6, 79.1, 79.3, 80.6, 91.5,
127.6, 127.7, 127.9, 128.0, 128.0(5), 128.1(1), 128.2, 128.3, 128.5, 128.6, 134.9,
137.5, 137.7, 138.3, 169.6

194

Benzyl(2S,3S,4S,5R)-3,4,5-tris(benzyloxy)-6-(*p*-tolylthio)tetrahydro-2*H*-pyran-2-car
boxylate



GD2

197

Same procedures as the preparation of **GD1**: treatment of **7b** (277 mg, 0.50 mmol) with acetic anhydride (2 mL) and triethylamine (2 mL) under ambient condition, smoothly afforded the corresponding acetyl glycoside **8b** within 4 h (the completion of the reaction was monitored by TLC). The reaction mixture was concentrated *in vacuo*, yielding a viscous residue, which was subjected to flash column chromatography (hexane/EtOAc = 5:1) to give **8b** as an oil liquid,. 204A dry CH₂Cl₂ was added to a Schlenk tube containing **8b** (ca. 0.50 mmol) and 205Tol-SH (74.0 mg, 0.60 mmol, 1.2 equiv.) under a N₂ atmosphere, and the reaction 206 mixture was cooled down to 0 °C before catalytic amount of BF₃⊕Et₂O 1M in Et₂O 207 (10 μ L, 0.01 mmol, 0.02 equiv.) was added dropwise to the mixture. The reaction 208was stirred for 3 h at RT and cooled to 0 °C before 100 μ L of triethylamine was 209 added slowly to guench the reaction. After that, the mixture was allowed to increase 210to RT and concentrated *in vacuo*, afforded a viscous residue, which was purified by silica gel column chromatography (hexane/EtOAc = 5:1) to afford GD2 as a 211212colorless viscous liquid as a mixture of two anomers (254 mg, 0.39 mmol, 77% yield 213in 2 steps).

 1 H NMR (CDCl₃, 400 MHz) δ 2.31 (s), 2.33 (s) (3H) (these two singlets are –Me in –

tolyl group of two anomers), 3.49 (t, J = 9.2 Hz), 3.69 (t, J = 9.2 Hz) (1H) (these two

triplets are one isomeric proton from two anomers) 3.81–3.86 (m, 1H), 3.89–3.95 (m,

217 1H), 4.49–4.62 (m, 1H), 4.67–4.97 (m, 6H), 5.09–5.22 (m, 2H), 5.51 (s, 1H), 7.02–

218 7.08 (m, 2H), 7.13–7.17 (m, 2H), 7.28–7.40 (m, 20H).

¹³C NMR (CDCl₃, 100 MHz) δ 21.1, 67.2, 67.3, 70.9, 72.6, 75.0, 75.1, 75.4, 75.8,

75.9, 78.1, 79.0, 79.1(5), 79.2(2), 80.1, 81.6, 85.9, 87.9, 88.3, 127.7, 127.7(7),
127.8(0), 128.0, 128.2, 128.3, 128.3(6), 128.4(4), 128.5, 128.6, 128.9, 129.5, 129.7,
132.8, 133.1, 135.1, 137.5, 137.7, 137.8, 137.9, 138.1, 138.4, 168.0, 169.1.

223

224 **Preparation of glycosyl donor GD3** (Fig. S2)

 $225 \qquad \mathsf{Benzyl}(2\mathsf{S}, 3\mathsf{S}, 4\mathsf{S}, 5\mathsf{R}) \text{-} 3, 4, 5 \text{-} \mathsf{tris}(\mathsf{benzyloxy}) \text{-} 6 \text{-} (p \text{-} \mathsf{tolylthio}) \mathsf{tetrahydro-} 2\mathsf{H} \text{-} \mathsf{pyran-} 2 \text{-} \mathsf{car}$

226 boxylate

OBn MeO BnO

6c

227

To a stirred solution of trityl ether **5a** [ref. S1] (828 mg, 2.0 mmol) in DMF (30 mL), a 60% dispersion of NaH in mineral oil (120 mg, 3.0 mmol, 1.5 equiv.) was added at 2300°C. After stirring the mixture for 30 min, benzyl bromide (355 μ L, 3.0 mmol, 1.5 231equiv.) was added, and the resultant mixture was stirred for a further 3 h at RT. 232MeOH (1.5 equiv.) was then added to stop the reaction. The reaction mixture was 233extracted three times with H₂O/EtOAc/hexane (2:1:1, 10 mL) to remove DMF, and 234dried over Na₂SO₄. After filtration of the organic layer, the mixture was concentrated 235in *vacuo*. The resultant crude product was purified by flash column chromatography 236(hexane/EtOAc= 3:1) to yield 6c as a colorless oil liquid (907 mg, 0.18 mmol, 90%) 237 vield).

²³⁸ ¹H NMR (CDCl₃, 400 MHz) δ 3.46 (s, 3H), 3.48–3.55 (m, 1H), 3.61–3.74 (m, 4H),

239 3.89 (t, *J* = 9.6 Hz, 1H), 3.98–4.03 (m, 1H), 4.13–4.18 (m, 1H), 4.49–4.65 (m, 3H),

240 4.74–4.81 (m, 3H), 4.88–4.96 (m, 1H), 5.10 (dd, J = 1.6, 1.0 Hz, 1H), 5.28–5.33 (m,

- 241 1 H), 5.88–5.98 (m, 1H), 7.27–7.40 (m, 15H)
- ¹³C NMR (CDCl₃, 100 MHz) δ 60.6, 68.1, 68.5, 70.2, 73.2, 73.4, 75.6, 79.4, 79.6,
 82.0, 95.6, 118.1, 127.5, 127.6, 127.8, 127.9, 128.0, 128.1, 128.2(5), 128.2(8),
 128.3, 133.7, 138.0, 138.2, 138.9
- 245

246 (3*R*,4*S*,5*R*,6*R*)-3,4-Bis(benzyloxy)-6-((benzyloxy)methyl)-5-methoxytetrahydro-2*H*-247 pyran-2-ol



7c

248

A mixture of **6c** (504 mg, 1.0 mmol), sodium acetate (205 mg, 2.5 mmol, 2.5 equiv.), $PdCl_2$ (212 mg, 1.2 mmol, 1.2 equiv.), and aqueous acetic acid (acetic acid/H₂O = 11:1, 12 mL) was stirred at RT for 48 h. After removing acetic acid and water in *vacuo*, the residue was subjected to silica gel column chromatography (hexane/EtOAc = 3:1) to afford **7c** as a white amorphous solid (348 mg, 0.75 mmol, 254 75% yield).

²⁵⁵ ¹H NMR (CDCl₃, 400 MHz) δ 3.28–3.38 (m, 1H), 3.47 (s, 3H), 3.51–3.55 (m, 1H),

3.63–3.74 (m, 2H), 3.85 (t, J = 9.6 Hz, 1H), 3.91–3.95 (m, 1H), 4.50–4.95 (m, 7H),
5.20 (d, J = 3.6 Hz, 1H), 7.29–7.39 (m, 15H).

¹³C NMR (CDCl₃, 100 MHz) δ 60.6, 68.6, 70.1, 73.1, 73.4, 75.5, 79.5, 79.6, 79.7,
81.5, 91.2, 127.5(7), 127.6(3), 127.8, 127.9, 128.0, 128.1, 128.3, 128.4, 130.1,
137.8, 138.6

261

264

262 (3R,4S,5R,6R)-3,4-Bis(benzyloxy)-6-((benzyloxy)methyl)-5-methoxy-2-(*p*-tolylthio)t
263 etrahydro-2*H*-pyran

OBn BnO BnO

GD3

Same procedures as the preparation of **GD1** and **GD2** (Fig. S1): treatment of **7c** (232 mg, 0.50 mmol) with acetic anhydride (2 mL) and triethylamine (2 mL) under ambient condition, smoothly afforded the corresponding acetyl glycoside **8c** within 4 h (the completion of the reaction was monitored by TLC). The reaction mixture was concentrated *in vacuo*, yielding a viscous residue, which was subjected to flash column chromatography (hexane/EtOAc = 5:1) to give **8c** as an oil liquid.

271A dry CH₂Cl₂ was added to a Schlenk tube containing 8c (ca. 0.50 mmol) and 272Tol-SH (74.0 mg, 0.60 mmol, 1.2 equiv.) under a N_2 atmosphere. The reaction 273mixture was cooled down to 0 °C before catalytic amount of BF₃⊕Et₂O 1M in Et₂O 274(10 μ L, 0.01 mmol, 0.02 equiv.) was added dropwise to the mixture. The mixture 275was stirred for 3 h at RT and cooled to 0 °C before 100 μ L of triethylamine was 276added slowly to quench the reaction. After that, the mixture was allowed to increase 277to RT and concentrated in vacuo to afford a viscous residue which was purified by 278silica gel column chromatography (hexane/EtOAc = 5:1) to afford GD3 as a 279 colorless viscous liquid as a mixture of two anomers (158 mg, 0.33 mmol, 65% yield 280 in 2 steps).

 1 H NMR (CDCl₃, 400 MHz) δ 2.29 (s), 2.31 (s) (3H) (these two singlets are –Me in –

tolyl group of two anomers), 3.49 (s), 3.51 (s) (3H) (these two singlets are –Me in 4– O-Me group of two anomers) 3.57–3.65 (m, 1H), 3.72–3.85 (m, 2H), 4.26 (dd, J =8.0, 2.0 Hz 1H), 4.45 (d, J = 12.4 Hz, 1H), 4.56–4.89 (m, 6H), 4.96 (d, J = 10.8 Hz, 1H), 5.55 (d, J = 5.2 Hz, 1H), 7.01–7.06 (m, 2H), 7.28–7.38 (m, 17H).

¹³C NMR (CDCl₃, 100 MHz) δ 21.3, 60.9, 68.8, 69.3, 71.2, 72.6, 73.5, 75.5, 75.8,
75.9, 79.2, 79.3, 79.7, 79.9, 80.7, 82.6, 86.8, 87.5, 87.7, 127.7, 127.9, 128.0, 128.1,
128.1(7), 128.2(4), 128.3, 128.4, 128.5, 128.6, 129.8, 130.8, 132.2, 132.8, 137.3,
137.9, 138.2, 138.6, 138.9

290

291 **Preparation of glycosyl donor GA1 and GA2** (Fig. S3)

292 ((2R,3S,4S,5R)-3,4,5,6-Tetrakis(benzyloxy)tetrahydro-2H-pyran-2-yl)methanol



293

Triphenylmethyl chloride (306 mg, 1.1 mmol) was added to a stirred solution of D-galactose (180 mg, 1.0 mmol) in pyridine (2 mL), and the mixture was heated at 50 °C for overnight to afford a yellow suspension. MeOH (1.5 equiv.) was added to quench the reaction, affording yellow solution, which was concentrated *in vacuo*. The resultant syrup was purified by flash column chromatography (EtOAc/MeOH = 4:1) to give crude trityl ether, a white fluffy solid, as the precursor of **GA1**, which can be used for next step.

A stirred suspension of crude trityl ether in dry DMF (5 mL) was added NaH (60% in mineral oil, 60.0 mg, 1.5 mmol) in 2 portions at 0 °C, and stirred for 30 min, followed by treatment of benzyl bromide (178 μ L, 1.5 mmol, 1.5 equiv.), stirred for 8 h at RT, MeOH (1.5 equiv.) was added to quench the reaction slowly until no gas was formed. After extracted 2 times with EtOAc/H₂O (1:1, 10 mL), the organic layer was concentrated *in vacuo*. The CH₂Cl₂/MeOH solution of the yielded mixture was added TsOH·H₂O (15 mol%) then it was stirred at RT for overnight. The solution was neutralized with NaHCO₃, and organic solvent was removed in *vacuo* to afford a crude viscous liquid. Column chromatography (hexane/EtOAc = 3:1) yielded glycosyl acceptor **GA1** (254 mg, 0.47 mmol, 47% yield) as a white solid.

¹H NMR (CDCl₃, 400 MHz) δ 2.13–2.16 (m, 1H), 3.58–3.67 (m, 2H), 3.72–3.78 (m,

- 312 1H), 4.06–4.10 (m, 2H), 4.34 (t, J = 7.2 Hz, 1H), 4.48 (dd, J = 12.4, 7.2 Hz, 2H),
- 313 4.56–4.65 (m, 3H), 4,71–4.83 (m, 3H), 4.98 (d, *J* = 2.0 Hz, 1H), 7.27–7.36 (m, 20H).
- ¹³C NMR (CDCl₃, 100 MHz) δ 61.5, 69.1, 72.3, 72.7, 81.0, 83.8, 98.7, 127.7, 127.9,
- 315 **128.0**, **128.2**, **128.4**, **137.3**, **137.4**, **137.7**, **138.3**.
- 316

318

317 ((2R,3R,4S,5R)-3,4,5,6-Tetrakis(benzyloxy)tetrahydro-2H-pyran-2-yl)methanol



GA2

GA2 as a white solid (270 mg, 0.50 mmol, 50% yield) was prepared from D-glucose
(180 mg, 1.0 mmol) in the same procedures as GA1 described above.

³²¹ ¹H NMR (CDCl₃, 400 MHz) δ 1.85 (t, J = 7.2 Hz, 1H), 3.34–3.39 (m, 1H), 3.49 (t, J = 322 8.0 Hz, 1H), 3.57 (t, J = 9.6 Hz, 1H), 3.67–3.73 (m, 2H), 3.85–3.90 (m, 1H), 4.56–4.88 (m, 6H), 4.91–5.00 (m, 3H), 7.25–7.38 (m, 20H).

- ¹³C NMR (CDCl₃, 100 MHz) δ 62.0, 71.6, 74.9(5), 75.0(2), 75.7, 77.5, 82.3, 84.5,
 102.8, 127.6, 127.7, 127.8(5), 127.9(1), 128.0, 128.1, 128.3, 128.4, 128.5, 137.2,
 137.9, 138.2, 138.4.
- 327

328 **Preparation of glycosyl donor GA3** (Fig. S4)

329 (3*R*,4*S*,5*R*,6*R*)-2-(Benzyloxy)-6-(hydroxymethyl)tetrahydro-2*H*-pyran-3,4,5-triol



9a

To a stirred solution of D-glucose [ref. S2] (1.8 g, 10.0 mmol) in DMF (30 mL), a 60% dispersion of NaH in mineral oil (400 mg, 10.0 mmol, 1.0 equiv.) was added at 0°C. After stirring the mixture for 30 min, benzyl bromide (BnBr: 140 μ L, 1.2 mmol, 1.2 equiv.) was added to the resultant mixture, stirred for a further 3 h at RT; MeOH (1.2 equiv.) was then added to stop the reaction. The reaction mixture was concentrated in *vacuo*, followed by flash column chromatography (EtOAc/MeOH = 4:1) to yield **9a** as an amorphous white solid (1.30 g, 4.8 mmol, 48% yield).

³³⁸ ¹H NMR (CD₃OD, 400 MHz) δ 3.30 (t, J = 1.2 Hz, 1H), 3.43–3.51 (m, 2H), 3.58 (dd, 339 J = 10.0, 8.0 Hz, 1H), 3.71–3.89 (m, 4H), 4.30 (d, J = 8.0 Hz, 1H), 4.65 (d, J = 12.0340 Hz, 1H), 4.83–4.94 (m, 3H), 7.25–7.33 (m, 3H), 7.41 (d, J = 7.2 Hz, 2H).

¹³C NMR (CD₃OD, 100 MHz) δ 62.5, 70.2, 71.6, 72.5, 74.9, 76.6, 103.8, 128.6,
129.1, 129.2, 139.1

343

346

344 (3a*S*,4*R*,7*R*,7a*R*)-6-(Benzyloxy)-4-(hydroxymethyl)-2,2-dimethyltetrahydro-4*H*-[1,3]
345 dioxolo[4,5-*c*]pyran-7-ol



 (\pm) -CSA (51.0 mg, 0.22 mmol, 0.05 equiv.) was added to a solution of **9a** (1.20 g, 4.4 mmol) in acetone dimethtylacetal (35 mL) and the solution was stirred overnight at room temperature. The reaction mixture was treated with triethylamine and concentrated to a residue, which was then dissolved in a mixture of MeOH/H₂O 10:1 (35 mL) and refluxed for 48 h. The reaction mixture was concentrated to a residue, which was subjected to silica gel column chromatography (hexane/EtOAc = 1:1) to yield **10a** as a white solid (1.27 g, 4.1 mmol, 93% yield).

354¹H NMR (CDCl₃, 400 MHz) δ 1.35 (s, 3H), 1.52 (s, 3H), 1.61 (d, J = 3.6 Hz, 1H),3552.04-2.08 (m, 1H), 2.44 (q, J = 2.0 Hz, 1H), 3.62 (td, J = 8.4, 2.4 Hz, 1H), 3.82–3.87356(m, 2H), 3.96–4.01 (m, 1H), 4.09 (dd, J = 7.2, 5.6 Hz, 1H), 4.15 (dd, J = 5.2, 2.0 Hz,

357 1H), 4.29 (d, J = 8.4 Hz, 1H), 4.66 (d, J = 12.0 Hz, 1H), 4.92 (d, J = 12.0 Hz, 1H),
358 7.30–7.38 (m, 5H).

¹³C NMR (CDCl₃, 100 MHz) δ 26.3, 28.1, 62.4, 71.3, 73.5, 73.6, 73.9, 78.8, 101.2,
110.4, 128.1, 128.2, 128.5, 136.8

361

364

362 (2*R*,3*R*,4*S*,5*R*)-5,6-Bis(benzyloxy)-2-((benzyloxy)methyl)tetrahydro-2*H*-pyran-3,4-d
 363 iol



11a

A stirred suspension of **10a** (1.24 g, 4.0 mmol) in dry DMF (10 mL) was added 365 NaH (60% in mineral oil, 400 mg, 10.0 mmol, 2.5 equiv.) in 2 portions at 0 °C, and 366 367 stirred for 2 h, followed by treatment of benzyl bromide (1.2 mL, 10.0 mmol, 2.5 368 equiv.), stirred for 8 h at RT, MeOH (1.5 equiv.) was added to quench the reaction 369 slowly until no gas was formed. After extracted 2 times with EtOAc/H₂O (1:1, 10 mL), 370 the organic layer was concentrated *in vacuo*. Aqueous acetic acid (60%) was added to the crude residue, stirred at 65°C for 2 h. Most solvent was removed in vacuo 371 372 before NaHCO₃ and MeOH was added to neutralize the residual HOAc. The 373 mixture was stirred for 30 min and then concentrated in vacuo to afford a crude 374 viscous, followed by column chromatography (hexane/EtOAc = 1:1), yielded **11a** (990 mg, 2.2 mmol, 55% yield) as a white solid. 375

 1 H NMR (CDCl₃, 400 MHz) δ 2.47 (d, J = 2.8 Hz, 1H), 2.57 (d, J = 2.8 Hz, 1H), 3.55–

377 3.59 (m, 2H), 3.63 (t, J = 5.6 Hz, 1H), 3.76–3.84 (m, 2H), 4.01 (s, 1H), 4.48 (d, J =

378 7.6 Hz, 1H), 4.61 (s, 2H), 4.66 (d, J = 11.6 Hz, 2H), 4.98 (dd, J = 11.6, 4.0 Hz, 2H),
379 7.29–7.39 (m, 15H).

¹³C NMR (CDCl₃, 100 MHz) δ 68.9, 69.3, 70.9, 73.1, 73.3, 73.7, 74.6, 79.1, 102.4,
127.6(9), 127.7(4), 127.8, 127.8(5), 127.9(1), 128.1, 128.4, 128.4, 128.5, 137.3,
137.8, 138.2.

386

384 (2*R*,3*S*,4*S*,5*R*)-5,6-Bis(benzyloxy)-2-((benzyloxy)methyl)-4-((4-methoxybenzyl)oxy)

385 tetrahydro-2*H*-pyran-3-ol



12a

387 A mixture of **11a** (900 mg, 2.0 mmol) and dibutyltin oxide (600 mg, 2.4 mmol, 1.2 equiv.) in methanol (10 mL) was refluxed for 10 h with stirring. The solvent was 388 389 removed and dried in vacuo. To the residue in dry benzene (10 mL) was added *para*-methoxy benzyl chloride (PMB-CI: 540 μ L, 4.0 mmol, 2.0 equiv.), MS4Å (0.4 g) 390 391 and tetrabutylammonium bromide (TBAB: 322 mg, 1.0 mmol, 0.5 equiv.), refluxed 392 for 6 h. The mixture was concentrated in *vacuo*, followed by column chromatography 393 (hexane/EtOAc = 3:1), yielded **12a** as a viscous oil liquid (1.01 g, 1.8 mmol, 89%) 394 vield).

³⁹⁵ ¹H NMR (CDCl₃, 400 MHz) δ 3.47 (dd, J = 9.6, 3.2 Hz, 1H), 3.57 (q, J = 5.2 Hz, 1H), 396 3.69 (dd, J = 9.6, 8.0 Hz, 1H), 3.76–3.84 (m, 5H), 4.00 (s, 1H), 4.46–4.47 (m, 1H), 4.61–4.74 (m, 7H), 4.90–5.00 (m, 2H), 6.84 (dd, J = 6.8, 2.0 Hz, 2H), 7.24–7.38 (m, 398 17H).

¹³C NMR (CDCl₃, 100 MHz) δ 55.2, 68.9, 70.9, 72.7, 73.5, 74.4, 75.2, 79.6, 82.0,
102.8, 113.7, 127.4(7), 127.5(2), 127.6, 127.8, 127.9, 128.1, 128.2, 128.3, 128.3(5),
128.4(1), 129.2, 130.6, 137.6, 137.9, 138.6, 138.7, 159.1.

402

403 (3*R*,4*S*,5*R*,6*R*)-2,3,5-Tris(benzyloxy)-6-((benzyloxy)methyl)tetrahydro-2*H*-pyran-4-404 ol



GA3

405

406 A stirred suspension of **12a** (620 mg, 2.0 mmol) in dry DMF (5 mL) was added NaH (60% in mineral oil, 200 mg, 5.0 mmol, 2.5 equiv.) in 2 portions at 0 °C, and stirred 407 408 for 2 h, followed by treatment of benzyl bromide (600 μ L, 5.0 mmol, 2.5 equiv.), 409 stirred for 8 h at RT, MeOH (1.5 equiv.) was added to guench the reaction slowly 410 until no gas was formed and organic solvent was removed in vacuo. The mixture 411 was dissolved in CH₂Cl₂ and washed with 2M HCI and then with saturated NaHCO₃ 412solution, dried over Na₂SO₄ and then concentrated. Water (1 mL) was added to the 413 resultant mixture in CH₂Cl₂ (15 mL) followed by addition of 414 2.3-dichloro-5.6-dicyanobenzoquinone (DDQ: 680 mg, 3.0 mmol 1.5 equiv.) under stirring. The mixture was stirred for 1 h at room temperature and the completion of 415416 the reaction was monitored by TLC. CH₂Cl₂ was filtered and the residue was 417 repeatedly washed with CH₂Cl₂. Filtrates were collected, washed with NaHCO₃ 418 solution, dried over Na₂SO₄ and then concentrated. The residue was subjected to 419 silica gel column chromatography (hexane/EtOAc = 5:1), yielded GA3 as an oil 420 liquid (775 mg, 1.4 mmol, 72% yield).

 1 H NMR (CDCl₃, 400 MHz) δ 2.26 (d, J = 3.2 Hz, 1H), 3.60–3.67 (m, 5H), 3.87 (s,

422 1H), 4.44–4.53 (m, 3H), 4.62–4.69 (m, 3H), 4.79 (d, J = 11.6 Hz, 1H), 4.96 (q, J =

423 6.0 Hz, 2H), 7.29–7.37 (m, 20H).

¹³C NMR (CDCl₃, 100 MHz) δ 68.7, 70.8, 73.5, 73.6, 74.0, 74.6, 74.9, 75.4, 79.6,
102.5, 127.6, 127.7, 127.8, 128.1, 128.2(5), 128.2(8), 128.4, 137.4, 137.8, 138.3,
138.4

- 427
- 428
- 429 **Preparation of glycosyl donor GA4** (Fig. S5)
- 430 (3*R*,4*S*,5*R*)-2-(Benzyloxy)tetrahydro-2*H*-pyran-3,4,5-triol

9b

431

432 To a stirred solution of D-xylose [ref. S2] (3.0 g, 20.0 mmol) in DMF (30 mL), a 433 60% dispersion of NaH in mineral oil (800 mg, 20.0 mmol, 1.0 equiv.) was added in 434 3 portions at 0°C. After stirring the mixture for 30 min, benzyl bromide (BnBr: 280 435 μ L, 2.4 mmol, 1.2 equiv.) was added to the resultant mixture, stirred for a further 3 h 436 at RT; MeOH (1.2 equiv.) was then added to stop the reaction. The reaction mixture 437 was concentrated in *vacuo*, followed by flash column chromatography 438 (EtOAc/MeOH = 4:1) to yield 9b as an amorphous white solid (2.16 g, 9.0 mmol, 439 45% yield).

¹H NMR (CDCl₃, 400 MHz) δ 2.10 (s, 1H), 3.18–3.23 (m, 1H), 3.39–3.46 (m, 1H), 3.58–3.69 (m, 1H), 3.84–3.99 (m, 1H), 4.33–4.37 (m, 1H), 4.45 (d, J = 12.0 Hz, 1H), 4.54 (d, J = 12.0 Hz, 1H), 4.71 (d, J = 12.0 Hz, 1H), 4.83 (d, J = 12.0 Hz, 1H), 5.30 (s, 1H), 7.27–7.33 (m, 5H).

¹³C NMR (CDCl₃, 100 MHz) δ 61.7, 65.2, 69.1, 69.5, 69.8, 70.9, 71.8, 73.0, 74.3,
76.0, 97.5, 102.1, 127.9, 128.0, 128.1, 128.4, 136.9.

446

447 (2R,3R,4aR,8R,8aR)-7-(Benzyloxy)-2,3-dimethoxy-2,3-dimethylhexahydro-5*H*-pyra
448 no[3,4-*b*][1,4]dioxin-8-ol



449

To a solution of **9b** [ref. S6] (1.92 g, 8.0 mmol) in MeOH (30 mL), trimethyl orthoformate (4.4 mL, 40.0 mmol, 5.0 equiv.), 2,3-butanedione (1 mL, 12.0 mmol, 1.5 equiv.), and (\pm)-CSA (200 mg) were added and the mixture was refluxed for 20 h under an argon atmosphere, cooled to room temperature and neutralized by the addition of triethylamine (1 mL). Solvents were evaporated under reduced pressure and the residue was purified by silica gel column chromatography (hexane/EtOAc = 456 3:1) to afford **10b** as a white solid (1.06 g, 3.0 mmol, 38% yield).

¹H NMR (CDCl₃, 400 MHz) δ 1.30 (s, 3H), 1.35 (s, 3H), 3.27 (s, 3H), 3.30 (s, 3H), 3.43 (t, J = 9.8 Hz, 1H), 3.57–3.67 (m, 2H), 3.78–3.84 (m, 1H), 3.93 (q, J = 5.2 Hz, 1H), 4.36 (d, J = 9.8 Hz, 1H), 4.60 (d, J = 7.6 Hz, 1H), 4.90 (d, J = 7.6 Hz, 1H), 7.31– 7.37 (m, 5H).

¹³C NMR (CDCl₃, 100 MHz) δ 17.5, 17.6, 47.9, 64.1, 65.8, 71.1, 71.3, 72.2, 99.4,
99.7, 103.2, 128.0, 128.2, 128.5, 136.8.

463

464 (3R,4S,5R)-5-(Allyloxy)-6-(benzyloxy)tetrahydro-2H-pyran-3,4-diol



465

466 To a stirred solution of **10b** (1.0 g, 2.8 mmol) in DMF (5 mL), a 60% dispersion of NaH in mineral oil (110 mg, 2.8 mmol, 1.0 equiv.) was added in 3 portions at 0°C. 467 After stirring the mixture for 30 min, allyl bromide (250 μ L, 3.0 mmol, 1.1 equiv.) was 468 469 added to the resultant mixture, stirred for a further 3 h at RT; MeOH (1.0 equiv.) was 470 then added to stop the reaction. The reaction mixture was concentrated in vacuo. 471 To a solution of the residue in dichloromethane (10 mL) was added 95% aqueous 472trifluoroacetic acid (5 mL) and the whole mixture was stirred for 15 min. Solvents 473 were evaporated in *vacuo* and the residue was immediately purified by column 474chromatography (hexane/EtOAc = 1:1) to afford **11b** as a white solid (196 mg, 0.70 mmol, 25% yield). 475

476 ¹H NMR (CDCl₃, 400 MHz) δ 3.35 (dd, *J* = 6.8, 4.8 Hz, 1H), 3.49 (dd, *J* = 12.0, 6.4

477 Hz, 1H), 3.71 (dd, J = 10.4, 4.8 Hz, 2H), 4.08–4.16 (m, 2H), 4.27 (dd, J = 12.8, 5.6

Hz, 1H), 4.59 (d, J = 11.6 Hz, 1H), 4.69 (d, J = 4.4 Hz, 1H), 4.86 (d, J = 12.0 Hz, 1H),
5.19–5.29 (m, 2H), 5.84–5.94 (m, 1H), 7.30–7.39 (m, 5H).

¹³C NMR (CDCl₃, 100 MHz) δ 63.3, 69.2, 70.4, 72.5, 77.9, 100.3, 118.0, 127.9,
128.0, 128.1, 128.5, 134.0, 136.7

483 (3*R*,4*S*,5*R*)-3-(Allyloxy)-2,4,5-tris(benzyloxy)tetrahydro-2*H*-pyran



484

To a stirred solution of **11b** (168 mg, 0.60 mmol) in DMF (5 mL), a 60% dispersion of NaH in mineral oil (60.0 mg, 1.5 mmol, 2.5 equiv.) was added at 0°C. After stirring the mixture for 30 min, benzyl bromide (BnBr: 180 μ L, 1.5 mmol, 2.5 equiv.) was added to the resultant mixture, stirred for a further 3 h at RT; MeOH (1.2 equiv.) was then added to stop the reaction. The reaction mixture was concentrated in *vacuo*, followed by flash column chromatography (EtOAc/MeOH = 5:1) to yield **12b** as a viscous oil liquid (215 mg, 0.47 mmol, 78% yield).

¹H NMR (CDCl₃, 400 MHz) δ 3.20 (dd, J = 11.6, 6.0 Hz, 1H), 3.05 (dd, J = 9.2, 8.0 Hz, 1H), 3.52 (t, J = 9.2 Hz, 1H), 3.57–3.63 (m, 1H), 3.94 (dd, J = 11.6, 5.2 Hz, 1H), 4.18–4.23 (m, 1H), 4.40 (d, J = 7.6 Hz, 2H), 4.62 (d, J = 12.0 Hz, 2H), 4.74 (d, J =11.6 Hz, 1H), 4.81–4.93 (m, 3H), 5.16 (dt, J = 10.4 Hz, 1.2 Hz, 1H), 5.26 (dt, J =496 17.2 Hz, 2.4 Hz, 1H), 5.89–5.97 (m, 1H), 7.28–7.38 (m, 15H).

¹³C NMR (CDCl₃, 100 MHz) δ 63.9, 71.0, 73.4, 73.8, 75.6, 77.7, 81.5, 83.7, 103.1,
116.9, 127.6, 127.7, 127.8, 128.0, 128.3, 128.3(6), 128.4(1), 135.0, 137.4, 138.2,
138.6.

500

502

501 (3*R*,4*R*,5*R*)-2,4,5-Tris(benzyloxy)tetrahydro-2*H*-pyran-3-ol



GA4

A mixture of **12b** (184 mg, 0.40 mmol), sodium acetate (82.0 mg, 1.0 mmol, 2.5 equiv.), $PdCl_2$ (85.0 mg, 0.48 mmol, 1.2 equiv.), and aqueous acetic acid (acetic acid/H₂O = 11:1, 6 mL) was stirred at RT for 48 h. After removing acetic acid and water in *vacuo*, the residue was subjected to silica gel column chromatography 507 (hexane/EtOAc = 3:1) to afford **GA4** as a white amorphous solid (138 mg, 0.33 508 mmol, 82% yield).

- $509 = {}^{1}\text{H NMR} (\text{CDCI}_{3}, 400 \text{ MHz}) \delta 2.74 (s, 1H), 3.35 (dd, J = 12.0, 8.0 \text{ Hz}, 1H), 3.55-3.63$
- 510 (m, 4H), 4.03 (dd, J = 11.6, 3.6 Hz, 1H), 4.46 (d, J = 6.0 Hz, 1H), 4.60–4.71 (m, 3H),
- 511 4.80–4.89 (m, 3H), 7.28–7.35 (m, 15H).
- ¹³C NMR (CDCl₃, 100 MHz) δ 62.4, 70.5, 72.1, 72.8, 74.1, 80.9, 101.6, 127.6, 127.8,
 127.9, 128.3(5), 128.4(0), 128.5, 137.2, 137.8, 138.4.
- 514
- 515

516 Typical procedure of glycosylation of GD and GA followed by deprotection for

- 517 **G1-G8 and G10** (Fig. S6)
- 518

519 **Glycosylation**

520 A 20 mL oven-dried Schlenk tube was charged with thioglycoside GD1 (25.0 mg, 5210.04 mmol) and glycosyl acceptor **GA1** (23.0 mg, 1.0 equiv.) and dry CH_2Cl_2 (2 mL). 522The mixed solution was cooled down to -40 °C, N-iodosuccinimide (NIS; 18.0 mg, 5230.08 mmol, 2.0 equiv.) was added until totally dissolved, followed by 2 μ L of TfOH. 524The reaction mixture was allowed to warm to RT slowly. After that, it was cooled to 5250 °C before triethylamine was added to neutralize the mixture. The mixture was 526concentrated in vacuo and purified by column chromatography (hexane/EtOAc = 5275:1) to yield the mixture of two anomers (28.1 mg, 0.03 mmol, 70% yield). After TLC 528isolation with CHCl₃/MeOH (40:1), the pure form of the β anomer (**G1**', precursor of 529**G1**) was isolated as the minor product, along with α anomer (**G2**', precursor of **G2**) 530as the major one.

531

532 **Deprotection**

533

534 A 20 mL Schlenk tube was charged with isolated **G1'** (7.0 mg) in MeOH/EtOAc 535 (1:1 v/v, 2 mL) under N₂, then added Pd(OH)₂/C (10%) (7.0 mg), H₂ gas was later 536 charged and replace the N₂ by air pump vacuum/H₂ exchange, the reaction mixture was stirred at room temperature for overnight, the completion of the reaction was monitored by LC-MS. The mixture was subjected to filtration by passing through a packed Celite® cake, to remove Pd(OH)₂/C, the crude residue was obtained after removing the solvent *in vacuo*, followed by reverse phase TLC (H₂O), afforded the corresponding product **G1** as a white solid (2.5 mg, 95%). In the same procedure, the reaction of **G2'** (α anomer of **G1'**, 7.0 mg) yielded **G2** as white solid (2.5 mg, 95%).

The NMR spectra of isolated disaccharide in D_2O was difficult to identify each signal to corresponding proton, nevertheless, the absence of aromatic proton in ¹H-NMR spectra was obviously observed which helped us to confirm the completion of the removal of all benzyl groups. The formation of desired disaccharide **G** was confirmed via HRMS showing the consistence of the found value with exact mass calculated.

550

551 Other disaccharides **G3–G8** and **G10** were synthesized in the same procedure 552 through a glycosylation of corresponding **GD** and **GA**, followed by a hydrogenative 553 deprotection.

554 The yield and additional information of each disaccharide was listed in Fig. S6. 555

556 Benzyl(2S,3S,4S,5R,6R)-4,5-bis(benzyloxy)-3-methoxy-6-(((2R,3S,4S,5R)-3,4,5,6-

557 tetrakis(benzyloxy)tetrahydro-2*H*-pyran-2-yl)methoxy)tetrahydro-2*H*-pyran-2-carbo

558 xylate



559

560 Glycosylation of **GD1** and **GA1** (β)

G1'

⁵⁶¹ ¹H NMR (CDCl₃, 600 MHz) δ 3.31 (s, 3H), 3.46 (t, J = 6.4 Hz, 1H), 3.50–3.51 (m,

562 2H), 3.67-3.70 (m, 1H), 3.75-3.78 (m, 2H), 3.98-4.02 (m, 2H), 4.09-4.11 (m, 1H), 563 4,35 (t, J = 7.2 Hz, 1H), 4.40 (d, J = 7.2 Hz, 1H), 4.45 (dd, J = 2.0, 12.8 Hz, 2H), 564 4.53-4.58 (m, 3H), 4.65-4.75 (m, 5H), 4.83 (d, J = 11.0 Hz, 1H), 4.88-5.02 (m, 2H), 565 5.14 (d, J = 8.0 Hz, 1H), 5.17 (d, J = 8.1 Hz, 1H), 7.19-7.29 (m, 35H).

¹³C NMR (CDCl₃, 150 MHz) δ 60.5, 67.1, 68.5, 69.9, 72.1, 72.3, 73.1, 74.3, 74.7,
75.5, 79.2, 80.1, 80.6, 81.1, 81.4, 83.6, 83.9, 98.1, 103.9, 127.3, 127.5, 127.6,
127.7, 127.8, 127.9, 128.2, 128.3, 128.4, 135.1, 137.4, 138.0, 138.1, 138.3, 138.4,
168.4.

- 570
- 571 (2S,3S,4R,5R,6R)-4,5-Dihydroxy-3-methoxy-6-(((2R,3R,4S,5R)-3,4,5,6-tetrahydrox
- 572 ytetrahydro-2*H*-pyran-2-yl)methoxy)tetrahydro-2*H*-pyran-2-carboxylic acid



- 573 **G1** 4-Me-GlcA- β (1,6)Gal
- 574 HRMS (ESI) m/z calcd for C₁₃H₂₁O₁₂ [M–H]⁻: 369.1028, found: 369.1022.
- 575
- 576 Benzyl(2S,3S,4S,5R,6S)-4,5-bis(benzyloxy)-3-methoxy-6-(((2R,3S,4S,5R)-3,4,5,6-t
- 577 etrakis(benzyloxy)tetrahydro-2*H*-pyran-2-yl)methoxy)tetrahydro-2*H*-pyran-2-carbox
- 578 ylate



579

580 Glycosylation of **GD1+GA1** (α)

G2'

⁵⁸¹ ¹H NMR (CDCl₃, 600 MHz) δ 3.33 (s, 3H), 3.37–3.41 (m, 2H), 3.50 (dd, J = 9.6, 3.4

582 Hz, 1H), 3.70 (dd, J = 9.6, 6.9 Hz, 1H), 3.85–3.88 (m, 2H), 3.98–4.00 (m, 4H), 4.34 583 (d, J = 6.9 Hz, 1H), 4.51–4.57 (m, 2H), 4.65–4.76 (m, 6H), 4.83–4.87 (m, 3H), 4.91 584 (d, J = 11.7 Hz, 2H), 5.17 (d, J = 12.4 Hz, 1H), 5.23 (d, J = 12.4 Hz, 2H) 7.24–7.41 585 (m, 35H)

- ¹³C NMR (CDCl₃, 150 MHz) δ 60.6, 67.1, 68.0, 68.9, 69.5, 70.3, 73.0, 73.4, 73.5,
 74.5, 75.3, 75.7, 76.4, 79.0, 79.1, 81.3, 81.8, 95.6, 97.7, 127.50, 127.55, 127.63,
 127.69, 127.77, 127.86, 127.9, 128.1, 128.21, 128.26, 128.34, 128.4, 128.5, 135.2,
 137.3, 138.0, 138.45, 138.5, 138.8, 169.8.
- 590
- 591 (2S,3S,4R,5R,6S)-4,5-Dihydroxy-3-methoxy-6-(((2R,3R,4S,5R)-3,4,5,6-tetrahydrox
- 592 ytetrahydro-2*H*-pyran-2-yl)methoxy)tetrahydro-2*H*-pyran-2-carboxylic acid



- 593 G2 4-Me-GlcA- α (1,6)Gal
- 594 HRMS (ESI) m/z calcd for C₁₃H₂₁O₁₂ [M–H]⁻: 369.1028, found: 369.1037.
- 595
- 596 Benzyl(2S,3S,4S,5R,6R)-4,5-bis(benzyloxy)-3-methoxy-6-(((2R,3R,4S,5R)-3,4,5,6-
- 597 tetrakis(benzyloxy)tetrahydro-2*H*-pyran-2-yl)methoxy)tetrahydro-2*H*-pyran-2-carbo
- 598 xylate



G3'

- 600 Glycosylation of **GD1+GA2** (β)
- 1 H NMR (CDCl₃, 400 MHz) δ 3.34 (s, 3H), 3.38–3.41 (m, 1H), 3.45–3.52 (m, 4H),

3.58 (t, J = 6.0 Hz, 1H), 3.62–3.68 (m, 1H), 3.76–3.79 (m, 1H), 4.02–4.09 (m, 1H),
4.36 (q, J = 4.8 Hz, 1H), 4.24–4.44 (m, 1H), 4.47–4.53 (m, 3H), 4.62–4.64 (m, 1H),
4.68–4.79 (m, 4H), 4.81–4.85 (m, 2H), 4.91–4.99 (m, 3H), 5.20–5.24 (m, 2H), 7.15–
7.39 (m, 35H).

- ¹³C NMR (CDCl₃, 100 MHz) δ 60.7, 67.3, 68.8, 68.9, 70.2, 71.2, 73.0, 74.5, 74.6,
 75.0, 75.0(5), 75.1(3), 75.8, 78.0, 78.3, 79.9, 81.1(6), 81.2(3), 81.4, 81.5, 82.0, 82.3,
 83.9, 84.0, 84.7, 95.2, 102.7, 104.1, 104.2, 127.6, 127.7, 127.7(8), 127.8(2), 127.9,
 128.0, 128.1, 128.2, 128.3, 128.4, 128.5(0), 128.5(4), 128.5(8), 128.6(0), 128.6(3),
 128.7, 128.8, 129.0, 135.3, 137.2, 137.6, 138.0, 138.2, 138.3, 138.3(6), 138.4(2),
 138.6, 139.0, 168.4, 168.6.
- 612
- 613 (2S,3S,4R,5R,6R)-4,5-Dihydroxy-3-methoxy-6-(((2R,3S,4S,5R)-3,4,5,6-tetrahydrox
- 614 ytetrahydro-2*H*-pyran-2-yl)methoxy)tetrahydro-2*H*-pyran-2-carboxylic acid



- 615 **G3** 4-Me-GlcA- β (1,6)Glc
- 616 HRMS (ESI) m/z calcd for C₁₃H₂₂O₁₂Na [M+Na]⁺: 393.1003, found: 393.0994.
- 617
- 618 Benzyl(2S,3S,4S,5R,6R)-4,5-bis(benzyloxy)-3-methoxy-6-(((3R,4S,5S,6R)-2,3,5-tri
- 619 s(benzyloxy)-6-((benzyloxy)methyl)tetrahydro-2*H*-pyran-4-yl)oxy)tetrahydro-2*H*-pyr
- 620 an-2-carboxylate



G4'

- 622 Glycosylation of **GD1+GA3** (β)
- 1 H NMR (CDCl₃, 400 MHz) δ 3.28 (s, 3H), 3.41 (t, J = 9.6 Hz, 1H), 3.47–3.60 (m,

624 5H), 3.67 (d, J = 10.0 Hz, 1H), 3.82–3.87 (m, 2H), 3.94 (t, J = 9.6 Hz, 1H), 4.36– 625 4.46 (m, 4H), 4.53–4.67 (m, 3H), 4.74–4.83 (m, 4H), 4.90 (t, J = 12.4 Hz, 2H), 4.99 626 (d, J = 11.2 Hz, 1H), 5.10–5.14 (m, 2H), 7.22–7.40 (m, 35H).

627 13 C NMR (CDCl₃, 100 MHz) δ 60.4, 66.8, 68.7, 70.5, 71.0, 73.3, 73.4, 74.2, 74.5, 628 74.7, 75.5, 79.2, 80.5, 81.2, 82.0, 97.1, 103.2, 127.2, 127.6, 127.7(5), 127.8(0), 629 127.9(0), 127.9(2), 128.0, 128.1, 128.2, 128.3, 128.3(7), 128.4(3), 135.3, 137.4, 630 137.7, 138.0, 138.3(7), 138.4(3), 169.9.

631

632 (2S,3S,4R,5R,6R)-4,5-Dihydroxy-3-methoxy-6-(((3R,4S,5S,6R)-2,3,5-trihydroxy-6-(

633 hydroxymethyl)tetrahydro-2*H*-pyran-4-yl)oxy)tetrahydro-2*H*-pyran-2-carboxylic acid



634 **G4** 4-Me-GlcA-*β*(1,3)Gal

- 635 HRMS (ESI) m/z calcd for C₁₃H₂₂O₁₂Na [M+Na]⁺: 393.1003, found: 393.1001.
- 636
- 637 Benzyl(2S,3S,4S,5R,6R)-4,5-bis(benzyloxy)-3-methoxy-6-(((3R,4S,5R)-2,4,5-tris(b
- 638 enzyloxy)tetrahydro-2*H*-pyran-3-yl)oxy)tetrahydro-2*H*-pyran-2-carboxylate



G5'

639

640 Glycosylation of **GD1+GA4** (β)

 1 H NMR (CDCl₃, 400 MHz) δ 3.35 (s, 3H), 3.39–3.54 (m, 3H), 3.59–3.64 (m, 2H),

- 642 3.67–3.73 (m, 1H), 3.79–3.81 (m, 1H), 3.94–4.05 (m, 1H), 4.53–4.58 (m, 2H), 4.60–
- 643 4.65 (m, 2H), 4.74–4.84 (m, 5H), 4.87–4.91 (m, 1H), 5.15–5.28 (m, 3H), 7.22–7.41
- 644 (m, 30H).
- 13 C NMR (CDCl₃, 100 MHz) δ 60.5, 63.2, 67.1, 70.0, 72.9, 74.2, 74.9, 75.0, 75.5,

646 75.6, 77.9, 78.4, 81.1, 82.1, 83.4, 83.9, 101.4, 102.7, 127.4, 127.5, 127.6, 127.8(7),
647 127.9(1), 128.1(5), 128.2(2), 128.3, 128.4, 128.5, 128.6, 128.7, 137.7, 137.9, 138.3,
648 168.3.

649

- 650 (2S,3S,4R,5R,6R)-4,5-Dihydroxy-3-methoxy-6-(((3R,4S,5R)-2,4,5-trihydroxytetrahy
- 651 dro-2*H*-pyran-3-yl)oxy)tetrahydro-2*H*-pyran-2-carboxylic acid



652 **G5** 4-Me-GlcA- β (1,2)Xyl

- 653 HRMS (ESI) m/z calcd for C₁₂H₂₀O₁₁Na [M+Na]⁺: 363.0898, found: 363.0888.
- 654
- 655 Benzyl(2S,3S,4S,5R,6R)-3,4,5-tris(benzyloxy)-6-(((2R,3S,4S,5R)-3,4,5,6-tetrakis(b
- 656 enzyloxy)tetrahydro-2*H*-pyran-2-yl)methoxy)tetrahydro-2*H*-pyran-2-carboxylate



657

658 Glycosylation of **GD2+GA1** (β) C₆₈H₆₈O₁₂

⁶⁵⁹ ¹H NMR (CDCl₃, 400 MHz) δ 3.50 (t, J = 8.8 Hz, 1H), 3.61 (t, J = 8.8 Hz, 1H), 3.69 660 (dd, J = 8.4, 7.6 Hz, 1H), 3.79–3.88 (m, 3H), 3.98–4.04 (m, 2H), 4.11 (dd, J = 10.8, 661 2.4 Hz, 1H), 4.36 (t, J = 7.2 Hz, 1H), 4.42–4.48 (m, 4H), 4.53–4.59 (m, 3H), 4.66– 662 4.76 (m, 6H), 4.85–4.93 (m, 3H), 5.09 (s, 2H), 7.09–7.30 (m, 40H).

¹³C NMR (CDCl₃, 100 MHz) δ 67.2, 68.6, 69.9, 72.2, 72.4, 73.1, 74.5, 74.8, 75.0,
75.7, 79.3, 80.1, 80.6, 81.6, 83.7, 84.0, 98.1,104.0, 127.4, 127.6, 127.6(8), 127.7(4),
127.8, 128.0, 128.2, 128.3, 128.3(5), 128.3(9), 128.5, 135.0, 137.4(5), 137.5(3),
137.8, 138.1, 138.3, 138.4, 168.4.

672

675

- 668 (2S,3S,4S,5R,6R)-3,4,5-Trihydroxy-6-(((2R,3R,4S,5R)-3,4,5,6-tetrahydroxytetrahy
- 669 dro-2H-pyran-2-yl)methoxy)tetrahydro-2H-pyran-2-carboxylic acid



670 **G6** GlcA- β (1,6)-Gal

- 671 HRMS (ESI) m/z calcd for C₁₂H₂₀O₁₂Na [M+Na]⁺: 379.0847, found: 379.0864.
- 673 Benzyl(2S,3S,4S,5R,6R)-3,4,5-tris(benzyloxy)-6-(((3R,4S,5R)-2,4,5-tris(benzyloxy)
- 674 tetrahydro-2*H*-pyran-3-yl)oxy)tetrahydro-2*H*-pyran-2-carboxylate



G7'

676 Glycosylation of **GD2+GA4** (β)

⁶⁷⁷ ¹H NMR (CDCl₃, 400 MHz) δ 3.18 (t, J = 10.0 Hz, 1H), 3.44 (dd, J = 10.0, 3.6 Hz, ⁶⁷⁸ 1H), 3.56–3.66 (m, 3H), 3.73 (t, J = 8.0 Hz, 1H), 3.86–3.91 (m, 2H), 4.30 (t, J = 11.6⁶⁷⁹ Hz, 2H), 4.39 (d, J = 11.6 Hz, 1H), 4.47–4.56 (m, 4H), 4.59–4.68 (m, 2H), 4.73–4.90 ⁶⁸⁰ (m, 5H), 4.98 (d, J = 2.4 Hz, 2H), 5.62 (d, J = 3.2 Hz, 1H), 6.95–7.30 (m, 35H).

¹³C NMR (CDCl₃, 100 MHz) δ 63.8, 66.8, 70.3, 70.7, 71.9, 73.2, 74.8, 75.2, 75.3,
75.7, 78.4, 78.9, 79.9, 80.8, 81.2, 95.4, 103.3, 127.2, 127.4, 127.5, 127.6, 127.7,
127.9, 128.0, 128.1, 128.2, 128.3, 128.3(5), 128.4(4), 128.7, 135.3, 137.1, 137.6,
137.9, 138.3, 138.5, 170.2.

685

 $686 \qquad (2S, 3S, 4S, 5R, 6R) - 3, 4, 5 - Trihydroxy - 6 - (((3R, 4S, 5R) - 2, 4, 5 - trihydroxytetrahydro - 2H - p)) - 2H - p)$

687 yran-3-yl)oxy)tetrahydro-2H-pyran-2-carboxylic acid



- 688 **G7** GlcA- β (1,2)-Xyl
- 689 HRMS (ESI) m/z calcd for C₁₁H₁₈O₁₁Na [M+Na]⁺: 349.0741, found: 349.0734.
- 690
- 691 (3*R*,4*S*,5*S*,6*R*)-2,3,4,5-Tetrakis(benzyloxy)-6-((((2*R*,3*R*,4*S*,5*R*,6*R*)-3,4-bis(benzylo
- 692 xy)-6-((benzyloxy)methyl)-5-methoxytetrahydro-2*H*-pyran-2-yl)oxy)methyl)tetrahydr
- 693 o-2*H*-pyran



G8'

694

695 Glycosylation of **GD3+GA1** (β)

696 ¹H NMR (CDCl₃, 400 MHz) δ 3.32 (d, J = 6.0 Hz, 2H), 3.42–3.52 (m, 5H), 3.68–3.71 697 (m, 1H), 3.80–3.89 (m, 2H), 4.01–4.07 (m, 3H), 4.15 (dd, J = 7.2, 3.2 Hz, 1H), 4.29 698 (d, J = 12.0 Hz, 1H), 4.38–4.43 (m, 2H), 4.46–4.53 (m, 4H), 4.58–4.65 (m, 2H), 699 4.67–4.78 (m, 4H), 4.86–4.94 (m, 3H), 5.14 (s, 1H), 7.20–7.35 (m, 35H).

¹³C NMR (CDCl₃, 100 MHz) δ 60.5, 68.4, 69.0, 70.3, 71.8, 72.0, 73.0, 73.3, 73.6,
75.4, 75.7, 79.2, 79.7, 80.7, 81.3, 81.8, 82.7, 88.3, 97.7, 105.0, 127.4(7), 127.5(2),
127.5(9), 127.6(3), 127.6(6), 127.7(4), 127.7(9), 127.8(4), 127.8(7), 127.9(3), 128.0,
128.2, 128.2(6), 128.3(2), 128.3(4), 128.4, 137.6, 137.7, 137.8, 138.0, 138.1.

704

705 (3*R*,4*S*,5*R*,6*R*)-6-(((((2*R*,3*R*,4*R*,5*S*,6*R*)-3,4-Dihydroxy-6-(hydroxymethyl)-5-methoxy

tetrahydro-2*H*-pyran-2-yl)oxy)methyl)tetrahydro-2*H*-pyran-2,3,4,5-tetraol



- 707 **G8** GlcA- β (1,6)-Gal
- 708 HRMS (ESI) m/z calcd for C₁₃H₂₄O₁₁Na [M+Na]⁺: 379.1211, found: 379.1205.
- 709
- 710 Benzyl(2S,3S,4S,5R,6R)-4,5-bis(benzyloxy)-3-methoxy-6-((tetrahydro-2H-pyran-2-
- 711 yl)methoxy)tetrahydro-2*H*-pyran-2-carboxylate



- 712 G10'
- 713 Glycosylation of **GD1+ROH** (tetrahydropyran-2-methanol) (β)
- ¹H NMR (CDCl₃, 400 MHz) δ 1.49–1.84 (m, 6H), 3.35 (s, 3H), 3.59–3.43(m, 5H),
- 715 3.79–3.91 (m, 2H), 3.97–4.00 (m, 1H), 4.46 (dd, J = 13.2, 7.6 Hz, 1H), 4.66–4.76 (m,
- 716 2H), 4.85 (dd, J = 10.8, 2.0 Hz, 1H), 4.96 (dd, J = 10.8, 4.0 Hz, 1H), 5.20–5.30 (m,
- 717 3H), 7.29–7.40 (m, 15H).
- ¹³C NMR (CDCl₃, 100 MHz) δ 23.21, 25.9, 28.1, 28.3, 60.6, 67.2, 68.2, 68.4, 73.4,
- 719 73.7, 74.4, 74.6, 74.8, 75.6, 76.2, 81.0, 81.3(8), 81.4(4), 83.7, 103.9, 104.4, 127.6,
- 720 127.9, 128.2, 128.3, 128.4, 128.5, 128.6, 129.0, 135.2, 138.4, 168.4, 168.5.
- 721
- 722 (2S,3S,4R,5R,6R)-4,5-Dihydroxy-3-methoxy-6-((tetrahydro-2*H*-pyran-2-yl)methoxy)
- tetrahydro-2*H*-pyran-2-carboxylic acid



724 **G10** 4-Me-GlcA- β -Pyran

725 HRMS (ESI) m/z calcd for C₁₃H₂₂O₈Na [M+Na]⁺: 329.1207, found: 329.1198.

- 726
- 727 **Preparation of G9** (Fig. S7)

To a stirred solution of the **G1** (1.5 mg) in 3 mL of toluene/MeOH (1:2), TMSCHN₂

729 (1.1–1.5 mmol) was added dropwise until yellow color persisted. The mixture was

stirred for 30 min at RT and concentrated to give the corresponding methyl ester **G9**

731 as white solid (1.6 mg, 95% yield).

- 732
- 733 Methyl(2S,3S,4R,5R,6R)-4,5-dihydroxy-3-methoxy-6-(((2R,3R,4S,5R)-3,4,5,6-tetra
- 734 hydroxytetrahydro-2*H*-pyran-2-yl)methoxy)tetrahydro-2*H*-pyran-2-carboxylate



735

- 736 HRMS (ESI) m/z calcd for C₁₄H₂₄O₁₂Na [M+Na]⁺: 407.1165, found: 407.1173.
- 737 738
- 739 **Preparation of G11** (Fig. S8)
- 740 (2R,4aR,6R,7R,8R,8aS)-6-Methoxy-2-phenylhexahydropyrano[3,2-d][1,3]dioxine-7,
- 741 **8-diol**



742

2-OMe

A mixture of commercially purchased 1-OMe-Glc (β) (776 mg, 4.0 mmol), benzaldehyde dimethyl acetal (1.8 mL, 12.0 mmol), and TsOH·H₂O (189 mg, 1.0 mmol, 0.25 equiv.) was heated at 50 °C for 3 h. The solution was neutralized with NaHCO₃, filtration and wash with MeOH. The filtrate was concentrated *in vacuo* and the residue was purified by flash column chromatography (EtOAc) and recrystallized from EtOAc/hexane (1:2) afforded an analytical sample of acetal **2-OMe** as a white solid (761 mg, 2.7 mmol, 68% yield).

- ⁷⁵⁰ ¹H NMR (CDCl₃, 400 MHz) δ 2.56 (s, 1H), 2.70 (s, 1H), 3.45–3.57 (m, 3H), 3.59 (s,
- 3H), 3.77–3.86 (m, 2H), 4.33–4.39 (m, 2H), 5.55 (s, 1H), 7.37–7.41 (m, 3H), 7.49–
 7.51 (m, 2H).

¹³C NMR (CDCl₃, 100 MHz) δ 57.5, 66.4, 68.7, 73.2, 74.5, 80.6, 101.9, 104.1, 126.3,
128.4, 129.3.

755

758

756 (2R,3R,4S,5R,6R)-4,5-Bis(benzyloxy)-2-(hydroxymethyl)-6-methoxytetrahydro-2H-

pyran-3-ol



3-OMe

759 To a stirred suspension of acetal **2-OMe** (676 mg, 2.4 mmol) in dry DMF (12 mL) was added NaH 60% in mineral oil (240 mg, 6.0 mmol, 2.5 equiv.) in 2 portions at 760 761 0 °C, and stirred for 1 h, followed by treated with benzyl bromide (BnBr: 720 μ L, 6.0 762mmol), stirred for 8 h at room temperature, and MeOH (1.0 equiv.) was added 763 slowly until no gas was formed to quench the reaction. After extracted with 764 EtOAc/H₂O = 1:1 (2 times), organic layer was concentrated *in vacuo* to remove 765solvents to give a crude oil mixture. To the CH₂Cl₂/MeOH solution of the yielded oil 766 mixture was added TsOH H_2O (10 mol%), and then the mixture was stirred at room 767 temperature for overnight. The solution was neutralized with NaHCO₃, and organic 768 solvent was concentrated in vacuo to afford a crude viscous liquid. Column 769 chromatography (hexane/EtOAc = 1:1) yielded dibenzyl **3-OMe** as a white solid

770 (664 mg, 1.8 mmol, 74% yield).

¹H NMR (CDCl₃, 400 MHz) δ 2.03 (t, J = 6.0 Hz, 1H), 2.70 (d, J = 2.4 Hz, 1H), 3.32– 3.37 (m, 1H), 3.39–3.47 (m, 2H), 3.54 (dd, J = 9.2, 2.0 Hz, 1H), 3.74–3.80 (m, 1H), 3.87–3.92 (m, 1H), 4.37 (d, J = 7.6 Hz, 1H), 4.69 (dd, J = 14.4, 11.2 Hz, 2H), 4.95 (dd, J = 17.6, 12.8 Hz, 2H), 7.29–7.39 (m, 10H).

- ¹³C NMR (CDCl₃, 100 MHz) δ 57.3, 62.5, 70.3, 74.6, 74.8, 75.2, 81.9, 83.8, 104.9,
- 776 127.7, 127.9, 128.1, 128.4, 128.6, 138.3, 138.4.
- 777

780

778 (2R,3R,4S,5R,6R)-4,5-Bis(benzyloxy)-6-methoxy-2-((trityloxy)methyl)tetrahydro-2H

779 -pyran-3-ol



4-OMe

4.0 2.5 781 Triphenylmethyl chloride (1.1)mmol, equiv.) and g, 782*N*,*N*-dimethyl-4-aminopyridine (DMAP: 20.0 mg, 0.16 mmol) were added to a stirred solution of dibenzyl **3-OMe** (600 mg, 1.6 mmol) in dry pyridine (10 mL) and the 783 784 reaction mixture heated at 60 °C for overnight. The reaction mixture was quenched 785 with MeOH (5 mL) and concentrated *in vacuo*. The resulting syrup was purified by 786short column chromatography (hexane/EtOAc = 1:1) to give the trityl ether 4-OMe 787 (523 mg, 0.85 mmol, 53% yield) as a viscous oil liquid.

¹H NMR (CDCl₃, 400 MHz) δ 2.46 (s, 1H), 3.35–3.47 (m, 5H), 3.60–3.66 (m, 4H),

4.34 (d, J = 7.6 Hz, 1H), 4.73 (t, J = 10.8 Hz, 2H), 4.92 (t, J = 10.8 Hz, 2H), 7.20–
7.38 (m, 20H), 7.42–7.48 (m, 5H).

¹³C NMR (CDCl₃, 100 MHz) δ 56.8, 64.0, 71.7, 73.9, 74.7, 75.4, 81.9, 84.1, 86.8,
104.6, 127.0, 127.6, 127.7(6), 127.8(3), 127.9(6), 128.0(4), 128.3, 128.5, 128.6,
138.5, 138.6, 143.7.

795 ((2R,3R,4S,5R,6R)-4,5-Bis(benzyloxy)-3,6-dimethoxytetrahydro-2H-pyran-2-yl)met

hanol

HO MeO OMe BnO ÖBn

5-OMe

797

798 To a stirred solution of trityl ether **4-OMe** (493 mg, 0.8 mmol) in DMF (8 mL) was 799 added 60% NaH dispersion in mineral oil (48.0 mg, 1.2 mmol, 1.5 equiv.) at 0 °C. 800 after 30 min, iodomethane (MeI: 75 μ L, 1.2 mmol) was added and the reaction 801 mixture was stirred for another 3 h at room temperature before 1.0 equiv. MeOH 802 was added to stop the reaction. The reaction mixture was extracted with 803 $H_2O/EtOAc/hexane = 2:1:1 (10 mL)$ for 2 times to remove DMF, dried over Na₂SO₄. 804 After filtration, the mixture was concentrated *in vacuo*. The yielded crude product 805 was dissolved in MeOH/CH₂Cl₂ (30 mL) to afford a yellow solution which was 806 treated with TsOH·H₂O (10 mol%) and stirred at RT, after 6 h, NaHCO₃ was added 807 to neutralize the reaction. The yellow solution was concentrated in vacuo, followed 808 by a flash column chromatography (hexane/EtOAc = 3:1) to yield alcohol **5-OMe** as a white solid (168 mg, 0.43 mmol, 54%). 809

¹H NMR (CDCl₃, 400 MHz) δ 2.03 (t, J = 6.4 Hz, 1H), 3.28–3.31 (m, 2H), 3.35 (t, J = 8.4 Hz, 1H), 3.56 (s, 3H), 3.57 (s, 3H), 2.03 (t, J = 6.4 Hz, 1H), 3.74 (dd, J = 9.2, 7.2 Hz, 1H), 3.90 (dd, J = 12.0, 3.6 Hz, 1H), 4.34 (d, J = 8.0 Hz, 1H), 4.70 (d, J = 11.2

813 Hz, 1H), 4.79 (d, J = 11.2 Hz, 1H), 4.89 (d, J = 11.2 Hz, 2H), 7.27–7.35 (m, 10H).

¹³C NMR (CDCl₃, 100 MHz) δ 57.2, 60.8, 61.9, 74.8, 75.0, 75.5, 79.5, 82.1, 84.2,
104.7, 127.6, 127.9, 128.0, 128.3, 138.4, 138.5

- 816
- 817 (2S,3S,4S,5R,6R)-4,5-Bis(benzyloxy)-3,6-dimethoxytetrahydro-2*H*-pyran-2-carbox
 818 ylic acid



6-OMe

819

820 To a 10 mL CH₂Cl₂ solution of alcohol **5-OMe** (116 mg, 0.30 mmol) and buffer (Na₂HPO₄/NaH₂PO₄, PH=7) was added iodobenzene diacetate (193 mg, 0.60 mmol, 821 822 2.0 equiv.) at 0 °C, followed by addition of 2-azaadamantane N-oxyl (AZADO, 4.0 823 mg, 0.03 mmol, 0.1 equiv.). The mixture was stirred at the same temperature. The 824 reaction was monitored by TLC, after 2 h, 5-OMe was consumed, the reaction 825 mixture was extracted by CH₂Cl₂ and water for 3 times, the organic layer was dried 826 over Na₂SO₄, solvent was removed in *vacuo* and the residue was purified by a silica 827 gel column chromatography (hexane/EtOAc = 5:1) to yield benzyl ether 6-OMe as 828 an amorphous solid (64.3 mg, 0.16 mmol, 53% yield).

¹H NMR (CDCl₃, 400 MHz) δ 3.38–3.56 (m, 9H), 3.75 (s, 1H), 4.30 (d, J = 5.6 Hz, 1H), 4.57 (d, J = 11.2 Hz, 1H), 4.75–4.80 (m, 4H), 7.22–7.23 (m, 10H).

¹³C NMR (CDCl₃, 100 MHz) δ 57.8, 60.4, 74.6, 75.4, 81.8, 82.2, 83.6, 104.5, 127.4,
127.7, 128.2, 138.4, 138.5, 174.2.

833

834 (2S,3S,4R,5R,6R)-4,5-Dihydroxy-3,6-dimethoxytetrahydro-2*H*-pyran-2-carboxylic
835 acid



836

A 20 mL Schlenk tube was charged with **6-OMe** (40.2 mg, 0.10 mmol) in MeOH/EtOAc = 1:1 solution (5 mL) under N₂, then added pre-activated (water-removed) Pd(OH)₂/C 20.0 mg, H₂ gas was later charged and replace the N₂ by air pump vacuum/H₂ exchange, the reaction mixture was stirred at room temperature for overnight, the completion of the reaction was monitored by LC-MS. The mixture was subjected to filtration by passing through a packed Celite® cake,

- 843 to remove Pd(OH)₂/C, the residue was obtained after removing the solvent *in vacuo*,
- followed by reverse phase TLC (MeOH), afford the corresponding product **G11** as
- an off-white solid (22.2 mg, 0.10 mmol, quant.).
- ¹H NMR (CD₃OD, 400 MHz) δ 3.13–3.21 (m, 1H), 3.24–3.25 (m, 2H), 3.40 (t, J = 9.2
- 847 Hz, 1H), 3.44 (s, 3H), 3.45 (s, 3H), 3.69 (d, J = 10.0 Hz, 1H), 4.14 (d, J = 8.0 Hz,
- 848 1H).
- ¹³C NMR (CD₃OD 100 MHz) δ 57.6, 60.8, 74.8, 75.8, 77.3, 83.0, 105.5 (carbon signal of C=O is missing for its low concentration in CD₃OD).
- ¹³C NMR (D₂O 100 MHz) δ 57.6 (the standard peak based on that observed in CD₃OD), 60.3, 72.9, 73.9, 75.1, 81.7, 103.4, 171.4.
- 853 HRMS (ESI) *m*/*z* calcd for C₈H₁₄O₇Na [M+Na]⁺: 245.0632 found: 245.0628. 854
- 855

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Biology

Table S2. AMOR activity results for synthetic AMOR and derivatives.

Compound	Concentration (μΜ)	Responsive Pollen Tubes (%)	S.D.	S.E.	P value (Scheffé test) Comparison with 4-Me-GIcA-β(1,6)-Gal (G1)
4-Me-GlcA-β(1,6)-Gal G1	1.35	80.0	10.0	5.780	
4-Me-GlcA-α(1,6)-Gal G2	1.35	33.3	5.68	3.286	0.0054475 (**P<0.01)
4-Me-GlcA-β(1,6)-Glc G3	1.35	56.7	15.2	8.829	0.4897411 (insignificant)
4-Me-GlcA-β(1,3)-Gal G4	1.35	40.0	10.0	5.78	0.0241134 (*P<0.05)
4-Me-GlcA-β(1,2)-Xyl G5	1.35	26.7	11.5	6.674	0.0011994 (**P<0.01)
GlcA-β(1,6)-Gal G6	1.35	3.3	5.8	3.337	7.87E-06 (**P<0.01)
GlcA-β(1,2)-Xyl G7	1.35	6.7	5.8	3.337	6.28E-05 (**P<0.01)
4-Me-Glc-β(1,6)-Gal G8	1.35	13.33	11.5	6.674	4.05E-06 (**P<0.01)
4,6-Me-GlcA-β(1,6)-G al G9	1.35	0	0	0	0 (**P<0.01)
4-Me-GlcA-β(1,6)-Prya n G10	1.35	83.3	11.5	6.674	0.9999993 (insignificant)
4-Me-GlcA-O-Me G11	1.35	23.3	5.8	3.337	0.0005656 (**P<0.01)