

1 **Supporting Information**

3 **Natriuretic peptide type C induces sperm attraction for fertilization in mouse**

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19 **Table S1. Compare of sperm motility in NPR2 heterozygote and mutant mice**

Treatment	NPR2 heterozygote	NPR2 mutant
Progressive (%)	21.3 ± 1.3	19.4 ± 0.7
VAP (μm/s)	113.1 ± 2.4	116.2 ± 4.2
VSL (μm/s)	74.2 ± 2.1	76.6 ± 4.9
VCL (μm/s)	214 ± 6.6	223.3 ± 5.5
ALH (μm)	8.9 ± 0.4	8.5 ± 0.2
BCF (HZ)	25.2 ± 0.9	23.9 ± 0.6
STR (%)	61.8 ± 1.4	63.9 ± 0.6
LIN (%)	35.2 ± 1.2	36.4 ± 0.5

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21 VAP, average path velocity; VSL, straight line velocity; VCL, curvilinear velocity;
 22 ALH, amplitude of lateral head displacement; BCF, beat cross frequency; STR,
 23 percentage of straightness; LIN, percentage of linearity; progressive motility (% of
 24 motile spermatozoa with VAP ≥ 50 μm/s and STR ≥ 80%). Bars indicate the mean ±
 25 SEM of three experiments; $n = 500$ for each sample.

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27 **Table S2. Sequence of PCR primers used for qRT-PCR**

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Primer	Forward	Reverse
<i>Nppa</i>	CCAGGCCATATTGGAGCAAAT	TTCTCCTCCAGGTGGTCTAGCA
<i>Nppb</i>	AGCTGCTTGGGCACAAGAT	CAGGCAGAGTCAGAAACTGGAGTCT

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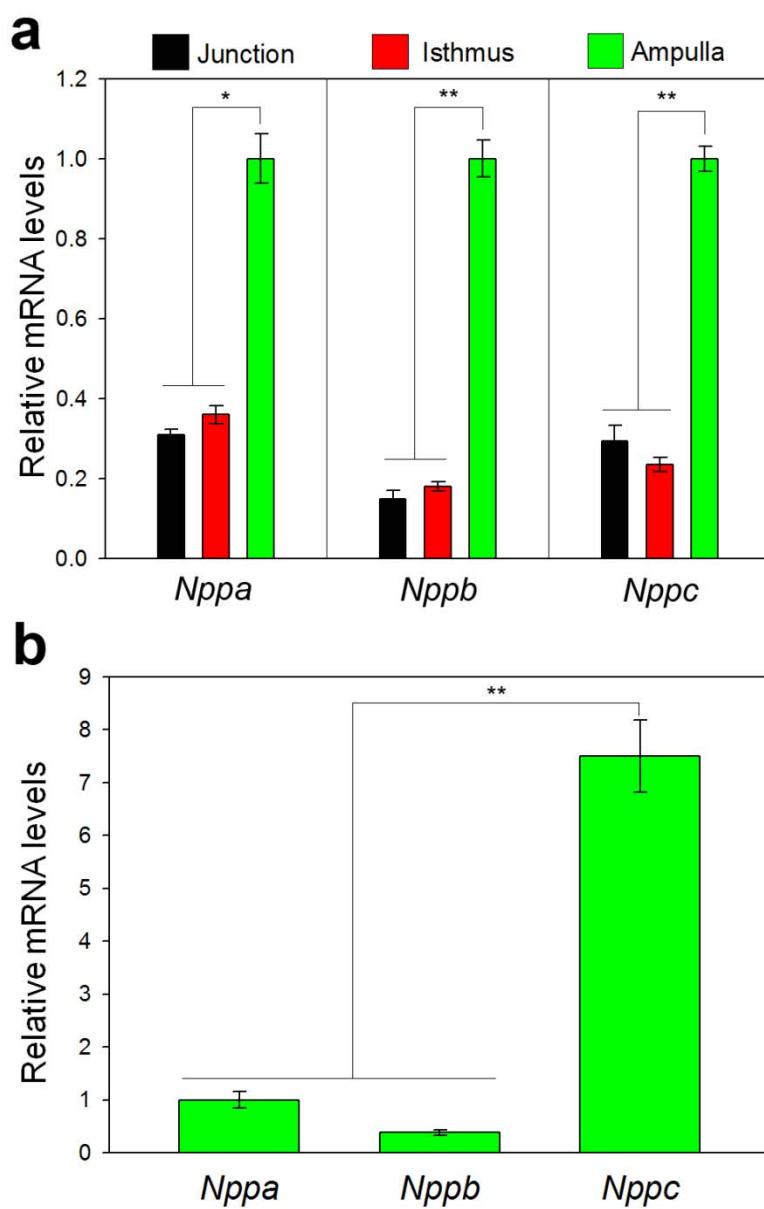
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 32 **Fig. S1. Comparison of amino sequences of mouse natriuretic peptides (NPs) and**
 33 **the confirmed chemoattractants.** Amino acid sequence analysis revealed that mouse
 34 natriuretic peptide type A (NPPA), type B (NPPB) and type C (NPPC) exhibited
 35 similar characters to the chemoattractant peptides resact and asterosap in marine
 36 invertebrates: there are glycine-rich peptides (red), circular by the formation of an
 37 intramolecular disulfide bond between cysteines (green), and binding to a
 38 receptor-type guanylyl cyclase (GC) to stimulate the synthesis of cGMP.

39	Species	Ligand	Amino acid sequence	Receptor	Signaling	Reference
40	<i>Arbacia punctulata</i>	Resact	A circular peptide chain with several red circles (glycine) and two green double-headed arrows (cysteines). It has a disulfide bond between the two cysteines.	A receptor-type guanylyl cyclase	cGMP	ref. 1
41	<i>Asterias amurensis</i>	Astersap	A circular peptide chain with several red circles and two green double-headed arrows. It has a disulfide bond between the two cysteines.	A single 130-kDa membrane protein	cGMP	ref. 2
42	Mouse	NPPA	A circular peptide chain with several red circles and two green double-headed arrows. It has a disulfide bond between the two cysteines.	Natriuretic peptide receptor 1	cGMP	ref. 3, 4
43	Mouse	NPPB	A circular peptide chain with several red circles and two green double-headed arrows. It has a disulfide bond between the two cysteines.	Natriuretic peptide receptor 1	cGMP	ref. 3, 4
44	Mouse	NPPC	A circular peptide chain with several red circles and two green double-headed arrows. It has a disulfide bond between the two cysteines.	Natriuretic peptide receptor 2	cGMP	ref. 3, 4
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53 References

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 55 sea urchin sperm. *Nat Cell Biol* **5**, 109-117 (2003).
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- 59 3 Potter, L. R., Abbey-Hosch, S. & Dickey, D. M. Natriuretic peptides, their
 60 receptors, and cyclic guanosine monophosphate-dependent signaling functions.
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 62 oocyte meiotic arrest via natriuretic peptide receptor 2 (NPR2) in cumulus cells.
Mol Reprod Dev **81**, 462-469 (2014).

65 **Fig. S2.** *Nppc* mRNA is expressed predominantly in the ampulla of estrous mice.
66 (a) Comparison of natriuretic peptides mRNA levels in the uterotubal junction
67 (Junction), isthmus and ampulla from the oviducts of estrous mice. The mean value in
68 ampulla group was set as 1. Bars indicate mean \pm SEM of four experiments. * $P <$
69 0.05; ** $P < 0.01$. (b) Comparison of *Nppa*, *Nppb* and *Nppc* mRNA levels in the
70 ampulla collected from estrous mice. Bars indicate mean \pm SEM of four experiments.
71 ** $P < 0.01$.

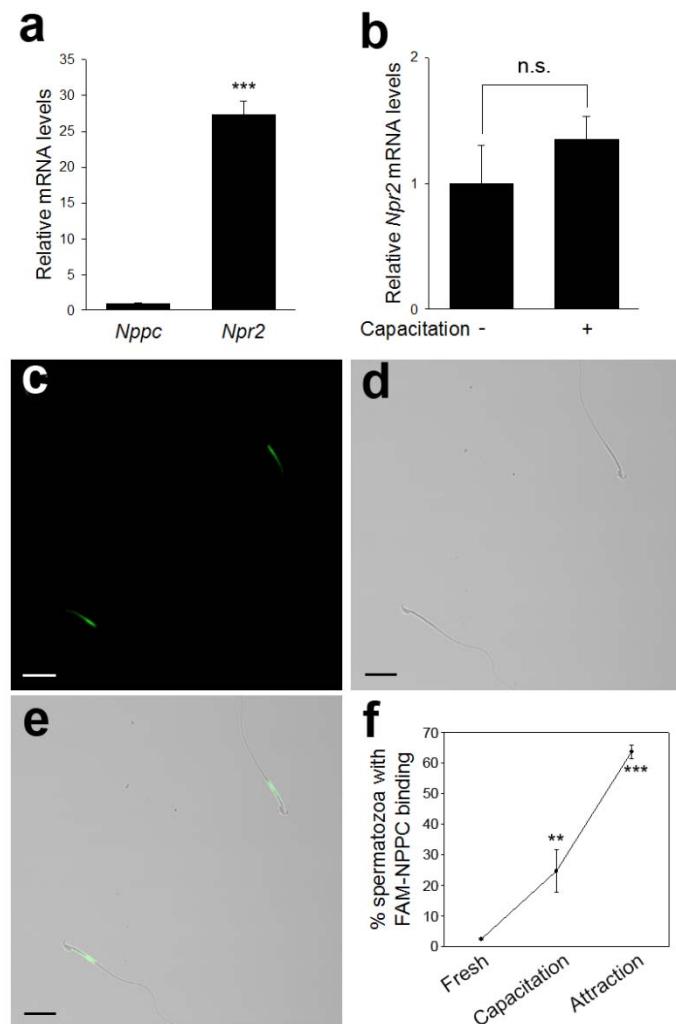


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94 **Fig. S3. NPR2 is expressed in spermatozoon.** (a) Comparison of *Nppc* and *Npr2*
 95 mRNA levels in fresh spermatozoa. Bars indicate mean \pm SEM of three experiments.
 96 *** $P < 0.001$. (b) Comparison of *Npr2* mRNA levels in fresh and capacitated
 97 spermatozoa. No obvious change was indicated. (c-e) Capacitated spermatozoa were
 98 incubated with 100 nM FAM-NPPC for 30 min, and the fluorescent ligand binding
 99 was assessed by a confocal laser-scanning microscope. Representative images showed
 100 fluorescence (c), bright (d) and merge (e) fields. Scale bars, 20 μ m. (f) Comparison of
 101 FAM-NPPC binding in fresh, capacitation and attraction spermatozoa. Bars indicate
 102 the mean \pm SEM of three experiments. $n = 500$ for each group. ** $P < 0.01$; *** $P <$
 103 0.001.

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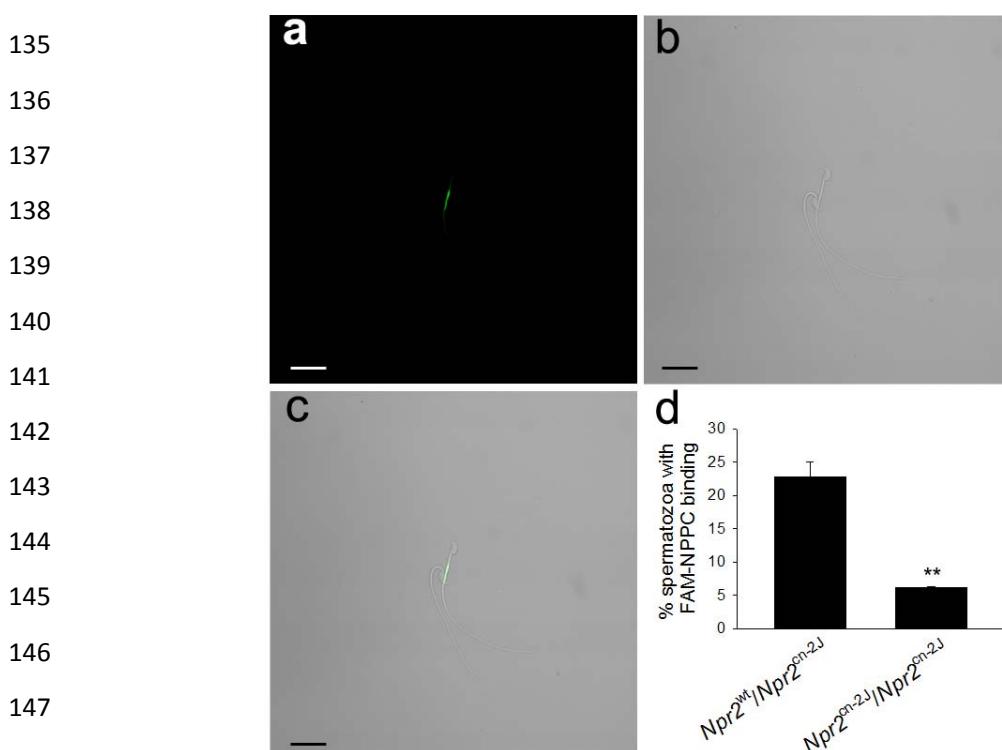
124 **Fig. S4. Failure in FAM-NPPC binding to spermatozoa from *Npr2* mutant mice.**

125 Capacitated spermatozoa from *Npr2* heterozygote and mutant mice were incubated
 126 with 100 nM FAM-NPPC for 30 min, and the fluorescent ligand binding was assessed
 127 by a confocal laser-scanning microscope. (a-c) Representative images showed
 128 fluorescence (a), bright (b) and merge (c) fields. Scale bars, 20 μ m. (d) Comparison
 129 of FAM-NPPC binding in capacitated spermatozoa from *Npr2* heterozygote and
 130 mutant mice. Bars indicate the mean \pm SEM of three experiments. $n = 500$ for each
 131 group. ** $P < 0.01$.

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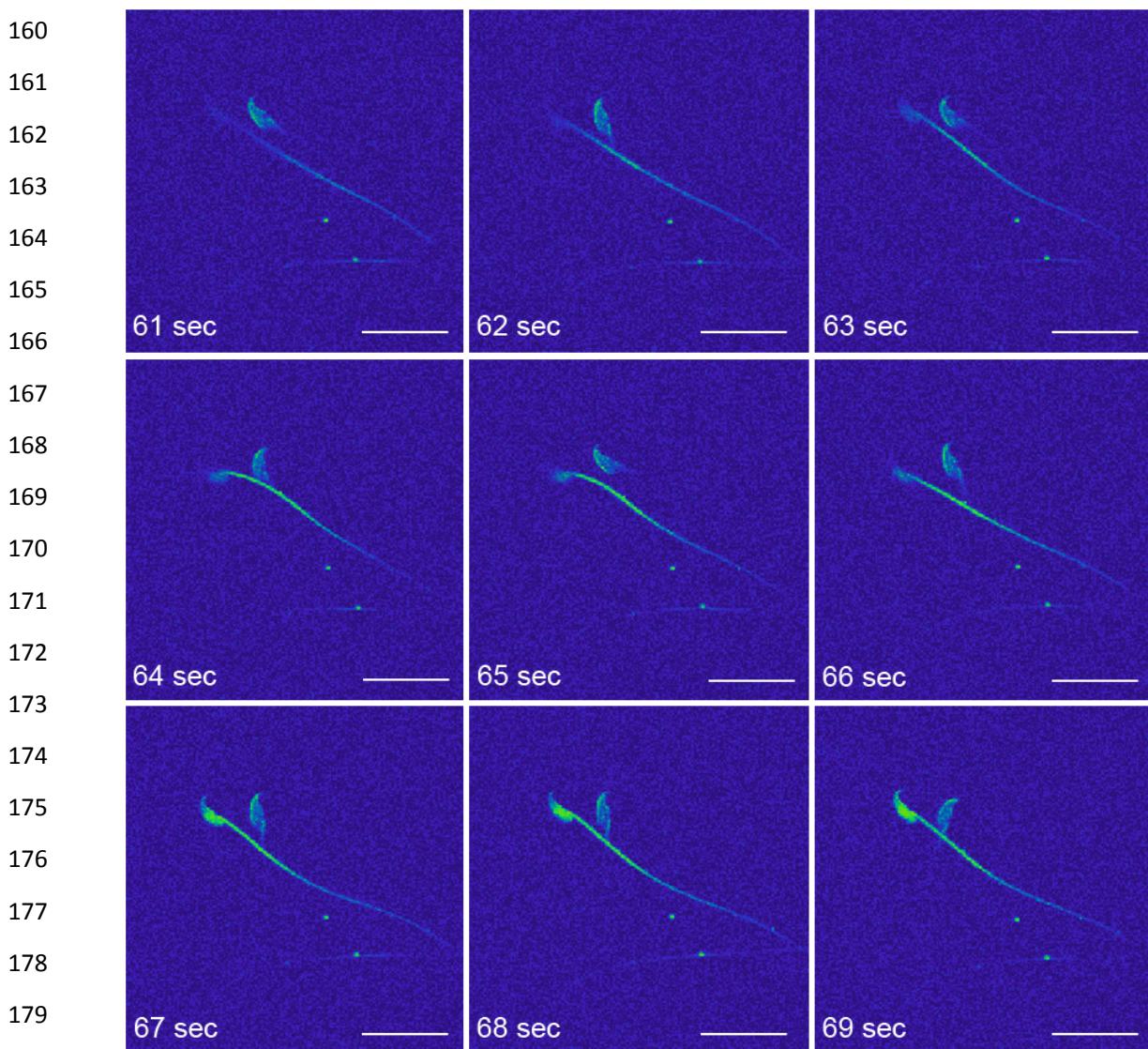
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154 **Fig. S5. Time lag in the Ca^{2+} influx induced by NPPC between flagellar midpiece**
155 **and head of mouse spermatozoon.** Time measurement analysis showing the
156 propagation of elevated Ca^{2+} from the midpiece of flagellum to the head after
157 application of 0.1 nM NPPC. The zero time was defined as the time of NPPC addition.
158 Representative images are presented in a pseudocolor format. Scale bars, 20 μm .

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185 **Fig. S6. *l*-cis-Diltiazem blocked NPPC-induced sperm accumulation.** (a) Sperm
186 accumulation in the capillaries in the groups of control, NPPC (0.1 nM),
187 *l*-cis-Diltiazem (*l*-cis-D, 50 μ M) and NPPC + *l*-cis-Diltiazem. Bars indicate the mean
188 \pm SEM of three experiments. (b) Sperm accumulation in the oviductal ampullae. Bars
189 indicate the mean \pm SEM of four experiments. *** $P < 0.001$.

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