

A.

	Weight (gram)		Count (10 ⁶)	
	Thymus	Spleen	Thymus	Spleen
WT (n=3)	0.08±0.02	0.08±0.03	67.3±12	44.6±18
PAXX ^{-/-} (n=4)	0.07±0.03	0.09±0.02	67.7±18	37.1±10

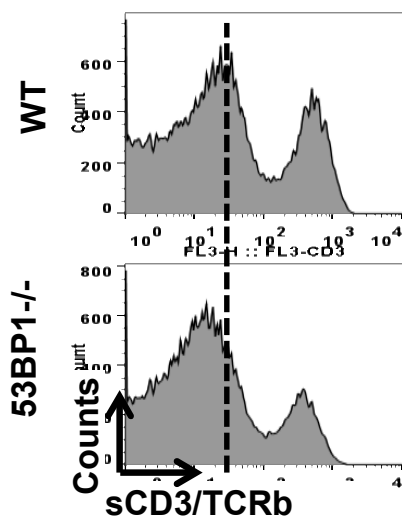
B.

x10 ⁶	Thymus				
	Total	DN	DP	CD4 SP	CD8 SP
WT (n=3)	67.3±12	1.8±1.2	54.2±7.0	8.7±2.2	2.5±1.2
PAXX ^{-/-} (n=4)	67.7±18	2.1±1.0	55.3±6.5	7.7±2.3	2.6±1.5

C.

	Bone Marrow			
	Pro-B%	Pre-B%	Pre/Pro Ratio	IgM+ B cells%
WT (n=3)	2.24±1.08	13.2±2.53	5.05±1.05	11.1±2.8
PAXX ^{-/-} (n=4)	2.27±1.05	14.5±3.28	6.38±1.58	10.8±3.7

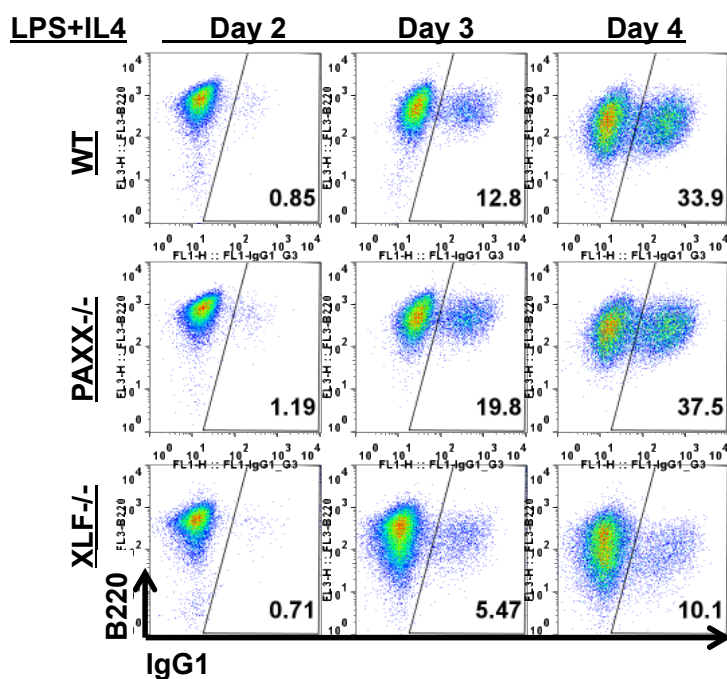
D.



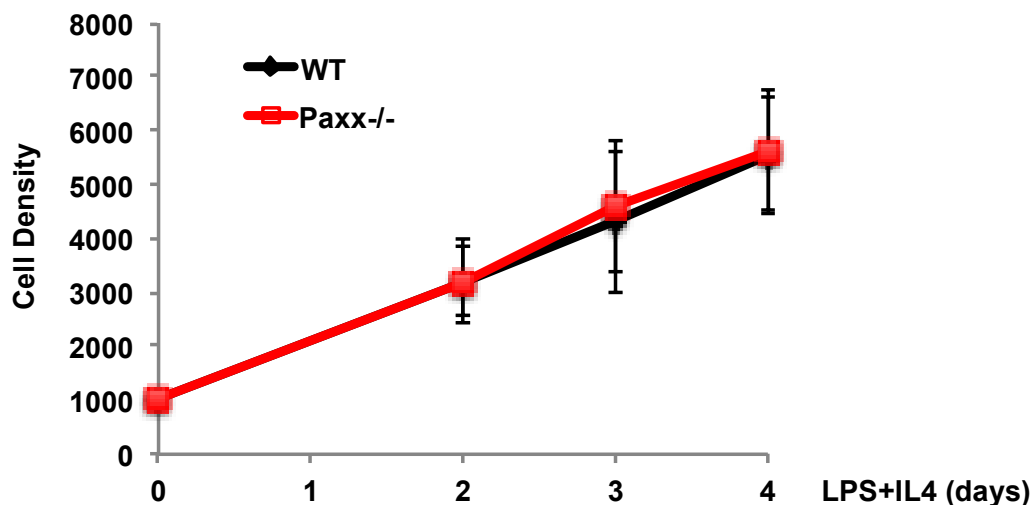
Supplementary Fig. 1 *Paxx*^{-/-} mice have normal lymphocyte development.

(A) The weight and total cellularity of the thymus and spleen from *Paxx*^{+/+} and *Paxx*^{-/-} mice. At least three mice at ~7 weeks of age for each genotype were analyzed and were used to calculate the average and standard deviations. (B) Total thymocyte counts and calculated CD4⁻CD8⁻ double negative (DN), CD4⁺CD8⁺ double positive (DP) thymocyte and single positive (SP) T cell numbers in the thymus. Each value represents the average ± standard deviation from at least three mice of ~7 weeks of age for each genotype. (C) The percentage of pro-B (B220⁺IgM⁻CD43⁺), pre-B (B220⁺IgM⁻CD43⁻) and mature B (B220⁺IgM⁺CD43⁻) cells in total bone marrow from different genotypes. Pre/Pro Ratio was calculated from each mouse. Each value represents the average ± standard deviation from at least three mice of ~7 weeks of age for each genotype. (D) Surface CD3/TCRβ expression in thymocytes from representative *53BP1*^{+/+} (WT) and *53BP1*^{-/-} mice. The vertical line marks the median of the surface CD3/TCRβ levels in WT thymocytes.

A.

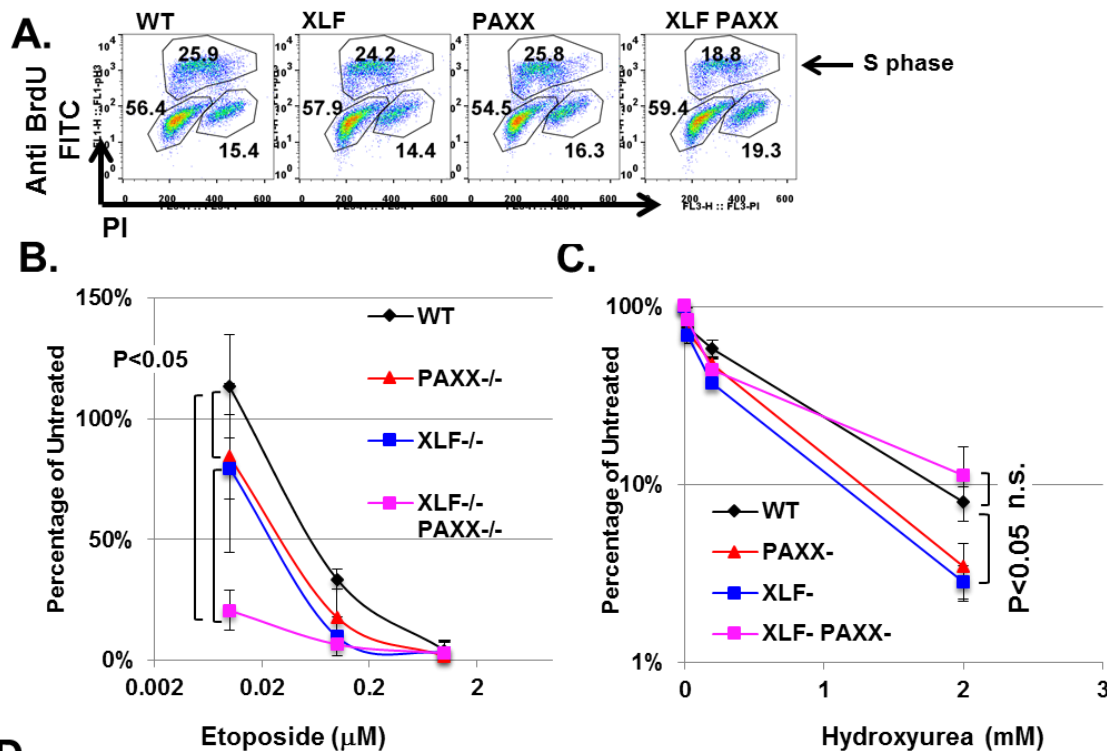


B.



Supplementary Fig. 2 Class switching recombination is normal in *Paxx*^{-/-} mice

(A) Representative flow cytometry analyses of LPS/IL4 stimulated CD43⁻ splenocytes derived from *Paxx*^{+/+} (WT), *Paxx*^{-/-} and *Xlf*^{-/-} mice at 2, 3 and 4 days after stimulation. (B) Proliferation of stimulated *Paxx*^{+/+} (WT) or *Paxx*^{-/-} B cells. The Y-axis indicates the cell density per μ l. Cell number was counted using standard hemocytometer.



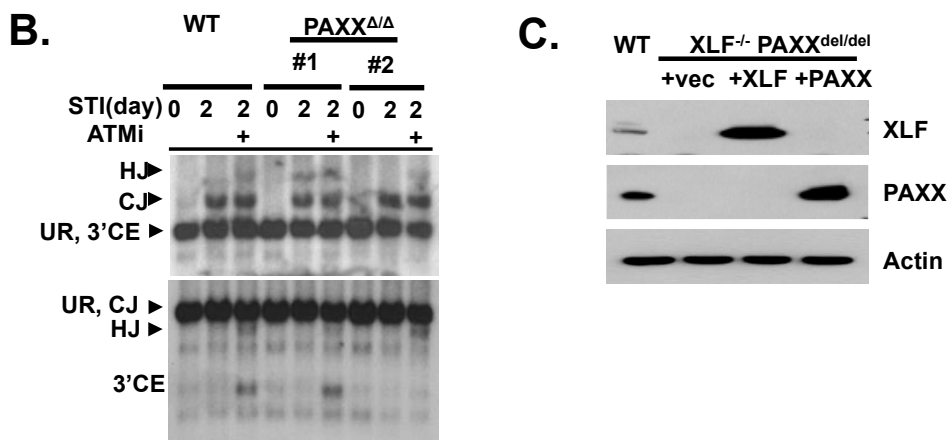
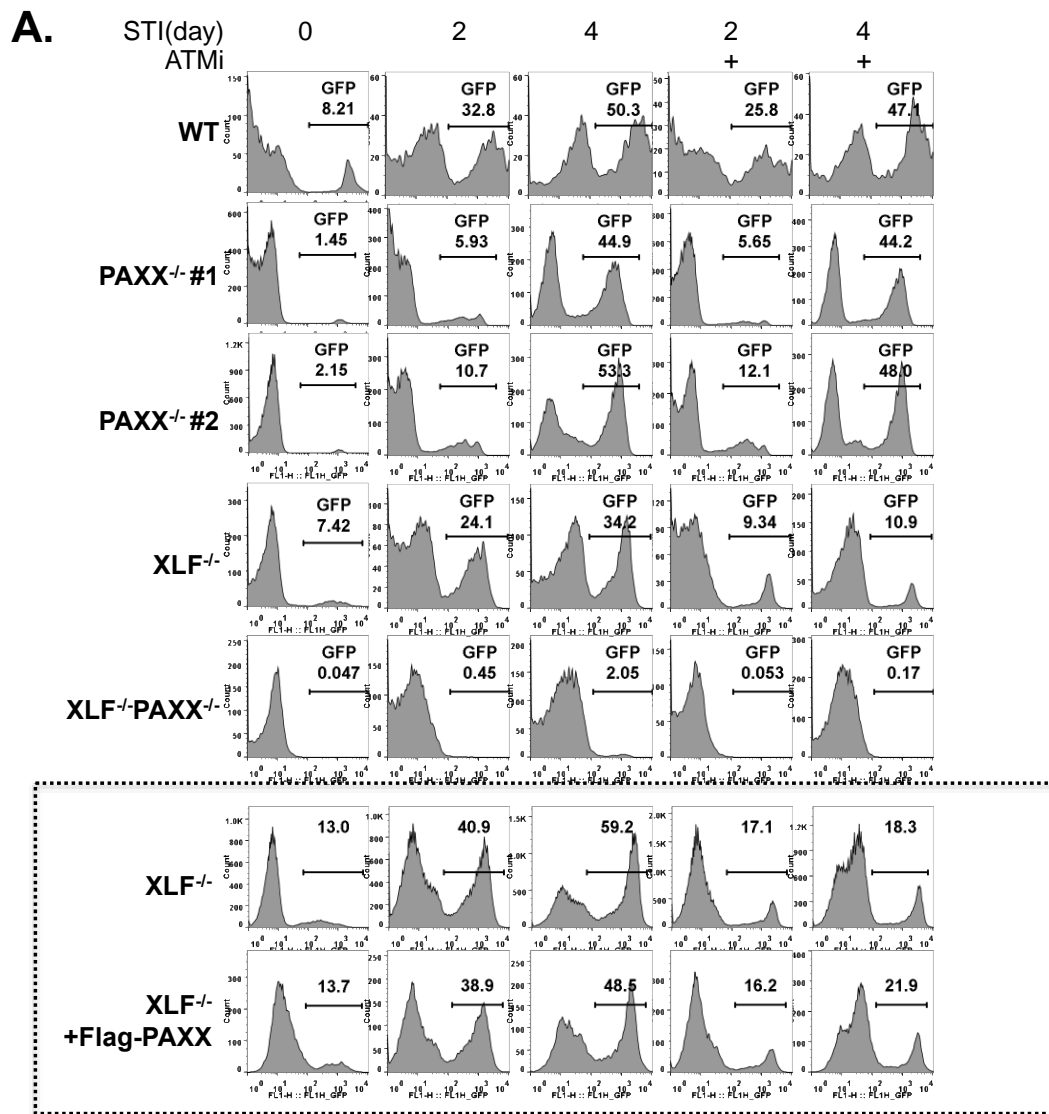
D.

	Total # MP	# abn. MP	% abn. MP	Total # abn.	Brks/MP	Csm. Brk	Csm. Brk/MP	Ctd. Brk	Ctd. Brk/MP
WT	144	3	2.04%±0.13%	3	0.020	2	0.015	1	0.005
XLF ^{-/-}	187	18	9.63%±0.01%	18	0.096	15	0.080	3	0.016
PAXX ^{-/-}	144	5	3.44%±0.33%	5	0.035	4	0.028	1	0.007
XLF ^{-/-} PAXX ^{-/-}	206	41	18.89%±2.60%	47	0.228	42	0.196	5	0.025

E.

	Total # MP	# abn. MP	% abn. MP	Total # abn.	Brks/M P	Csm. Brk	Csm. Brk/MP	Ctd. Brk	Ctd. Brk/MP
WT	151	8	5.28%±0.56%	8	0.053	7	0.046	1	0.007
XLF ^{-/-}	150	15	10.00%±1.15%	16	0.107	13	0.087	3	0.020
XLF ^{-/-} PAXX ^{-/-}	169	22	13.02%±2.01%	24	0.142	18	0.137	2	0.005

Supplementary Fig. 3 Increased genomic instability in *Xlf^{-/-}Paxx^{-/-}* MEF cells and v-abl transformed pre-B cells (A) Cell cycle distribution of the indicated P1 primary MEF cells. P1 MEFs were incubated with BrdU for 30 min before collected for PI and FITC anti-BrdU staining. (B) Etoposide or (C) Hydroxyurea (HU) sensitivity assay in P1 primary MEF cells. Student's t test was used to compare the percentage of survival between different groups. n.s. not significant. The percentage (D) and (E) Summary of cytogenetic abnormalities in WT, *Xlf^{-/-}*, *Paxx^{-/-}* and *Xlf^{-/-}Paxx^{-/-}* primary MEF cells (P1) and v-abl transformed pre-B cells. Abbreviations: MP- metaphase, Abn- Abnormal, Csm- Chromosomal, Ctd- Chromatid, Brk- Break. The data summarizes the results from two or more independent experiments using at least two independently derived cell lines of each genotype. All images were obtained and processed with Metafer (Metasystem Inc).



Supplementary Fig. 4 *Xlf* and *Paxx* double deficiency abrogated chromosomal V(D)J recombination
 (A) Flow cytometry analyses for GFP+ cells after STI571 treatment (3 μ M) (with or without ATM inhibitor, KU55933, 15 μ M, added 1hr before STI571) of WT, two independent *Paxx* ^{Δ/Δ} clones, *Xlf*^{-/-} and *Xlf*^{-/-} *Paxx* ^{Δ/Δ} cells. In parallel (lower two rows), another *Xlf*^{-/-} cells with or without ectopic overexpression of PAXX were also treated with STI571 in the presence or the absence of ATM kinase inhibitor. Gated hCD4+ cells were plotted in the histogram. (B) Southern blot analyses of the V(D)J recombination substrates, intermediates and products of representative *Paxx*^{+/+} (WT) and *Paxx* ^{Δ/Δ} cells with or without ATM kinase inhibitor (KU55933, 15 μ M, added 1hr before STI571 at 3 μ M). See Fig.6A for the location for probes and digestion sites. (C) Western blot for XLF, Flag-PAXX and β -actin in WT and *Xlf*^{-/-} *Paxx* ^{Δ/Δ} cells with or without ectopic expression of XLF or PAXX.

Fig 5B

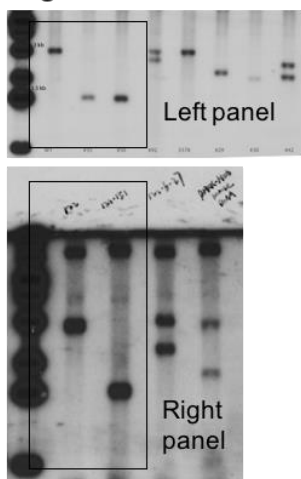


Fig 5C

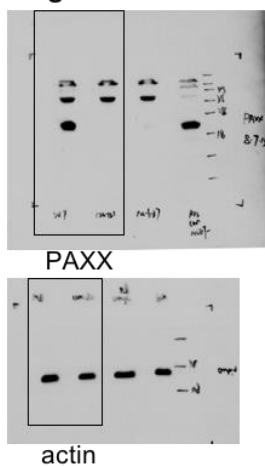


Fig 6B

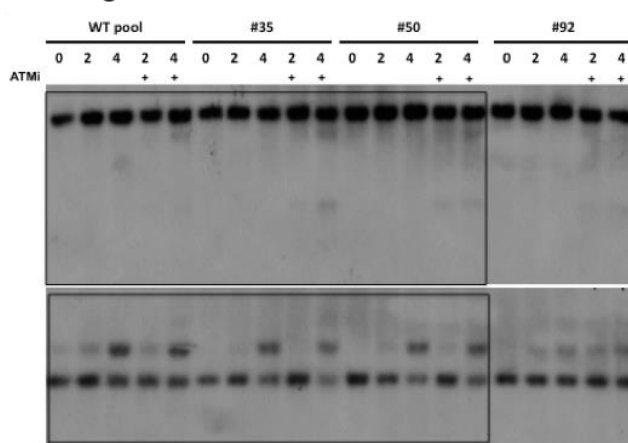


Fig 6C hCD4 probe

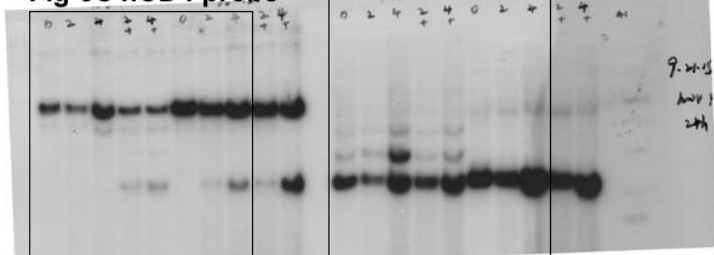


Fig 6C GFP probe

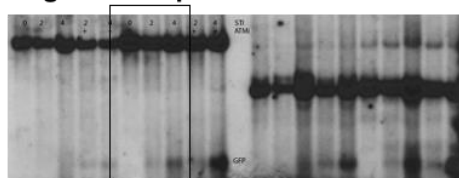


Fig 6D

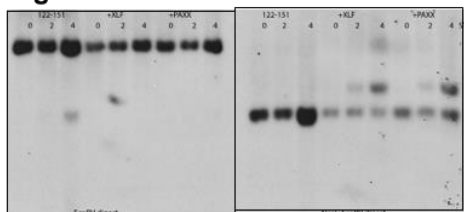


Fig 6E

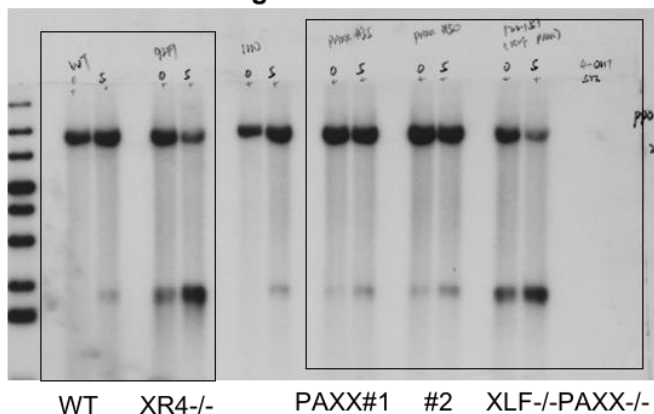


Fig 7A

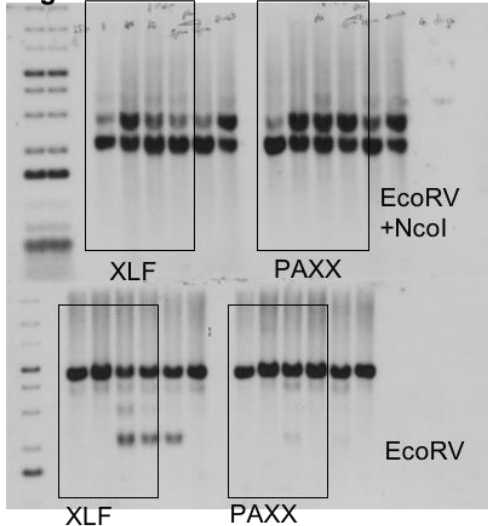
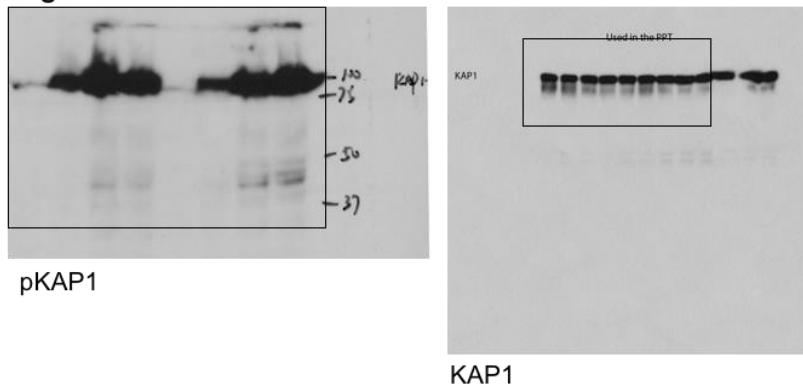


Fig 7B



Supplementary Fig. 6 Original images of Southern blots and Western blots.

Fig 5B, 6B, 6C, 6D, 6E and 7A are original images of Southern blots hybridized with probes mentioned in paper. Fig 5C and 7B are original images of Western blots.

Supplementary Table 1 Endogenous V(D)J recombination junction at Vβ14-Dβ-Jβ1.1/2 from WT or Paxx^{-/-} thymocytes. P:

Palindromic elements, N: non-template nucleotide addition. The total number of nucleotide deletion from V-D or Paxx^{-/-} thymocytes were calculated and listed on the right.

Vβ14	P	N	P	Db1	P	N	P	Jb1.1 or Jb 1.2	VD del (nt)	DJ del (nt)
Paxx+/+								Jb1.1 (NTEVFFGKGR)		
TGGCTTACCTCTGTGCCCTGGAGTCT				GGGACAGGGGGC				CAACACAGAAGTCTTCTTTGGTAAAGGAACCAGACT		
TGGCTTACCTCTGTGCCCTGGAGTC	C		C	GGGACAGGG	C			ACACAGAAGTCTTCTTTGGTAAAGGAACCAGACT	1	6
TGGCTTACCTCTGTGCCCTGGGA			C	AC	G	ACGG	TG	CAACACAGAAGTCTTCTTTGGTAAAGGAACCAGACT	7	7
TGGCTTACCTCTGTGCCCTGGAGTCT			C	GGGGG				AAACACAGAAGTCTTCTTTGGTAAAGGAACCAGACT	6	2
TGGCTTACCTCTGTGCCCTGGAGTCT	A	ATG		GGGGG		A		ACAGAAGTCTTCTTTGGTAAAGGAACCAGACT	6	5
TGGCTTACCTCTGTGCCCTGGAGTCT				AGG	C	AG		AAACACAGAAGTCTTCTTTGGTAAAGGAACCAGACT	5	5
TGGCTTACCTCTGTGCCCTGGAGTCT				AC	G			ACAGAAGTCTTCTTTGGTAAAGGAACCAGACT	10	12
TGGCTTACCTCTGTGCCCTGGAGTC		A		GACAGGGG	C	GG		ACACAGAAGTCTTCTTTGGTAAAGGAACCAGACT	3	5
TGGCTTACCTCTGTGCCCTGGAGTCT				ACAGGGG				ACACAGAAGTCTTCTTTGGTAAAGGAACCAGACT	3	5
TGGCTTACCTCTGTGCCCTGGAGTCT		GA		GGACAGGGGG		G		ACAGAAGTCTTCTTTGGTAAAGGAACCAGACT	1	6
								Jb1.2 (NSDYTFGSGRLLV)		
TGGCTTACCTCTGTGCCCTGGAGTCT				GGGACAGGGGGC				CAAACTCCGACTACACCTTCGGCTCAGGGACCAGG		
TGGCTTACCTCTGTGCCCTGGAGTC		AGGG		AGGGGG		G		AACTCCGACTACACCTTCGGCTCAGGGACCAGG	6	3
						TGAGAG				
TGGCTTACCTCTGTGCCCTGGAG		AGACA		GGG		G		ACTCCGACTACACCTTCGGCTCAGGGACCAGG	9	6
TGGCTTACCTCTGTGCCCT		C		ACAGGGGG		TCTG		ACTCCGACTACACCTTCGGCTCAGGGACCAGG	10	3
TGGCTTACCTCTGTGCCCT	A	CCG	CC	GGGACAGGGGGC	G	GG		TCCGACTACACCTTCGGCTCAGGGACCAGG	7	5
TGGCTTACCTCTGTGCCCTGGAGTCT				AGGGGG		GAAT		CAAACTCCGACTACACCTTCGGCTCAGGGACCAGG	5	1
TGGCTTACCTCTGTGCCCTGG		G	CC	GGG				ACTCCGACTACACCTTCGGCTCAGGGACCAGG	11	5
TGGCTTACCTCTGTGCCCTGGAG		GCGA		GGGACAGGG	C			CCGACTACACCTTCGGCTCAGGGACCAGG	3	9
TGGCTTACCTCTGTGCCCTGGAGTCT		C		ACAGGG				AACTCCGACTACACCTTCGGCTCAGGGACCAGG	3	5
TGGCTTACCTCTGTGCCCTGGAG	C		C	GA		G		ACTCCGACTACACCTTCGGCTCAGGGACCAGG	5	11
TGGCTTACCTCTGTGCCCTGGAGTCT		GC		CA		CC	TG	CAAACTCCGACTACACCTTCGGCTCAGGGACCAGG	4	6
TGGCTTACCTCTGTGCCCTGGAGTCT				AGG		AGA		AAACTCCGACTACACCTTCGGCTCAGGGACCAGG	5	5
TGGCTTACCTCTGTGCCCTGGAGTC			C	GGACAG			TG	CAAACTCCGACTACACCTTCGGCTCAGGGACCAGG	2	5
TGGCTTACCTCTGTGCCCTGGAGTCT	A	AA		CAGGGG		AGAAGG		AACTCCGACTACACCTTCGGCTCAGGGACCAGG	4	4
								Average	5.27	5.50
								Stdev	2.85	2.56

Supplementary Table 2 Primers used in the study.

Primer Name	Seq 5'-3'	Note
c9orf142_DNA_H3F16758	AGAAGCTTAGAATGCCCAAGTCAGTACAC	5' ARM
c9orf142_DNA_H3R20016	CAAAGCTTAAAGCGGCAGCGACAA	5'ARM
C90rf142_DNA_F21421	CTAGGCCAGACTTGAGTTGTGA	3'ARM
C90rf142_DNA_R24826	GAAACGCAGGCCGAGATA	3'ARM
C9orf142_F24844	ACAGCAGCAGCCATCTAGG	3' probe
C9orf142_R25126	TCCAGGTTCCAAGGATATCAG	3' probe
mPAXX probe F	CACGCAAAGGCTCTAGTCC	mPAXX probe
mPAXX probe R	CGGTCTTCTTTCCTATCTATCCC	mPAXX probe
C90rf142_DNA_F19601	ATTGAAGAGCGGCAGATATGT	Genotype = KO specific (WT=2Kb, KO=0.7kb)
C90rf142_DNA_R21583	ACGCAGAATCAACACAGTAGGT	Genotype =common
C9orf142_F20809	TTGAGTGAGATGCCACGACT	Genotype =WT specific (WT=0.8Kb)