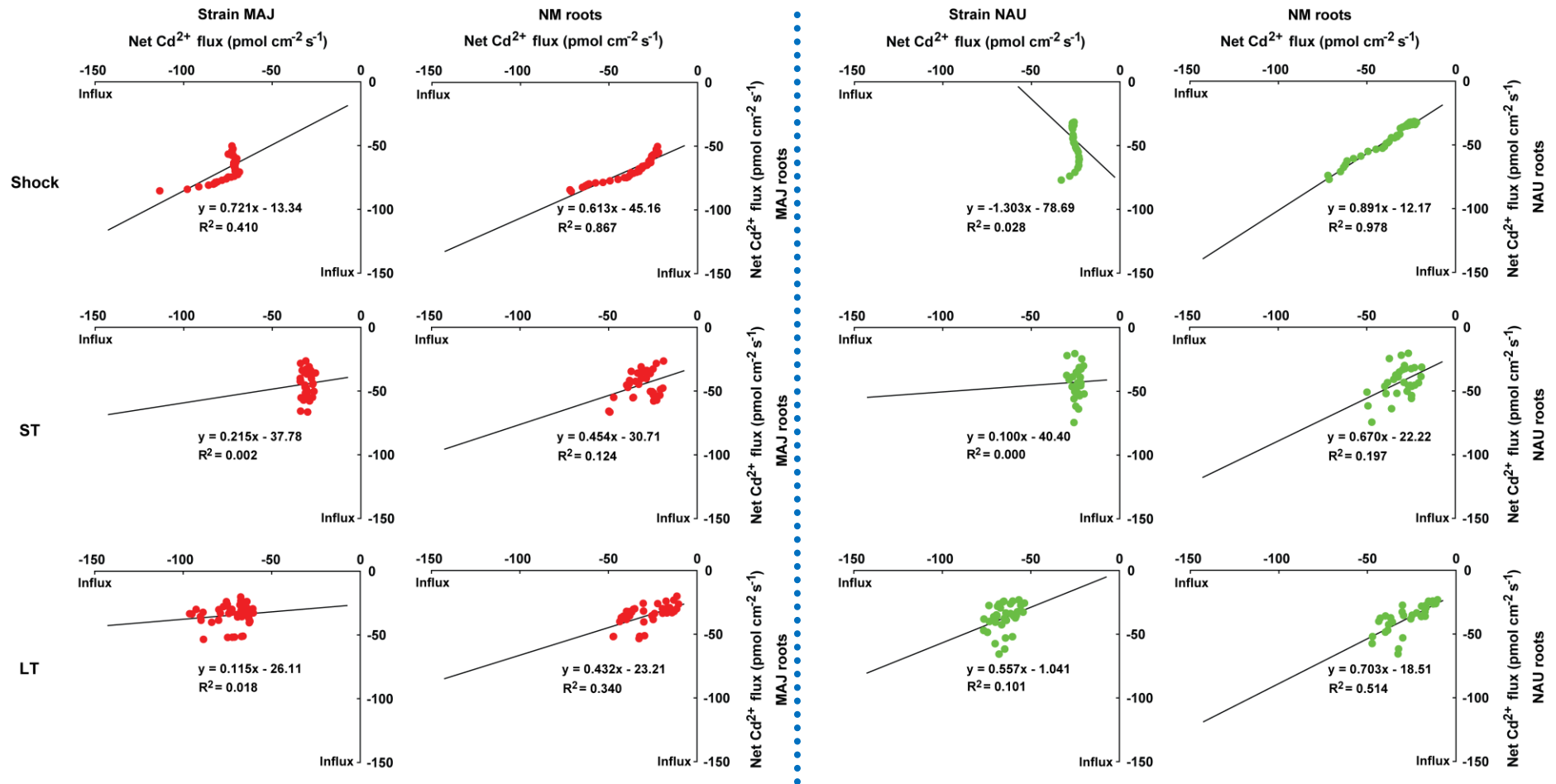


**Supplemental FIGURE S1. Effects of CdCl<sub>2</sub> on steady Cd<sup>2+</sup> and Ca<sup>2+</sup> fluxes in *Populus × canescens* roots inoculated with or without *Paxillus involutus* strains.** *P. × canescens* roots were cultured without or with the *P. involutus* strains MAJ and NAU for 7 days, respectively. Fungal-inoculated and non-inoculated *P. × canescens* plants were subjected to short-term (ST, 24 h) exposure to 50 μM CdCl<sub>2</sub>. Control roots were well fertilized but treated without CdCl<sub>2</sub>. Cd<sup>2+</sup> (A) and Ca<sup>2+</sup> (B) fluxes were measured along root axis, 100–2,300 μm from the apex, at intervals of 200 to 300 μm. Each point is the mean of four to five individual

plants, and bars represent the standard error of the mean. Asterisks denote significant difference at  $P < 0.05$  between treatments. The mean values of  $\text{Cd}^{2+}$  and  $\text{Ca}^{2+}$  flux before (-Cd) and after (+Cd) the addition of  $\text{CdCl}_2$  are shown in the right panel. Columns represent the mean of four to five individual plants and bars represent the standard error of the mean. Different letters, a, b, c and d, indicate significant difference at  $P < 0.05$  between treatments.

MAJ roots-Strain MAJ MAJ roots-NM roots

NAU roots-Strain NAU NAU roots-NM roots

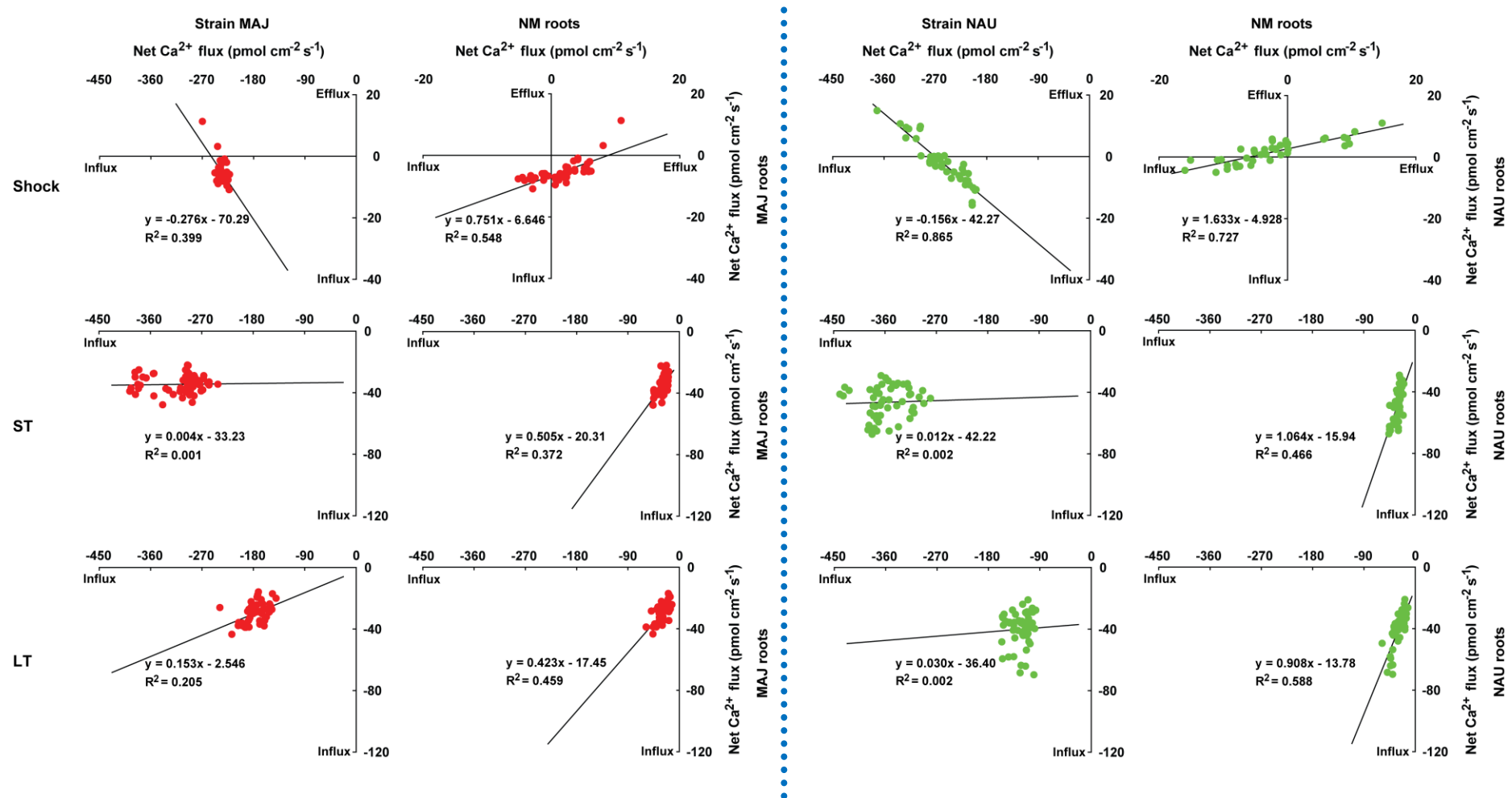


Supplemental FIGURE S2. The correlations of Cd<sup>2+</sup> fluxes between ectomycorrhizal *Populus × canescens* roots, *Paxillus involutus* strains, and non-mycorrhizal (NM) roots under Cd<sup>2+</sup> stress. *P. × canescens* roots (NM, MAJ, and NAU) and *P. involutus* isolates MAJ and NAU were subjected to CdCl<sub>2</sub>

(50  $\mu\text{M}$ ) shock, short-term (ST, 24 h), and long-term (LT, 7 d) treatments, respectively. For shock treatment, transient kinetics of  $\text{Cd}^{2+}$  in roots (100  $\mu\text{m}$  from the root tip) and axenic mycelia were recorded for a period of 40 min after the required amount of 50  $\mu\text{M}$   $\text{CdCl}_2$  was introduced into the measuring solution. The correlations between ectomycorrhizal (EM) roots, NM roots, and *P. involutus* strains were established in terms of measuring time after the flux recordings were started. Under ST and LT treatments, steady fluxes of  $\text{Cd}^{2+}$  in *P. × canescens* roots (NM, MAJ, NAU) were measured along root axis, 100–2,300  $\mu\text{m}$  from the apex, at intervals of 200 to 300  $\mu\text{m}$ .  $\text{Cd}^{2+}$  fluxes of *P. involutus* isolates MAJ and NAU were measured around the surface of pelleted hyphae over a recording period of 30 min. Each point is the mean of four to five individual plants or axenic EM cultures (pelleted hyphae). The correlations between EM roots and NM roots were established in terms of measuring sites; the correlations between EM roots and *P. involutus* strains were established in terms of measuring time after the flux recordings were started.

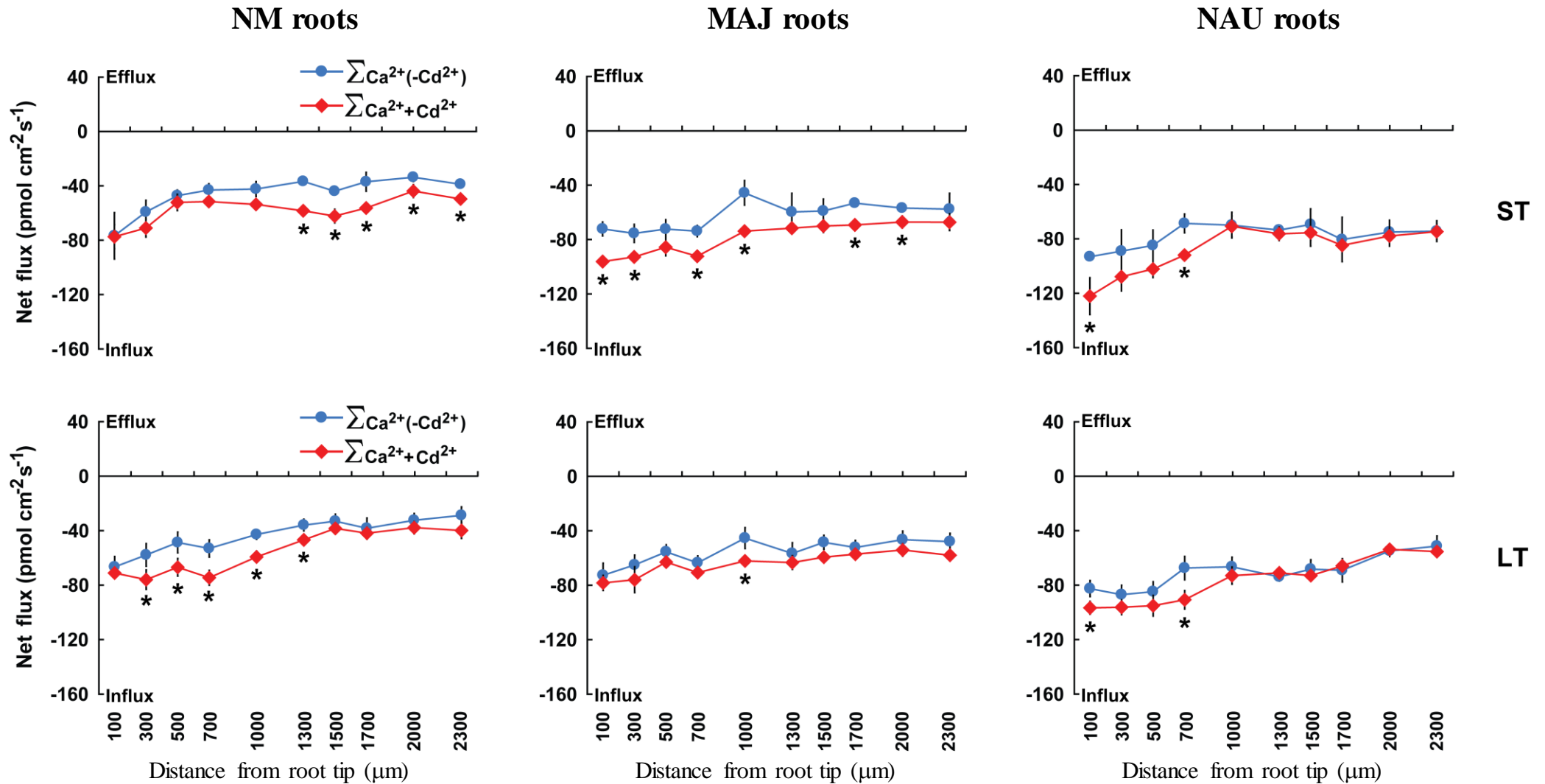
MAJ roots-Strain MAJ MAJ roots-NM roots

NAU roots-Strain NAU NAU roots-NM roots



Supplemental FIGURE S3. The correlations of  $\text{Ca}^{2+}$  fluxes between ectomycorrhizal *Populus \times canescens* roots, *Paxillus involutus* strains, and non-mycorrhizal (NM) roots under  $\text{Cd}^{2+}$  stress. *P. \times canescens* roots (NM, MAJ, and NAU) and *P. involutus* isolates MAJ and NAU were subjected to  $\text{CdCl}_2$

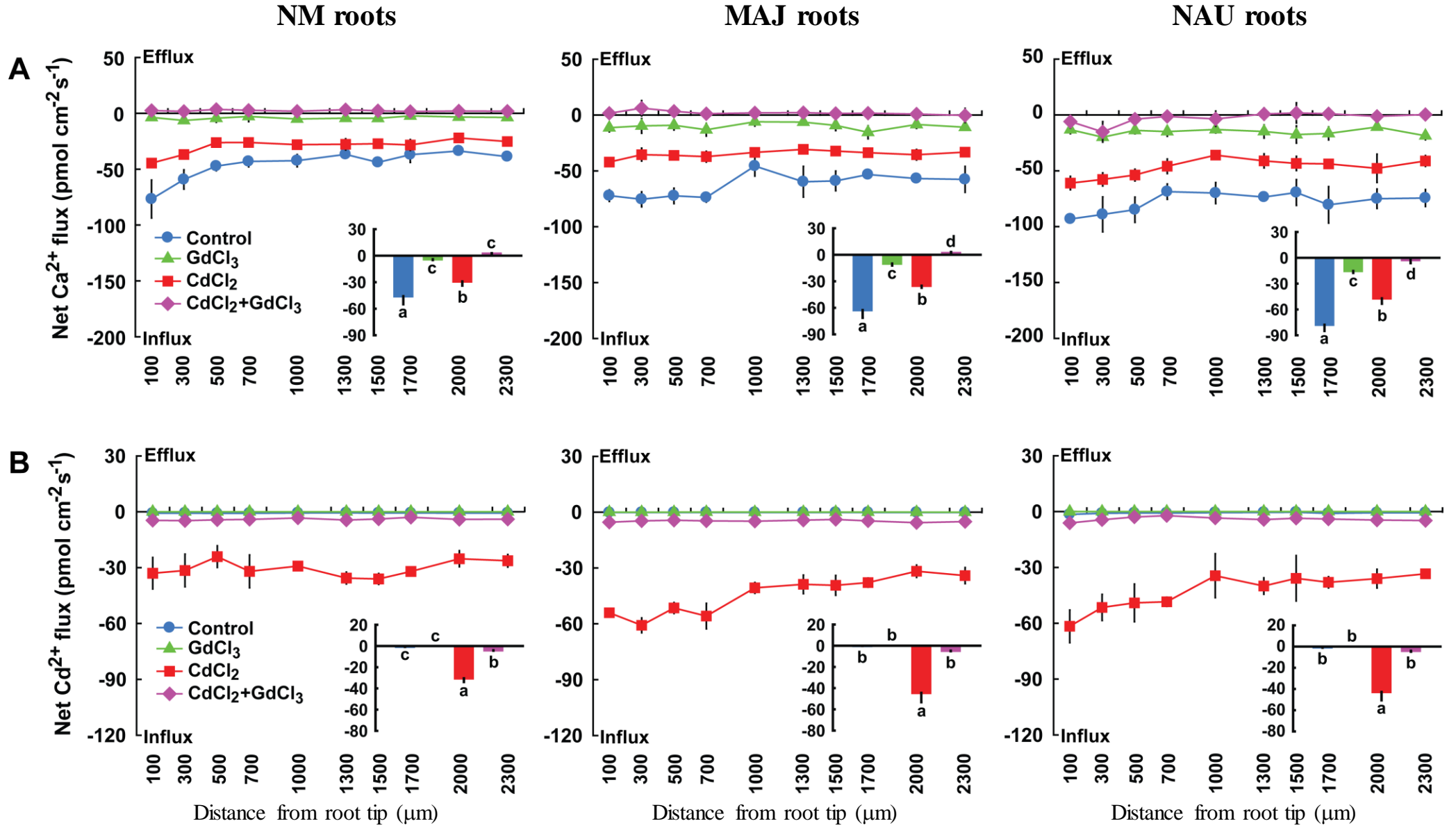
(50  $\mu\text{M}$ ) shock, short-term (ST, 24 h), and long-term (LT, 7 d) treatments, respectively. For shock treatment, transient kinetics of  $\text{Ca}^{2+}$  in roots (100  $\mu\text{m}$  from the root tip) and axenic mycelia were recorded for a period of 40 min after the required amount of 50  $\mu\text{M}$   $\text{CdCl}_2$  was introduced into the measuring solution. The correlations between ectomycorrhizal (EM) roots, NM roots, and *P. involutus* strains were established in terms of measuring time after the flux recordings were started. Under ST and LT treatments, steady fluxes of  $\text{Ca}^{2+}$  in *P. × canescens* roots (NM, MAJ, NAU) were measured along root axis, 100–2,300  $\mu\text{m}$  from the apex, at intervals of 200 to 300  $\mu\text{m}$ .  $\text{Ca}^{2+}$  fluxes of *P. involutus* isolates MAJ and NAU were measured around the surface of pelleted hyphae over a recording period of 30 min. Each point is the mean of four to five individual plants or axenic EM cultures (pelleted hyphae). The correlations between EM roots and NM roots were established in terms of measuring sites; the correlations between EM roots and *P. involutus* strains were established in terms of measuring time after the flux recordings were started.



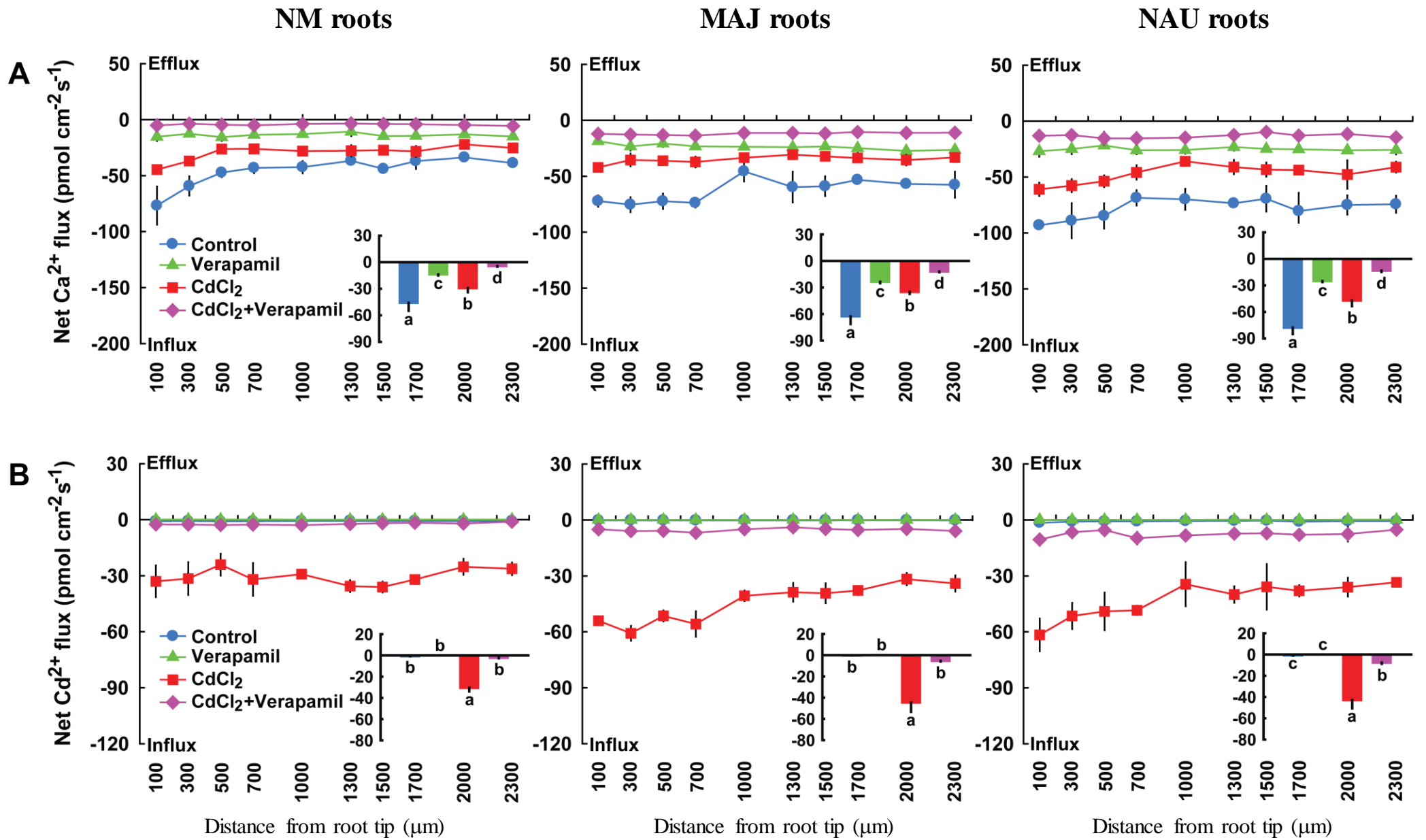
Supplemental FIGURE S4. Total fluxes of  $Ca^{2+}$  and  $Cd^{2+}$  ( $\Sigma Ca^{2+}+Cd^{2+}$ ) in the presence of  $CdCl_2$  ( $50 \mu M$ ,  $+Cd^{2+}$ ) and  $Ca^{2+}$  flux [ $\Sigma Ca^{2+}(-Cd^{2+})$ ] in the absence of  $CdCl_2$  ( $0 \mu M$ ,  $-Cd^{2+}$ ) in roots of ectomycorrhizal (MAJ and NAU) and non-mycorrhizal (NM) *Populus × canescens*. Ectomycorrhizal (MAJ and NAU) and

NM *P. ×canescens* plants were subjected to short-term (ST, 24 h) and long-term (LT, 7 d) exposure to 50  $\mu\text{M}$   $\text{CdCl}_2$ , respectively. Control roots were well fertilized but treated without  $\text{CdCl}_2$ .  $\text{Ca}^{2+}$  and  $\text{Cd}^{2+}$  fluxes were measured along root axis, 100–2,300  $\mu\text{m}$  from the apex, at intervals of 200 to 300  $\mu\text{m}$ . Each point is the mean of four to five individual plants, and bars represent the standard error of the mean. Asterisks denote significant difference at  $P < 0.05$  between treatments.

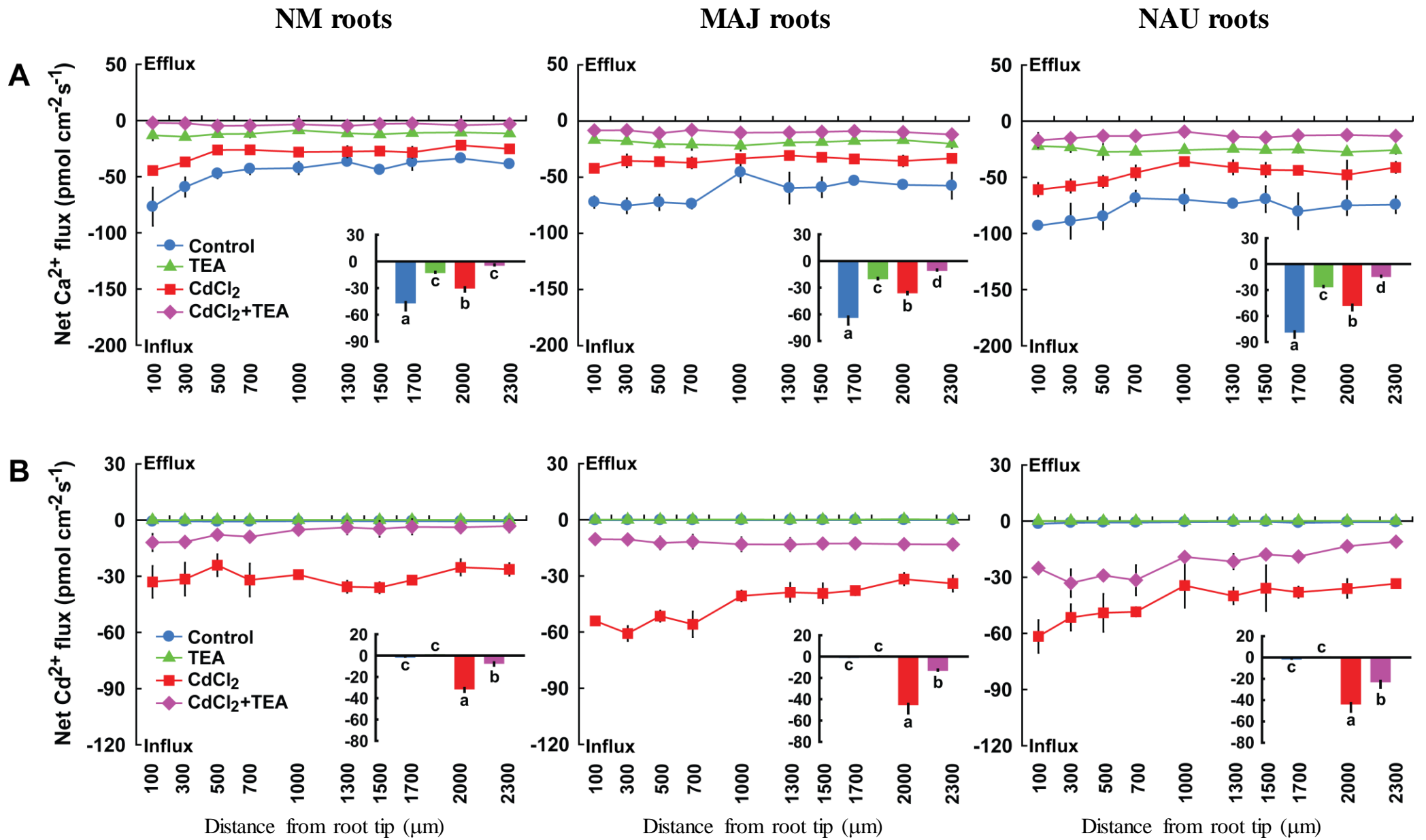




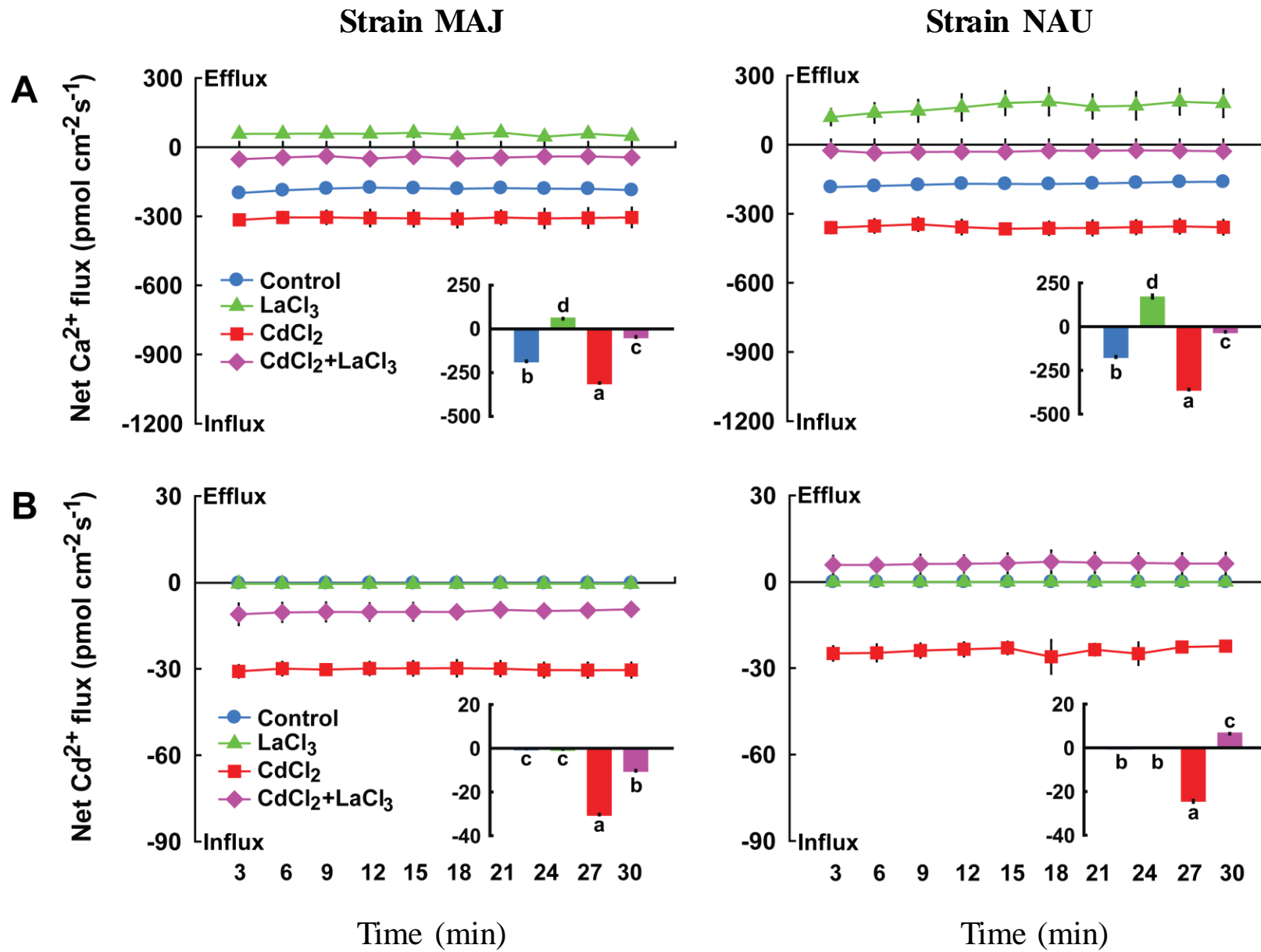
**Supplemental FIGURE S5. Effects of GdCl<sub>3</sub> on steady Ca<sup>2+</sup> and Cd<sup>2+</sup> fluxes in roots of ectomycorrhizal (MAJ and NAU) and non-mycorrhizal (NM) *Populus × canescens* under Cd<sup>2+</sup> stress.** Ectomycorrhizal (MAJ and NAU) and NM *P. × canescens* plants were subjected to 0 or 50 μM CdCl<sub>2</sub> for 24 h in the presence and absence of 500 μM GdCl<sub>3</sub>. Ca<sup>2+</sup> (A) and Cd<sup>2+</sup> (B) fluxes were measured along root axes, 100–2,300 μm from the apex, at intervals of 200 to 300 μm. Each point is the mean of four to five individual plants and bars represent the standard error of the mean. Inserted sections show the mean flux rates and different letters, a, b, c, and d, indicate significant difference at  $P < 0.05$  between treatments.



**Supplemental FIGURE S6. Effects of verapamil on steady  $\text{Ca}^{2+}$  and  $\text{Cd}^{2+}$  fluxes in roots of ectomycorrhizal (MAJ and NAU) and non-mycorrhizal (NM) *Populus × canescens* under  $\text{Cd}^{2+}$  stress.** Ectomycorrhizal (MAJ and NAU) and NM *P. × canescens* plants were subjected to 0 or 50  $\mu\text{M}$   $\text{CdCl}_2$  for 24 h in the presence and absence of 20  $\mu\text{M}$  verapamil.  $\text{Ca}^{2+}$  (A) and  $\text{Cd}^{2+}$  (B) fluxes were measured along root axes, 100–2,300  $\mu\text{m}$  from the apex, at intervals of 200 to 300  $\mu\text{m}$ . Each point is the mean of four to five individual plants and bars represent the standard error of the mean. Inserted sections show the mean flux rates and different letters, a, b, c, and d, indicate significant difference at  $P < 0.05$  between treatments.

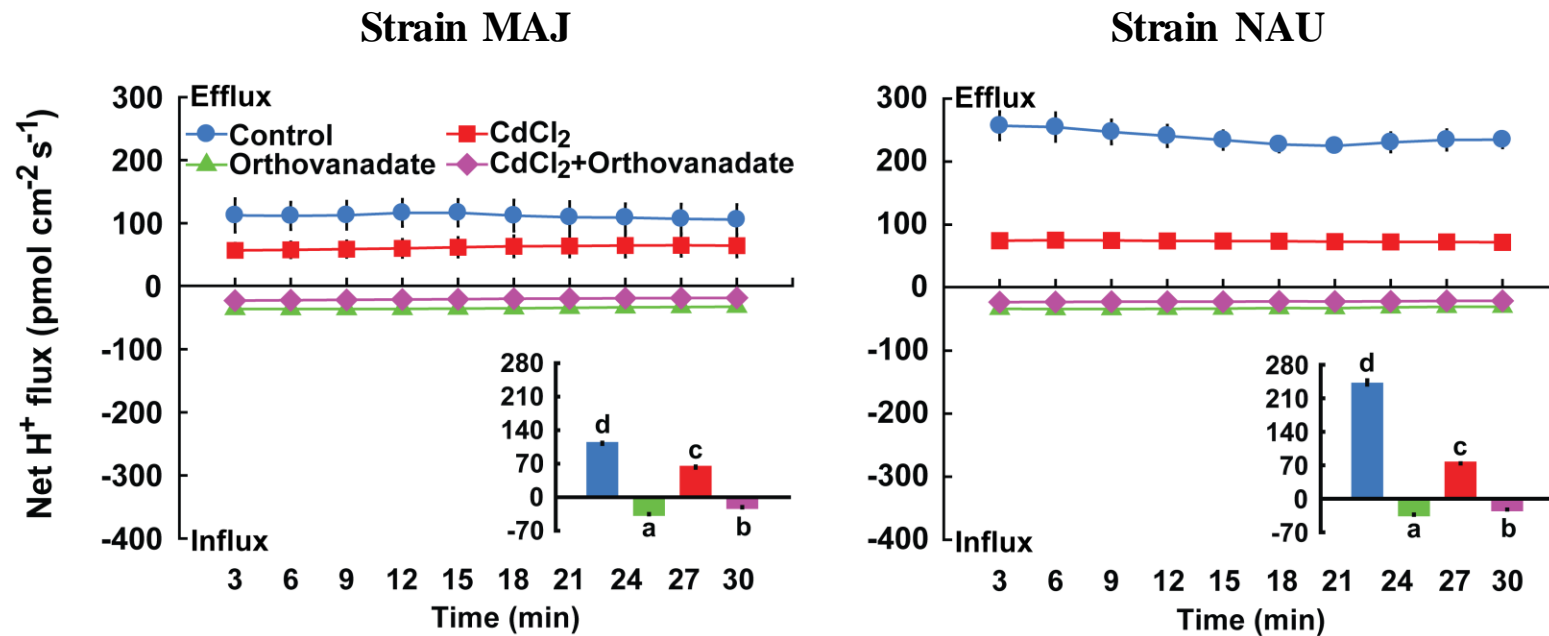


**Supplemental FIGURE S7. Effects of TEA on steady  $\text{Ca}^{2+}$  and  $\text{Cd}^{2+}$  fluxes in roots of ectomycorrhizal (MAJ and NAU) and non-mycorrhizal (NM) *Populus × canescens* under  $\text{Cd}^{2+}$  stress.** Ectomycorrhizal (MAJ and NAU) and NM *P. × canescens* plants were subjected to 0 or 50  $\mu\text{M}$   $\text{CdCl}_2$  for 24 h in the presence and absence of 50  $\mu\text{M}$  TEA.  $\text{Ca}^{2+}$  (A) and  $\text{Cd}^{2+}$  (B) fluxes were measured along root axes, 100–2,300  $\mu\text{m}$  from the apex, at intervals of 200 to 300  $\mu\text{m}$ . Each point is the mean of four to five individual plants and bars represent the standard error of the mean. Inserted sections show the mean flux rates and different letters, a, b, c, and d, indicate significant difference at  $P < 0.05$  between treatments.



**Supplemental FIGURE S8. Effects of LaCl<sub>3</sub> on steady Ca<sup>2+</sup> and Cd<sup>2+</sup> fluxes in *P. involutus* strains MAJ and NAU under Cd<sup>2+</sup> stress.** *P. involutus* isolates MAJ and NAU were subjected to 0 or 50 μM CdCl<sub>2</sub> for 24 h in the presence and absence of 5 mM LaCl<sub>3</sub>. Control axenic mycelia were treated without CdCl<sub>2</sub> or LaCl<sub>3</sub>. Ca<sup>2+</sup> (A) and Cd<sup>2+</sup> (B) fluxes were measured along the surface of pelleted hyphae over a recording period of 30 min. Each point is the mean of four to five axenic EM cultures and bars represent the standard error of the mean. Inserted sections show the mean flux rates and different letters, a, b, c and d, indicate significant difference at  $P < 0.05$  between treatments.





**Supplemental FIGURE S9. Effects of sodium orthovanadate on steady H<sup>+</sup> fluxes in *P. involutus* strains MAJ and NAU under Cd<sup>2+</sup> stress.** *P. involutus* isolates MAJ and NAU were subjected to 0 or 50 μM CdCl<sub>2</sub> for 24 h in the presence or absence of 500 μM sodium orthovanadate. Control axenic mycelia were treated without CdCl<sub>2</sub> or sodium orthovanadate. H<sup>+</sup> flux was measured along the surface of pelleted hyphae over a recording period of 30 min. Each point is the mean of four to five axenic EM cultures and bars represent the standard error of the mean. Inserted sections show the mean flux rates and different letters, a, b, c and d, indicate significant difference at *P* < 0.05 between treatments.