

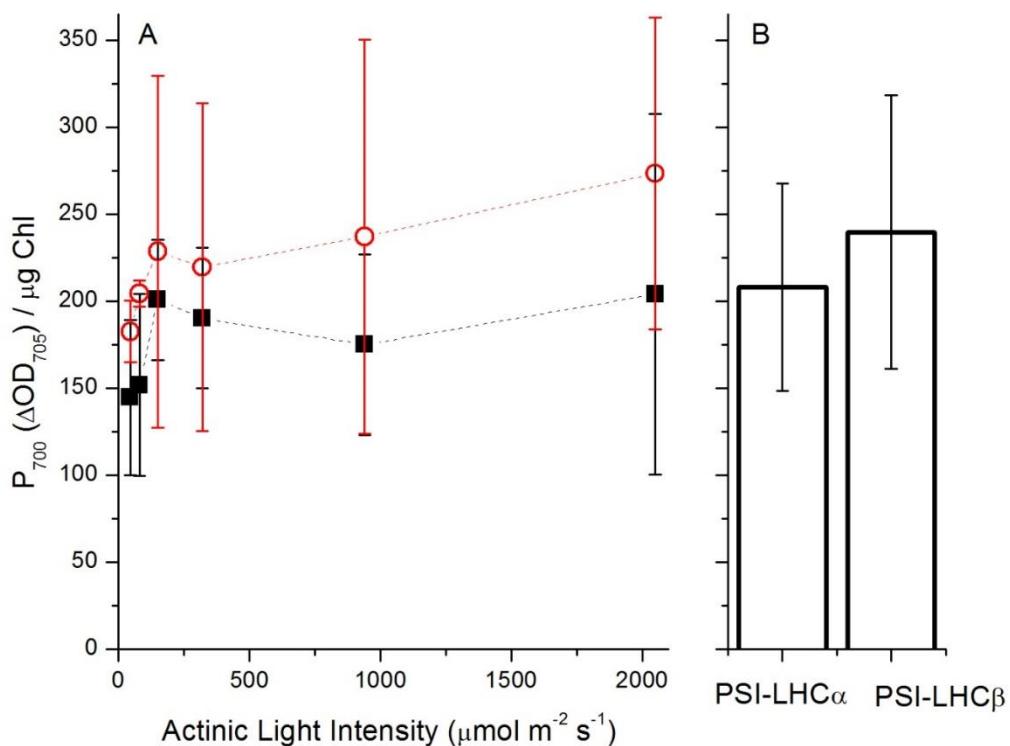
**Conservation of core complex subunits shaped the structure and function of photosystem I  
in the secondary endosymbiont alga *Nannochloropsis gaditana***

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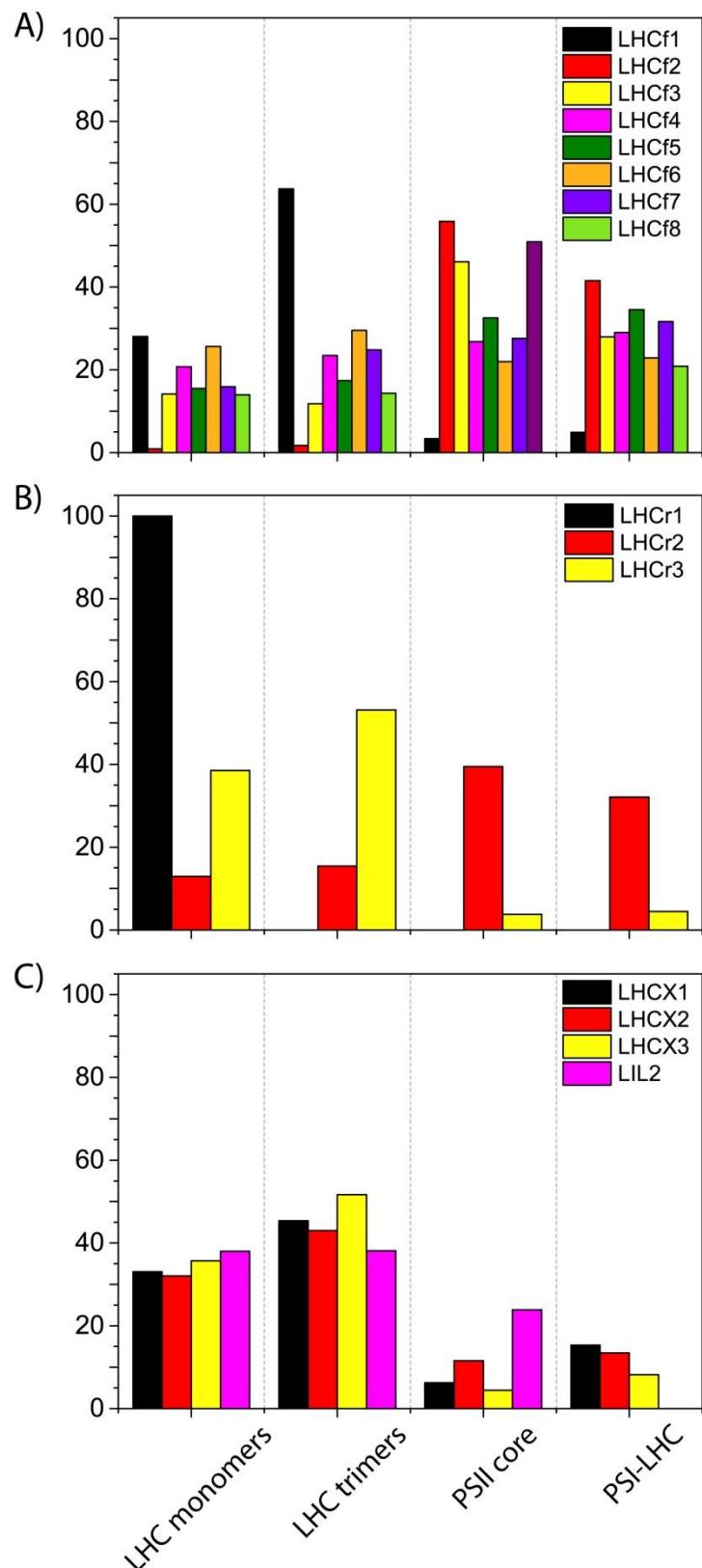
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**SUPPLEMENTARY MATERIAL**

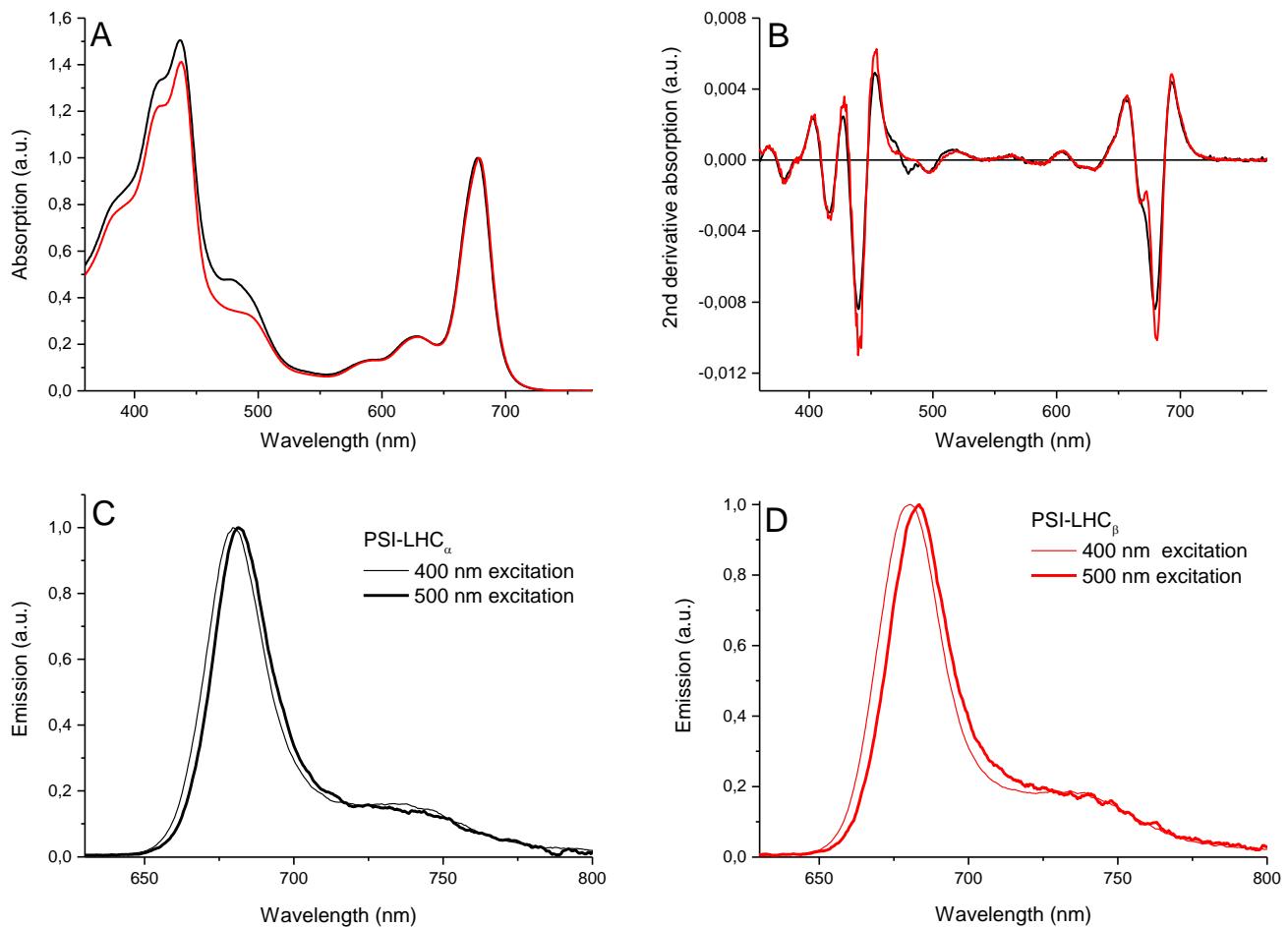
**FIGURE S1. Quantification of P700 content in PSI-LHC isolated from *Nannochloropsis gaditana*.** The  $\Delta OD_{705}$  signal measured with a JTS10 spectrophotometer (Biologic, France) was used as a quantification of the total P700 amount present in the sample after normalization to the precise Chl content of each sample ( $\approx 10 \mu\text{g}$  of Chl). To obtain the maximal P700 oxidation, P700 of isolated PSI-LHC was fully reduced by incubating the samples in the dark for 5 minutes with  $375 \mu\text{M}$  Methylviologen and  $2 \text{ mM}$  Ascorbate. After incubation, the sample was illuminated with an actinic red light (630 nm). A) Signal from PSI-LHC $\alpha$  (black squares) and PSI-LHC $\beta$  (red circles) depending on the actinic light intensity ( $n = 3$ ). B) Average over the different light intensities ( $150-2050 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$ ). The average values are not statistically different ( $n = 12$ ). Error bars represent the standard deviation.



**Figure S2. Protein distribution in the different fractions collected from a sucrose gradient loaded with thylakoids of *N. gaditana* solubilized with  $\alpha$ -DM.** Only LHC subunits not specifically enriched in PSI fraction are reported here, divided in LHCF (A), LHCR (B) and LHCX and LHC-like (C). The sucrose density gradient fractions F2-F5 corresponds to LHC monomers, LHC oligomers, PSII core and PSI-LHC, as reported in Figure 3A.



**FIGURE S3. Spectroscopic characterization of *N. gaditana* PSI-LHC at room temperature.** Absorption spectra (A) of PSI-LHC<sub>α</sub> (black) and PSI-LHC<sub>β</sub> (red), normalized to the maximum in the Q<sub>y</sub> region, and their second derivative (B). Fluorescence emission spectra upon 400 nm excitation (narrow line) or 500 nm excitation (thick line) of PSI-LHC<sub>α</sub> (C) and PSI-LHC<sub>β</sub> (D). Spectra are normalized to the maximum emission.



**Table S1.** NCBI Sequences AC numbers of identified PSI core subunits. \* indicates that more than one isoform has been identified in the genome (Jensen *et al.*, 2007; Busch *et al.*, 2013). ND stands for not determined sequence.

	<b>subunit</b>	<i>A. thaliana</i>	<i>O. sativa</i>	<i>P. patens</i>	<i>C. reinhardtii</i>	<i>Galactaria sulphuraria</i>	<i>Cyanidioschyzon merolae</i>	<i>Phaeodactylum tricornutum</i>	<i>Thalassiosira pseudonana</i>	<i>Ectocarpus siliculosus</i>	<i>Nannochloropsis gaditana</i>	<i>Emiliania huxleyi</i>
Present in cyanobacteria	<b>PsaA</b>	NP_0510 59.1	NP_0393 83.1	NP_9042 02.1	NP_9583 75.1	ADO3296 5.1	NP_8490 44.1	YP_87435 9.1	YP_87449 0.1	YP_00328 9256.1	YP_00731 6990.1	YP_27731 1.1
	<b>PsaB</b>	NP_0510 58.1	NP_0393 82.1	NP_9042 03.1	NP_9584 04.1	XP_00570 5021.1	NP_8490 45.1	YP_87435 8.1	YP_87449 1.1	YP_00328 9257.1	YP_00731 6991.1,	YP_27731 2.1
	<b>PsaC</b>	NP_0511 10.1	NP_0394 45.1	NP_9042 39.1	NP_9584 23.1	ADO3296 7.1	NP_8489 82.1	YP_87448 7.1	YP_87457 6.1	YP_00328 9280.1	YP_00731 7105.1; AGI98658 .1	YP_27734 6.1
	<b>PsaD</b>	NP_1921 86.1 (D1); *	NP_0010 62511.1	XP_00175 1403.1; *	XP_00169 7722.1	XP_00570 5023.1	NP_8490 51.1	YP_87439 4.1	YP_87454 7.1	YP_00328 9238.1	YP_00731 7065.1	YP_27739 8.1
	<b>PsaE</b>	NP_5678 18.2 (Ea); *	BAC8408 8.1	XP_00176 8221.1; *	XP_00170 2611.1	ADO3296 9.2	NP_8490 39.1	YP_87442 8.1	XP_00229 7543.1; *	YP_00328 9176.1	YP_00731 7004.1	XP_00578 4629.1; *
	<b>PsaF</b>	NP_1744 18.1	AAC7810 6.1	XP_00178 2922.1; *	XP_00169 6798.1	XP_00570 5045.1	NP_8491 04.1	YP_87436 1.1	YP_87456 2.1	YP_00328 9261.1	YP_00731 7046.1	YP_27734 8.1
	<b>Psal</b>	NP_0510 69.1	NP_0393 95.1	NP_9041 92.1	XP_00170 3367.1	ADO3297 1.1	NP_8491 27.1	YP_87437 8.1	YP_87454 1.1	YP_00328 9284.1	AGI98707 .1	YP_27739 4.1
	<b>PsaJ</b>	NP_0510 79.1	NP_0394 06.1	NP_9041 80.1	NP_9584 17.1	ADO3297 2.1	NP_8491 05.1	YP_87436 0.1	YP_87456 3.1	YP_00328 9262.1	YP_00731 7104.1	YP_27734 9.1
	<b>PsaK</b>	NP_1743 27.1	NP_0010 58895.1	XP_00175 2725.1; *	XP_00169 7230.1	ADO3297 3.1; P31567.1	NP_8489 78.1	ND	ND	ND	ND	ND
	<b>PsaL</b>	NP_1930 16.1	NP_0010 66659.1	XP_00175 4840.1	XP_00169 1084.1	ADO3297 4.1	NP_8491 35.1	YP_87436 6.1	YP_87455 7.1	YP_00328 9277.1	YP_00731 7052.1	YP_27735 2.1
Only eukaryotic	<b>PsaM</b>	ND	ND	NP_9042 14.1	ND	ADO3297 5.1	NP_8489 56.1	YP_87438 9.1	YP_87453 3.1	YP_00328 9254.1	ND	YP_27739 6.1
	<b>PsaG</b>	NP_1759 63.1	NP_0010 63493.1	XP_00176 5720.1; *	P14224.1	ND	ND	ND	ND	ND	ND	ND
	<b>PsaH</b>	NP_1882 35.1 (H1); *	NP_0010 56304.1	XP_00176 6961.1; *	XP_00169 0629.1	ND	ND	ND	ND	ND	ND	ND
	<b>PsaN</b>	AT5G640 40	EEC68974 .1	Nd	AAK0677 4.1	ND	ND	ND	ND	ND	ND	ND
	<b>PsaO</b>	NP_5638 15.1	NP_0010 52755.1	XP_00177 8960.1; *	XP_00170 0109.1	XP_00570 4648.1	XP_00553 7773.1	ND	ND	ND	ND	ND
	<b>PsaP</b>	AT2G468 20;	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND

**Table S2.** Identity matrix for PsaA protein obtained with MUSCLE tool (EMBL-EBI). Conservation versus *A. thaliana* and *N. gaditana* are highlighted in green and yellow, respectively.

PsaA	<i>Emiliania huxleyi</i>	<i>Nannochloropsis gaditana</i>	<i>Ectocarpus siliculosus</i>	<i>Thalassiosira pseudonana</i>	<i>Phaeodactylum tricornutum</i>	<i>P. patens</i>	<i>A. thaliana</i>	<i>O. sativa</i>	<i>C. reinhardtii</i>	<i>Cyanidioschyzon merolae</i>	<i>Galdieria sulphuraria</i>
<i>Emiliania huxleyi</i>	100	82.95	84.25	85.77	85.11	83.31	80.24	79.71	82.42	82.75	81.52
<i>Nannochloropsis gaditana</i>	82.95	100	85.91	85.5	83.76	82.68	79.6	78.52	82.39	83.06	83.09
<i>Ectocarpus siliculosus</i>	84.25	85.91	100	85.98	85.45	82.44	79.22	78.42	82.89	82.68	81.98
<i>Thalassiosira pseudonana</i>	85.77	85.5	85.98	100	95.48	82.78	79.71	78.5	82.16	81.82	82.31
<i>Phaeodactylum tricornutum</i>	85.11	83.76	85.45	95.48	100	82.11	79.04	77.84	81.62	80.48	81.12
<i>P. patens</i>	83.31	82.68	82.44	82.78	82.11	100	92.67	90.67	89.71	84.89	84.25
<i>A. thaliana</i>	80.24	79.6	79.22	79.71	79.04	92.67	100	95.47	86.36	82.49	81.31
<i>O. sativa</i>	79.71	78.52	78.42	78.5	77.84	90.67	95.47	100	85.03	80.75	80.51
<i>C. reinhardtii</i>	82.42	82.39	82.89	82.16	81.62	89.71	86.36	85.03	100	85.41	84.42
<i>Cyanidioschyzon merolae</i>	82.75	83.06	82.68	81.82	80.48	84.89	82.49	80.75	85.41	100	87.3
<i>Galdieria sulphuraria</i>	81.52	83.09	81.98	82.31	81.12	84.25	81.31	80.51	84.42	87.3	100

**Table S3. LHC proteins correspondence between *N. gaditana* and *N. oceanica*.** The UniProt ID allowed the identification of the corresponding *N. oceanica* Gene ID. Proposed names of *N. oceanica* CCMP1779 are also reported (Litvín *et al.*, 2016). \* indicates proposed attribution to PSI.

	<b><i>N. gaditana</i></b>	<b>Gene ID</b>	<b>UniProt ID</b>	<b><i>N. oceanica</i> CCMP1779</b>
LHCf	NgLHCf1	Naga_100012g50	W7T4V5	LHCv2
	NgLHCf2	Naga_100005g99	W7TY83	LHCv9
	NgLHCf3	Naga_100157g5	K8Z8N4	LHCv5
	NgLHCf4	Naga_100168g14	W7TFG9	LHCv7
	NgLHCf5	Naga_100017g59	W7TRI0	LHCv4
	NgLHCf6	Naga_100004g86	W7TXE6	LHCv8
	NgLHCf7	Naga_100013g28	W7U405	LHCv3
	NgLHCf8	Naga_100027g19	W7TCK1	LHCv6
LHCr	NgLHCr1	Naga_100002g18	K8YRV9	LHCr5
	NgLHCr2	Naga_100168g13	W7TZB5	LHCr7
	NgLHCr3	Naga_100018g45	K8ZB38	LHCr6
	NgLHCr4*	Naga_100056g15	W7TJ16	LHCr1*
	NgLHCr5*	Naga_100092g17	W7TX20	LHCr3*
	NgLHCr6*	Naga_100434g4	W7TJ16	LHCr4*
	NgLHCr7*	Naga_100641g3	W7T8I0	LHCr2
	NgLHCr8*	Naga_100017g83	W7UAI7	LHCr11
LHCx	NgLHCx1	Naga_100173g12	K8YWB4	LHCx1
	NgLHCx2	Naga_100056g42	K8YZX9	LHCx4
	NgLHCx3	Naga_101036g3	W7TIB0	LHCx3
LHC-like	LHC-like – LIL1*	Naga_100030g5	W7TTN9	Red-CAP1*
	LHC-like – LIL2	Naga_101227g1	W7TI84	LHCv10
				LHCv1
				LHCx2*

**Table S4.** Maximum absorption of PSI-LHC<sub>α</sub> and PSI-LHC<sub>β</sub> in the Q<sub>y</sub> and absorption forms visible in the second derivative of the absorption spectra, at 77K and RT.

	Maximum absorption in the Q <sub>y</sub>		2nd Derivative in the Chls region		2nd Derivative in the Car region	
	77K	RT	77K	RT	77K	RT
PSI-LHC <sub>α</sub>	677.5	677	669, 679.5, 685, 697.5	668.5, 679.5	486.5, 503	480, 497
PSI-LHC <sub>β</sub>	678.5	678.5	669, 679.5, 685, 697.5	667.5, 680.5	486.5, 503	479.5, 497

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