

Supplemental Materials

Molecular Biology of the Cell

Levin et al.

SUPPLEMENTARY FIGURES

Figure S1. **2xP4M expression does not affect Golgi morphology.** **A)** Representative confocal micrograph of a RAW264.7 cell co-expressing the PtdIns4P-biosensor GFP-2xP4M and the Golgi-marker mCh-sialyltransferase. Main panel shows mCh-sialyltransferase fluorescence; the cell outline is drawn in yellow. Bottom left inset shows GFP-2xP4M fluorescence. **B)** Representative confocal micrograph of a RAW cell expressing only the Golgi-marker mCh-sialyltransferase. Top right insets show magnifications of the corresponding dotted white boxes. Scale bars = 5 μ m.

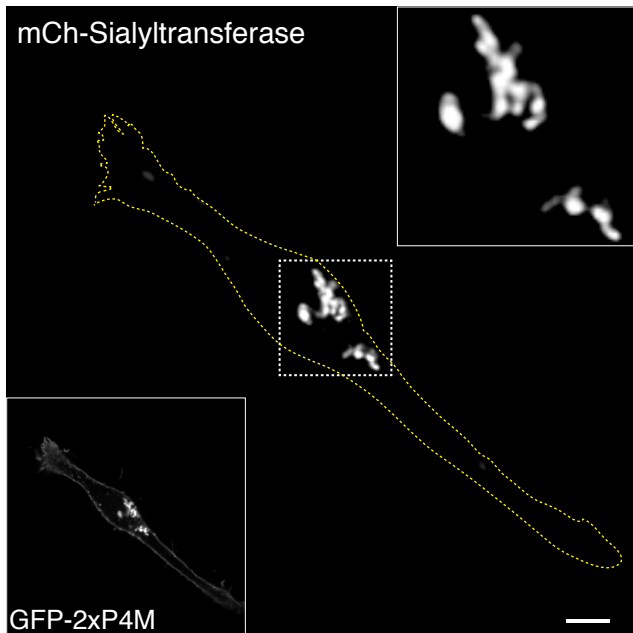
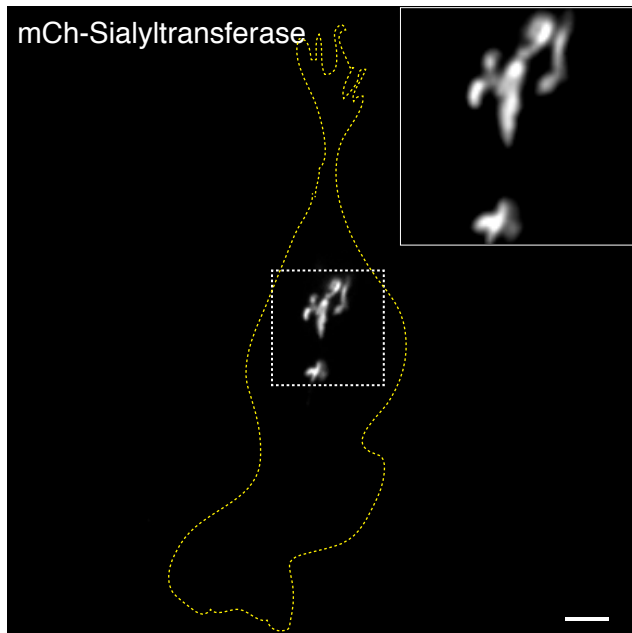
Figure S2. **Phosphoinositide metabolism during the early phagocytic stages.** **A)** Time-lapse gallery of confocal micrographs of RAW264.7 cells transiently co-expressing mCh-2xP4M and GFP-PH-Gab2 [a PtdIns(3,4,5)P₃-specific biosensor] during phagocytosis of IgG-SRBCs. The Roman numerals refer to the stages defined on Figure 3. Images are representative of at least 10 similar determinations. **B)** Schematic diagram illustrating possible enzymatic reactions whereby phagosomes can be depleted of PtdIns4P. **C)** Confocal micrographs of RAW264.7 cells co-expressing GFP-PH-TAPP1 [a PtdIns(3,4)P₂ biosensor] and mCh-2xP4M during the phases of initial increase (left) and disappearance of PtdIns4P (right); PtdIns(3,4)P₂ was not detected in phagosomes during PtdIns4P disappearance. **D)** Confocal micrographs acquired during phagocytosis of 1.6 μ m TMR-IgG-coated latex beads by LY294002-treated (50 μ M) RAW264.7 cells expressing GFP-2xP4M. Representative images of the phases of initial PtdIns4P increase (left) and its subsequent disappearance (right) are shown. LY294002 did not prevent PtdIns4P disappearance. Insets show magnifications of dotted white boxes. **E)** RAW264.7 cells were challenged with 3.87 μ m IgG-coated latex beads. After 15 min, cells were washed and fixed, permeabilized and immunostained using anti-PI4KA antibodies. Confocal sections are shown. A forming phagosome is indicated with a yellow arrow while a maturing phagosome is indicated with a white arrow. Images are representative of at least 10 similar determinations. Scale bars = 5 μ m.

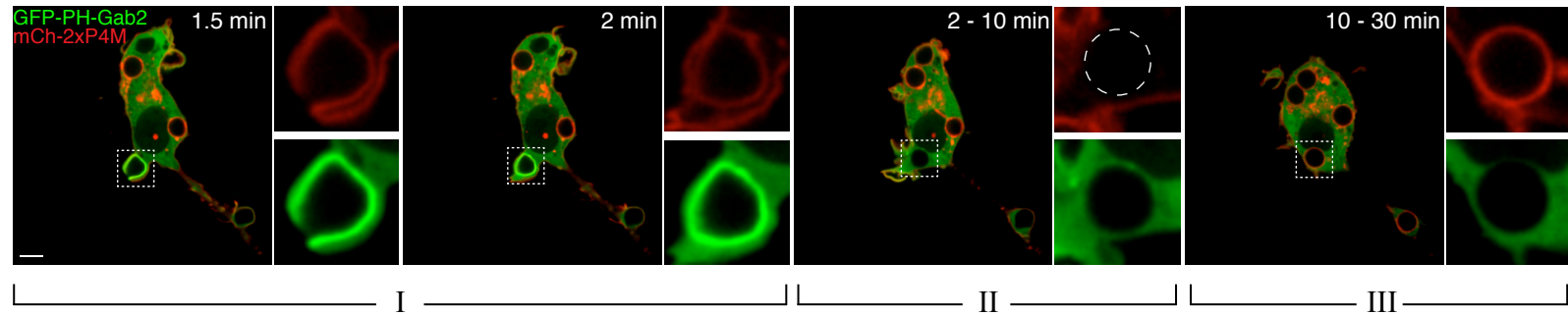
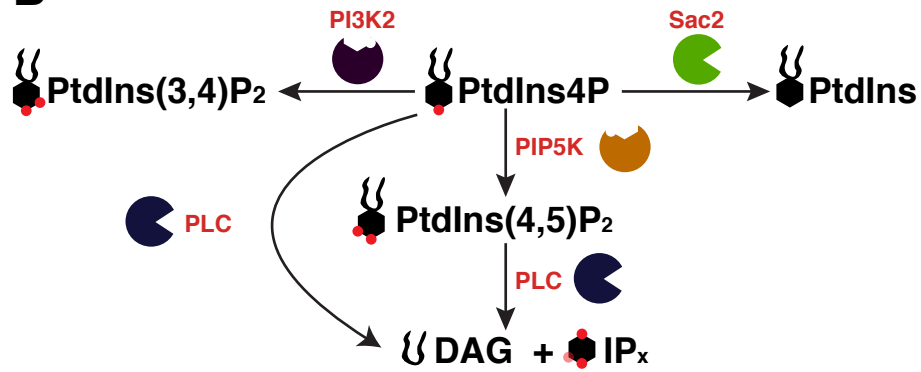
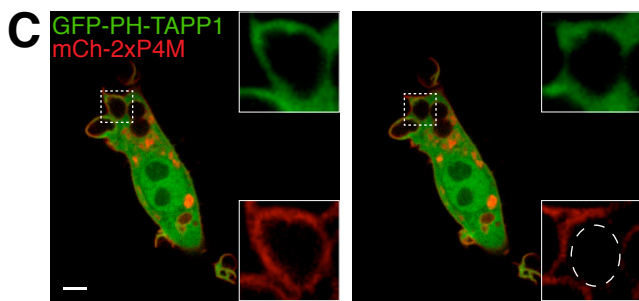
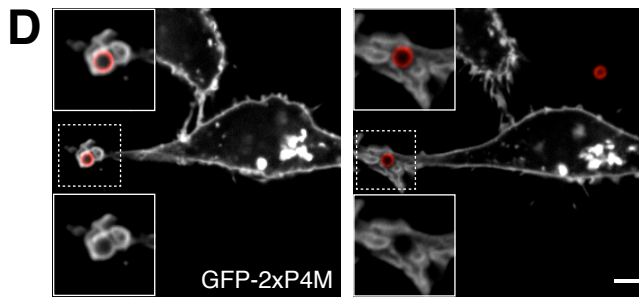
Figure S3. **PtdIns4P dynamics in COS-1-Fc γ RIIa cells.** **A)** Time-lapse gallery of confocal micrographs acquired during phagocytosis of TAMRA-labeled IgG-opsonized sheep red blood cells (IgG-SRBCs; shown in red) by COS-1-Fc γ RIIa cells transiently expressing GFP-2xP4M (green). The panels indicate the time elapsed from the moment the target was engaged. Insets show magnifications of the area delimited by dotted white boxes. **B)** Summary of the changes in PtdIns4P content of phagosomes following engagement of IgG-SRBCs. The intensity of GFP-2xP4M in phagosomes was quantified and normalized to that of the plasmalemma. The three phases defined in RAW264.7 cells were recapitulated in COS-1-Fc γ RIIa cells, with slower dynamics. Data are expressed relative to the maximum value attained during the initial transient increase, which was defined as time "0". Values are means \pm SEM of 4 independent experiments. Scale bars = 5 μ m.

Figure S4. **Cresyl violet co-localizes with GFP-Rab7, but not with Rab5.** **A)** Representative confocal section of RAW264.7 cells transiently expressing GFP-Rab5 and treated with cresyl violet (1 μ M for 2 min; shown in red). **B)** Single confocal section of

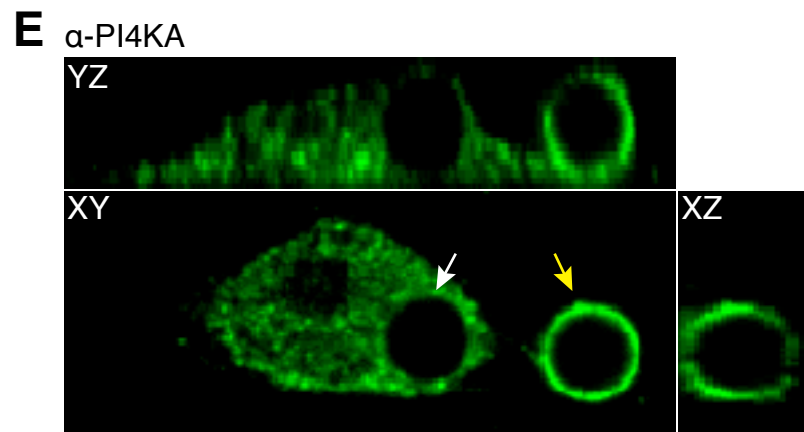
RAW264.7 cells transiently expressing GFP-Rab7 treated with cresyl violet as above. Insets show magnifications of dotted white boxes for each channel. Scale bars = 5 μ m.

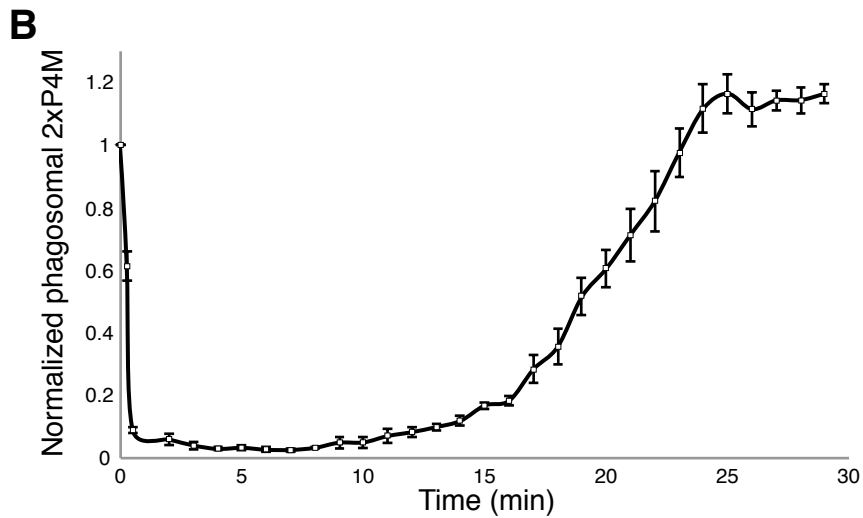
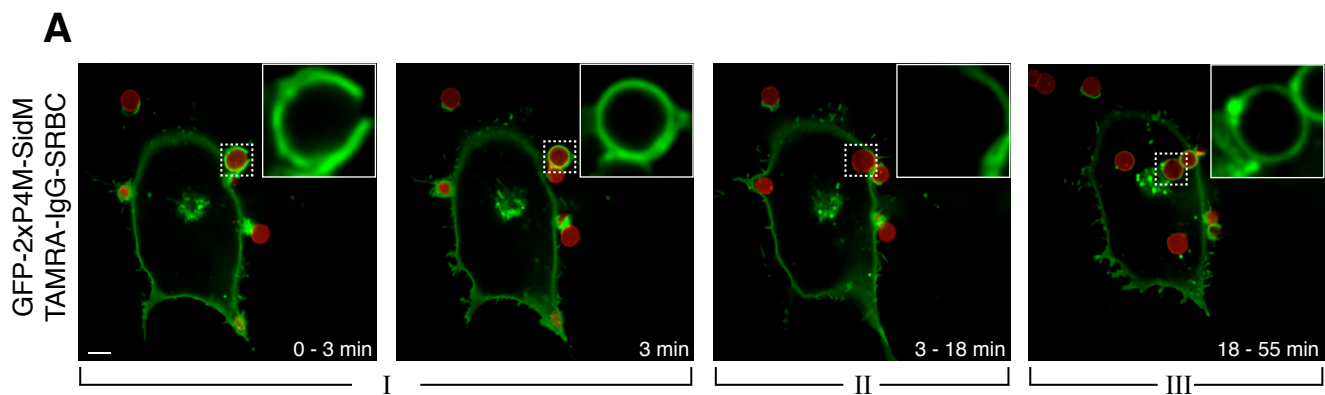
Figure S5. **Diagrammatic representation of the changes undergone by phosphoinositides during phagosome formation and maturation.** The changes in PtdIns4P (shades of blue) are compared with those undergone by PtdIns(4,5)P₂ (green), PtdIns(3,4,5)P₃ (red) and PtdIns3P (purple). Phagosome formation is divided into receptor engagement, pseudopod extension and vacuole sealing stages, while maturation is divided into early and late/lysosomal stages.

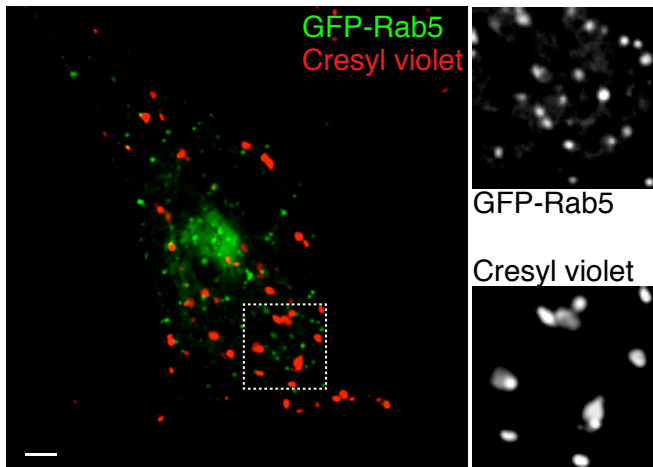
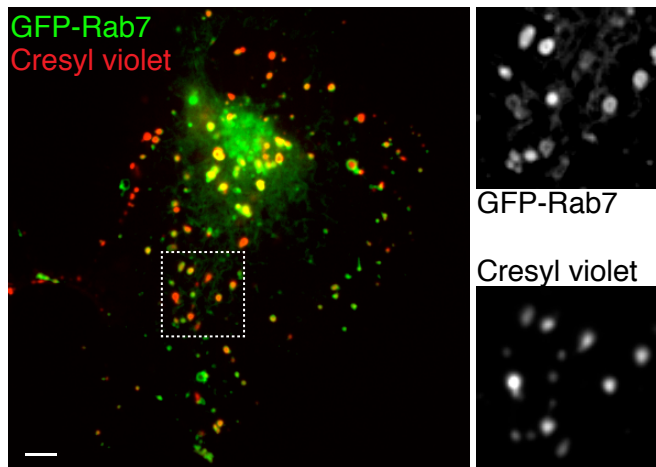
A**B**

A**B****C****D**

LY294002

E



A**B**

Phagosome formation

Phagosome maturation

Engagement

Extension

Sealing

Early maturation

Late maturation /
phagolysosome

