

**Supplemental table 1. AA, EPA and DHA bioactive metabolome:
CLP infectious exudates from WT and DRV2-KO.**

LM levels in mouse exudate (pg/ml)														
AA bioactive metabolome	Q1	Q3	WT veh.			WT RvD2			DRV2-KO veh.			DRV2-KO RvD2		
LXA ₄	351	115	8.0	±	4.5	16.1	±	14.2	9.3	±	6.3	6.9	±	2.9
LXB ₄	351	221	82.9	±	24.3	156.5	±	40.7	162.4	±	37.3	177.1	±	30.4
5S,15S-diHETE	335	235	53.0	±	14.1	52.1	±	10.8	63.4	±	30.9	77.4	±	13.3
AT-LXA ₄	351	115	13.7	±	5.5	17.1	±	9.4	14.3	±	3.6	21.0	±	4.6
AT-LXB ₄	351	221	14.4	±	9.9	45.6	±	38.7	23.1	±	15.0	28.7	±	14.8
LTB ₄	335	195	857.3	±	227.3	903.3	±	343.8	1825.3	±	296.2	2459.0	±	629.4
PGD ₂	351	233	312.9	±	107.4	329.1	±	66.3	389.0	±	116.0	558.7	±	158.7
PGE ₂	351	189	1326.1	±	595.5	2554.3	±	615.3	2800.8	±	1079.5	3490.1	±	889.0
PGF _{2α}	353	193	1358.2	±	540.4	608.3	±	115.5	1817.8	±	996.1	2050.9	±	1046.0
TXB ₂	369	169	589.4	±	72.7	755.1	±	162.5	1128.9	±	237.8	1492.0	±	206.1
DHA bioactive metabolome														
RvD1	375	121	15.0	±	9.8	7.6	±	2.3	5.7	±	1.9	8.9	±	3.4
RvD2	375	175	12.0	±	5.6	196.3	±	81.2	8.9	±	2.9	121.0	±	41.4
RvD3	375	147	0.4	±	0.1	1.2	±	0.5	0.5	±	0.2	1.0	±	0.3
RvD4	359	255	30.4	±	5.4	43.8	±	17.2	54.8	±	28.0	43.0	±	11.4
RvD5	359	199	65.1	±	19.9	74.9	±	22.9	68.9	±	36.8	115.6	±	43.4
AT-RvD1	375	121	30.3	±	12.9	35.0	±	15.2	17.3	±	9.7	21.6	±	7.1
AT-RVD3	375	147	0.7	±	0.2	3.4	±	1.5	1.4	±	0.8	2.6	±	1.3
PD1	359	153	2.8	±	0.5	4.2	±	1.1	4.1	±	0.6	5.7	±	1.2
10S,17S-diHDHA	359	153	3.9	±	0.4	2.3	±	0.4	3.9	±	0.7	2.6	±	0.5
AT-PD1	359	153	11.0	±	5.8	8.3	±	1.3	17.2	±	13.2	15.0	±	4.7
MaR1	359	221	12.4	±	6.3	6.7	±	2.2	26.8	±	18.0	21.6	±	7.2
7S,14S-diHDHA	359	221	33.0	±	18.5	30.3	±	6.9	125.6	±	93.3	99.3	±	33.5
4S,14S-diHDHA	359	101	527.9	±	193.9	126.3	±	38.9	278.1	±	80.6	155.9	±	54.9
EPA bioactive metabolome														
RvE1	349	195	15.1	±	3.5	45.5	±	20.9	47.3	±	22.8	34.7	±	12.0
RvE2	333	253	4.4	±	1.1	4.9	±	2.2	6.4	±	1.5	5.7	±	2.2
RvE3	333	201	4.7	±	1.5	3.0	±	0.5	3.3	±	1.3	2.2	±	0.4

Exudates from mouse after CLP were collected as described then were extracted and LM levels investigated using LM metabolipidomics (see materials and methods for details). Values are express as mean ± SEM.

Supplemental Table 2. Proteome profiling of CLP infectious exudates from WT and DRV2-KO with RvD2 treatment

Cytokines, chemokines and growth factor	<i>P</i> value		<i>P</i> value		<i>P</i> value
Adiponectin/Acrp30	0.431	DPPIV/CD26	0.175	IL-27	0.193
Amphiregulin	0.055	EGF	0.159	IL-28	0.093
Angiopoietin-1	0.170	Endoglin/CD105	0.258	IL-33	0.109
Angiopoietin-2	0.206	Endostatin	0.477	LDL R	0.193
Angiopoietin-like 3	0.128	Fetuin A/AHSG	0.376	Leptin	0.251
BAFF/BLyS/TNFSF13B	0.200	FGF acidic	0.249	LIF	0.119
C1q R1/CD93	0.239	FGF-21	0.086	Lipocalin-2/NGAL,	0.176
CCL2/JE/MCP-1	0.293	Flt-3 Ligand	0.062	LIX	0.007 *
CCL3/CCL4 MIP-1 alpha/beta	0.242	Gas6	0.305	M-CSF	0.124
CCL5/RANTES	0.242	G-CSF	0.447	MMP-2	0.147
CCL6/C10	0.241	GDF-15	0.158	MMP-3	0.002 *
CCL11/Eotaxin	0.145	GM-CSF	--	MMP-9	0.016 *
CCL12/MCP-5	0.411	HGF	0.258	Myeloperoxidase	0.057
CCL17/TARC	0.079	ICAM-1/CD54	0.334	Osteopontin (OPN)	0.204
CCL19/MIP-3 beta	0.223	IFN-gamma	--	Osteoprotegerin/TNFRSF11B	0.383
CCL20/MIP-3 alpha	0.170	IGFBP-1	0.402	PD-ECGF/Thymidine phosphorylase	0.189
CCL21/6CKine	0.120	IGFBP-2	0.471	PDGF-BB	0.052
CCL22/MDC	0.105	IGFBP-3	0.251	Pentraxin 2/SAP	0.036 *
CD14	0.071	IGFBP-5	0.084	Pentraxin 3/ TSG-14	0.038 *
CD40/TNFRSF5	0.083	IGFBP-6	0.283	Periostin/OSF-2	0.101
CD160	0.192	IL-1 alpha/IL1F1	0.326	Pref-1/DLK-1/FA1	0.085
Chemerin	0.184	IL-1 beta/IL-1F2	0.089	Proliferin	0.086
Chitinase 3-like 1	0.460	IL-1ra/IL-1F3	0.351	Protein Convertase 9/PCSK9	--
Coagulation Factor III/Tissue Factor	0.111	IL-2	--	RAGE	0.438
Complement Component C5/C5a	0.067	IL-3	--	RBP4	--
Complement Factor D	0.230	IL-4	0.231	Reg3G	0.018 *
C-Reactive Protein/CRP	0.308	IL-5	0.095	Resistin	0.309
CX3CL1/Fractalkine	0.085	IL-6	0.338	E-Selectin/CD62E	0.099
CXCL1/KC	0.405	IL-7	--	P-Selectin/CD62P	0.126
CXCL2/MIP-2	0.479	IL-10	0.238	Serpin E1/PAI-1	0.282
CXCL9/MIG	0.198	IL-11	0.225	Serpin F1/PEDF	0.019 *
CXCL10/IP-10	0.238	IL-12p40	0.167	Thrombopoietin	0.158
CXCL11/I-TAC	0.096	IL-13	0.107	TIM-1/KIM-1/HAVCR	0.086
CXCL13/BLC/BCA-1	0.350	IL-15	0.091	TNF-alpha	0.158
CXCL16	0.109	IL-17A	0.323	VCAM-1/CD106	0.261
Cystatin C	0.282	IL-22	0.036 *	VEGF	0.205
Dkk-1	0.199	IL-23	0.112	WISP-1/CCN4	0.182

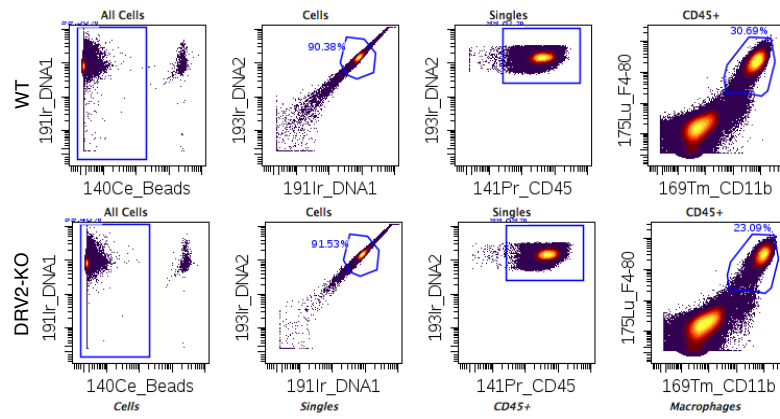
**p*<0.05; -- not detected

Peritoneal lavages were collected at 12h post CLP. Cell-free supernatants were collected by centrifugation. A 1:10 dilution of the supernatant (150µl) were incubated with the pre-coated Proteome Profiler array membranes (ARY028, Mouse Cytokine Antibody array kit; R&D Systems) and processed according to the manufacturer's instructions. Densitometric analysis of dot blots was performed using Image J software (National Institute of Health, Bethesda, MD, USA). *P* values were obtained by comparing WT+ RvD2 vs. DRV2-KO + RvD2 groups using 2-tailed Student's *t* test. **p*<0.05; -- not detected.

Supplemental Table 3. Antibodies used for mass cytometry

Target epitope	Clone	Isotopic label	Supplier
CD45	30-F11	Pr 141	LMA CyTOF
pAKT (S473)	D9E	Sm 152	Fluidigm
p-p38 (T180/Y182)	D3F9	Gd 156	Fluidigm
pStat3 (Y705)	4	Gd 158	Fluidigm
pStat1 (Y701)	58D6	Gd 160	LMA CyTOF
pNF- κ b p65 (S529)	K10-895.12.50	ER166	Fluidigm
pEER1/2 (T202/Y204)	D13.14.4E	Er 167	Fluidigm
CD11b	M1/70	Tm 169	LMA CyTOF
pS6 (S235/S236)	N7-548	Yb 172	Fluidigm
pStat5 (Y694)	D47/E7 XP	Yb 174	LMA CyTOF
F4/80	BM8	Lu 175	LMA CyTOF
pCREB (S133)	87G3	Yb 176	Fluidigm

LMA CyTOF (Longwood Medical Areas CyTOF Antibody Resources Core; Brigham and Women's Hospital)



Supplementary Figure 1. Sequential gating strategy used to identify resident peritoneal macrophages (CD11b⁺F4/80⁺) for CyTOF