

SUPPLEMENTARY METHODS

Transfections

Cells were seeded on a 6 well plate. The next day, cells were transfected using 2 µg of hCD24 and 4 µl of lipofectamine 2000 (Invitrogen) with Opti-MEM medium (Gibco), according to manufacturer instructions. The cell culture medium was changed 4 hr after transfection and cells were lysed 24 hours post-transfection, using 100 µl of RA lysis buffer (Macherey-Nagel).

RNA extraction and qRT-PCR

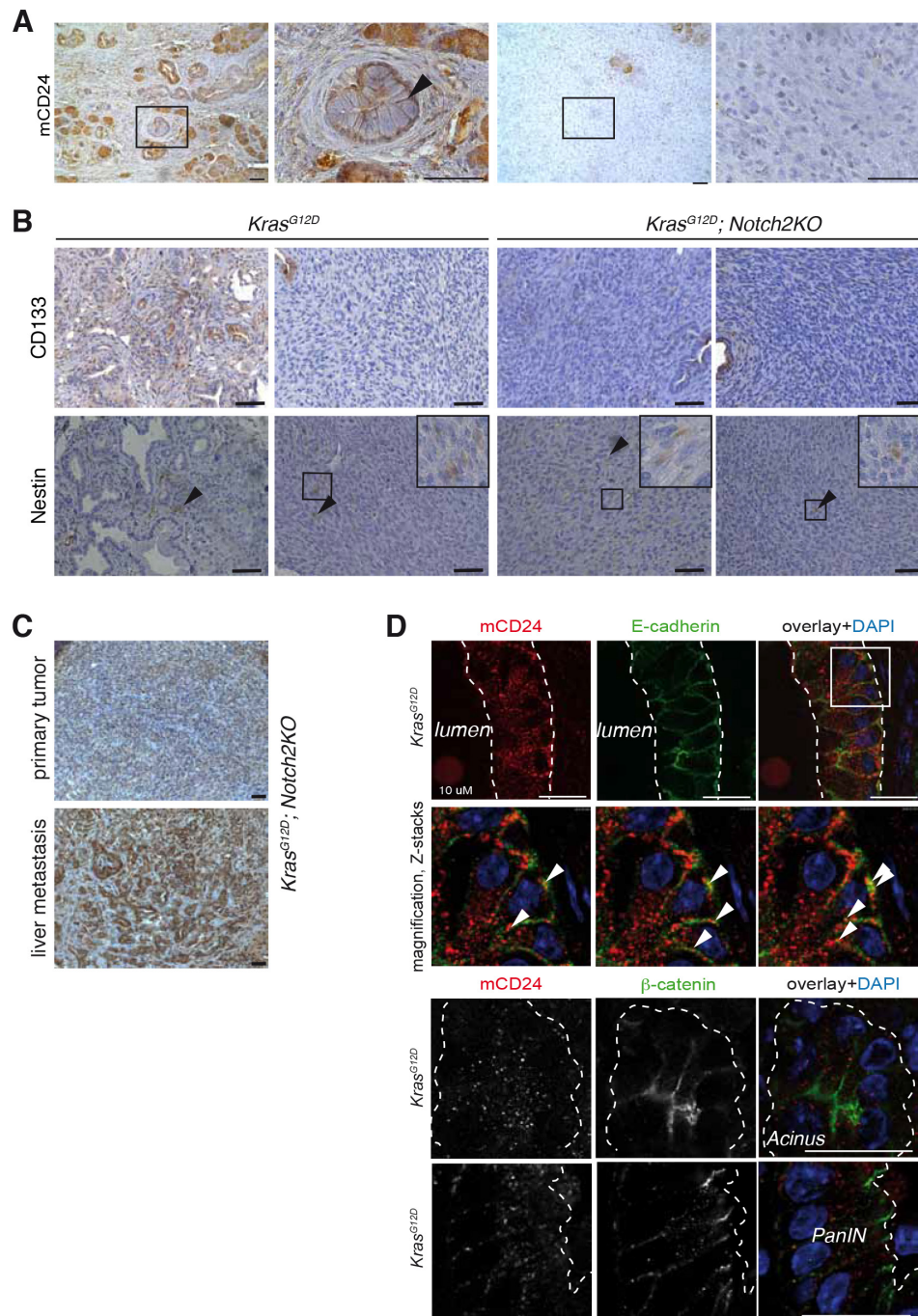
Total RNA was extracted from cells using the Maxwell 16 LEV simply RNA kit (Promega), according

to manufacturer's instructions. A total of 2 µg of RNA was reverse transcribed to cDNA, in 20 µl reactions, using Super Script II RT (Applied Biosystems) and following manufacturer's instructions. The resultant cDNA was diluted 10-fold and 5 µl was used as template for quantitative real time PCR (qRT-PCR). qRT-PCR was performed in triplicate, in a total volume of 20 µl, using the primers listed below, and 480 SYBR Green I Master (Roche) according to manufacturer's instructions. The relative amount of each gene of interest was normalized to the reference gene GAPDH using the $\Delta\Delta CT$ method (Livak, K. J. & Schmittgen, T. D. Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta CT}$ Method. (*Methods* 25, 402-408, 2001)).

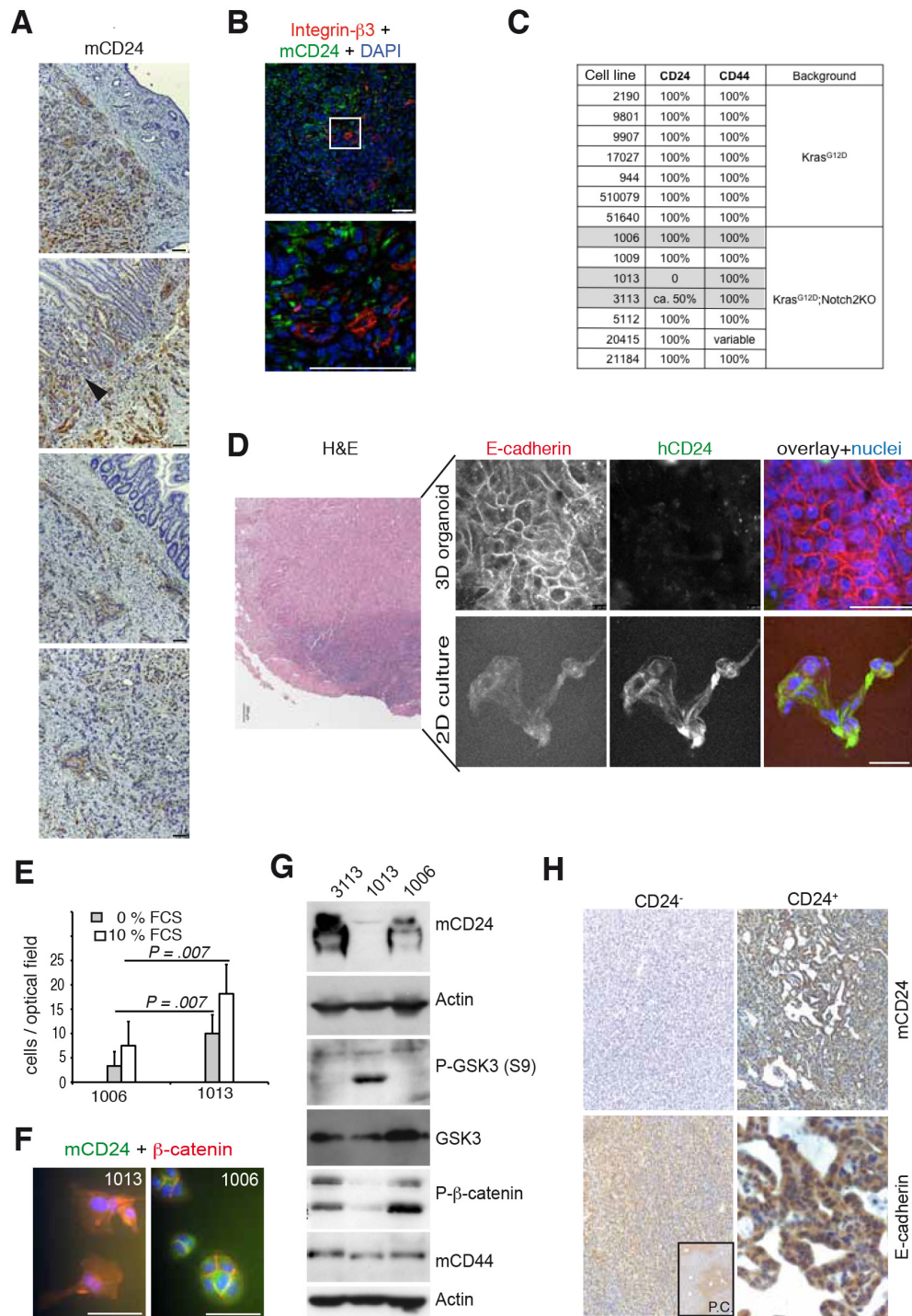
Primer list for qRT-PCR experiments

Primer name	Sequence
RT-hCtnnB_F	AGCTGACCAGCTCTCTCTCA
RT-hCtnnB_R	CCAATATCAAGTCCAAGATCAGC
RT-hSnail1_F	GCACATCCGAAGCCACAC
RT-hSnail1_R	GGAGAAGGTCCGAGCACA
RT-hCdh1_F	CCCGGGACAACGTTTATTAC
RT-hCdh1_R	GCTGGCTCAAGTCAAAGTCC
RT-hCd24_F	GACATGGGCAGAGCAATGGTGGC
RT-hCd24_R	GAGTGAGACCACGAAGAGACTGGC
RT-hTwist1_F	TCGGACAAGCTGAGCAAGAT
RT-hTwist1_R	CTCCATCCTCCAGACCGAG
RT-hZeb1_F	TCAAAAGGAAGTCAATGGACAA
RT-hZeb1_R	GTGCAGGAGGGACCTCTTTA
RT-hGapdh_F	GCTCTCTGCTCCTCCTGTTC
RT-hGapdh_R	ACGACCAAATCCGTTGACTC
RT-CyclophilinD1 F	ATGGTCAACCCACCGTG
RT-CyclophilinD1 R	TTCTGCTGTCTTTGGAACCTTTGTC
RT-Amylase F	TGGTCAATGGTCAGCCTTTTTTC
RT-Amylase R	CACAGTATGTGCCAGCAGGAAG
RT-Axin2 F	GCCAATGGCCAAGTGTCTCT
RT-Axin2 R	GCGTCATCTCCTTGGGCA
RT-p21 F	CACAGCGATATCCAGACATTCAG
RT-p21 R	CGGAACAGGTCCGACATCA
RT-CyclinD1 F	CTGTGCCACAGATGTGAAGTTC
RT-CyclinD1 R	AGTCCGGGTCACACTTGATGA
RT-mCd24 F	GCTCCTACCCACGCAGATTTA
RT-mCd24 R	CCCTCTGGTGGTAGCGTTACTT

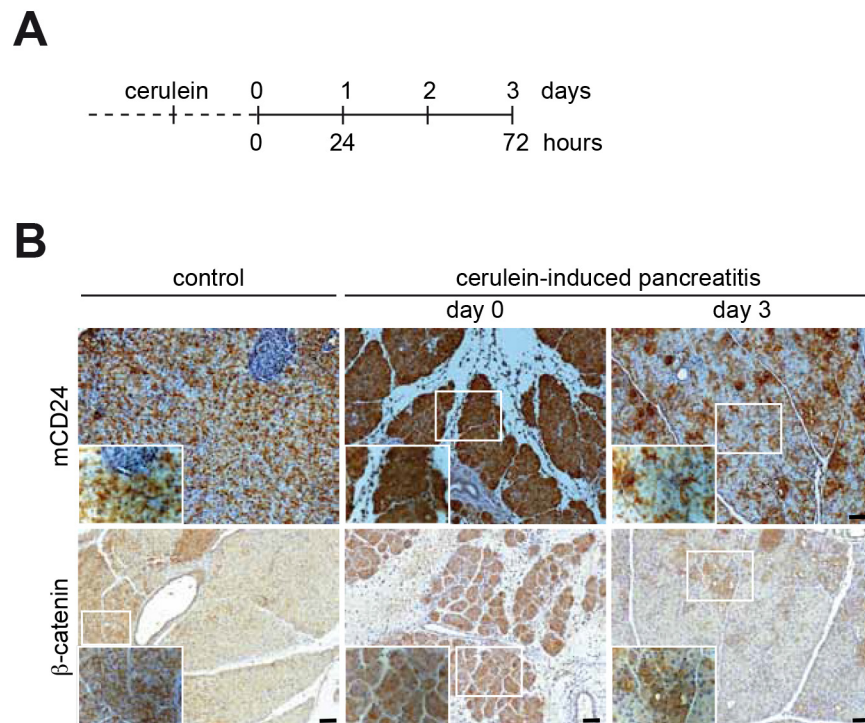
SUPPLEMENTARY FIGURES



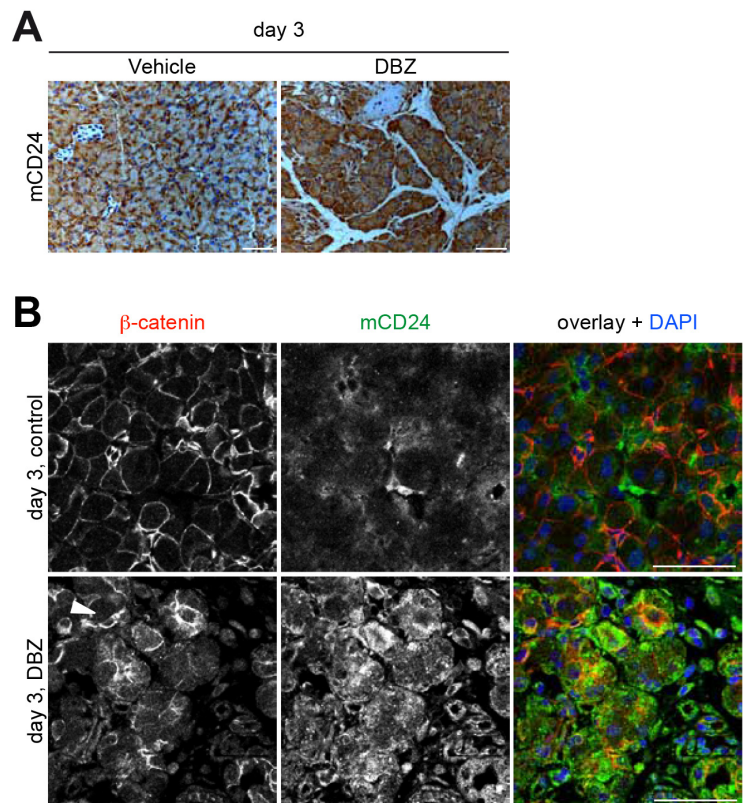
Supplementary Figure S1: CD24 localization. **A.** Immunohistological staining for mCD24 in pancreatic sections of *Kras^{G12D}* mice. Scale bars = 50 μ M. **B.** Immunohistological staining for cancer stem cell markers in pancreatic sections of mice of the indicated genotypes. Scale bars = 50 μ M. **C.** Immunohistological staining for mCD24 of a primary tumor and matched liver metastasis. **D.** Confocal analysis shows vesicular localization of CD24 in the pancreas of 3-6 months old *Kras^{G12D}* mice. Part of CD24+ vesicles colocalize with E-cadherin and β -catenin at adherent junctions (arrows). Images of the single channels were taken sequentially. Scale bars = 10 μ M.



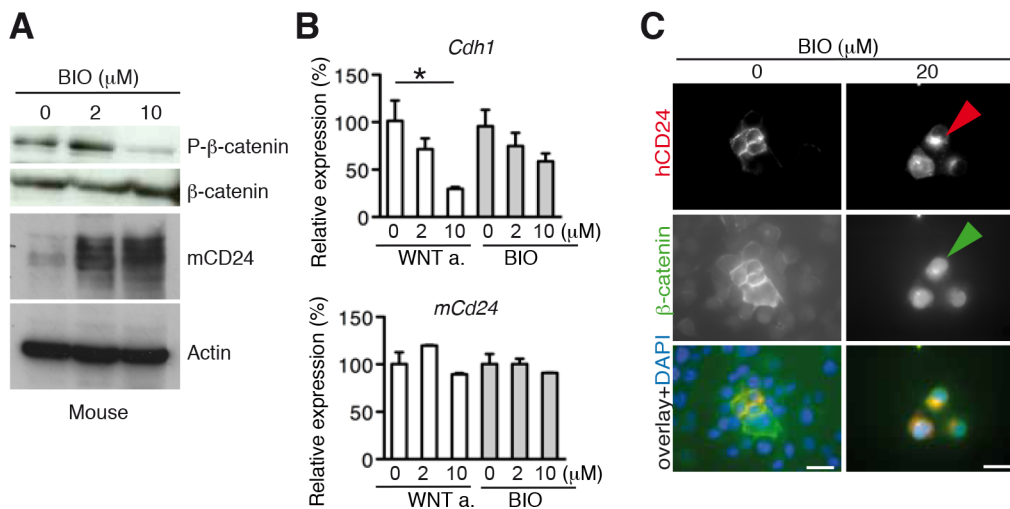
Supplementary Figure S2: CD24 expression. **A.** Immunohistological staining in tissue sections from murine invasive PDAC. **B.** Integrin-β3 and mCD24 immunofluorescence staining of murine invasive PDAC. **C.** Tumor cells derived from murine PDACs were analyzed for CD24 and CD44 expression by FACS analysis. **D.** Organoids were generated from human tissue and cultivated in a 3-dimensional matrigel matrix (3D) or on conventional 2 dimensional plastic dish (2D). H&E staining of the original tissue. 3D and 2D cultures were analyzed by immunofluorescence staining. Scale bars = 50 μM. **E.** migration assay. Cells that were either CD24 positive (#1006) or CD24 negative (#1013). Mean ± SD. Quantification of migrated cells was performed 48 hours later. **F.** Immunofluorescence staining of murine pancreatic tumor cell lines. **G.** Western blot analysis of murine pancreatic tumor cell lines. **H.** Immunohistochemistry of orthotopic tumors. Scale bars = 50 μM.



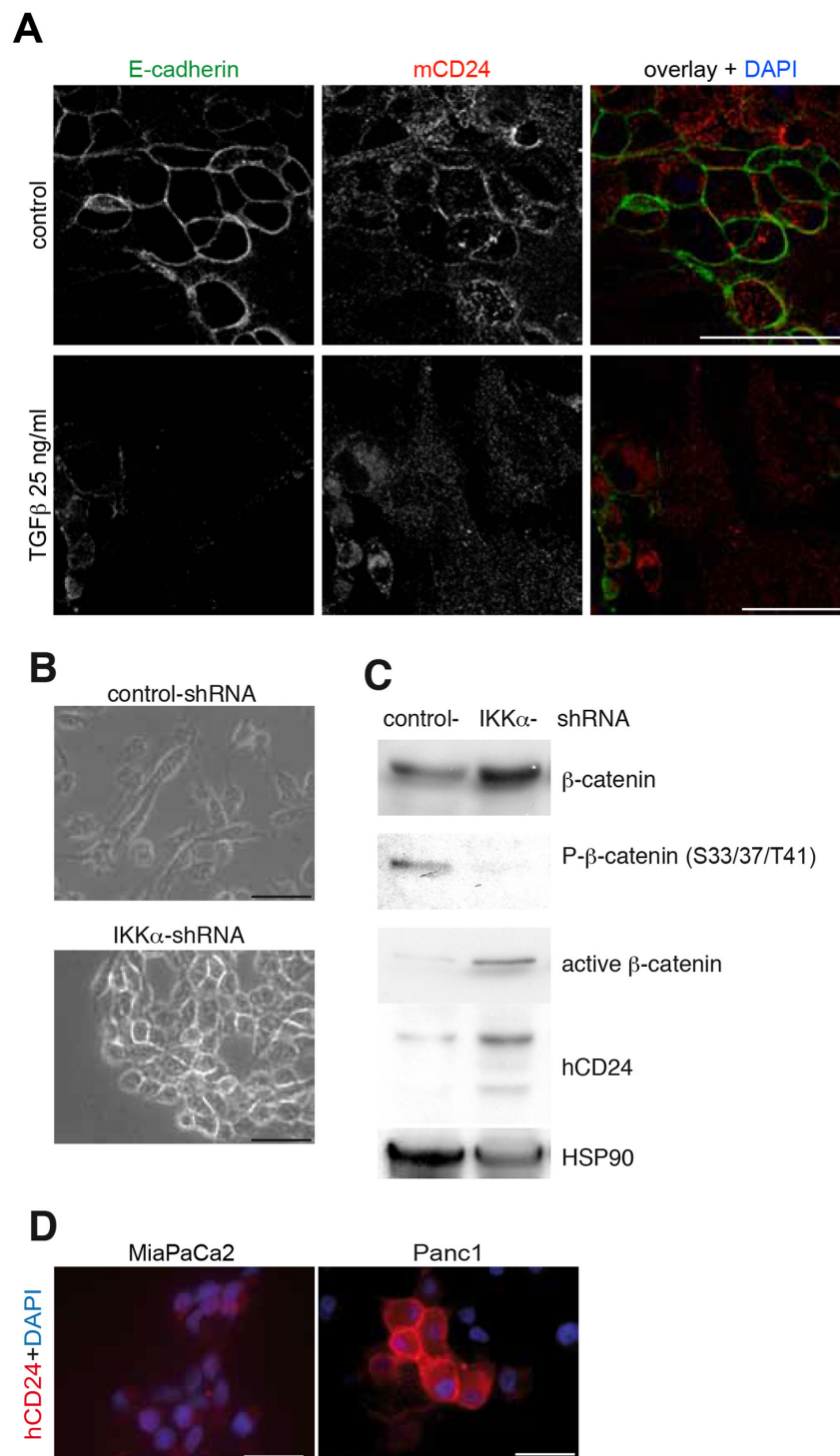
Supplementary Figure S3: CD24 expression in a mouse model for acute pancreatitis. A. Experimental design used for the induction of acute pancreatitis. B. Pancreatic tissue of mice treated as described in A was stained for mCD24 and β -catenin. CD24 expression was strongly upregulated after induction of acute pancreatitis in concomitance with increased cytoplasmic β -catenin expression. Scale bars = 50 μ M.



Supplementary Figure S4: CD24 expression in a mouse model for acute pancreatitis treated with DBZ. A, B. Acute pancreatitis was induced in wild type mice. Inhibition of γ -secretase with DBZ strongly enhanced CD24 expression. B, confocal pictures of immunofluorescence stainings. Mice were treated as described in A. Images of the single channels were taken sequentially. Scale bars = 50 μ M.



Supplementary Figure S5: Activation of the WNT pathway. A. Murine pancreatic cells (#1006) were stimulated with BIO for 24 hours. Western blot analysis shows a decrease in β -catenin phosphorylation (S33/37/T41) indicating successful activation of the Wnt pathway in concomitance with increase of CD24 expression. B. Pancreatic cells were treated as described in A and real-time qRT-PCR analysis for the indicated genes was performed. C. pancreatic cells were treated with BIO for 24 hours as described above, immunofluorescence staining shows intracellular accumulation of CD24 (red arrow) after β -catenin activation (green arrow). Scale bars = 50 μ M.



Supplementary Figure S6: CD24 expression during EMT. **A.** Murine pancreatic tumor cells were treated with TGFβ for 48 hours and analyzed by immunofluorescence staining. Downregulation of E-cadherin (green) expression indicates induction of EMT. **B.** The mesenchymal pancreatic cell line MiaPaCa2 acquires an epithelial phenotype upon stable transfection with IKKα siRNA. **C.** Western blot analysis shows upregulation of total β-catenin in concomitance with dephosphorylation of S33/37/T41. **D.** Immunofluorescence staining for CD24 in two human pancreatic cancer cell lines. Scale bars = 50 μM.