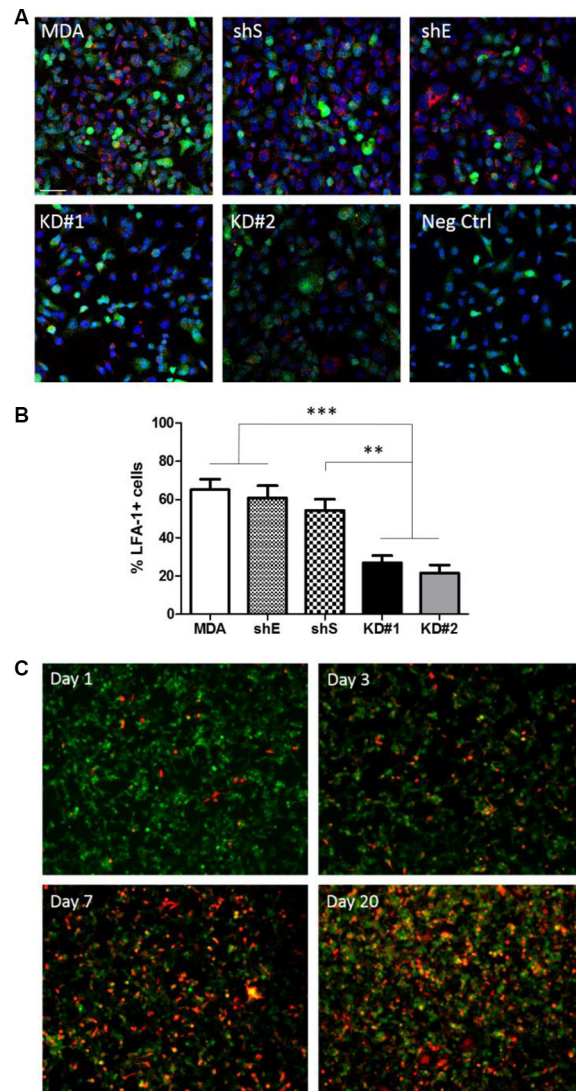
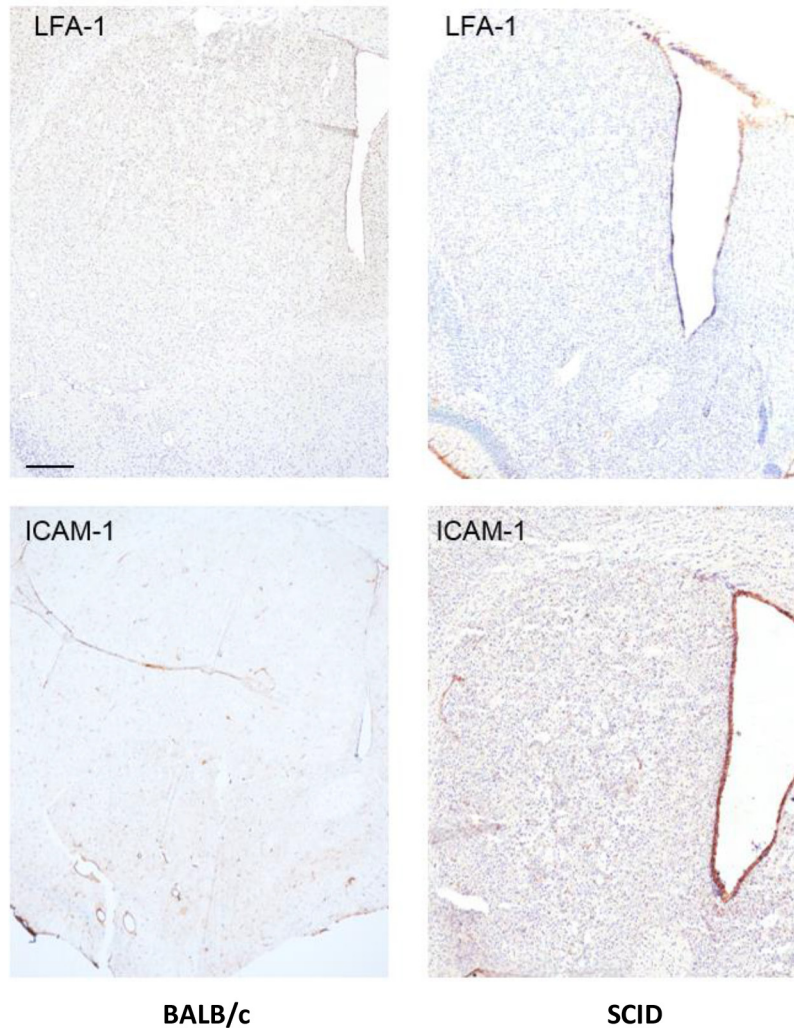


## Disruption of tumour-host communication by downregulation of LFA-1 reduces COX-2 and e-NOS expression and inhibits brain metastasis growth

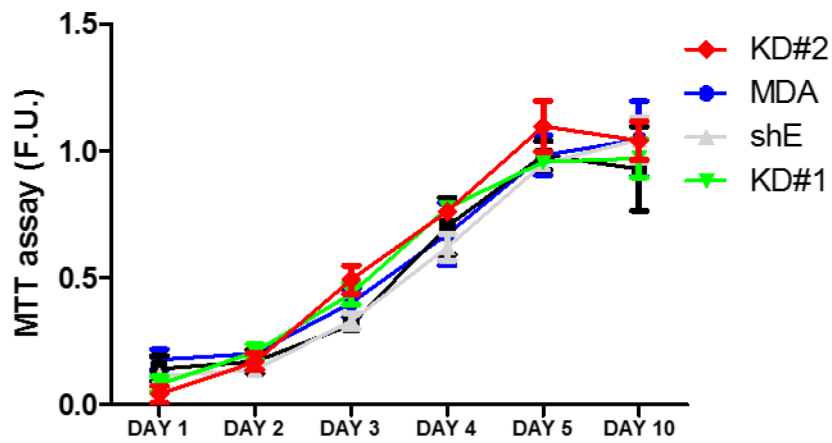
### Supplementary Materials



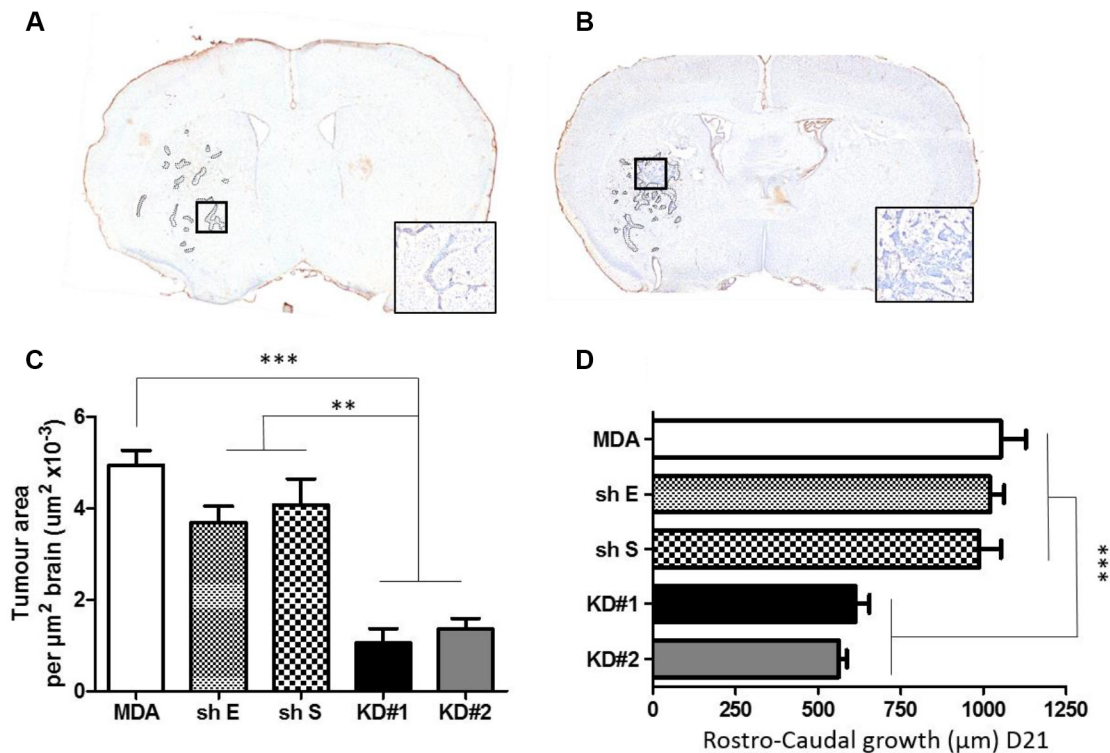
**Supplementary Figure S1: (A) Assessment of cell surface expression of LFA-1.** Cells were coated for 30 min with anti-LFA-1 antibody and then fixed for further staining. Immunofluorescent (red; Cy3 fluorophore) expression was evident on the cell surface of all the different clones used *in vivo* (MDA, shS and shE), but to a much lesser extent in the LFA-1 knock-down clones (KD#1 and KD#2). **(B)** Quantitation of LFA-1 positive cells per field of view; 4 different ROIs with > 300 cells were quantified for each group. **(C)** Assessment of shRNA transfection stability in MDA231Br-GFP cells (green). The shRNA plasmid against LFA-1 was coupled with red fluorescent protein (RFP, red) and puromycin resistance. Presence of the shRNA plasmid was determined at days 1, 3, 7 and 20 after the onset of transfection. The cells showed stable presence of the shRNA plasmid (red and green co-localisation) from day 7 to day 20 post-transfection. For the *in vivo* studies, transfected cells were injected intracerebrally at day 7 after transfection and tumour growth assessed 14 days later ( $\approx$  day 20 post-transfection). These data suggest, therefore, that the cells should maintain stable plasmid transfection throughout the *in vivo* time course studied.



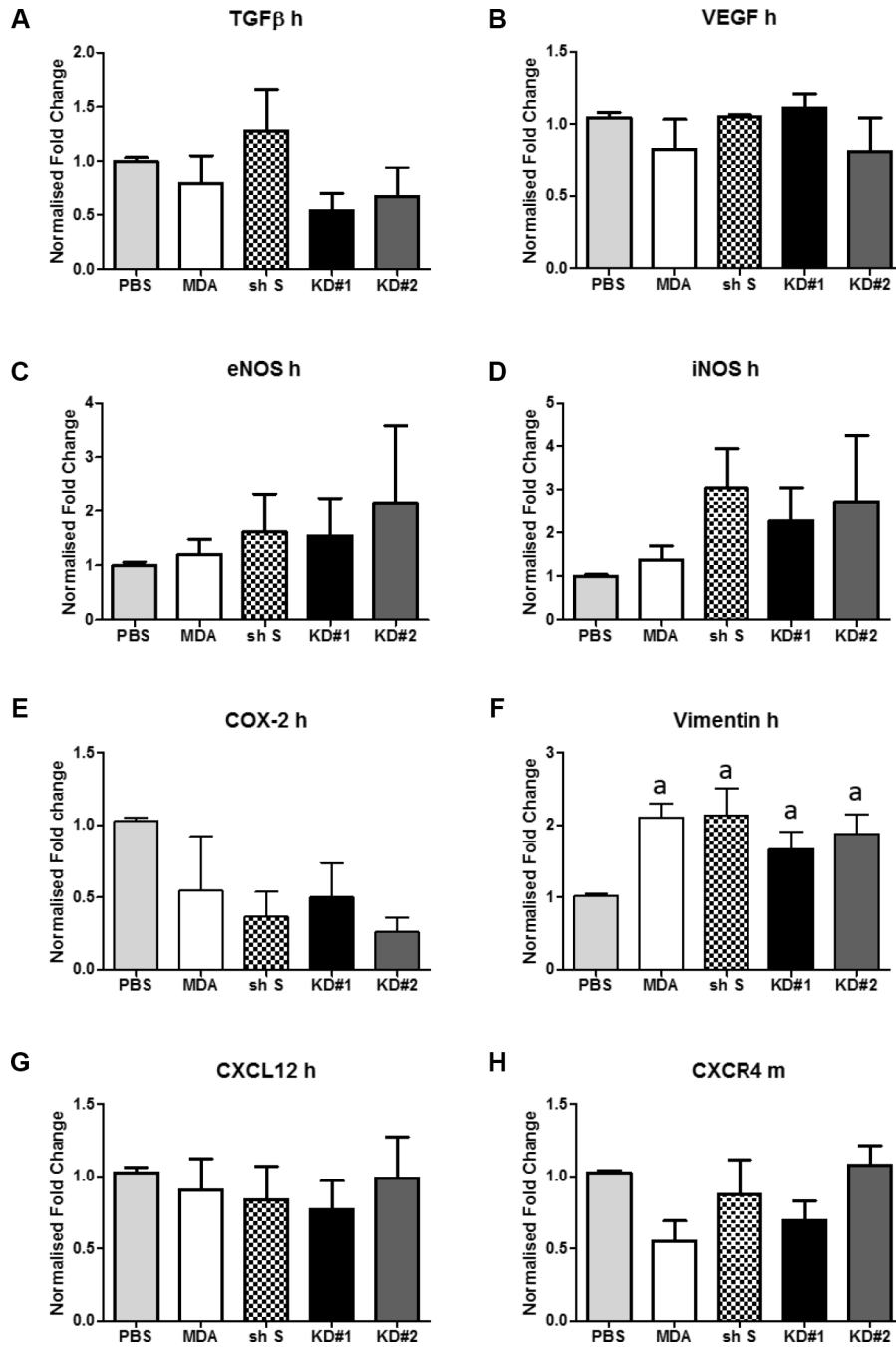
**Supplementary Figure S2: Photomicrographs from the left striatum of BALB/c (left panels) and SCID (right panels) mice 10 and 14 days post-injection with PBS, respectively.** Sections were stained against LFA-1 (top) and ICAM-1 (bottom). Basal levels of expression of both CAMs were significantly lower than the experimental groups and largely undetectable. Scale bar = 300  $\mu$ m.



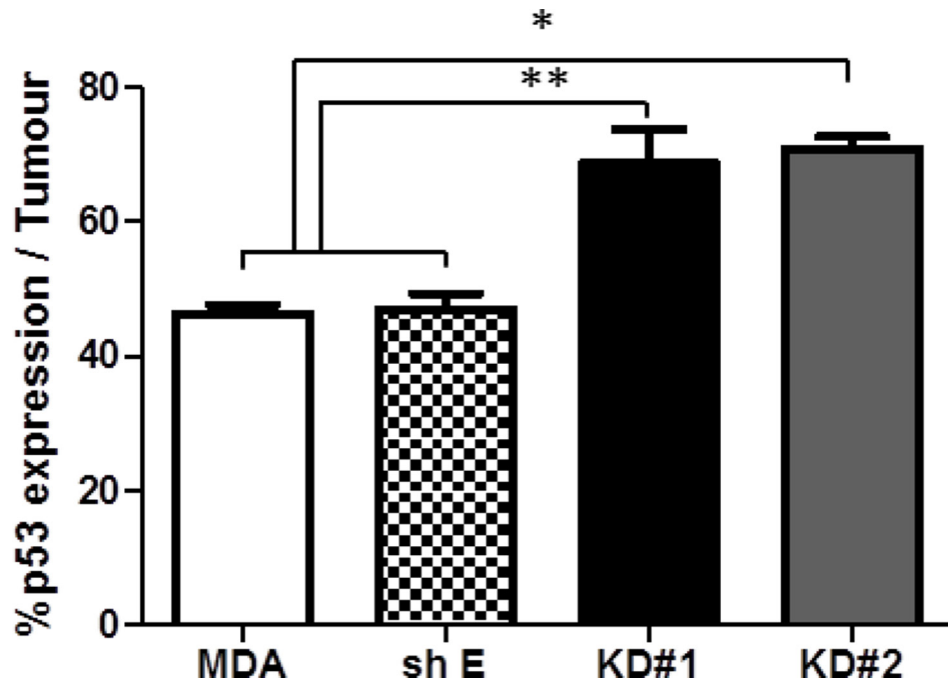
**Supplementary Figure S3: MTT assay study of four different MDA231BR-GFP clones used in the *in vivo* study.** No differences in cell activity were found between the parental cells and the LFA-1 knock-down clones.



**Supplementary Figure S4: (A–B)** Photomicrographs of SCID mice 21 days after injection of KD#1 (a) and MDA (b) tumour cells, respectively. (C) Following the previous study of tumour growth at day 14 after tumour induction (Figure 3), quantitation of tumour growth at a later time point (day 21) was performed in animals injected intrastriatally with parental MDA231Br-GFP cells (MDA), control knock down cells (empty cassette, shE; and scramble cassette, shS) or LFA-1 knock down cells (KD#1 and KD#2) ( $n = 4$  per group). Statistical significance was determined by one-way ANOVA, with Tukey's post-hoc tests.  $**p < 0.01$ ;  $***p < 0.005$  (D) Quantitation of rostro-caudal tumour growth throughout the striatum in the same animals as for (C). Statistical significance determined by one-way ANOVA, with Tukey's post-hoc tests.  $***p < 0.005$ .

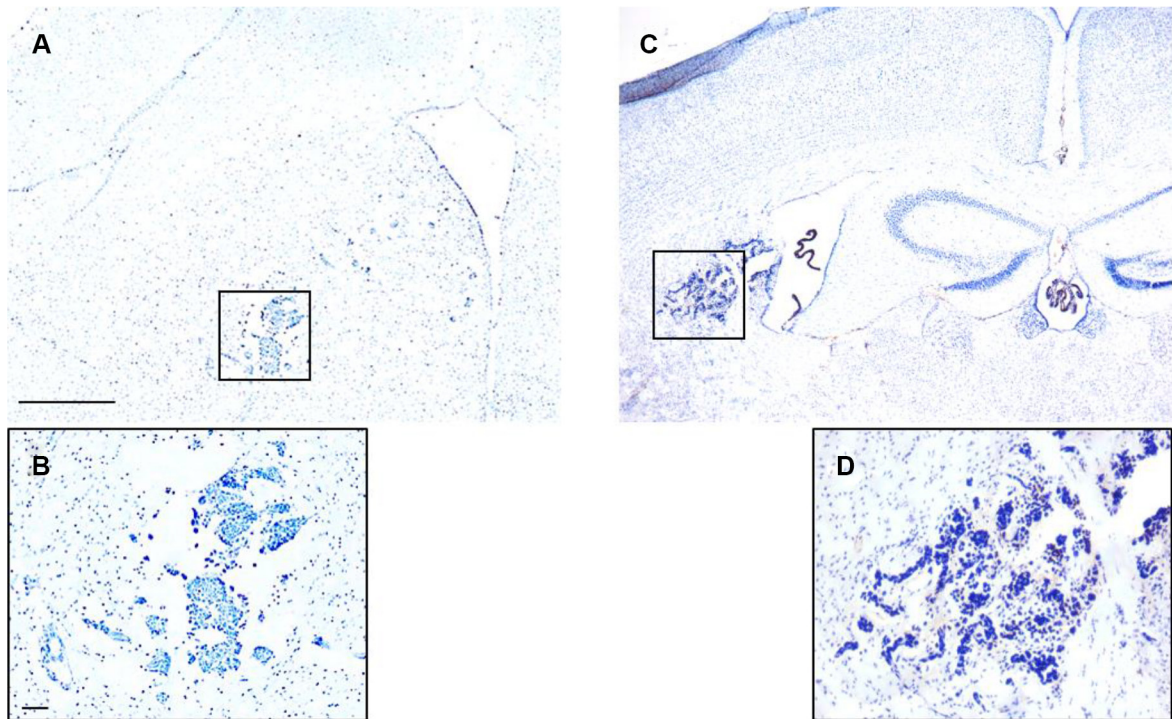


**Supplementary Figure S5: Quantitative reverse transcription PCR analysis of (A–G) human and mouse (H) gene expression in the striatum of mice injected with either PBS or one of the MDA231Br-GFP clones ( $n = 5$  per group). Statistical significance determined by one-way ANOVA, with Tukey post-hoc tests. Multiple Comparison Test: a= significant compared to the PBS group.  $p < 0.05$**



Supplementary Figure S6: Quantitation of p53 expression levels within tumour colonies 21 days after intracerebral injection of MDA231Br-GFP cells (MDA), control knock-down cells (shE) or LFA-1 knockdown cells (KD#1 and KD#2). Statistical significance was determined by one way ANOVA followed by Tukey's post-hoc tests; \* $p < 0.05$  and \*\* $p < 0.01$ .





**Supplementary Figure S7: MDA231Br-GFP cells injected into the left striatum of the brain (A), or left ventricle of the heart (C) in SCID mice.** Insets show a similar pattern of growth in the tumour colonies induced either by direct intracerebral injection (B) or via haematogenous dissemination to the brain (D). Scale bar 500  $\mu\text{m}$  and 50  $\mu\text{m}$  (inset)