

**Table 1. Primer description and thermal profiles for PCR amplification of *amoA***

Primer pair*	Sequence (5'-3')	Round of nested PCR	Thermal profile <sup>†</sup>	Molecular analysis
<b>A189 (1)</b>	<b>GGNGACTGGGACTTCTGG</b>	1	94°C, 30 s	T-RFLP
amoA-2R (2)	CCCCTCKGSAAAGCCTTCTTC		62-52°C, <sup>‡</sup> 30 s 72°C, 45 s 30 cycles	
<b>amoA-1F (2)</b>	<b>§GGGGTTTCTACTGGTGGT</b>	2	94°C, 60 s	
amoA-2R	CCCCTCKGSAAAGCCTTCTTC		55°C, 90 s 72°C, 90 s 18 cycles	
<b>A189</b>	<b>GGNGACTGGGACTTCTGG</b>	1	94°C, 45 s	Real-time PCR <sup>  </sup>
amoA-2R	CCCCTCKGSAAAGCCTTCTTC		56°C, 90 s 72°C, 180 s 13 cycles	
<b>amoA-1F</b>	<b>GGGGTTTCTACTGGTGGT</b>	2 (Real-time PCR)	94°C, 45 s	
amoA-2R	CCCCTCKGSAAAGCCTTCTTC		55°C, 30 s 72°C, 180 s Plate read at 83°C 30 cycles Melting curve <sup>¶</sup>	

\*Primers in bold represent the 5'-primer.

<sup>†</sup>All PCR profiles started with an initial denaturation at 94°C for 3 min and ended with a final elongation step at 72°C for 10 min, prior to holding the temperature at 4°C.

<sup>‡</sup>Annealing temperature was decreased 0.5°C after each cycle until it reached 52°C.

<sup>§</sup>Primer labeled with 5-carboxyfluorescein.

<sup>¶</sup>Melting curve analysis: 65-98°C, 0.2°C per read, 1-s hold, final elongation step at 72°C for 10 min, prior to holding the temperature at 10°C.

<sup>||</sup>Control reactions with the primers M13F (5'-GTAAAACGACGGCCAG-3') and M13R (5'-CAGGAAACAGCTATGAC-3') were performed with identical thermal profiles.

1. Holmes, A. J., Costello, A., Lidstrom, M. E. & Murrell, J. C. (1995) *FEMS Microbiol. Lett.* **132**, 203-208.

2. Rotthauwe, J., Witzel, K. & Liesack, W. (1997) *Appl. Environ. Microbiol.* **63**, 4704-4712.