

# Supporting Information

## Cellulose Structural Polymorphism in Plant Primary Cell Walls Investigated by High-Field 2D Solid-State NMR Spectroscopy and Density Functional Theory Calculations

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**Table S1.** Structural parameters of non-equivalent glucose units used in the DFT calculation. U and D annotate the two non-equivalent glucose units within the same glucan chain in the I $\alpha$  allomorph, while C and O denote to center and origin chains of the I $\beta$  allomorph. Apostrophe denotes the atoms in the adjacent glucose residue.

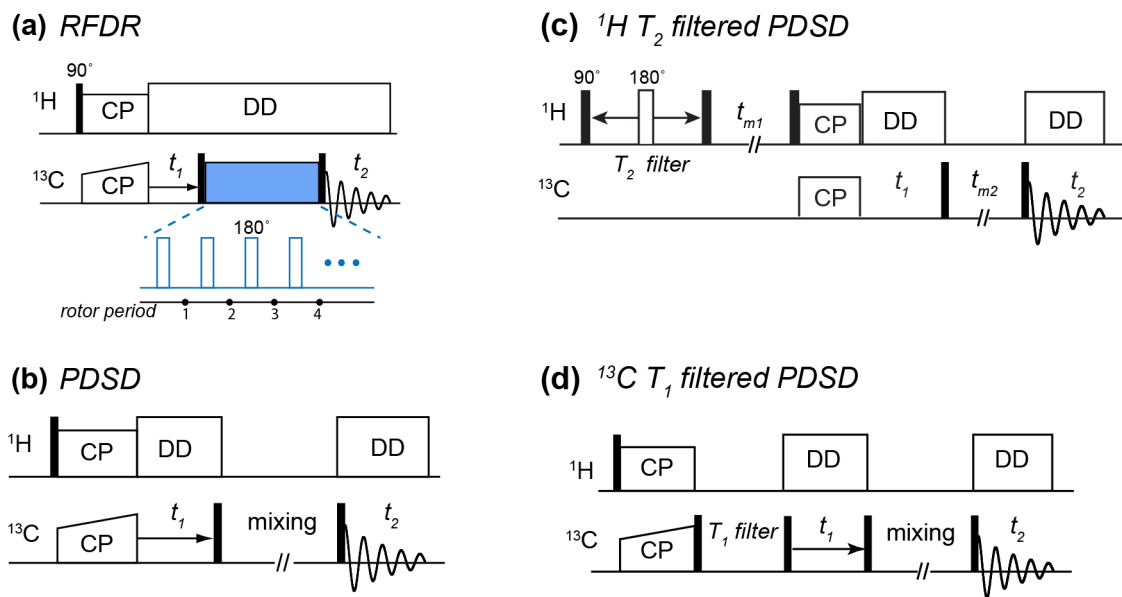
		H---C distance (Å)		H---H distance (Å)			Torsion angles (°)					
		H <sub>6a</sub> -C <sub>4</sub>	H <sub>6b</sub> -C <sub>4</sub>	H <sub>6a</sub> -H <sub>4</sub>	H <sub>6b</sub> -H <sub>4</sub>	H' <sub>1</sub> -H <sub>4</sub>	$\theta$ (H <sub>6a</sub> -C <sub>6</sub> •C <sub>4</sub> -H <sub>4</sub> )	$\theta$ (H <sub>6b</sub> -C <sub>6</sub> •C <sub>4</sub> -H <sub>4</sub> )	X <sub>1</sub> (O <sub>5</sub> -C <sub>5</sub> -C <sub>6</sub> -O <sub>6</sub> )	X <sub>2</sub> (C <sub>4</sub> -C <sub>5</sub> -C <sub>6</sub> -O <sub>6</sub> )	$\Phi$ (C <sub>5</sub> -C <sub>4</sub> -O <sub>4</sub> -C <sub>1</sub> )	$\Psi$ (C <sub>4</sub> -O <sub>4</sub> -C <sub>1</sub> '-O <sub>5</sub> ')
I $\beta$	C	2.7	3.5	2.4	3.7	2.1	0.7	114.2	167.9	-74.3	-143.2	-92.7
	O	2.7	3.5	2.5	3.7	2.2	-6.6	106.7	165.1	-76.4	-145.0	-94.2
I $\beta$	C	2.8	2.8	3.0	2.5	2.1	-101.9	6.3	58.1	178.1	-145.1	-93.2
	O	2.9	2.8	3.2	2.6	2.1	-110.8	-2.2	52.1	170.4	-150.7	-92.2
I $\beta$	C	3.5	2.8	3.7	3.0	2.1	137.8	-99.6	-63.9	57.2	-140.5	-86.9
	O	3.5	2.9	3.8	3.3	2.1	115.5	-115.9	-67.3	54.0	-155.7	-85.4
I $\alpha$	U	2.7	3.5	2.4	3.6	2.1	-3.3	109.3	165.3	-76.8	-143.1	-92.6
	D	2.7	3.5	2.4	3.7	2.2	-5.7	107.8	165.2	-76.0	-145.6	-95.0
I $\alpha$	U	2.8	2.7	3.1	2.5	2.2	-109.6	-2.6	58.4	177.0	-156.1	-89.4
	D	2.7	2.9	2.9	2.7	2.2	-96.1	12.8	77.2	-163.9	-155.9	-85.4
I $\alpha$	U	3.5	2.8	3.7	3.1	2.2	130.4	-107.4	-62.7	57.8	-140.0	-81.3
	D	3.5	2.8	3.8	3.2	2.2	118.4	-116.4	-67.9	52.1	-160.2	-92.9

**Table S2.** Estimated fractions of different glucose conformations in crystalline I $\alpha$  (A and A') and I $\beta$  (B and B') allomorphs. The estimation is based on spectral deconvolution results of Kono and Numata<sup>1</sup>, with an additional assumption that A and A' have equal populations.

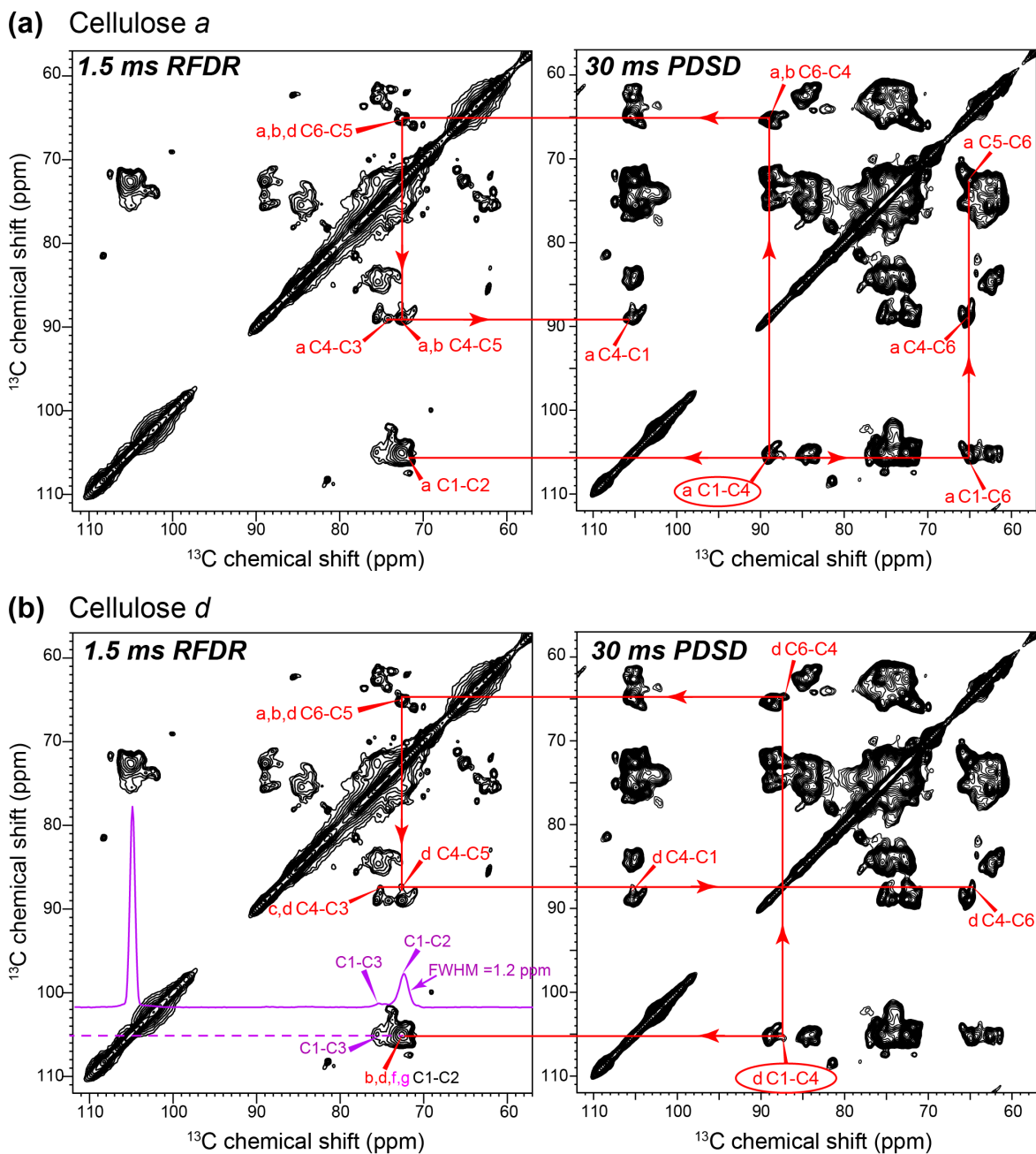
Cellulose	Organism	A	A'	B	B'
I $\alpha$ dominant	<i>Cladophora</i>	32%	32%	17%	18%
I $\beta$ dominant	tunicate	11%	11%	38%	41%

**Table S3.** <sup>13</sup>C chemical shift RMSDs (ppm) between measured primary wall cellulose and DFT calculated chemical shifts for different crystalline allomorphs, hydroxymethyl conformations, and glucose residue location. C and O denote center and origin sheets in the I $\beta$  allomorph while U and D denote the two non-equivalent glucose units in an I $\alpha$  chain. The five types of crystalline cellulose signals observed in this study are denoted as *a*, *b*, *c*, *d*, *e*.

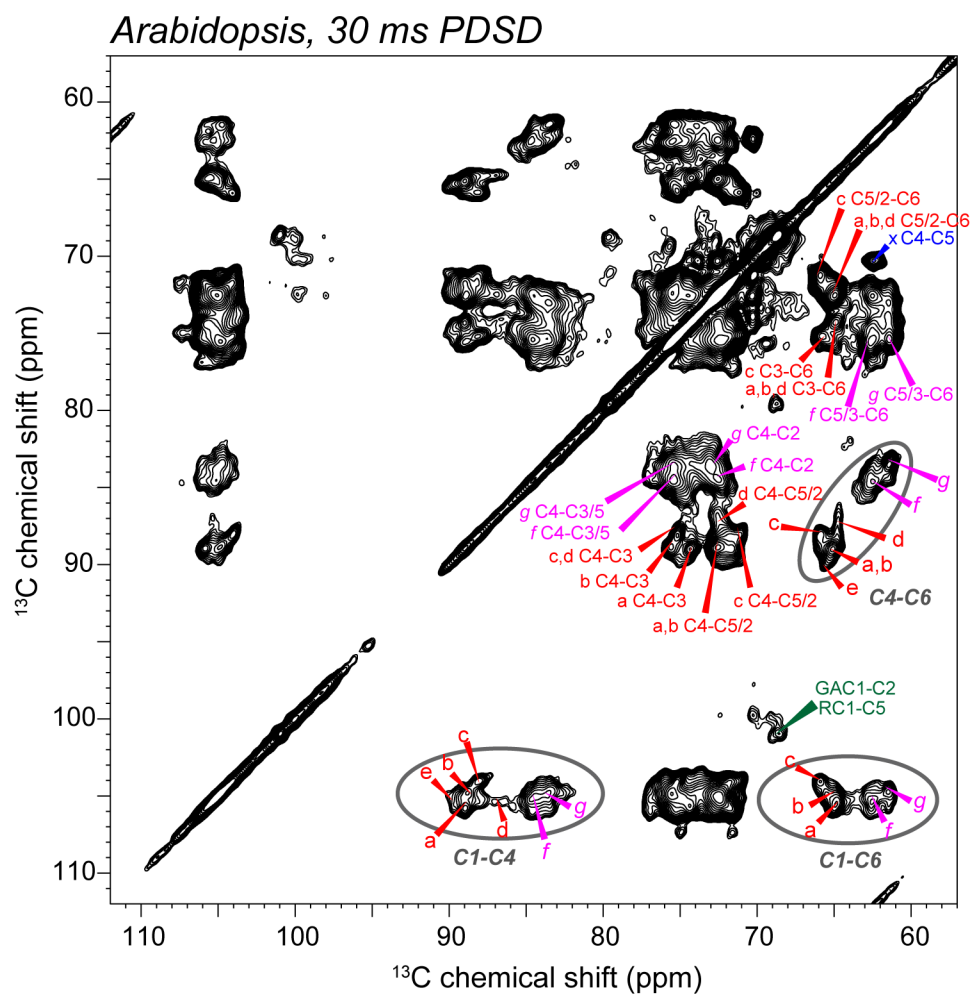
		<i>a</i>	<i>b</i>	<i>c</i>	<i>c</i>	<i>e</i>
I $\beta$ allomorph	gg_C	5.5	5.3	5.2	4.7	5.8
	gg_O	4.5	4.5	4.4	3.8	4.9
	gt_C	4.3	4.0	3.9	3.6	4.3
	gt_O	2.3	2.5	2.5	1.9	2.7
	tg_C	2.4	2.6	2.0	2.1	2.4
I $\alpha$ allomorph	tg_O	1.9	2.3	2.0	1.8	2.1
	gg_U	5.3	5.2	5.0	4.5	5.6
	gg_D	5.6	5.5	5.5	4.9	6.0
	gt_U	4.3	4.3	4.4	3.6	4.7
	gt_D	4.1	3.8	3.7	3.5	4.2
	tg_U	2.0	2.2	1.9	1.7	2.2
	tg_D	2.1	2.5	2.0	2.2	1.9



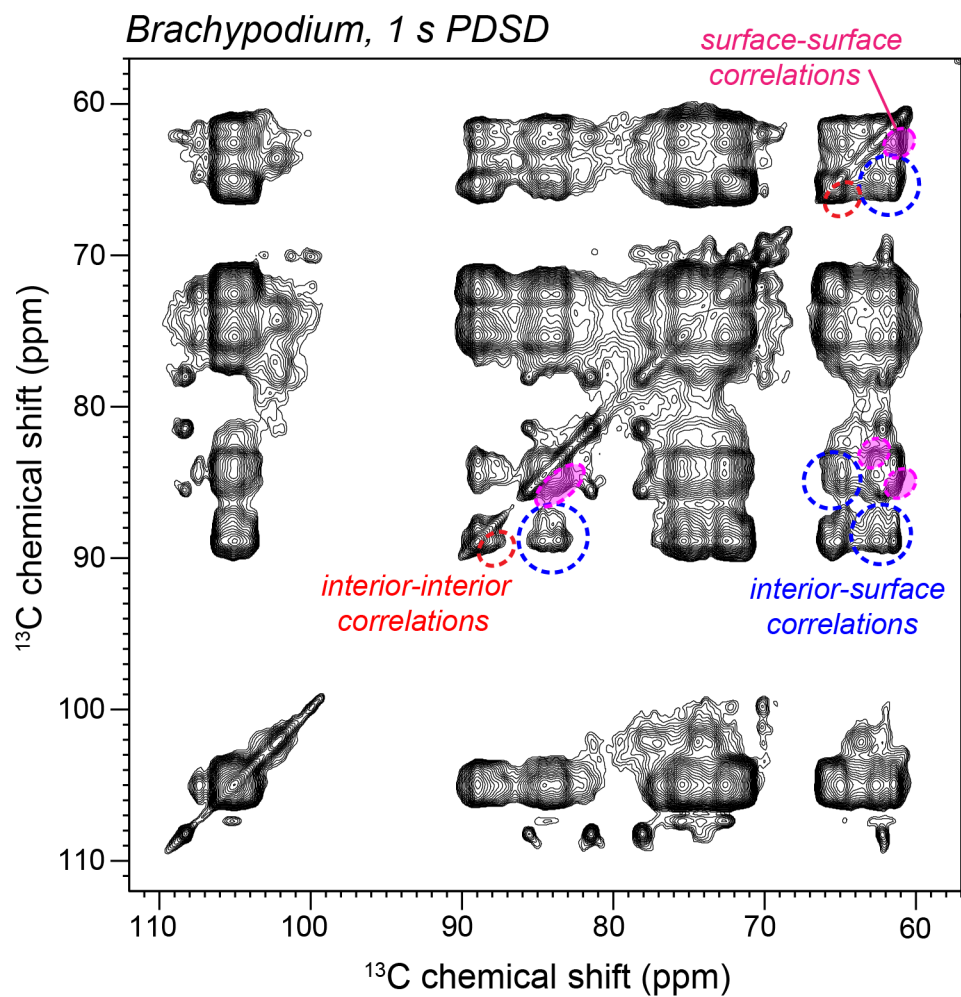
**Figure S1.** Different 2D  $^{13}\text{C}$ - $^{13}\text{C}$  correlation pulse sequences used to measure cellulose chemical shifts, structure, and dynamics. (a) Radio-frequency-driven recoupling (RFDR) experiment. (b) Proton-driven  $^{13}\text{C}$  spin diffusion (PDS) experiment. (c)  $^1\text{H}$   $T_2$  filtered PDS experiment for determining the water contact of wall polysaccharides. (d)  $^{13}\text{C}$   $T_1$  filtered PDS experiment for measuring the  $^{13}\text{C}$   $T_1$  relaxation rates of wall polysaccharides.



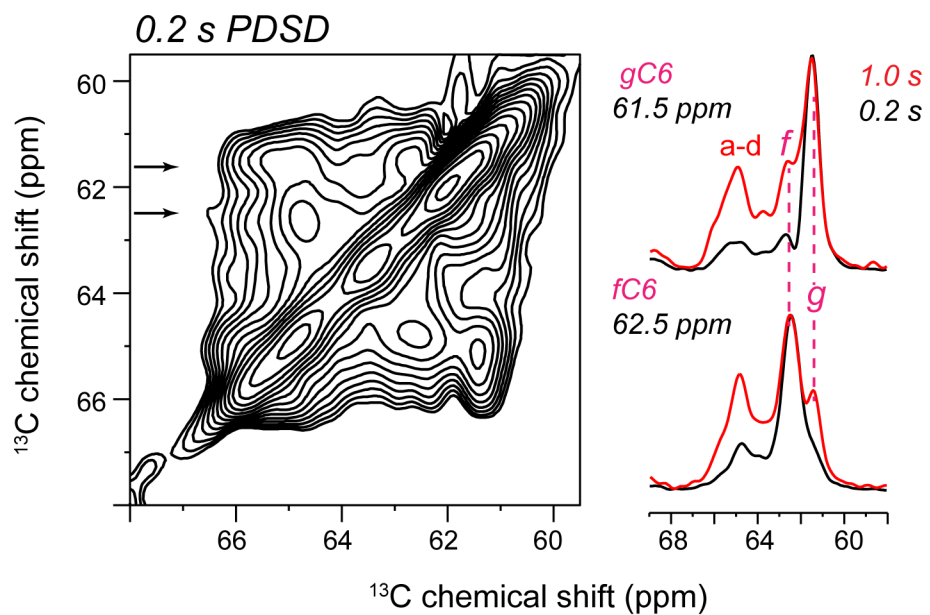
**Figure S2.** Assignment of (a) cellulose *a* and (b) cellulose *d*  $^{13}\text{C}$  chemical shifts of the *Brachypodium* cell wall using 2D RFDR and PDSM spectra. The C1-C4 cross peaks serve as the starting points for assignment.



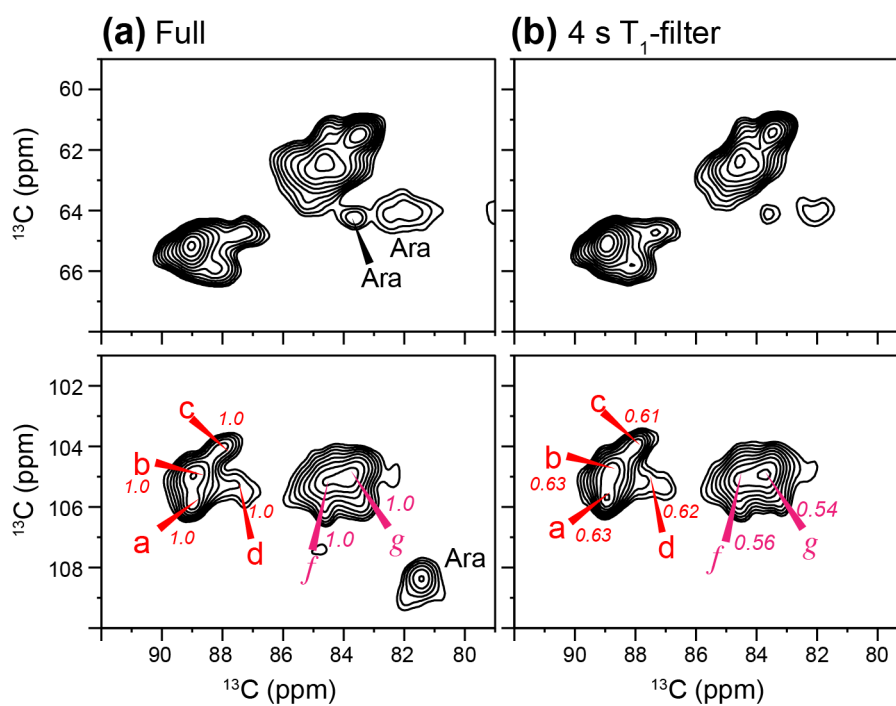
**Figure S3.** 2D  $^{13}\text{C}$ - $^{13}\text{C}$  PDSD spectrum of the *Arabidopsis* cell wall. The spectrum was measured with 30 ms mixing at 296 K on an 800 MHz spectrometer. The characteristic C1-C4, C1-C6 and C4-C6 cross peaks are highlighted.



**Figure S4.** 2D PDSD spectrum of the *Brachypodium* cell wall with 1 s mixing. The spectrum was measured at 296 K on an 800 MHz spectrometer. Regions of long-range cross peaks are highlighted.



**Figure S5.** 0.2 s PDSD spectrum of the *Brachypodium* cell wall measured on an 800 MHz spectrometer. The surface-surface and surface-interior cellulose cross peaks show similar intensities.



**Figure S6.** <sup>13</sup>C T<sub>1</sub> filtered 2D PDSD spectra of the *Brachypodium* primary cell wall. (a) Full 2D spectrum. (b) 2D spectrum with a 4 s T<sub>1</sub> relaxation filter. The relative intensity of each cellulose cross peak is indicated. Interior cellulose signals decay to ~62% while surface cellulose signals decay to ~55%, indicating that surface cellulose is slightly more dynamic on the nanosecond timescale.

### Supplementary References

1. Kono, H.; Numata, Y., Structural investigation of cellulose I<sub>a</sub> and I<sub>β</sub> by 2D RFDR NMR spectroscopy: determination of sequence of magnetically inequivalent D-glucose units along cellulose chain. *Cellulose* **2006**, *13*, 317-326.