

Supplementary Figures:

Table 1. Affinity of hLXR α for non-fluorescent ligands determined by quenching of hLXR α aromatic amino acid fluorescence (K_d) and ligand efficiencies determined by displacement of hLXR α -bound BODIPY C16-CoA (K_i).

Ligand	Chain length: double bonds (position)	K_d (nM)	K_i (nM)
Palmitoleic acid	C16:1 (n-7)	113±64	N.D.
Palmitoleoyl-CoA	C16:1 (n-7)	46±16	N.D.
Stearic acid	C18:0	25±7	N.D.
Stearoyl-CoA	C18:0	>197	N.D.
Oleic acid	C18:1 (n-9)	45±15	N.D.
Oleoyl-CoA	C18:1 (n-9)	>68	N.D.
Linoleic acid	C18:2(n-6)	21±7	N.D.
Linoleoyl-CoA	C18:2(n-6)	>61	N.D.
Arachidonic acid	C20:4 (n-6)	31±9	N.D.
Arachidonoyl-CoA	C20:4 (n-6)	20±9	N.D.
Docosahexanoic acid	C22:6	>80	N.D.
Docosahexaenoyl-CoA	C22:6	25±3	N.D.
22 (R) Hydroxycholesterol		N.D.	1.2±0.3
25-Hydroxycholesterol		17±4	N.D.

Values represent the mean ± S.E. (n ≥ 3). ND, not determined.

Table 2. Secondary structures of hLXR α protein in the presence of fatty acids and fatty acyl-CoAs

Ligand	α -helix regular H(r)%	α -helix distort H(d)%	β -sheet regular S(r)%	β -sheet distort S(d)%	Turns T%	Unordered U%
Ethanol	13.9 ± 0.4	11.9 ± 0.2	13.3 ± 0.3	8.8 ± 0	19.6 ± 0.8	32.4 ± 1.5
C16:1	14.0 ± 1.0	11.3 ± 0.9	17.0 ± 3	10.1 ± 0.9	20.9 ± 0.5*	26.5 ± 2.4
C16:1-CoA	14.3 ± 0.6	11.6 ± 0.6	13.7 ± 1.4	9.3 ± 0.3	20.6 ± 0.6	30.2 ± 1.1
C18:0	16.4 ± 3.2	11.7 ± 0.8	10.1 ± 5.1	9.2 ± 0.4	16.9 ± 3.4	35.4 ± 4.8
C18:0-CoA	10.3 ± 1.3	9.7 ± 0.7	19.0 ± 2.4	10.4 ± 0.5	20.1 ± 1.1	30.4 ± 2.0
C18:1	14.0 ± 1	11.3 ± 0.9	17.0 ± 3	10.1 ± 0.9	20.9 ± 0.5	26.6 ± 2.4
C18:1-CoA	14.0 ± 0.2	10.9 ± 0.8	12.0 ± 6	9.0 ± 0.1	18.9 ± 0.2	35.2 ± 0.6
C18:2	14.1 ± 1.1	11.6 ± 1.2	15.0 ± 2	9.4 ± 1.1	20.9 ± 1.1	28.9 ± 1.3
C18:2-CoA	7.4 ± 0.3	8.5 ± 0.1	28.2 ± 9.8	10.8 ± 0.1	25.4 ± 0.4	19.4 ± 9.7
C20:4	13.2 ± 0.2	10.3 ± 0.1	16.1 ± 2.9	10.1 ± 0.1	21.3 ± 0.5	28.7 ± 0.2
C20:4-CoA	13.9 ± 0.6	12.1 ± 0.3	14.21 ± 0.9	8.9 ± 0.2	20.9 ± 0.3	30.0 ± 0.5
C20:5	13.1±0.1	11.6±0.2	14.8±0.8*	9.1±0.1	20.8±0.1	30.5±0.7
C22:5	15.7±1.9	12.2±0.4	10.1±4	8.3±0.6	19.8±1.5	33.8±4
C22:6	15.6±1.1	12.2±1	14±3	9.5±0.8	21±0.4	27.5±1.8
KH ₂ PO ₄	12.3±0.1	9.8±0.3	16.5±1.8	9.7±0.3	19.2±0.3	32.4±0.7
C20:5-CoA	14.2±0.7	12±0.5	14±0.1	8.8±0.4	21.1±0.5	29.9±0.1
C22:5-CoA	13.4 ± 1.1	11.6 ± 0.5	14.9 ± 1.7	9.2 ± 0.3	20.6 ± 0.6	30.2 ± 1.1
C22:6-CoA	14.3 ± 0.4	12.4 ± 0.2	13.2 ± 0.8	8.5 ± 0.1	20.6 ± 0.5	30.9 ± 0.9

Significant difference between hLXR α with solvent compared to the absence or presence of fatty acids or fatty acyl-CoA dissolved in either ethanol or in KH₂PO₄ were determined by t-test * = P<0.05, ** = P<0.01, *** = P<0.001.

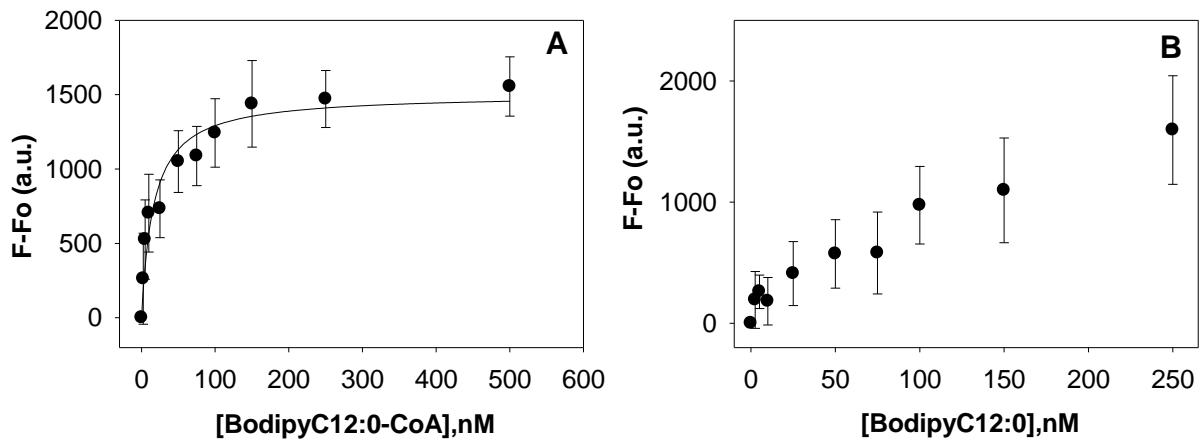


Fig. 1: Forster resonance energy transfer (FRET). Unlabeled LX α (donor) (excitation wavelength 280 nm) was titrated against increasing concentrations of BODIPY C12:0 or BODIPY C12:0-CoA (acceptor) (emission wavelength 300-540 nm). Changes in the fluorescence intensity at 341 nm wavelength were plotted as a function of ligand conc. to determine apparent dissociation constant (K_d) values.

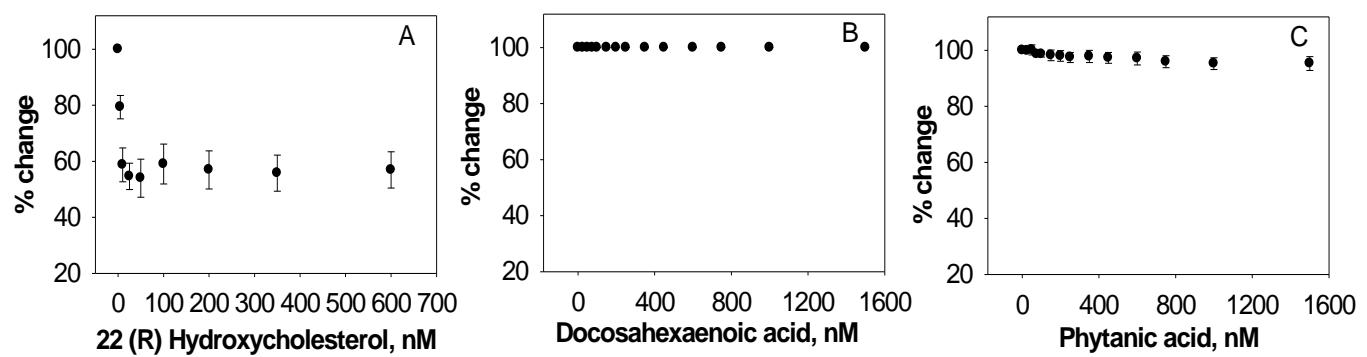


Fig. 2: Displacement assay of BODIPY C16:0-CoA bound LXR α . BODIPY C16:0-CoA bound to LXR α was displaced with LXR α endogenous ligand 22 (R) Hydroxycholesterol, but not with long chain fatty acids docosahexaenoic acid and phytanic acid.

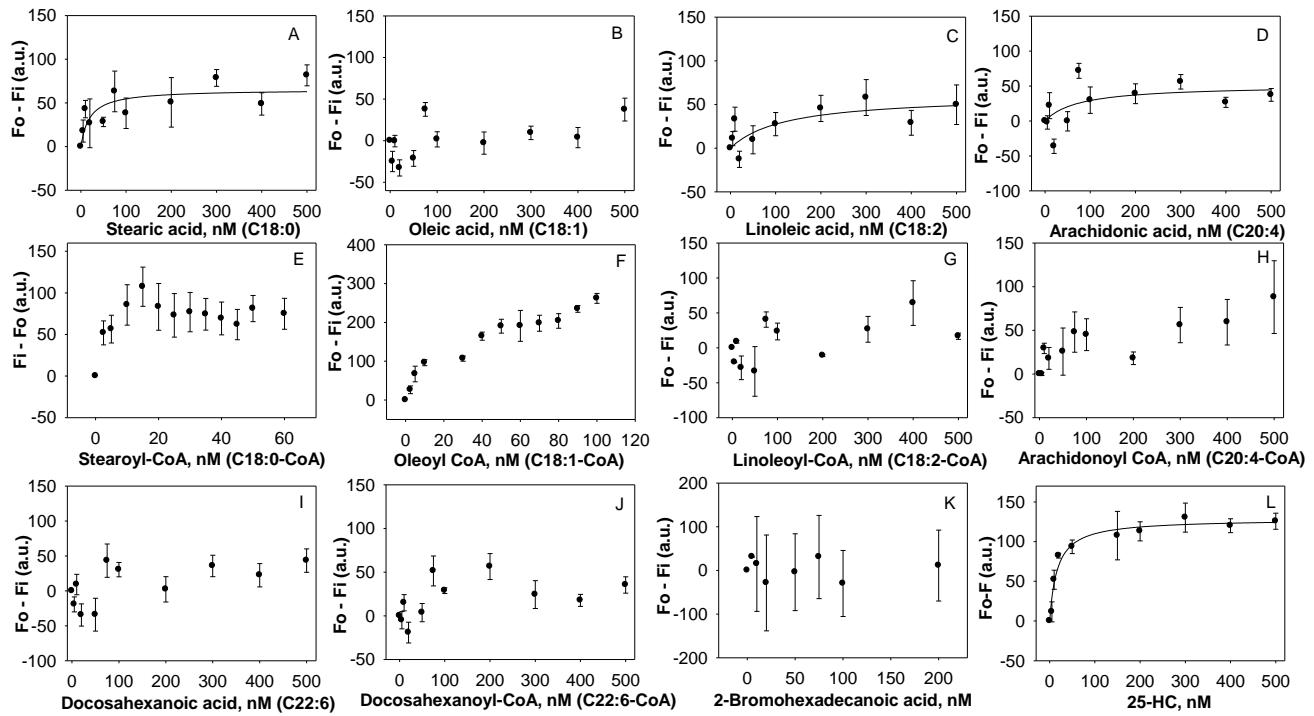


Fig. 3: Direct binding assay based on quenching of LX α aromatic amino acid fluorescence emission when titrated with the following ligands (A) Stearic acid, (B) Oleic acid, (C) Linoleic acid, (D) Arachidonic acid, (E) Stearyl-CoA, (F) Oleoyl-CoA, (G) Linoleoyl-CoA, (H) Arachidonoyl-CoA, (I) Docosahexanoic acid, (J) Docosahexanoyl-CoA, (K) 2-Bromohexadecanoic acid, and (L) 25-Hydroxycholesterol

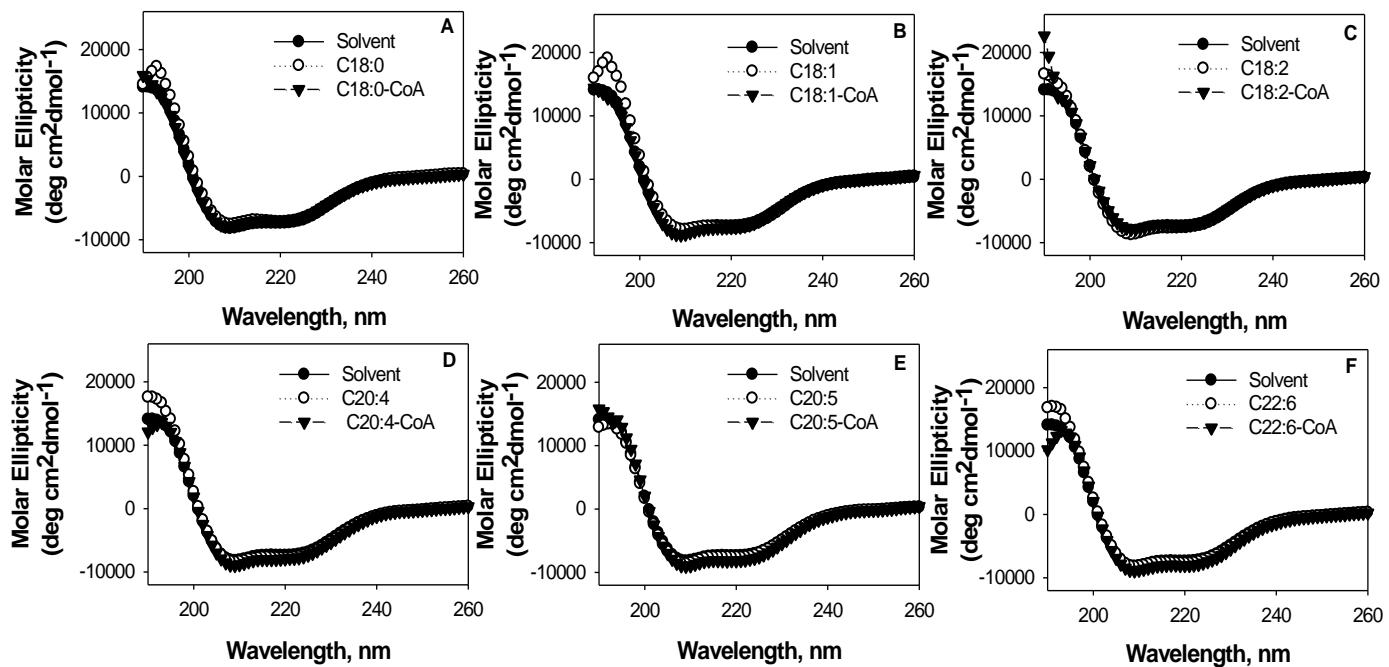


Fig. 4: Far UV circular dichroic spectra of LXR α in the absence (filled circles) and presence of added ligand: (A) C18:0 FA (open circles) or C18:0-CoA (filled triangles); (B) C18:1 FA (open circles) or C18:1-CoA (filled triangles); (C) C18:2 FA (open circles) or C18:2-CoA (filled triangles); (D) C20:4 FA (open circles) or C20:4-CoA (filled triangles); (E) C20:5 FA (open circles) or C20:5-CoA (filled triangles), and (F) C22:6 FA (open circles) or C22:6-CoA (filled triangles). Each spectrum represents an average of 10 scans for a given representative spectrum from at least three replicates.