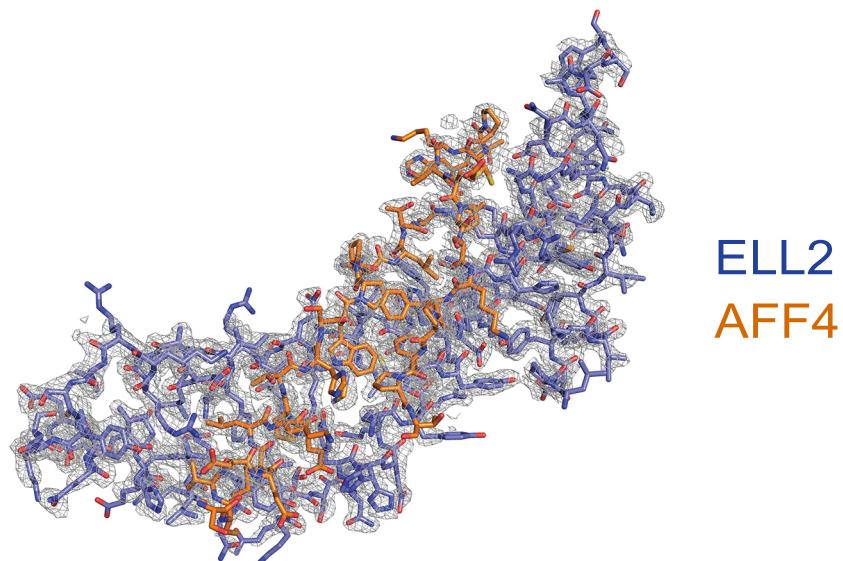
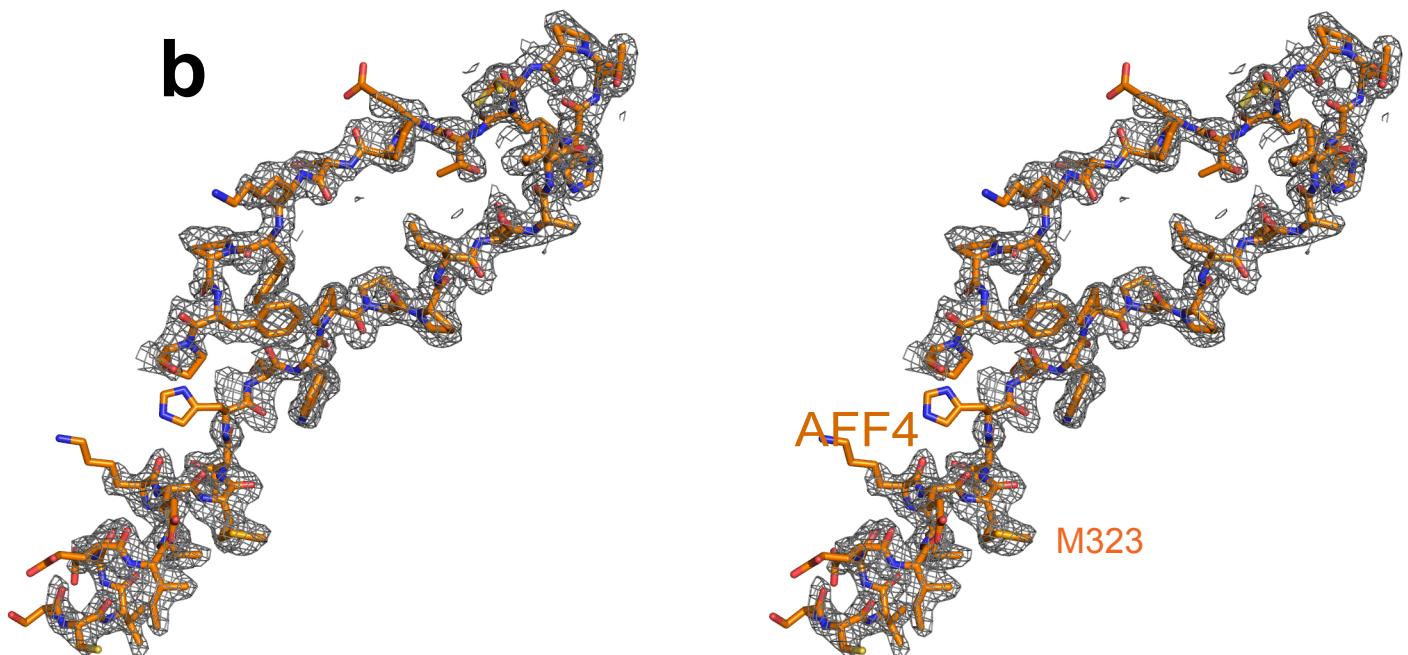
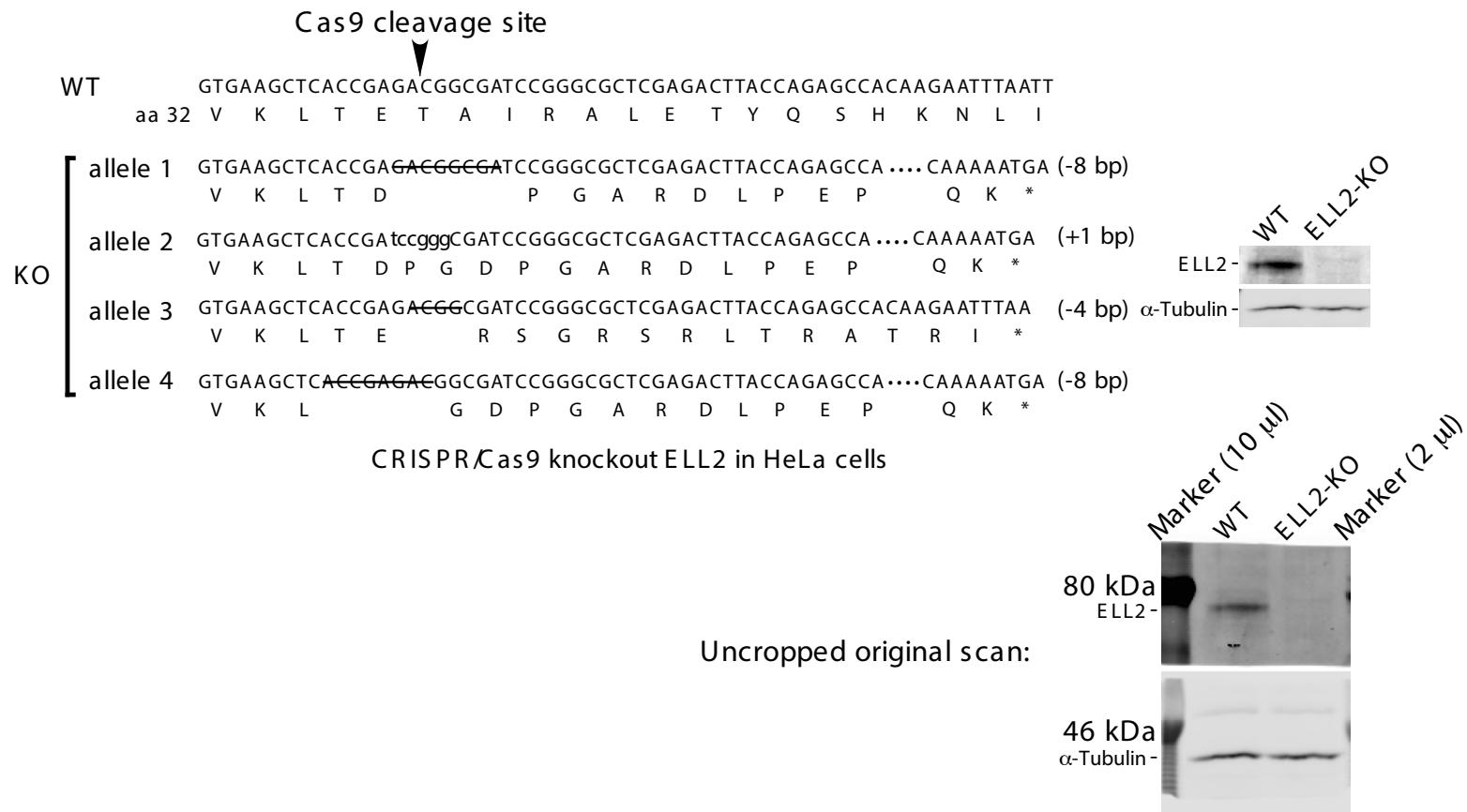


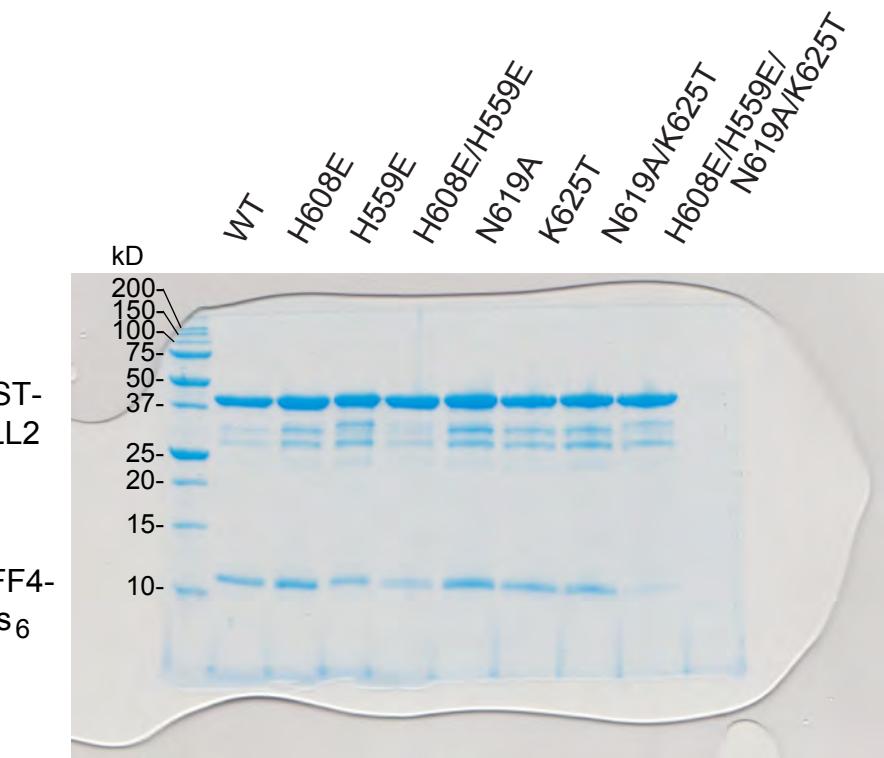
Supplementary Figure 1. a. The unfused ELL2Occ:AFF4ELLBow complex is monomeric in solution. b. The ELL2Occ-(Gly-Ser)4- AFF4ELLBow was eluted at 62.45ml on the Hiload 16/60(GE) while AFF4ELLBow-(Gly-Ser)4-ELL2Occ was eluted at 73.22 ml, which correspond to a dimer and a monomer, respectively. Red line: ELL2Occ-(Gly-Ser)4- AFF4ELLBow , Blue line: AFF4ELLBow-(Gly-Ser)4-ELL2Occ . c. Schematic of hypothesis for ELL2Occ-(Gly-Ser)4- AFF4ELLBow dimerization in solution while AFF4ELLBow-(Gly-Ser)4-ELL2Occ was monomeric. ELL2Occ is shown in blue. AFF4ELLBow is shown in orange. N, C represent amino termini and carboxyl termini respectively.

a**b**

Supplementary Figure 2. a. The overall experimental electron density map after density modification is displayed at a contour level of 2σ (gray), with AFF4ELLBow and ELL2Occ shown as stick in orange and light blue, respectively. b. Stereo view of the portion of the map from (A) corresponding to AFF4ELLBow, displayed at a contour level of 2σ (gray), with AFF4ELLBow shown in a stick model. Met323 is highlighted.

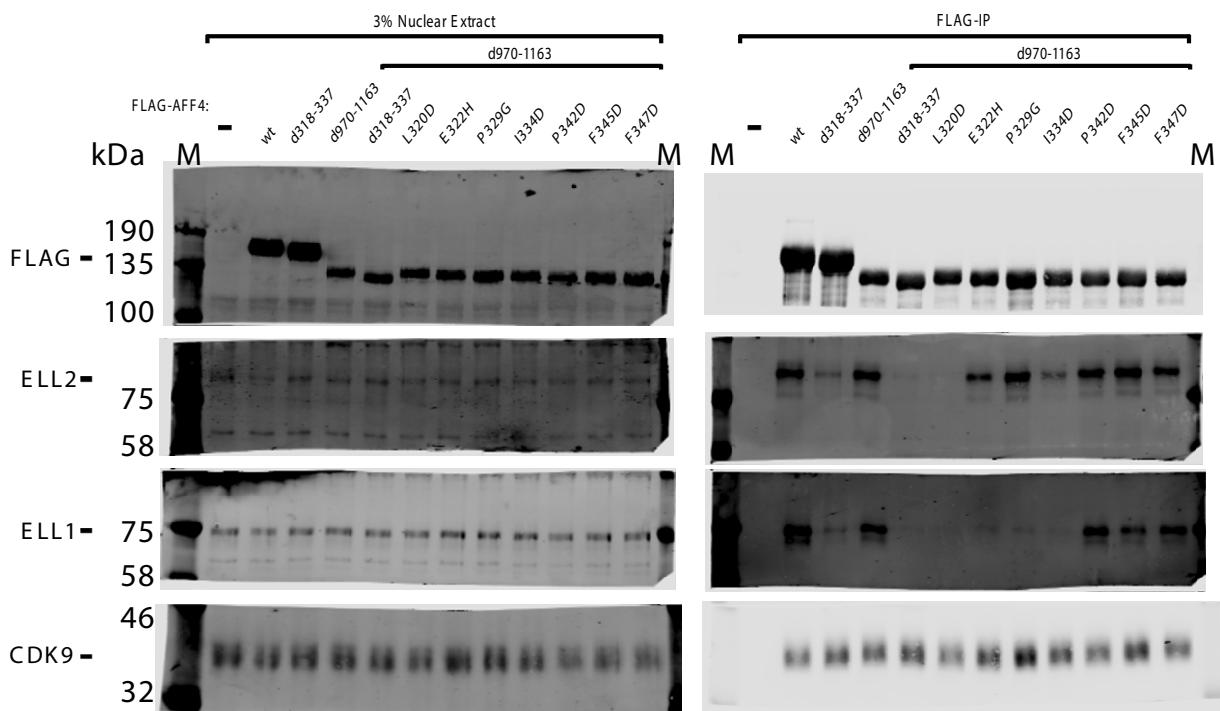


Supplementary Figure 3. Verification of disruption of the ELL2 gene by CRISPR-Cas9 in the HeLa-derived ELL2 knockout cell line Δ ELL2. DNA and predicted amino acid sequences surrounding the intended Cas9 cleavage site (arrowhead) in the wild-type ELL2 gene and its mutant alleles generated by CRISPR-Cas9 are shown. Insertions of extra nucleotides are indicated by lowercase letters, deletions are indicated by capital letters containing strike-throughs, and the omitted nucleotides are marked by four consecutive dots. Premature stop codons as a result of frame-shift mutations are indicated by stars. The loss of ELL2 protein expression in Δ ELL2 was confirmed by immunoblotting with the indicated antibodies.

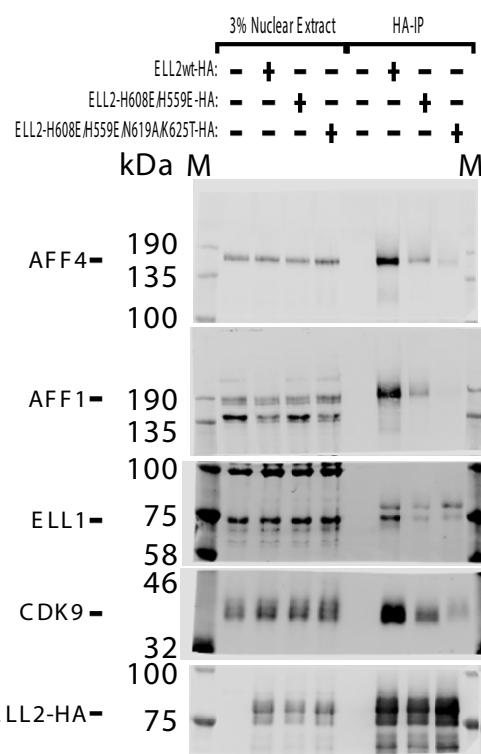


Supplementary Figure 4. Uncropped version of gel shown in Figure 5c.

A

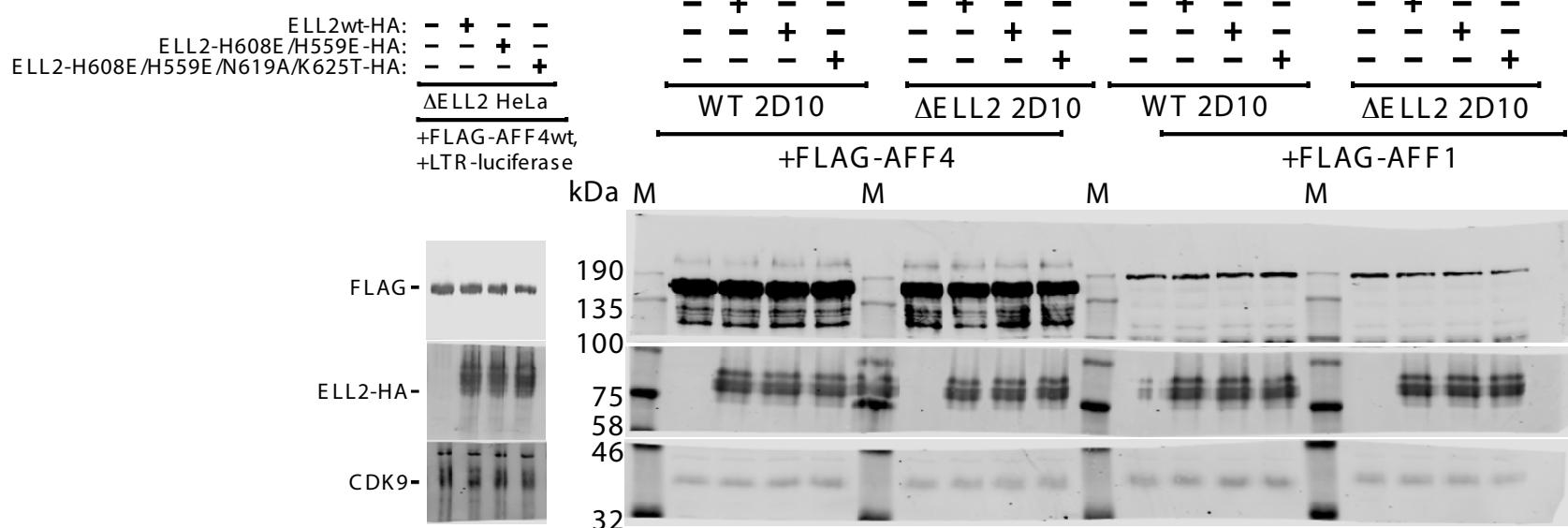
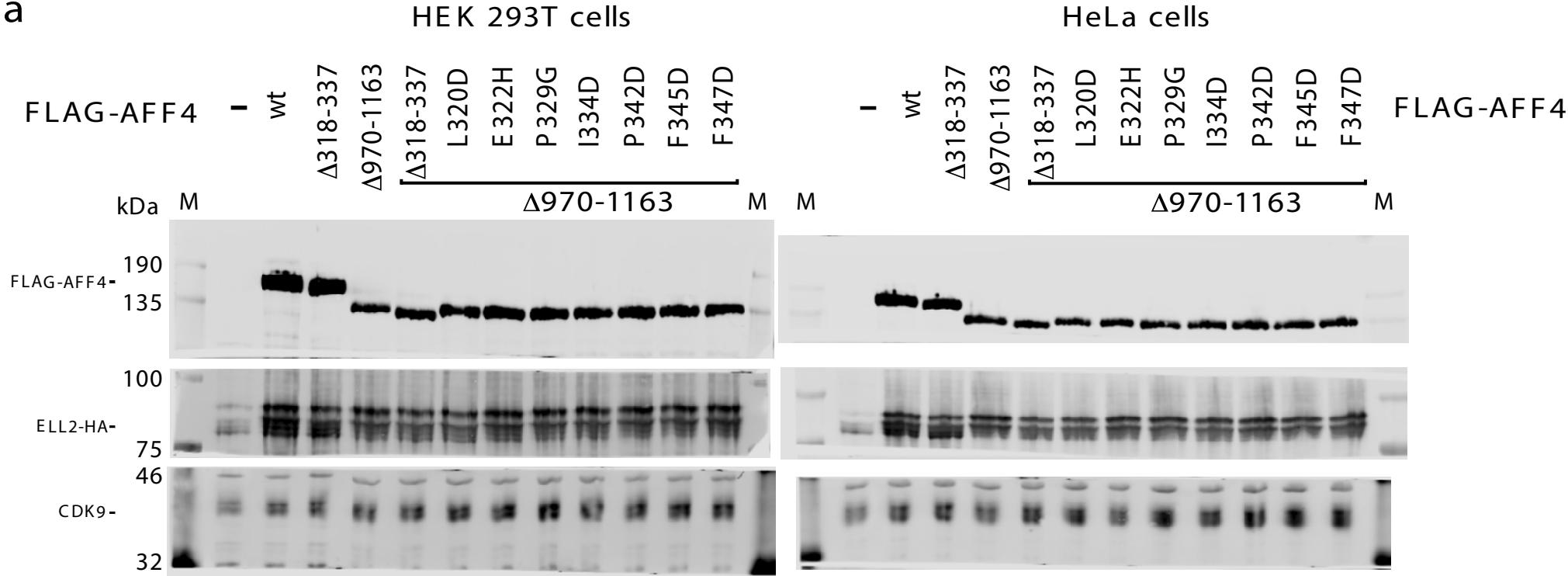


B



Supplementary Figure 5. Uncropped versions of the gels shown in Figure 6.

a



Supplementary Figure 6. Uncropped versions of the gels shown in Figure 7.

Supplementary Table 1. K_d values for AFF4-ELL2 binding as determined by fluorescence anisotropy. Fluorescence anisotropy data in Figure 5a, b were fitted with a single site binding model. NA means fitting was not applicable because of very low affinity binding.

AFF4 construct/mutation	K_d (μM)	R^2
WT, 300-350 +His ₆ -tag	0.086 + 0.024	0.97
WT, 318-350 peptide	4.0 + 1.57	0.91
V316D	0.23 + 0.048	0.99
I319D/L320D	5.4 + 1.87	0.92
M323D	2.7 + 1.14	0.88
D317P/E318P	0.80 + 0.18	0.99
K321P	0.24 + 0.06	0.98
E322H	0.13 + 0.033	0.99
W327D	0.37 + 0.086	0.96
P328G/P329G	0.27 + 0.098	0.97
L331D	1.6 + 0.31	0.98
T332K	0.069 + 0.037	0.97
I334D	0.67 + 0.21	0.95
L331D/I334D	1.9 + 0.51	0.97
P342D	0.2 + 0.05	0.99
F345D/F347D	0.13 + 0.058	0.97
I319D/L232D/M323D	NA	
D317P/E318P/E322H	NA	
M323D/L331D/I334D	NA	
W327D/ L331D/I334D	NA	

Supplementary Table 2. Primers used for PCR and mutagenesis in this study.

PCR for Recombinant expression		
Gene	Forward Primer	Reverse Primer
ELL2(519-640)	GTTGTTGGATCCGAACCTTCAGCAATTGAAC	CCTTCACTCGAGTTATGCTGGTGTCAAATT
AFF4(300-350)	GTGTTGGATCCCCCTCCCAACCCTG	CCTTCACTCGAGTTACTCCTAGTTGGAAAAG
ELL2(519-640)-4GS-AFF4(300-350)	CAAGCAGACTCATGGTCCGgttagtggtagttgttagtgc	CAGTGGTTGGGAAGGAcactaccactaccactaccGG
	TTCCAACCCTG	ACCATGACTCTGCTTG
AFF4(300-350)-4GS-ELL2(519-640)	CTTTCCAACAATAGGAGgttagtggtagttgttagtgc	CAATTGCTGAAGGTTActaccactaccactaccCT
	CCTCAGCAATTG	CCTTAGTTGGAAAAG
mutations		
ELL2		
V593D	CATGAAGAAGaaCTTACAAGAATATC	TTCTTGTAAGCTTCTTCATGAAC
H608E	CCCAATTACgaaGAAGAAAAATAC	TTTTCTTCttcGAATTGGGACTAG
F548E	AAGGTGACgaCAATGCAGAGTATG	CTCTGCATGtcGTCATCCTATAATT
Y555E	TAIGATGAGGaaCAGAGCTTGCATG	CAAAGCTCTGtcCTCATCATACTC
H559E	AGAGCTTGGaaGCCAGGTGGAG	CATCTGGCttcAAAGCTCTGTAC
D632A	GAATTGcaAACAGCAAGCAG	CTGTTGtgCAAATTACACCTATTAG
N619A	CTTCATgcCAAGCTGGCACATC	CAGCTTGgcATGAAGATATTACATC
K625T	CACATCacAAGGATGATAGGT	CATCCTTgtGATGTGAGGCCAGCTTG
AFF4		
V316D	TGTGAGCTGTGatGATGAAATC	TTTCATCatCACAGCTCACATC
I319/320D	ATGAAgaCgtAAAGAGATGAC	TCTTtatGATTCATCCACACAG
M323D	CTAAAGAGGatACGCATTATG	TGAATGCGTateCTCTTTAGGATITC
317D318E	AGCTGTGTGccTccAACTCTAAAGAGATG	TTTTAGGATTggAggCACACAGCTCAC
K321P	GAAATCCTAccAGAGATGACGGCATC	CGTCATCTTggTAGGATTTCATC
E322H	ATCCCTAAAcacATGCCATTATG	ATGGCTCATgttTTAGGATTTC
W327D	ACGGATTCAgtCTCCCCCTCTAAC	AGGGGGAGGAtcTGAATGGCTCATCTC
L331D	CCTCCCCCTgtatACGGTATTCTAC	AATAGCGTateAGGGGGAGGCCATG
D317A	TGTGTGGtGAAATCTTAAAC	GATTTCAGCACACAGCTCACATC
T332K	CCTCTAAaGGGTATTCTACACCATG	AATAGCCTTAGAGGGGGAGGCCATG
I334D	CTAACGGGTgtATCACACCATG	TGGTGTATGAtcAGCGTTAGAG
P342D	AAAACAGAAgaTCCAAATITC	AAATTGGAAtcTTCTGTTTGCATG
P3282930G	CATTCATGGgtGcgTCTAACGGCTATTCTCATGGgg	AGCCGTTAGAccGecAccCCATGAATGCGTCCGTT
	TggCggTCTAACGGCTATTTC	AGAccGecAccCCATGAATGGGTATC
F345347D	CCCTCCAAAGaTCTGtaTCCAACTAAGGAG	CTTAGTTGGAtcAGGAtcTTGGAGGTT
V316D/I319/320D/M323D/317D318E	CTGTGatGATGAGacgtAAAGAGgtACGCATTATG	TGAATGCGTateCTCTTategtcTTCATCatCACAGC
	GTGGATGAAgagatAAAGAGgtACGCATTATG	TCACATCATGCGTateCTCTTateGcTTCATCCAC
	TC	ACAGCTCAC
L331D/I334D	CCCTgtatACGGCTgtatCATAACCATG	GTATGAtcAGCCGtateAGGGGGAGGCCATG
PCR for <i>In Vivo</i> Assay		
Gene	Forward Primer	Reverse Primer
AFF4-d318-337	GCTCTGGTGAATGGAGCTGTGGATTGCAAAC	GGTTCTGTTTGCATCCACACAGCTC
	AGAACCTTCCAAATTCTTTTC	
AFF4-d970-1163	GAATCCAATCCCCATTCCCTATGtaTCGAGCATGC	CTCGATCACATAGGAAATGGGG
	ATCTAGAGGGC	
AFF4-L320D	ATGTGAGCTGTGGATGAAATC GAC AAAGAGATG	TCATCTTTGTCGATTTCATCCAC
	ACGGATTATGCC	
AFF4-E322H	AGCTGTGTGGATGAAATCTAAACACATGACGCA	TGCGTCATGTGTTAGGATTTCATC
	TTCATGGCCTCCC	
AFF4-P329G	AGAGATGACGCAATTCTGGCT GT CCTCTAACGGC	CGTTAGAGGACCAGGCCATGAATG
	TATTCATACACC	
AFF4-I334D	GGCCCTCCCCCTCTAACGGCT GAC CATACACCATGCA	ATGGTGTAGGTAGCCGTTAGAG
	AAACAGAACCC	
AFF4-P342D	GCTATTCTACACCATGCAAACAGAA GACT CAA	AAAGGAAATTGGAGTCTCTGTTTG
	ATTTCCTTTCAACTAAGG	
AFF4-F345D	CCATGCAAACAGAACCTCCAAAG GAC CTTTCC	GTTGGAAAAGGGTCTTGAAGGTT
	AACTAAGGAGTCTCA	
AFF4-F347D	CAAAACAGAACCTCCAAATTCT GAC CCAAC	TCCTTAGTGGGTAGGAAATTGG
	AGGAGTCTCAGCAGT	
ELL2-N619A/K625T	GAAAAATACAGATGTGAATATCTCATgcaAAGCTGG	GTGATGTGAGCCAGCTTGCATG
	CTCACATCaccAGGCTAATAGGTGAATTGACCAAC	
ELL2-H559E	GAGTATGATGAGTACAGACTTGGagGCCAGGATG	CATCTGGCCTCCAAAGCTCTG
	GAGACTGTAGCTAG	
ELL2-H608E	GATAAAGCAGTCTAGTCCAAATTACgagGAAGAAA	GTATTTCTCCTCGTAATTGGACTAG
	ATACAGATGTGAATATCTTC	