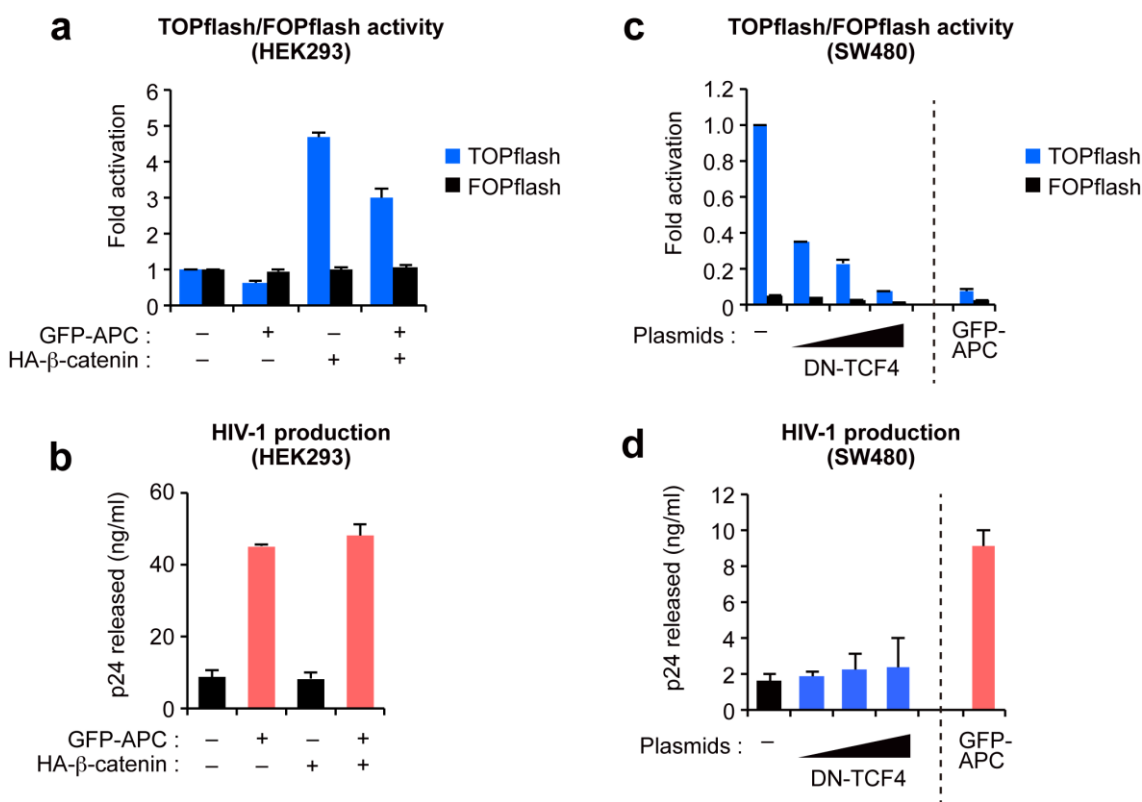


## Supplementary Figure 1

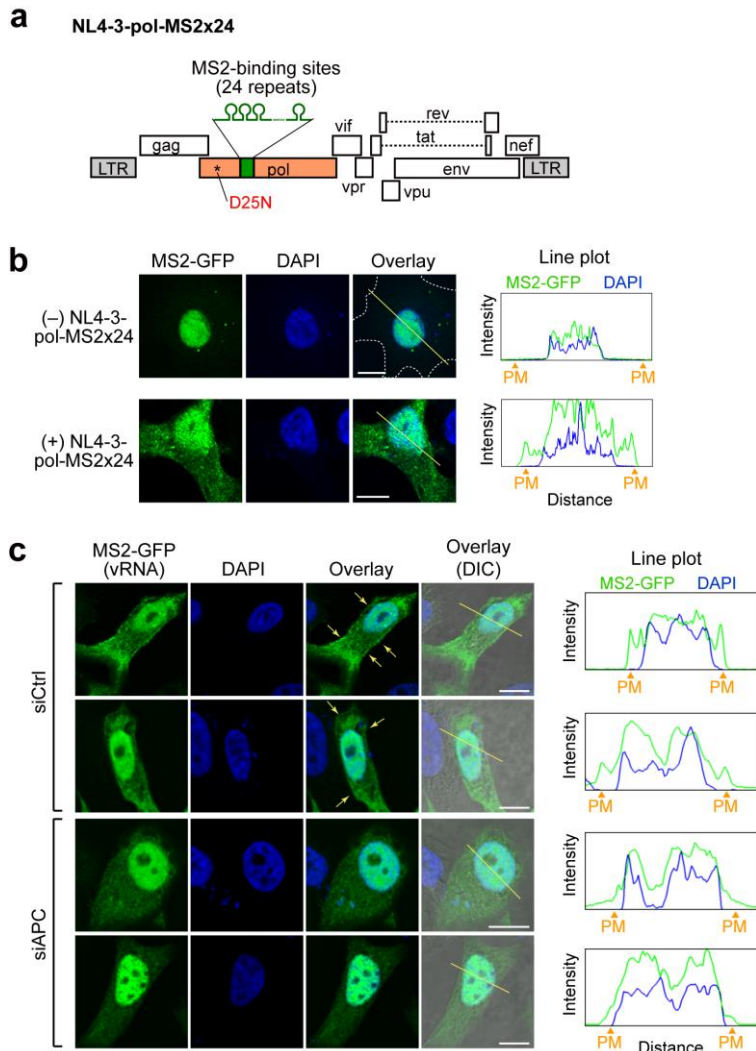


### Supplementary Figure 1. Wnt/ $\beta$ -catenin signaling does not affect APC function in HIV-1 production.

(a, b) TOPflash (TCF4-luc) reporter assay (a) and HIV-1 production assay (b) of HEK293 cells expressing  $\beta$ -catenin and APC. FOPflash vector (TCF4-mut-luc) was also used as a negative control. HEK293 cells were co-transfected with pNL4-3 and GFP-APC in the presence or absence HA-tagged  $\beta$ -catenin. The bar chart in (b) indicates the viral p24 antigen levels in the culture supernatants. Note that the expression of  $\beta$ -catenin had no observable effects on APC-mediated HIV-1 production while it significantly increased TCF4-dependent transcriptional activity.

(c, d) SW480 cells were transfected with either TOPflash or FOPflash and co-transfected with different amounts of dominant-negative (dn)TCF4 or GFP-APC, followed by a gene reporter assay (c). SW480 cells were co-transfected with pNL4-3 together with different amounts of dnTCF4 or GFP-tagged APC. The bar chart in (d) indicates the viral p24 antigen levels in the culture supernatants. Note that the expression of dnTCF4 in SW480 cells (APC-mutated colorectal cancer cells) had no observable effects on the HIV-1 production, whereas it significantly reduced TCF4-dependent transcriptional activity. All graphs are presented as a mean  $\pm$  s.d. ( $n = 3$ ).

## Supplementary Figure 2



### Supplementary Figure 2. vRNA labeling system used in this study.

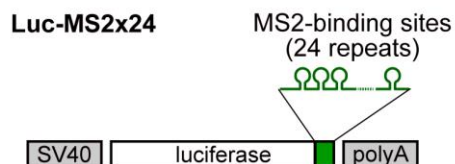
(a) Schematic representation of the HIV-1 gene tagged with twenty-four repeats of MS2-binding sites (MBS) used in this study. MBS were inserted into a Pol gene with no protease activity due to a D25N mutation in the protease region.

(b) HeLa cells were co-transfected with the MS2-GFP expression plasmid with or without pNL4-3-pol-MS2x24. Cells were stained with anti-GFP antibody to highlight MS2-GFP signals, and the nuclei were counterstained with DAPI (blue). The dotted line denotes the cell border. Scale bar, 10  $\mu$ m. Line plots in the right panels indicate the fluorescence intensity of MS2-GFP (green) and DAPI (blue) within regions of the plasma membrane (PM).

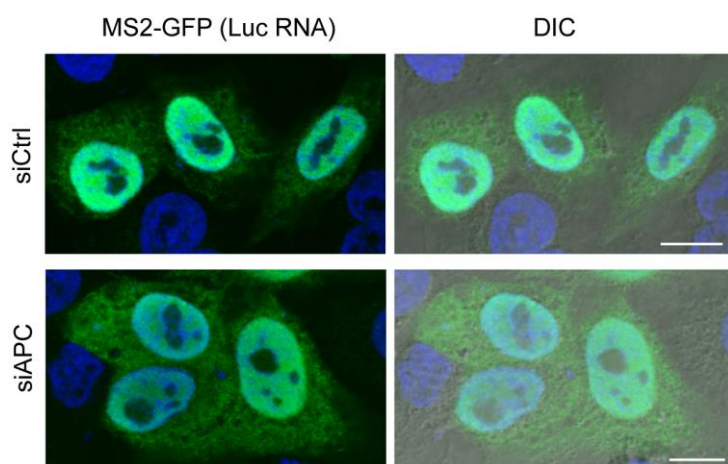
(c) Other representative example of cell images shown in Figure 6d.

### Supplementary Figure 3

**a**



**b**

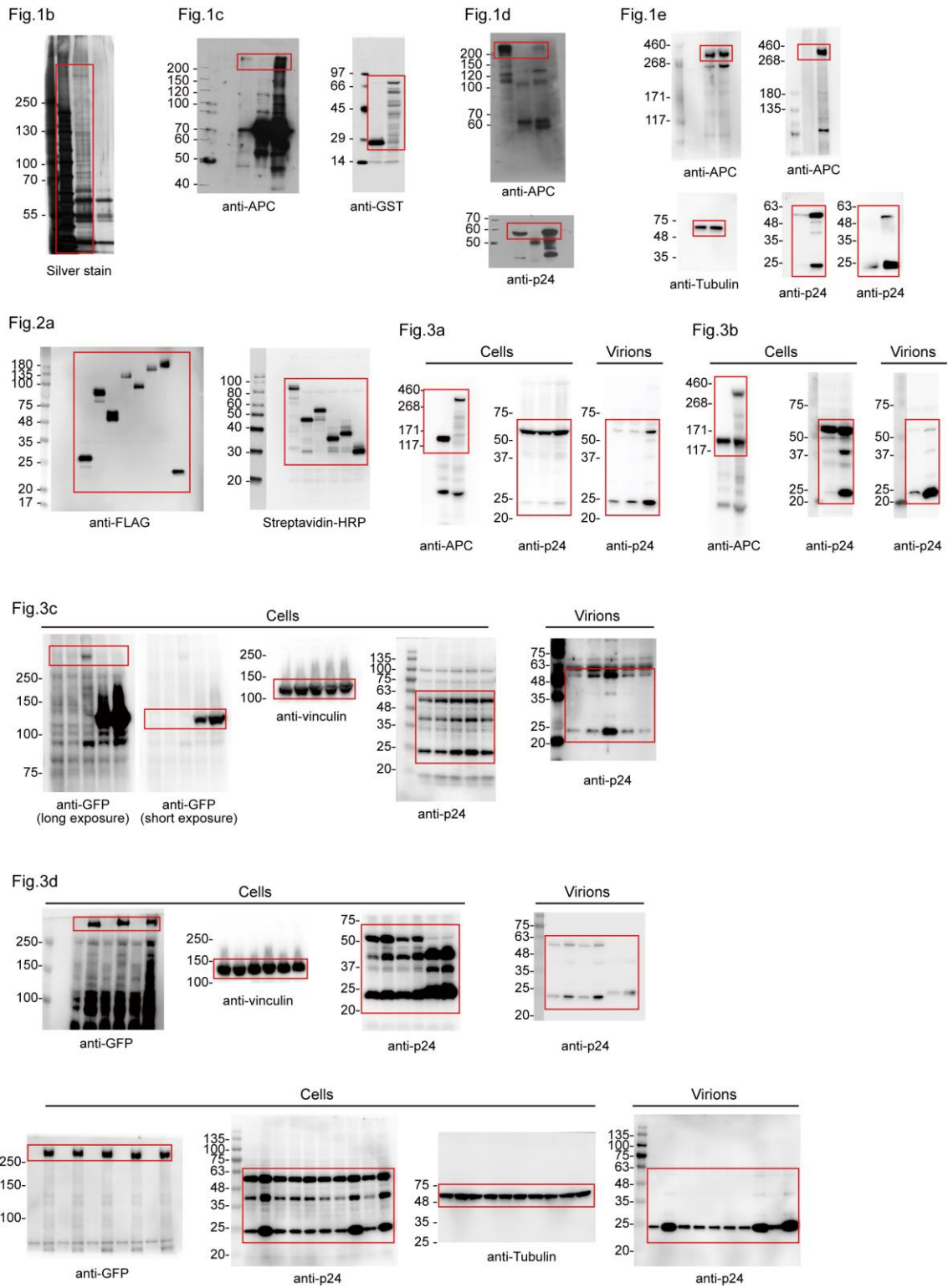


#### Supplementary Figure 3. APC does not affect non-retroviral RNA.

(a) Schematic representation of a non-viral (Renilla luciferase) RNA construct tagged with twenty-four repeats of MS2-binding sites (MBS). MBS were inserted at the end of luciferase gene of pGL4.73 (SV40-driven luciferase expression vector, obtained from Promega).

(b) HeLa cells were treated with control siRNA (siCtrl) or APC-targeted siRNA mix (siAPC) for 24 h prior to co-transfection with a pLuc-MS2x24 and MS2-GFP expression plasmid. Cells were stained with anti-GFP antibody and with DAPI (blue). Scale bar, 10  $\mu$ m.

## Supplementary Figure 4



Supplementary Figure 4. Full images of the immunoblots presented in Figures 1-3.

## Supplementary Figure 5

Fig.4a

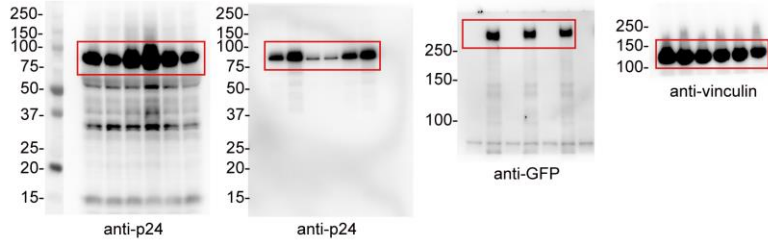


Fig.5a

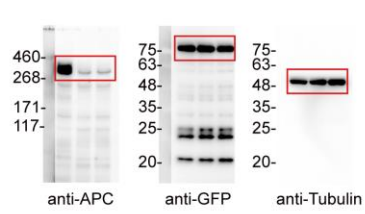


Fig.5c

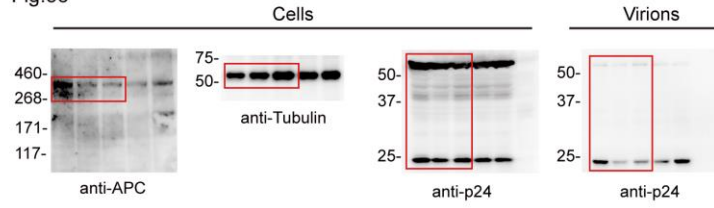


Fig.5d,e

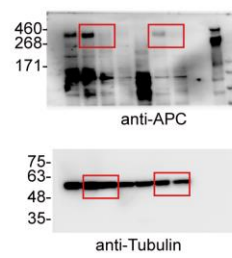


Fig.5d

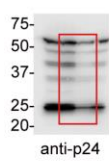


Fig.5e

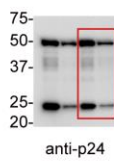
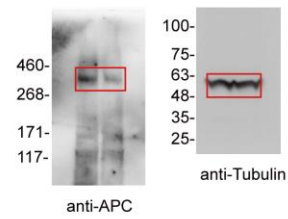


Fig.7f



Supplementary Figure 5. Full images of the immunoblots presented in Figures 4-7.

## Supplementary Table 1

Gel slice <sup>a</sup>	Gene symbol	M.W. <sup>b</sup>	Protein score <sup>c</sup>	Protein matches <sup>d</sup>	Peptide matches <sup>e</sup>	Previously reported HIV interactions <sup>f</sup>	Reference
<b>#1</b>	<b>APC</b>	<b>311.6</b>	<b>20</b>	<b>7</b>	<b>1</b>	–	
	LAMA3	366.7	17	6	2	Env, Tat	
<b>#2</b>	<b>NUMA1</b>	<b>238.1</b>	<b>614</b>	<b>44</b>	<b>18</b>	Gag-Pol, Tat	1
	MYH10	228.9	76	16	2	Gag, Tat	2
	BAZ2A	211.1	44	13	3	Vif	
	SEC16A	233.5	33	11	1	–	
	PARP12	226.3	19	4	1	–	
<b>#3</b>	<b>CLTC</b>	<b>191.5</b>	<b>674</b>	<b>32</b>	<b>24</b>	Gag, Pol, Env, Vpr	3
	CHD1	196.6	66	15	4	–	
	NUP188	196.0	62	7	3	–	
	ARMS	196.4	41	6	2	Tat	
	DNMT1	183.1	38	12	3	Tat	
	TAB182	181.7	26	16	3	–	
<b>#4</b>	<b>AMOT</b>	<b>118.0</b>	<b>1039</b>	<b>66</b>	<b>41</b>	Gag	4
	UBAP2L	114.5	313	10	9	Gag	2
	NAT10	115.7	275	21	12	Gag, Rev	2, 5
	DHX36	114.7	88	5	3	Rev	
	HLTF	113.9	81	9	4	Nef	
	CNTN1	113.3	49	5	2	–	
	DDX46	117.4	24	8	1	Tat	
	PSD3	116.0	23	8	1	–	
	RNF20	116.0	21	5	2	Gag, Env, Nef	3
<b>#5</b>	<b>HSP90B1</b>	<b>92.4</b>	<b>62</b>	<b>4</b>	<b>2</b>	–	
	MOGS	91.9	39	4	2	Gag, Pol, Env, Vpr, Nef	6
	KANK2	91.2	29	2	1	–	
	MCM6	92.8	25	4	1	–	
	AMPD3	88.8	16	7	1	–	
	ZNF616	90.3	21	10	1	–	

<sup>a</sup>Gel slice numbers indicated in Figure 1b. <sup>b</sup>Molecular weight (kD).

<sup>c</sup>MASCOT protein score. <sup>d</sup>Number of assigned spectra.

<sup>e</sup>Total numbers of peptides sequenced.

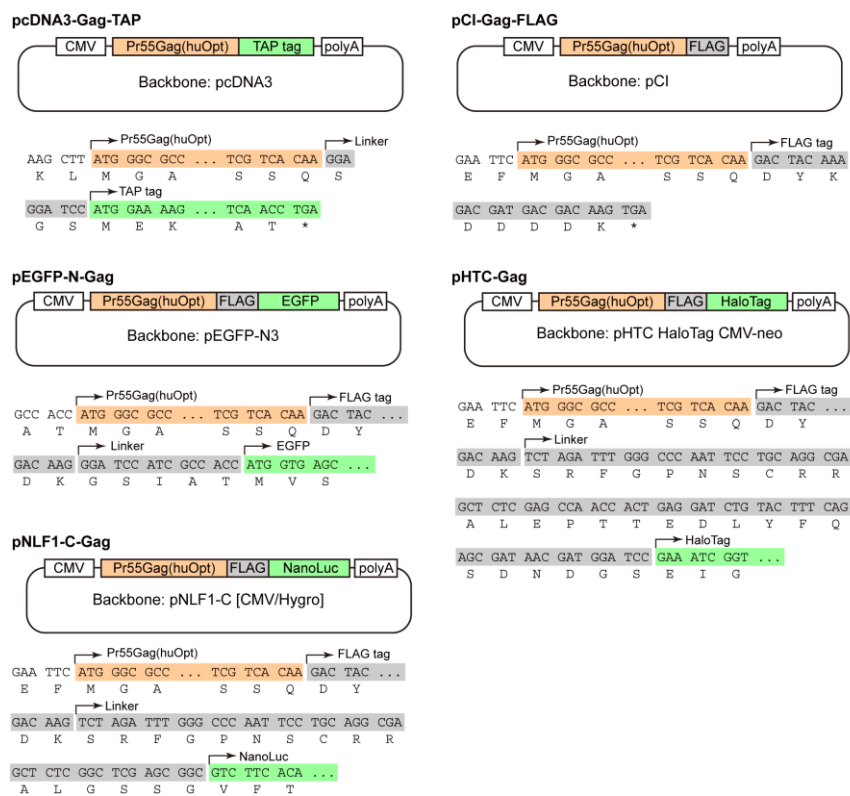
<sup>f</sup>Data collected from the HIV-1 Human Interaction Database ([www.ncbi.nlm.nih.gov/genome/viruses/retroviruses/hiv-1/interactions/](http://www.ncbi.nlm.nih.gov/genome/viruses/retroviruses/hiv-1/interactions/)).

### Supplementary Table 1. Gag-associated proteins identified by mass spectrometry.

## Supplementary Table 2

Plasmids	Gene/Insert	References
pcDNA3-Gag-TAP	Gag(huOpt*)-TAP	See below
pCI-Gag-FLAG	Gag(huOpt)-FLAG	See below
pEGFP-N-Gag	Gag(huOpt)-FLAG-GFP	See below
pGEX-Gag	Gag(huOpt)-GST	7
pHTC-Gag	Gag(huOpt)-FLAG-HaloTag	See below
pNLF1-C-Gag	Gag(huOpt)-FLAG-NanoLuc	See below
pNL4-3	All HIV-1 proteins	8
pNL4-3/Fyn(10)fullMA	Fyn(10)**-Gag and the other viral proteins	9
pNL4-3/Fyn(10) $\Delta$ MA	Fyn(10)-Gag(MA-deficient) and the other viral proteins	9
pNL4-3/Fyn(10)fullMA/Gag-YFP	Fyn(10)-Gag-YFP, Tat, Rev and Nef	9, 10
pNL4-3/Fyn(10)-Gag(6A2T)-YFP	Fyn(10)-Gag(6A2T)-YFP, Tat, Rev and Nef	11
pNL4-3/Fyn(10)-Gag(RKswitch)-YFP	Fyn(10)-Gag(RKswitch)-YFP, Tat, Rev and Nef	11
pNL4-3/Gag-EGFP	Gag-EGFP ( $\Delta$ Pol) and the other viral proteins	12

\*Human codon-optimized. \*\*Fyn(10): N-terminal 10 amino acid derived from Fyn kinase.



Supplementary Table 2. Gag fusion constructs and HIV-1 molecular clones used in this study.

### Supplementary Table 3

Antibodies/Reagents	Source (Catalog number)	Dilution
APC	Santa Cruz Biotechnology (#sc-896)	1:100 (IF)
APC	Abcam (#ab58)	1:1000 (WB)
$\alpha$ -Tubulin	Sigma-Aldrich (#T6199)	1:10000
Vinculin	Sigma-Aldrich (#V9264)	1:1000
HA	Roche (#11867423001)	1:1000
FLAG	Sigma-Aldrich (#F3165)	1:10000
GST	Santa Cruz Biotechnology (#sc-138)	1:10000
GFP	Wako (#012-20461)	1:100 (IF)
GFP	Roche (#11814460001)	1:1000 (WB)
HIV-1 Gag p24	NIH AIDS Reagent Program (#3537)	1:1000
Streptavidin-HRP Conjugate	GE Healthcare (#RPN1231)	1:5000
HRP-conjugated anti-mouse IgG	GE Healthcare (#NA931)	1:10000
HRP-conjugated anti-rabbit IgG	GE Healthcare (#NA934)	1:10000
Cholera Toxin Subunit B, Alexafluor594-conjugated	Thermo Fisher Scientific (#C34777)	1:200

**Supplementary Table 3. Antibodies and reagents used in this study.**



## Supplementary References

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