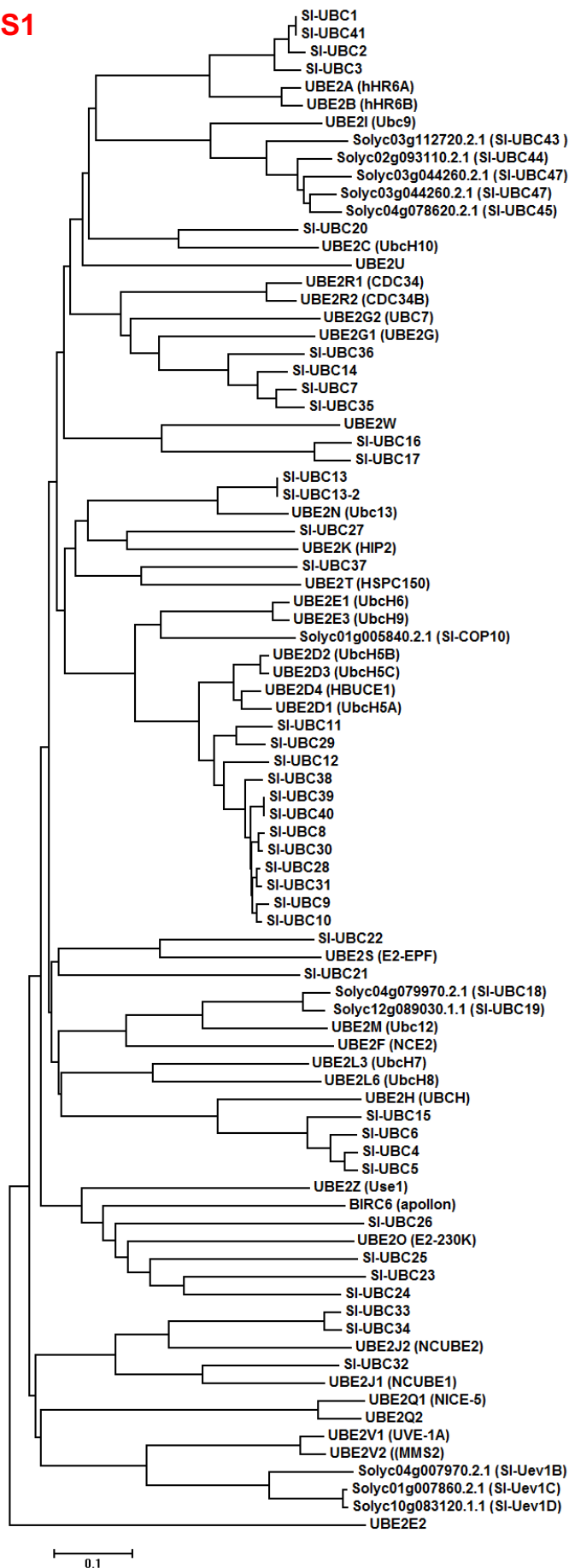


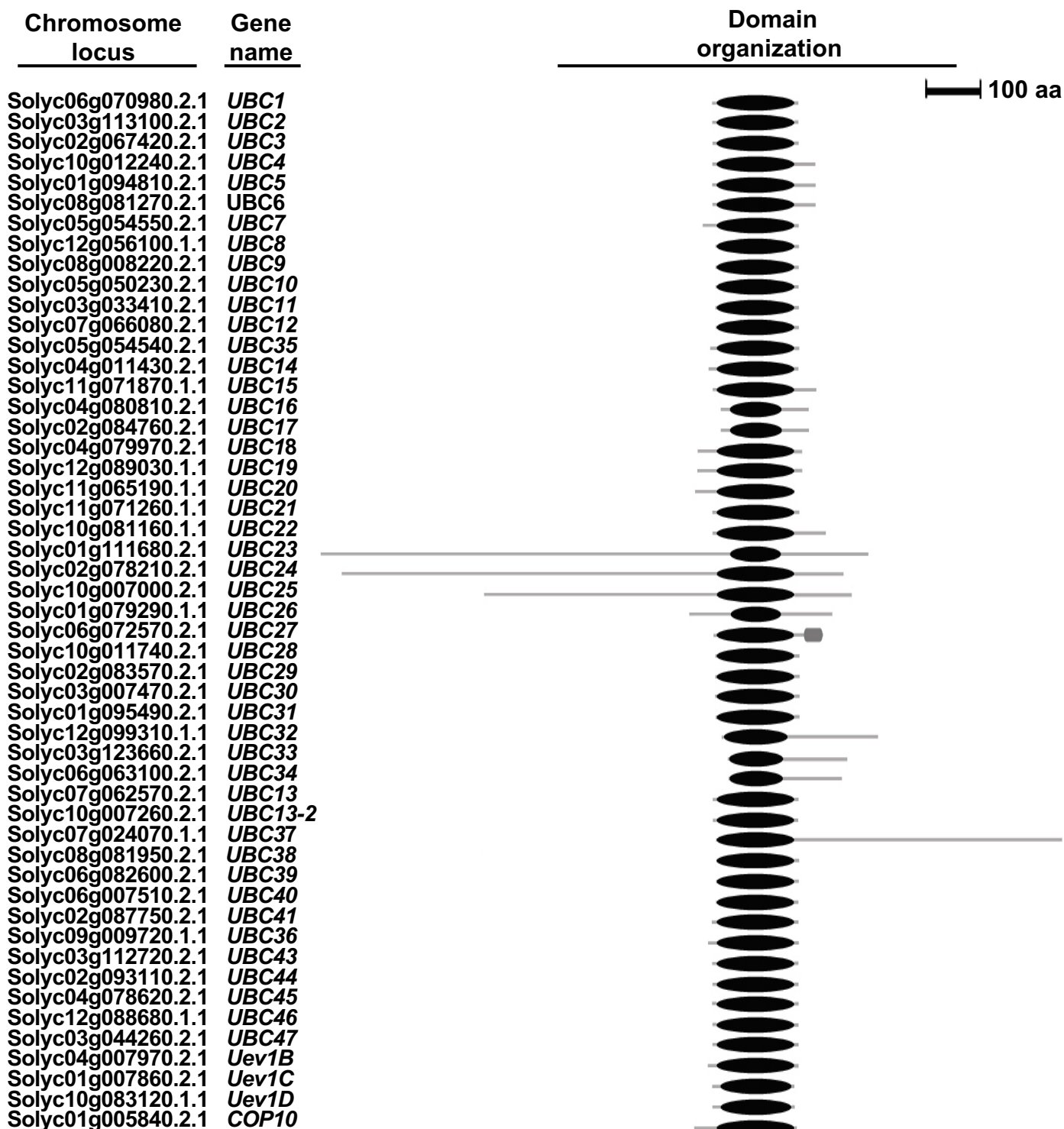
Supplemental Fig. S1



0.1

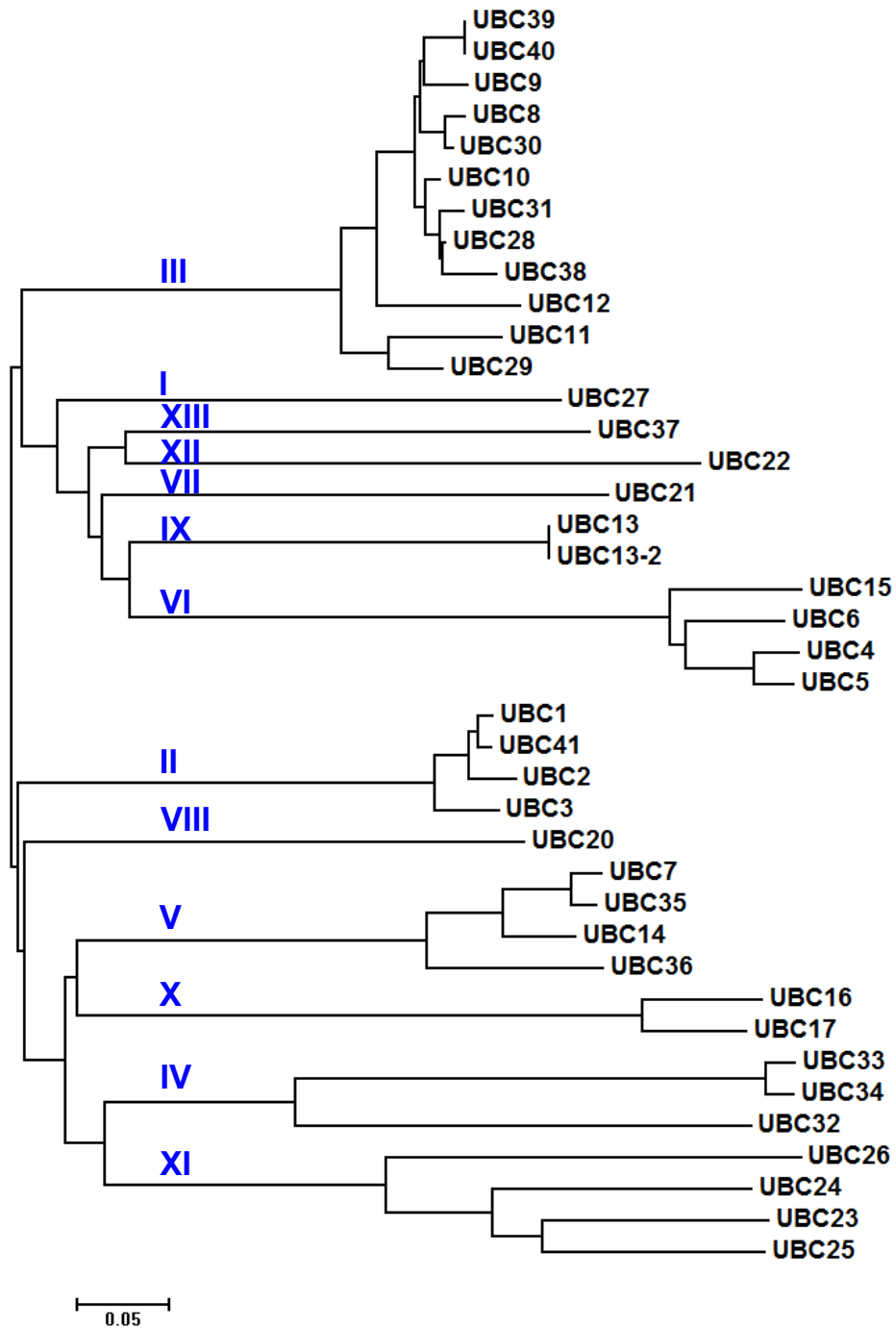
Supplemental Figure S1. Phylogeny of the human and tomato UBC domain-containing proteins. Phylogenetic analysis of the human and tomato UBC domain-containing proteins was performed using the same method and bootstrap trials as described for Figure 1. The phylogenetic tree is drawn to scale, with branch lengths in the same units as those of the evolutionary distances used to infer the phylogenetic tree. Accession numbers of the tomato and human UBC domain-containing proteins are shown in the Supplemental Table I.

Supplemental Fig. S2



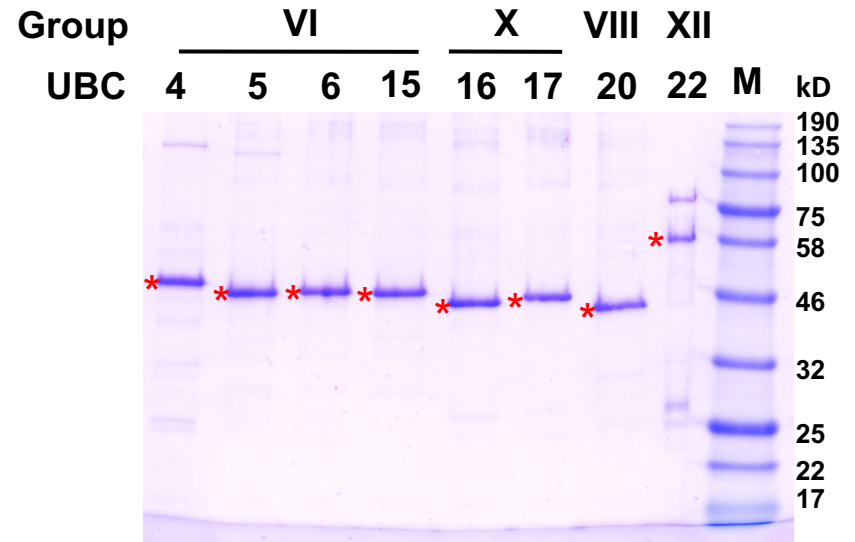
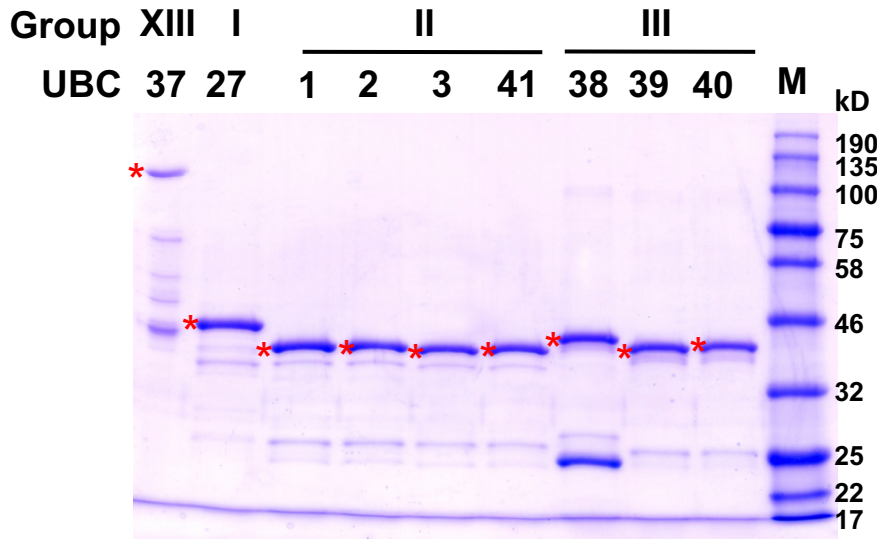
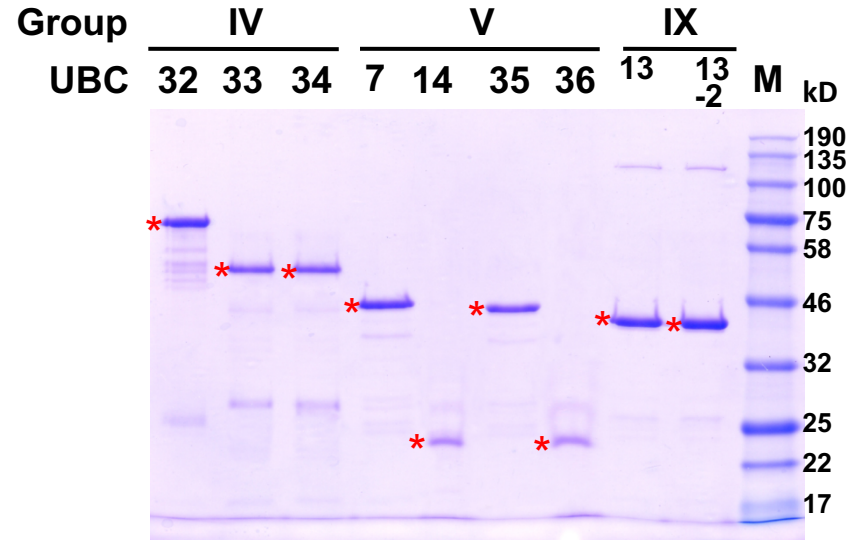
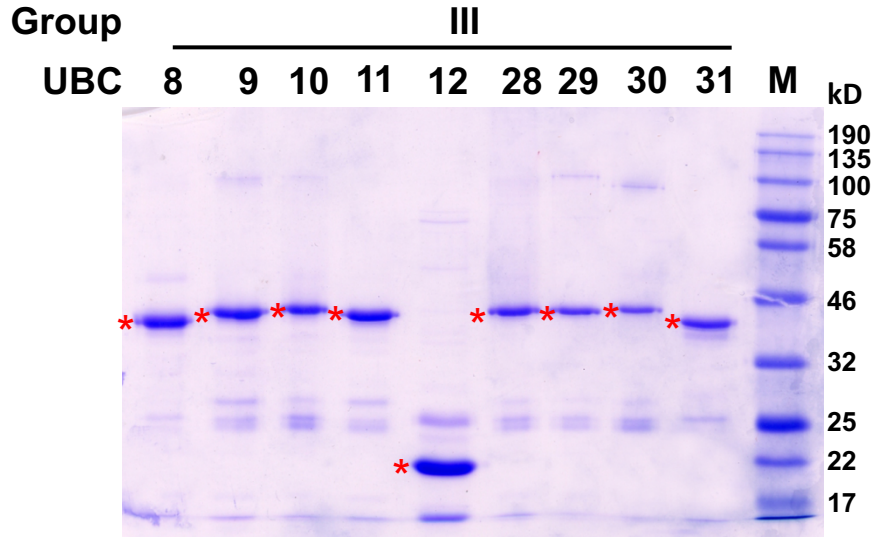
Supplemental Figure S2. Schematic representation of domain organization in the tomato UBC domain-containing proteins. The chromosomal loci and given gene names encoding the UBC domain-containing proteins are shown on the left side. The UBC fold and the extension of the UBC domain-containing proteins are represented as dark ellipse and light-gray line, respectively, both of which are drawn in scale to their length in the number of amino acids. The UBA domain of SI-UBC27 is indicated as a dark-gray rectangle. The scale bar represents length of protein in amino acids.

Supplemental Fig. S3



Supplemental Figure S3. The tomato ubiquitin E2 enzymes are classified into thirteen subgroups. Numbering of the groups was based on both the phylogenetic analysis of tomato ubiquitin E2s and the previously reported classification of Arabidopsis ubiquitin E2s (Zhao et al., 2013). Phylogenetic analysis of the forty tomato ubiquitin E2s using the amino acid sequence of core UBC domain was performed using the same method and bootstrap trials as described for Figure 1. The Roman numerals designate the different E2 groups.

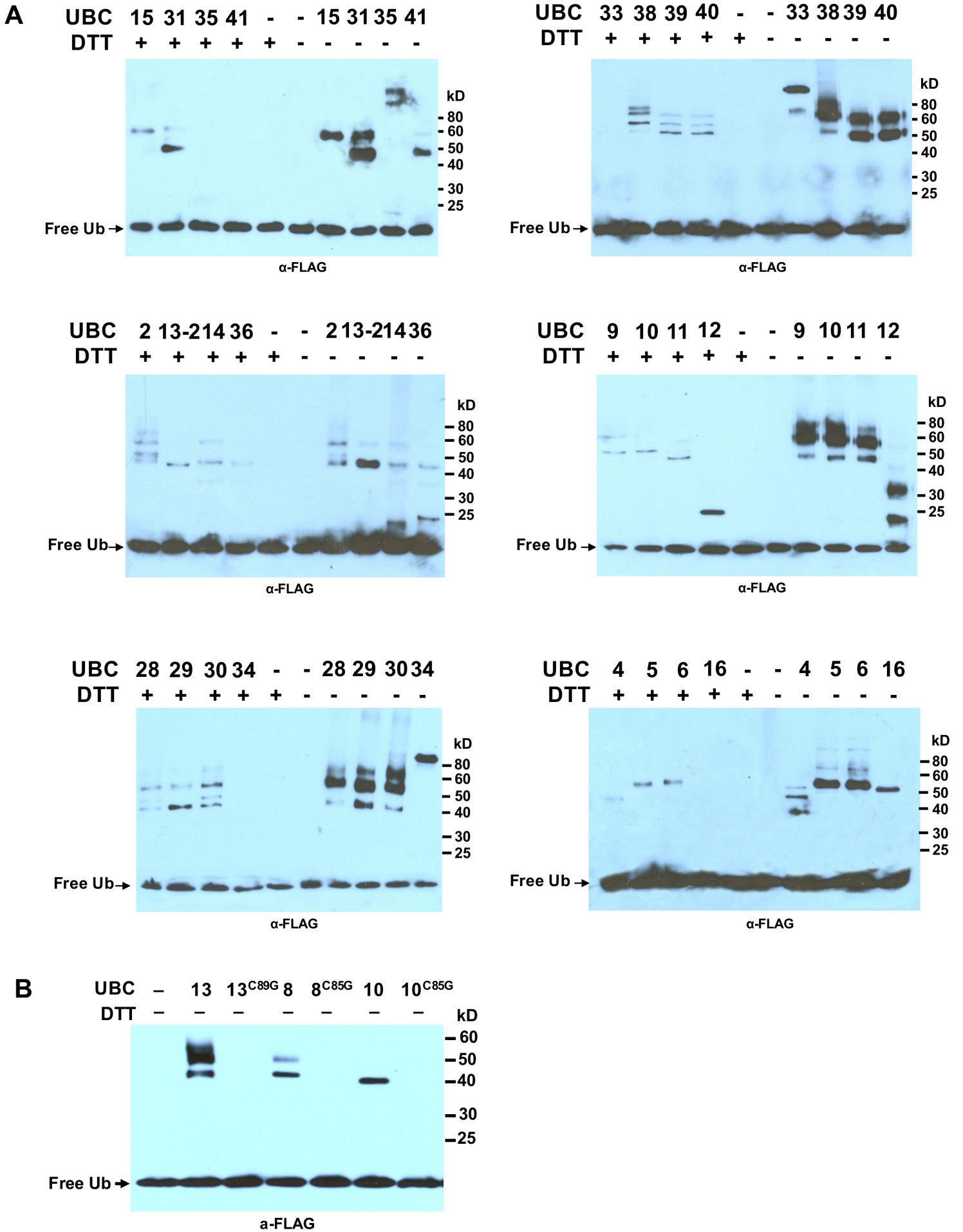
Supplemental Fig. S4



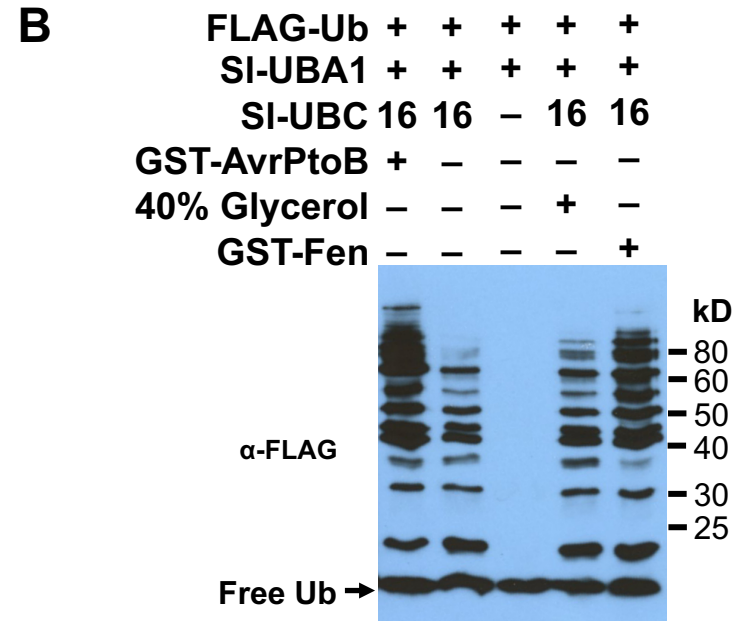
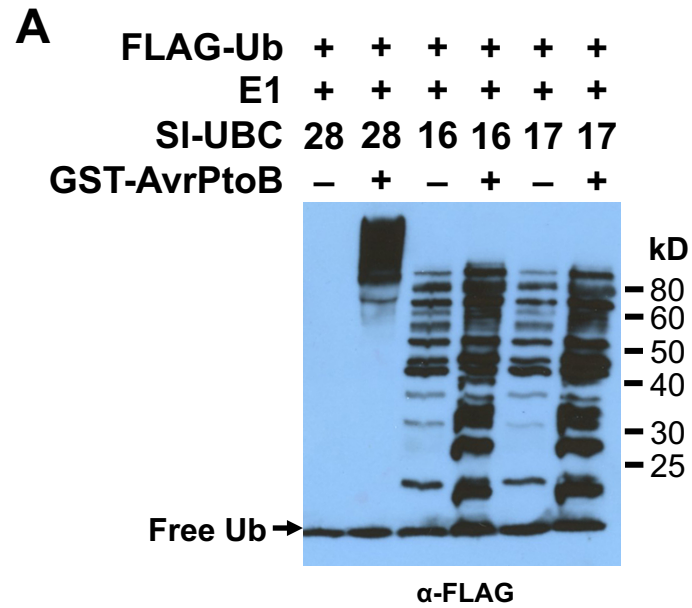
Supplemental Figure S4. Purified tomato E2 proteins as shown by SDS-PAGE.

Approximately 3 μ g of thirty-five purified E2 proteins were separated by 10% SDS-PAGE and stained with Coomassie Brilliant Blue. The Roman numerals designate the group into which the E2s are classified. M represents the molecular weight markers. The numbers on the right denote the molecular mass of marker proteins in kD. The asterisks denote the band of corresponding purified E2 proteins. Except for UBC12, 14 and 36 that are 6xHis-tagged, all E2s are fused to GST.

Supplemental Fig. S5

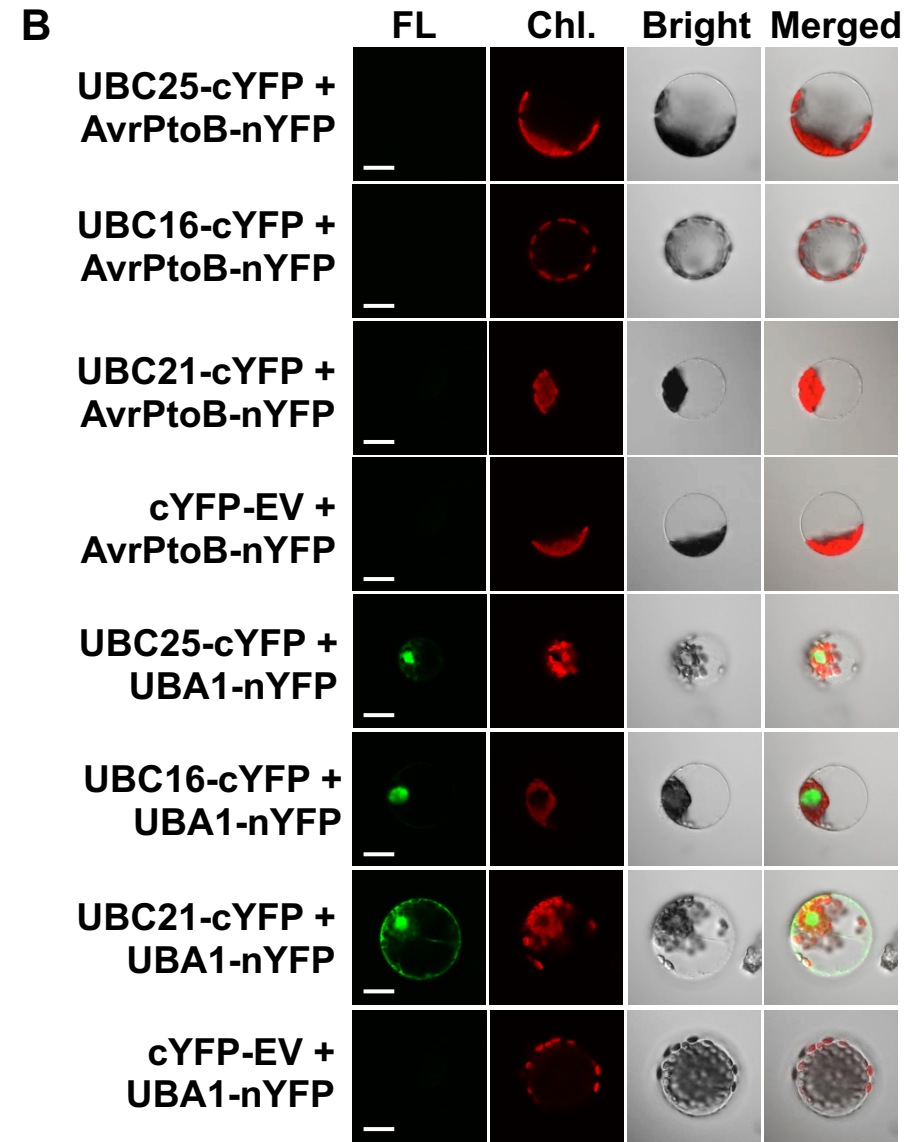
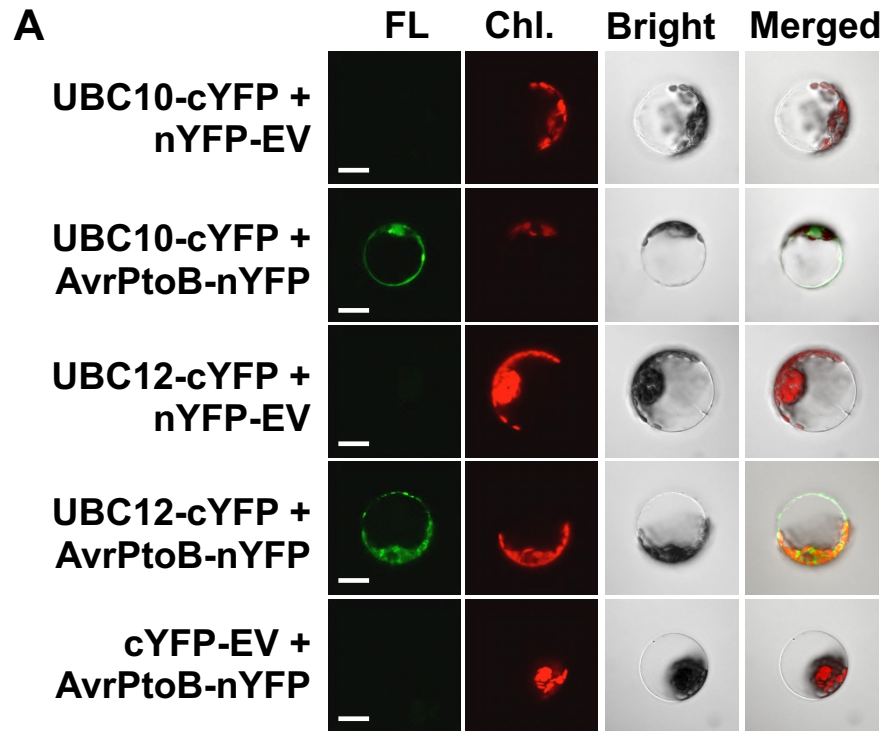


Supplemental Figure S5. Examination of the ubiquitin-conjugating activity of tomato E2s by thioester formation assay. (A) Anti-FLAG Western blots were performed following thioester assay of different tomato ubiquitin E2s. The reactions of the assay were terminated by adding SDS sample loading buffer in the presence of 100 mM DTT (DTT +) or 4M Urea (DTT -). The formation of DTT-sensitive ubiquitin adducts by tomato E2s is shown as charged E2. The numbers on the right denote the molecular mass of marker proteins in kD. The experiment was repeated two times with similar results. (B) Tomato E2 mutants in which the cysteine residue at the active site mutated lost ubiquitin-conjugating activity. Thioester assay was performed using wild type tomato ubiquitin E2s and E2 mutants. The reactions of the assay were terminated by adding SDS sample loading buffer in the presence of 4M Urea (DTT-). The numbers on the right denote the molecular mass of marker proteins in kD. The experiment was repeated two times with similar results.



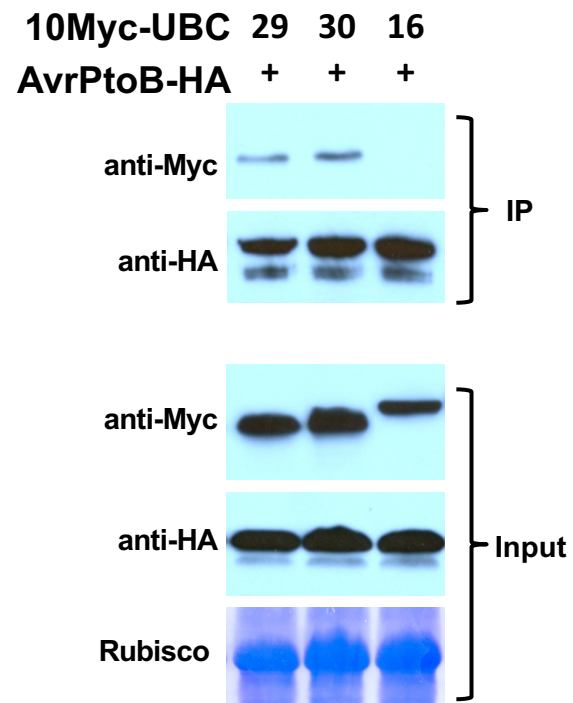
Supplemental Figure S6. AvrPtoB shows no specificity towards the tomato ubiquitin E2 enzymes SI-UBC16, and 17. (A) AvrPtoB shows no specificity towards the tomato E2s SI-UBC16 and 17 in *in vitro* ubiquitination assay. The E2s SI-UBC16 and 17 demonstrated auto-ubiquitin-conjugation activity in the absence of an E3 ligase (lane 3 and 5). The presence of AvrPtoB enhanced their conjugation activity but did not alter the pattern of conjugates formed (lane 4 and 6). SI-UBC28 was included as a control. The numbers on the top mark the lanes/reactions. (B) The enhancement of the conjugation activity of SI-UBC16 by AvrPtoB is non-specific. The presence of a non-E3 ligase protein, GST-Fen also enhanced the auto-ubiquitin-conjugation activity of SI-UBC16 (lane 5, as compared to lane 2), which is comparable to the effect of AvrPtoB (lane 1). In the absence of SI-UBC16, no ubiquitin conjugation was observed (lane 3). The numbers on the right denote the molecular mass of marker proteins in kD.

Supplemental Fig. S7



Supplemental Figure S7. AvrPtoB interacts with tomato group III E2 members but not with E2s from other groups in BiFC assays. (A) Members of the group III E2s interact with AvrPtoB in *N. benthamiana* protoplasts. (B) Members of group VII, X, and XI tomato E2 did not interact with AvrPtoB in the BiFC assay. Tomato ubiquitin E1 enzyme UBA1 was used as positive control. Different construct pairs were transiently co-expressed in protoplasts isolated from *N. benthamiana* leaves. Cells were viewed with a confocal microscope under bright or laser light to detect cells and green fluorescence, respectively. The empty vector expressing N- and C-terminus of YFP (nYFP-EV and cYFP-EV) were used as negative control. EV, empty vector; FL, fluorescence; Chl., chlorophyll autofluorescence; Bright, bright field image. Scale bar = 20 μm .

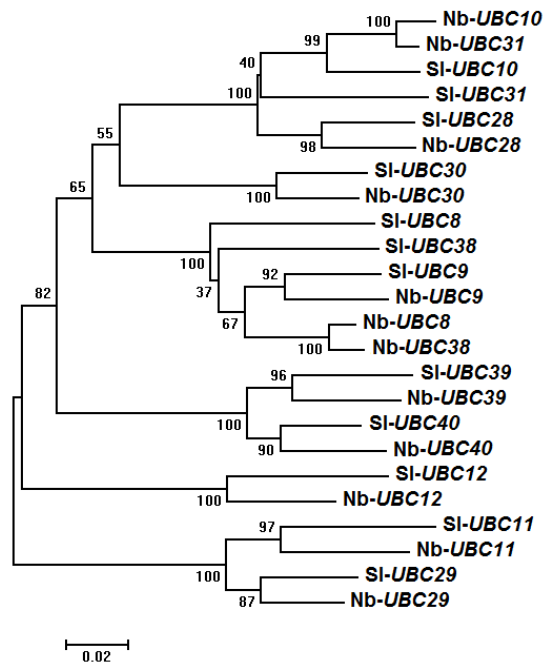
Supplemental Fig. S8



Supplemental Figure S8. Members of Group III E2 interacted with AvrPtoB in co-immunoprecipitation (Co-IP) assay. Group III members Ubc29 and 30 were randomly selected for the assay. AvrPtoB-HA and 10Myc-tagged E2s were transiently co-expressed in *N. benthamiana* leaves. The Co-IP was carried out with an anti-HA antibody (IP: anti-HA). The presence of corresponding proteins (Top panel) and the input (bottom panel) were detected by Western blot using anti-HA antibody for AvrPtoB-HA and anti-Myc antibody for 10Myc-tagged E2s.

Supplemental Fig. S9

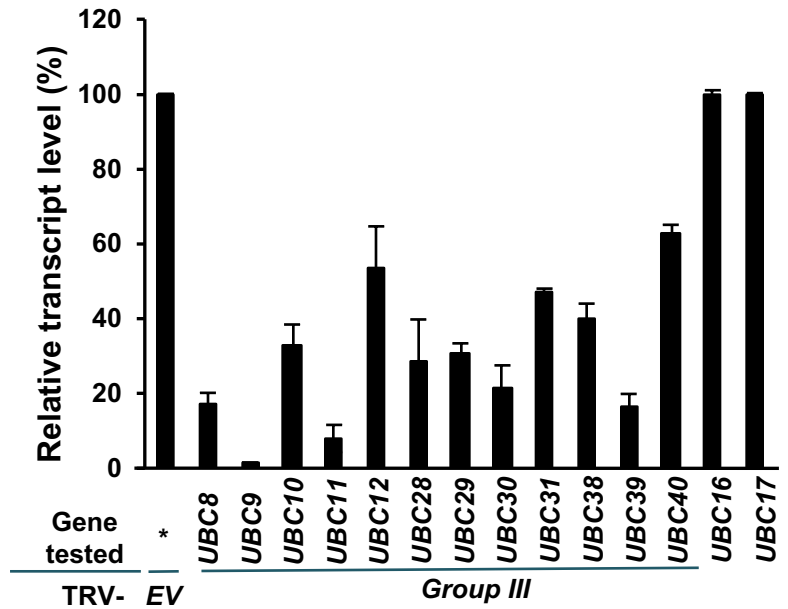
A



B



C



Supplemental Figure S9. Knocking down group III E2 genes in *N. benthamiana* by Virus-Induced Gene Silencing (VIGS). (A) The *N. benthamiana* group III E2 genes are highly homologous to their counterparts in tomato. The DNA sequences of *N. benthamiana* and tomato group III E2 genes' open reading frame (ORF) were used to generate the phylogenies. The same method and bootstrap trials as described for Figure 1 were employed for the phylogenetic analysis. The sequence IDs of the *N. benthamiana* ubiquitin E2 genes identified from the Sol Genomics Network database (SGN; <http://solgenomics.net/>) are shown in Supplemental Table I. The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (1000 replicates) is shown next to the branches. (B) Phenotypes of the *N. benthamiana* plants in which the group III E2 genes are silenced. The non-silenced TRV empty-vector (TRV) was used as a control. Photographs were taken 4 weeks after the approximately 3-week old seedlings were infiltrated with TRV or TRV-*group III* E2 genes constructs. The upper panel shows the top view of the plants while the lower panel shows the side view. (C) The group III ubiquitin E2 genes are efficiently and specifically silenced in *N. benthamiana* using the TRV vector. The transcript level of group III E2 genes and closely related E2 genes Nb-*UBC16* and *17* (outside the group III) in non-silenced TRV control (TRV) and group III ubiquitin E2 genes-silenced (TRV-*group III*) *N. benthamiana* plants was examined as described (Mural et al., 2013). Nb-*EF1 α* was used as an internal reference for the determination of the amount of cDNA template to be used. Experiments in (B) and (C) were repeated at least two times with similar results.

Supplemental Fig S10

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SI-UBC8	AAGCAACAATATATGGGTCCGCTGACAGCCCTTATGCTGGCGGAGTGTTCCTGTTACCAATTCATTTTCCACCTGACTATCCATTAAAGCCACC	200
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SI-UBC8	AGCTTTTCCAGCAAAAGGTTTCCACCCAAACATCAAAGCAATGGTAGCATTTGCCTCGACATTTGAAGGAACAATGGAGTCCGGCCTTACAAATCTCC	300
Nb-UBC8	AAGGTAATGCTGCTCAATCTGTTCTCTGTGACAGACCCCTAATCCGATGATCCATTGGTGCCGAGATTGCTCATATGTACAAAGACTGATAAAGCAAGT	400
SI-UBC8	AAGGTAATGCTGCTCAATCTGTTCTCTGTGACAGACCCCTAATCCGATGATCCATTGGTGCCGAGATTGCTCATATGTACAAAGACTGATAAAGCAAGT	400
Nb-UBC8	ACGAAACAACCTGCCCGGAGCTGGACTCAAAAATATGCCATGGGTAG	447
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SI-UBC9	ACGAAGCCTACTGCTCGGAGCTGGACCAAAAATATGCTATGGGTAG	447
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SI-UBC38 ACGAAACAACCGCTCGAGCTGGACTCAAAGTATGCCATGGGTTAG 447

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SI-UBC39 ATGGCGTCGAAGCGCATATTGAAAGAGCTCAAGGATTGTCAGAAGGATCCTCCGACATCATGCAGCGCTGGTCCAGTCTGCTGAGGATATGTTCCATGGC 100

Nb-UBC39 AAGCAACATCATGGGGCTACGATAGCCCTTATGCAAGGAGGTGATTTTTGGTTCAATCTCATTTCCTCTCTGATTATCCTTTCAAGCCTCAAAGGT 200
SI-UBC39 AAGCAACATCATGGGGCTACGATAGCCCTTATGCAAGGAGGTGATTTTTGGTTCAATCTCATTTCCTCTCTGATTATCCTTTCAAGCCTCAAAGGT 200

Nb-UBC39 TGCATTTAGAACTAAAGTTTTCCATCCCAACATCAATAGCAATGGAAGTATATGTTAGATATCTTAAAGACAGTGGAGTCCAGCTTTGACCATATCT 300
SI-UBC39 TGCATTTAGAACTAAAGTTTTCCATCCCAACATCAATAGCAATGGAAGTATATGTTAGATATCTTAAAGACAGTGGAGTCCAGCTTTGACCATATCT 300

Nb-UBC39 AAGGTCCTGTTGTCATCTGTTCTCTGTTGACAGATCCAAATCCAGACGATCCACTTGACCAGAAATGCTCATATGTACAAGACTGACAGGGCCAAAT 400
SI-UBC39 AAGGTCCTGTTGTCATCTGTTCTCTGTTGACAGATCCAAATCCAGACGATCCACTTGACCAGAAATGCTCATATGTACAAGACTGACAGGGCCAAAT 400

Nb-UBC39 ACGAGGCCACTGCTCGTAGCTGGACACAGAAATATGCTATGGGATGA 447
SI-UBC39 ACGAGGCCACTGCTCGTAGCTGGACTCAAAAATATGCTATGGGATGA 447

Nb-UBC40 ATGGCGTCGAAGGATATGAAAGGAGCTCAAGGATCTGTCAGAAGGATCCTCCACATCATGCAGTCTGGTCCAGTGGCAGAGGATATGTTCCATTGGC 100
SI-UBC40 ATGGCGTCGAAGGATATGAAAGGAGCTCAAGGATCTGTCAGAAGGATCCTCCACATCATGCAGTCTGGTCCAGTGGCAGAGGATATGTTCCATTGGC 100

Nb-UBC40 AAGCAACAATCATGGGGCTACAGATAGCCCTTATGCGGGAGTGTATTTTTGGTTCAATCTTCCCTCCGATTATCCTTTCAAGCCTCAAAGGT 200
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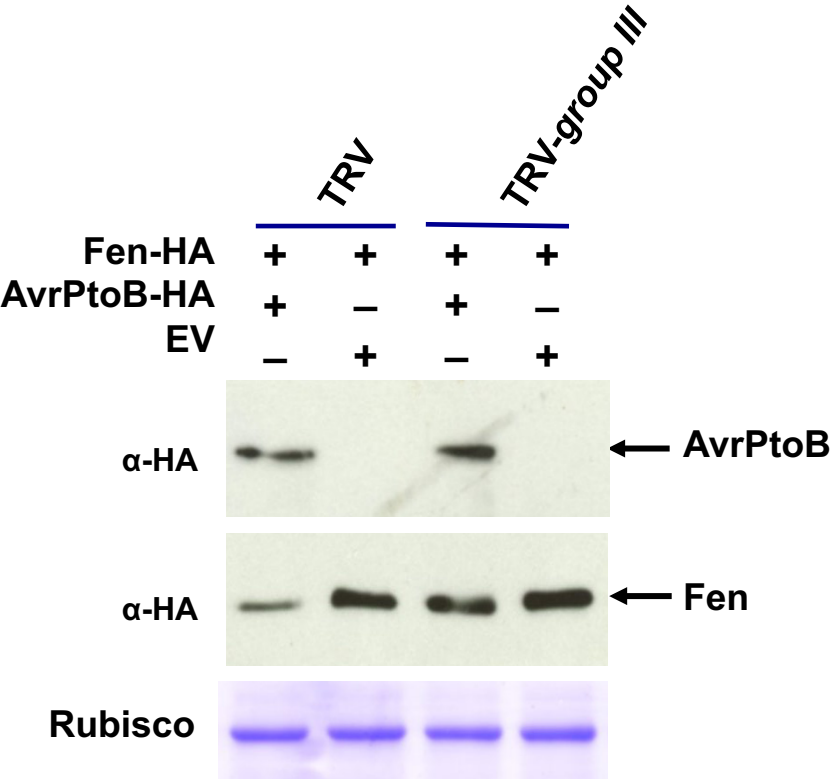
Nb-UBC40 TGCTTTAGAACTAAGTTTTCCATCCCAACATCAATAGCAATGGAAGTATGTTAGATATCTTAAAGACAGTGGAGTCCAGCTTTAACCATATCT 300
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Nb-UBC40 AAGGTCCTGCTCTCCATCTGCTCTCTGTTGACAGATCCAAATCCAGACGATCCACTTGACCAGAAATGCTCATATGTACAAGACTGACAGGGCCAAAT 400
SI-UBC40 AAGGTCCTGCTCTCCATCTGCTCTCTGTTGACAGATCCAAATCCAGACGATCCACTTGACCAGAAATGCTCATATGTACAAGACTGACAGGGCCAAAT 400

Nb-UBC40 ACGAGGCCACTGCTCGTAGCTGGACACAGAAATATGCTATGGGATGA 447
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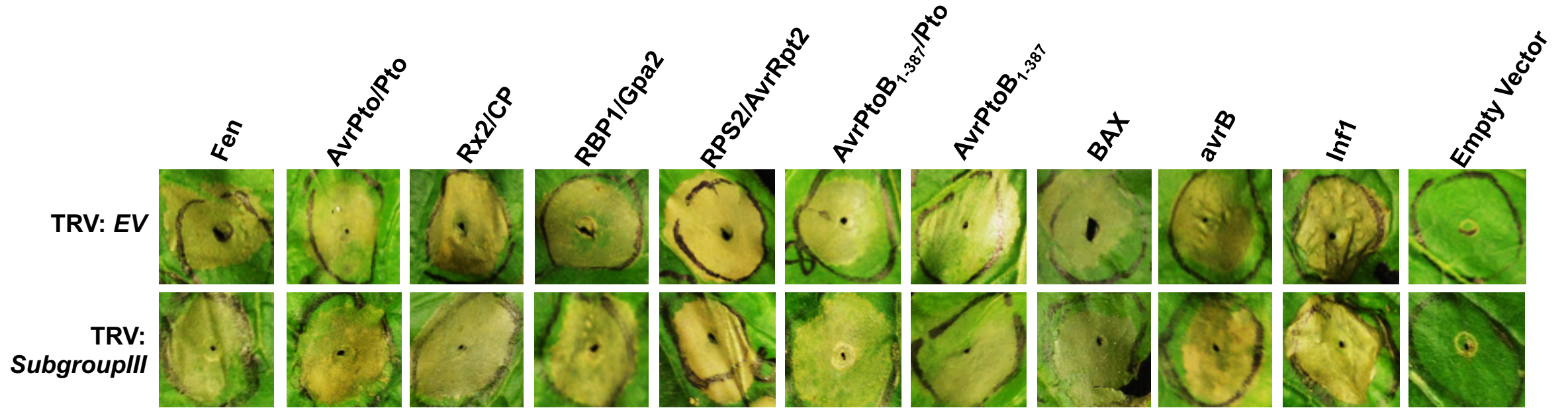
Supplemental Figure S10. Homologs of the group III E2 genes from *N. benthamiana* share high nucleotide sequence identity to their counterpart of tomato. The group III E2 genes from *N. benthamiana* were searched using the BLAST algorithm (Camacho et al., 2009) against the Sol Genomics Network (SGN, <http://solgenomics.net>) database with the counterparts from tomato as the queries. The Clustal X algorithm (Larkin et al., 2007) was used for the sequence alignment. The sequences underlined red were the fragments used for building the TRV-*group III* VIGS construct. The DNA fragment from Nb-*UBC9* was designed for silencing Nb-*UBC8*, *9* and *38*; the DNA fragment from Nb-*UBC28* was designed for silencing Nb-*UBC10*, *28* and *31*; the fragment from Nb-*UBC11* for silencing Nb-*UBC11* and *29*; and the fragment from Nb-*UBC39* for silencing Nb-*UBC39* and *40*. The blue-underlined were sequences that are putatively targeted in VIGS by the red-underlined sequence of the corresponding gene from the same clade in the phylogenetic tree (Figure S6A).

Supplemental Fig. S11



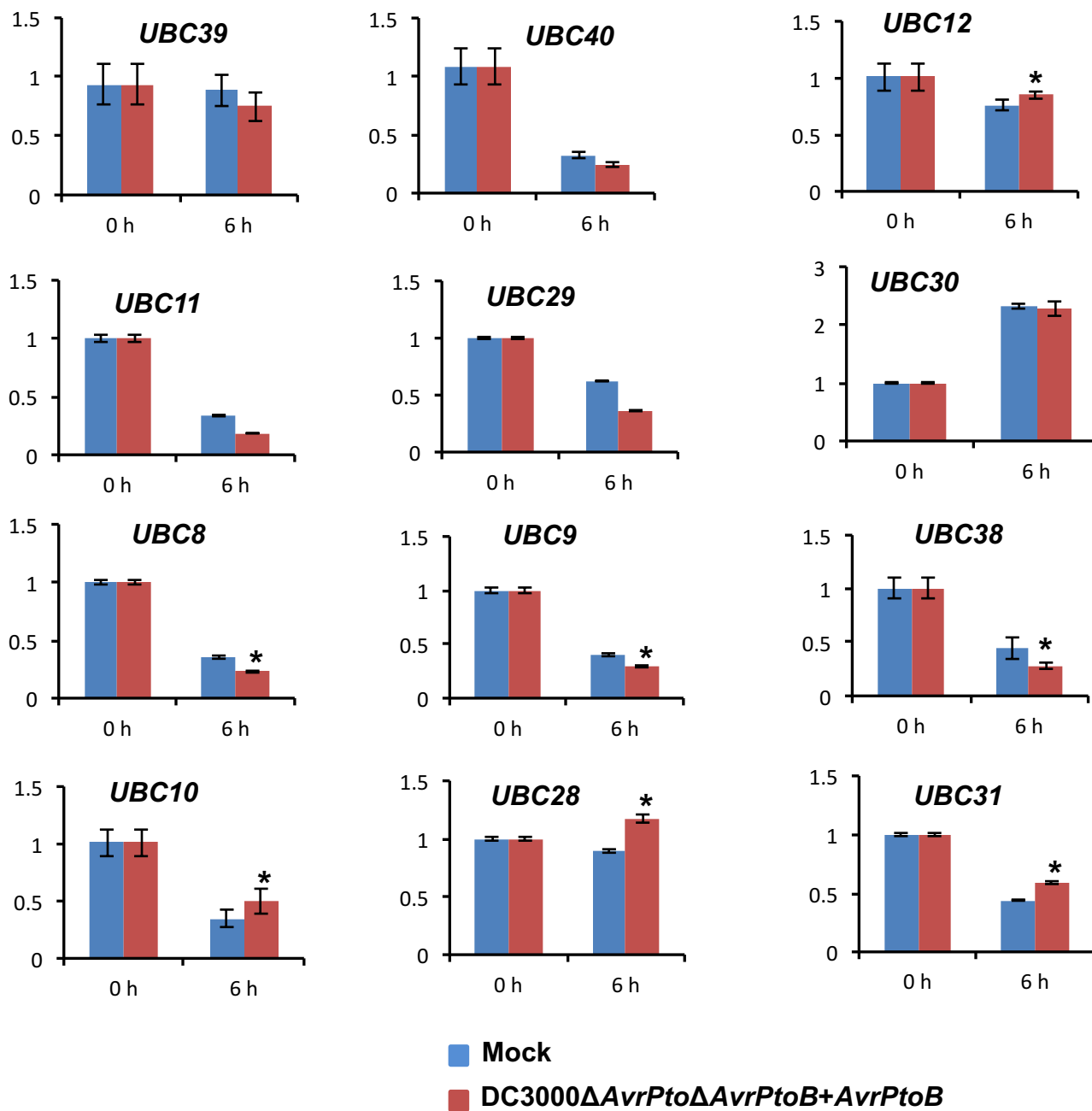
Supplemental Figure S11. Silencing group III E2 genes diminished AvrPtoB-promoted degradation of Fen in *N. benthamiana* protoplasts. AvrPtoB-HA and Fen-HA were transiently co-expressed in protoplasts isolated from leaves of group III E2 genes-silenced (TRV-*group III*) and non-silenced TRV control (TRV) *N. benthamiana* plants. Protoplasts were harvested and lysed at 21 h after the protoplasts were transfected with DNA carrying corresponding genes to isolate total proteins. Western blot was performed using anti-HA antibody for detecting Fen-HA and AvrPtoB-HA. Staining of ribulose 1, 5-bisphosphate carboxylase–oxygenase (Rubisco) subunits by Coomassie blue demonstrated equal loading. Marker minus (-) denotes the corresponding gene was not transfected into the protoplasts. The experiment was repeated two times with similar results.

Supplemental Fig S12



Supplemental Figure S12. Silencing group III E2 genes does not influence multiple ETI elicitors-triggered programmed cell death (PCD). *Agrobacterium*-mediated transient expression of Fen, AvrPto/Pto, Rx2/CP, RBP1/Gpa2, RPS2/AvrRpt2, AvrPtoB1-387/Pto, AvrPtoB1-387, BAX, avrB and Inf1 were performed in group III ubiquitin E2 genes-silenced (TRV-*group III*) and non-silenced TRV control (TRV) *N. benthamiana* plants as described (Mural et al., 2013). *Agrobacterium*-mediated transient expression of empty vector (EV) was performed as the control. At least three spots of infiltration were performed for each elicitor or EV on four different plants with typical result being shown. Photographs were taken on day four after infiltration. The experiment was repeated at least two times with similar results.

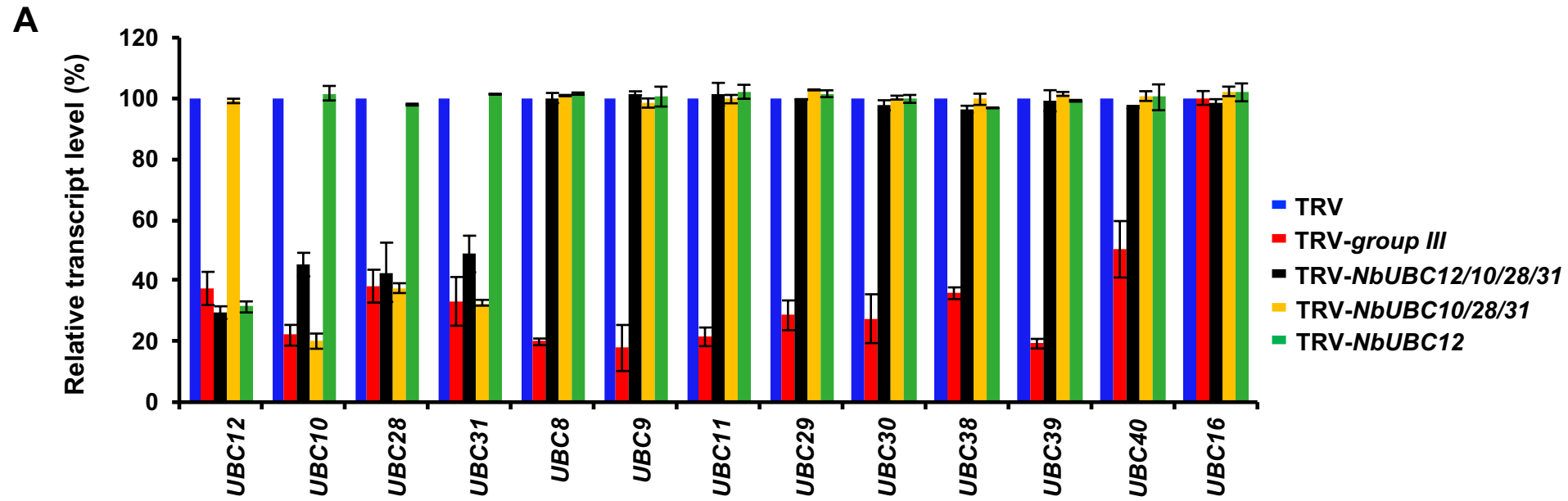
Supplemental Fig S13



Supplemental Figure S13. Effect of AvrPtoB on the expression of group III E2 genes.

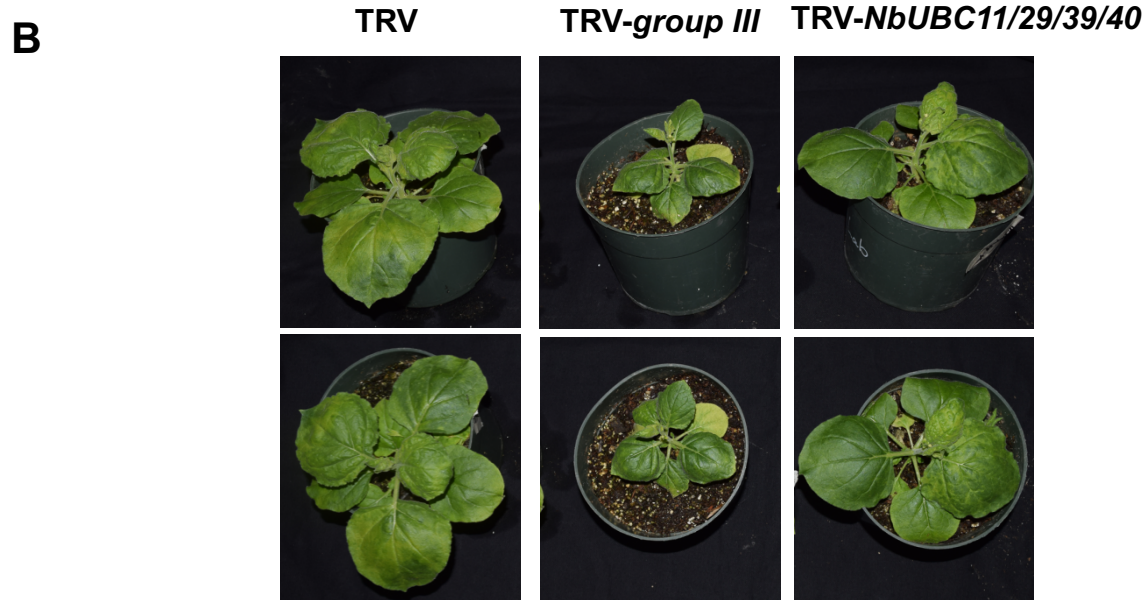
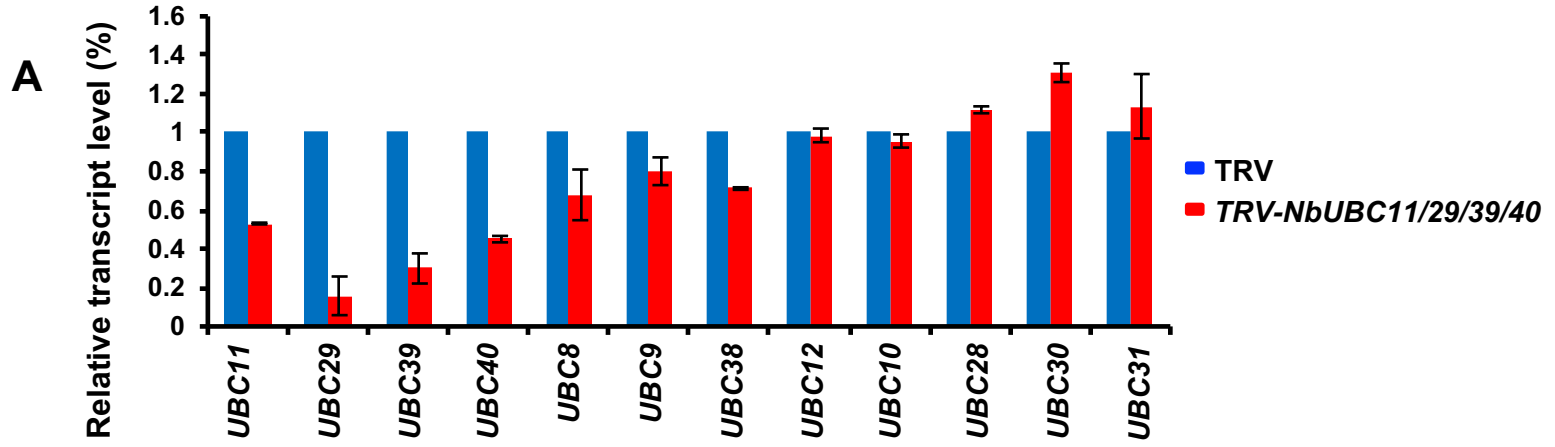
Real time PCR analysis of the transcript level of group III E2 genes in *Pst*- or mock-treated tomato plants. Tomato RG-pto11 plants were inoculated with *Pst* strain DC3000 Δ *avrPto* Δ *avrPtoB* expressing AvrPtoB or mock (10 mM MgCl₂). Samples were collected at 0 and 6 h post inoculation. The Y-axis depicts the relative expression of the gene being tested. The experiment was performed using three technical repeats in each of the three biological replicates. Asterisks denote significant difference (P<0.05) in the expression of the E2 gene on the plants inoculated with *Pst* strain DC3000 Δ *avrPto* Δ *avrPtoB* expressing AvrPtoB compared with mock inoculation.

Supplemental Fig S14



Supplemental Figure S14. Specific silencing of E2 genes Nb-UBC12 alone, Nb-UBC10, 28 and 31 together, or Nb-UBC10, 12, 28 and 31 together in *N. benthamiana* by Virus-Induced Gene Silencing. (A) Nb-UBC10, 12, 28 and 31 genes were specifically and efficiently silenced in *N. benthamiana* by TRV-based VIGS. The transcript level of group III E2 genes and a closely-related E2 gene outside the group III, Nb-UBC16 in various VIGS-treated *N. benthamiana* plants was examined as described (Mural et al., 2013). Nb-*EF1 α* was used as an internal reference for the determination of the amount of cDNA template to be used. The experiment was repeated three times with similar results. (B) Phenotypes of the *N. benthamiana* plants in which Nb-UBC12 alone, three E2 genes Nb-UBC10, 28 and 31, or Nb-UBC10, 12, 28 and 31 were specifically silenced. The non-silenced TRV-infected plant and *group III*-silenced plant were included as control. Photographs were taken 4 weeks after the ~ 3-week-old seedlings were infiltrated with TRV vector-based VIGS constructs. The upper panel shows the side view of the plants while the lower panel shows the top view.

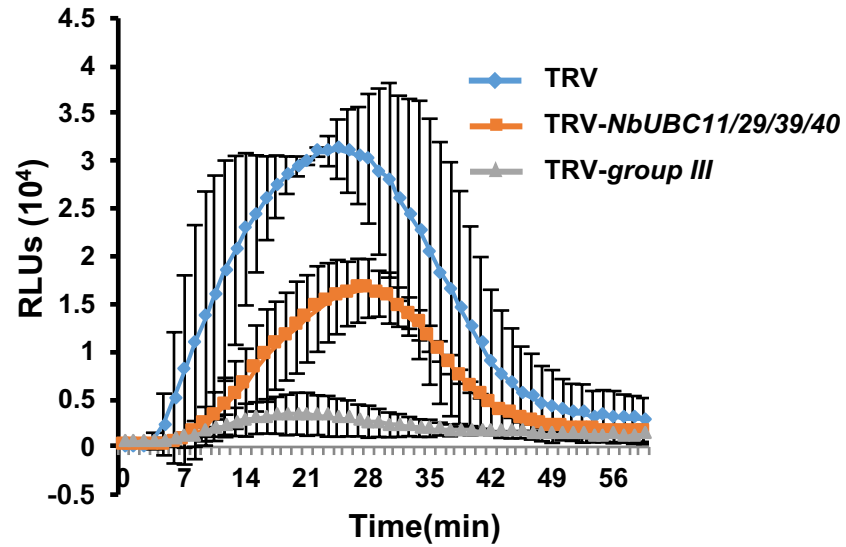
Supplemental Fig S15



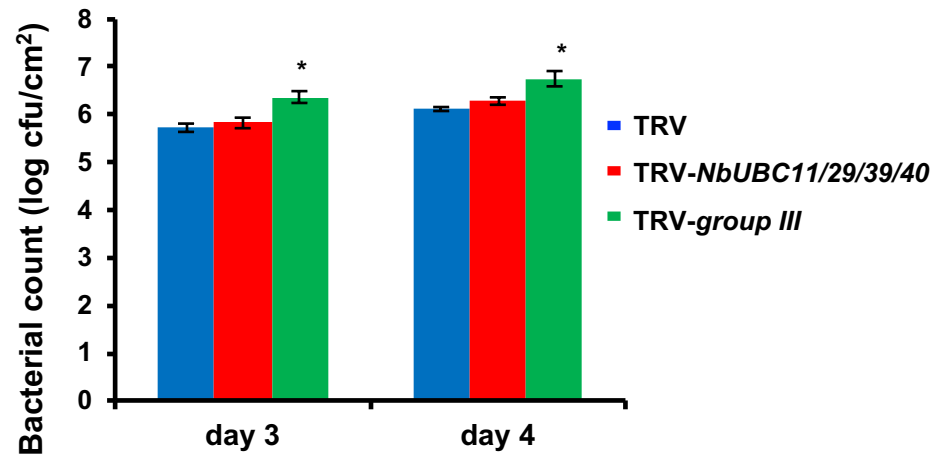
Supplemental Figure S15. Specific silencing of E2 genes Nb-UBC11, 29, 39 and 40 together in *N. benthamiana* by Virus-Induced Gene Silencing. (A) Nb-UBC11, 29, 39 and 40 genes were efficiently silenced in *N. benthamiana* by TRV-based VIGS. The Nb-UBC8, 9 and 38 were also very slightly knocked down in the plants. The transcript level of group III E2 genes in VIGS-treated *N. benthamiana* plants was examined as described (Mural et al., 2013). Nb-EF1 α was used as an internal reference for the determination of the amount of cDNA template to be used. The experiment was repeated three times with similar results. (B) Phenotypes of the *N. benthamiana* plants in which Nb-UBC11, 29, 39 and 40 were silenced. The non-silenced TRV-infected plant and *group III*-silenced plant were included as control. Photographs were taken 4 weeks after the ~ 3-week-old seedlings were infiltrated with TRV vector-based VIGS constructs. The upper panel shows the side view of the plants while the lower panel shows the top view.

Supplemental Fig S16

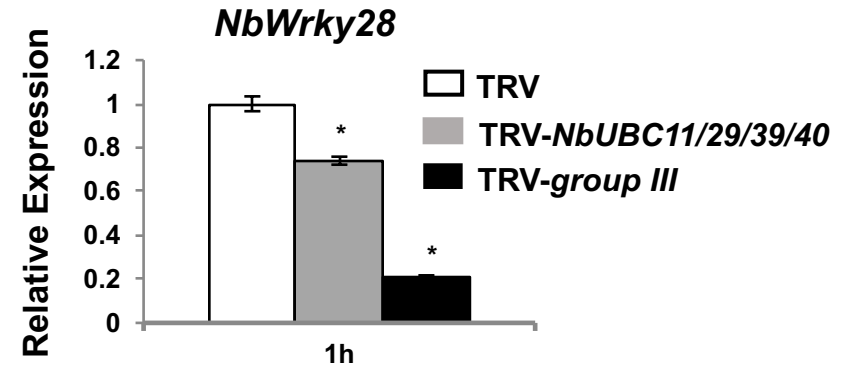
A



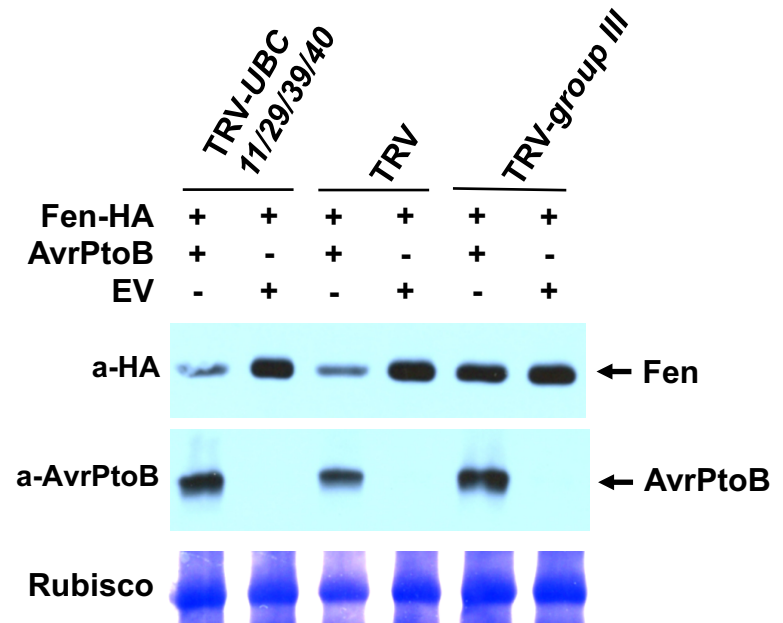
C



B



D



Supplemental Figure S16. UBC11, 29, 39 and 40 of group III play a more important role in PTI. (A) Silencing the E2 genes *UBC11*, *29*, *39* and *40* resulted in reduced ROS production induced by flg22 in a chemiluminescence assay. The diminishment of ROS on the *UBC11/29/39/40*-silenced plants was to a less extent than that on the group III-knocked down plants. (B) Knocking down the E2 genes *UBC11*, *29*, *39* and *40* down-regulates the induction of PTI reporter gene *Wrky28* by flg22. The expression of *N. benthamiana* PTI marker gene *Wrky28* was performed as described in Figure 6. The experiment was performed with three technical repeats in each of the three biological replicates. Error bars indicate standard deviation. Asterisks mark significant reduction of the expression of *Wrky28* in group III E2 genes-silenced and *UBC11/29/39/40*-silenced plants compared to non-silenced TRV control plants ($P < 0.05$). (C) Bacterial populations of the *Pst* strain DC3000 Δ *hopQ1-1* on leaves of various VIGS-treated plants. Experiments were performed as described in Figure 8D and repeated three times with similar results. Asterisks indicate significantly increased bacterial growth on *group III*-silenced plants compared to the non-silenced control plants based on the one-way ANOVA ($P < 0.01$). (D) No effect on the degradation of Fen caused by AvrPtoB was observed on *N. benthamiana* plants in which the expression of Nb-*UBC11*, *29*, *39* and *40* was knocked down. The experiment was performed as shown in Figure 5B and was repeated two times with similar results.

Supplemental Table I. List of UBC domain-containing proteins from tomato, *Arabidopsis*, *N. benthamiana*, and human.

Supplemental Table II. List of primers used in this study.

REFERENCES

- Camacho, C., Coulouris, G., Avagyan, V., Ma, N., Papadopoulos, J., Bealer, K., and Madden, T.L.** (2009). BLAST+: architecture and applications. *BMC Bioinformatics* **10**, 421.
- Larkin, M.A., Blackshields, G., Brown, N.P., Chenna, R., McGettigan, P.A., McWilliam, H., Valentin, F., Wallace, I.M., Wilm, A., Lopez, R., Thompson, J.D., Gibson, T.J., and Higgins, D.G.** (2007). Clustal W and Clustal X version 2.0. *Bioinformatics* **23**, 2947-2948.
- Mural, R.V., Liu, Y., Rosebrock, T.R., Brady, J.J., Hamera, S., Connor, R.A., Martin, G.B., and Zeng, L.** (2013). The tomato Fni3 lysine-63-specific ubiquitin-conjugating enzyme and SUV ubiquitin E2 variant positively regulate plant immunity. *Plant Cell* **25**, 3615-3631.
- Zhao, Q., Tian, M., Li, Q., Cui, F., Liu, L., Yin, B., and Xie, Q.** (2013). A plant-specific in vitro ubiquitination analysis system. *Plant J* **74**, 524-533.

**Supplemental Table I List of UBC domain-containing proteins from
tomato, Arabidopsis, *N. benthamiana*, and human**

Arabidopsis UBCs

NCBI Accession number of genes	Accession number of proteins	Gene locus	Protein name
NM_101307.4	NP_563951.1	AT1G14400	AtUBC1
NM_126331.3	NP_565289.1	AT2G02760	AtUBC2
NM_125648.3	NP_568956.1	AT5G62540	AtUBC3
NM_123499.4	NP_568589.1	AT5G41340	AtUBC4
NM_105055.2	NP_564817.2	AT1G63800	AtUBC5
NM_130166.3	NP_566062.1	AT2G46030	AtUBC6
NM_125320.1	NP_568902.1	AT5G59300	AtUBC7
NM_180783.1	NP_851114.1	AT5G41700	AtUBC8
NM_118934.2	NP_567791.1	AT4G27960	AtUBC9
NM_124709.2	NP_568788.1	AT5G53300	AtUBC10
NM_111703.3	NP_566331.1	AT3G08690	AtUBC11
NM_111704.1	NP_566332.1	AT3G08700	AtUBC12
NM_114513.5	NP_566884.1	AT3G46460	AtUBC13
NM_115396.3	NP_567020.1	AT3G55380	AtUBC14
NM_103582.3	NP_564493.1	AT1G45050	AtUBC15
NM_106198.4	NP_565110.1	AT1G75440	AtUBC16
NM_119804.2	NP_568004.1	AT4G36410	AtUBC17
NM_123665.5	NP_568619.1	AT5G42990	AtUBC18
NM_112897.3	NP_566653.1	AT3G20060	AtUBC19
NM_103932.3	NP_564572.1	AT1G50490	AtUBC20
NM_122477.2	NP_568476.1	AT5G25760	AtUBC21/PEX4
NM_120590.2	NP_568148.1	AT5G05080	AtUBC22
NM_127245.2	NP_179284.1	AT2G16920	AtUBC23
NM_179887.2	NP_850218.1	AT2G33770	AtUBC24/PHO2
NM_112402.2	NP_188154.1	AT3G15355	AtUBC25
NM_104180.1	NP_175710.1	AT1G53020	AtUBC26
NM_124465.5	NP_199900.1	AT5G50870	AtUBC27
NM_105097.8	NP_564828.1	AT1G64230	AtUBC28
NM_127226.2	NP_565391.1	AT4G27960	AtUBC29
NM_124997.3	NP_568835.1	AT5G56150	AtUBC30
NM_103322.2	NP_564472.1	AT1G36340	AtUBC31
NM_112576.2	NP_566563.1	AT3G17000	AtUBC32
NM_124425.3	NP_199854	AT5G50430	AtUBC33
NM_001084085.1	NP_001077554.1	AT1G17280	AtUBC34
NM_106535.3	NP_565192.1	AT1G78870	AtUBC35
NM_101550.3	NP_564011.1	AT1G16890	AtUBC36
NM_113362.1	NP_566751.1	AT3G24515	AtUBC37
NM_001160817.1	NP_001154289.1	AT4G36800	AtRCE1
NM_127416.3	NP_565440.1	AT2G18600	AtRCE2/AtUB12L
NM_112075.2	NP_566423.1	AT3G12400	AtELC

NM_121389.2	NP_196890.1	AT5G13860	AtELCL(ELC-Like)
NM_115649.2	NP_191346.1	AT3G57870	AtSCE1
NM_102517.4	NP_564289.1	AT1G27530	AtUfc1
NM_128839.2	NP_565754.1	AT2G32790	AtUBCD/E
NM_112201.4	NP_566459.2	AT3G13550	AtCOP10
NM_180353.2	NP_850684.1	AT3G52560	AtUEV1D
NM_105734.3	NP_564994.1	AT1G70660	AtUEV1B
NM_129165.3	NP_565834	AT2G36060	AtUEV1C
NM_102175.3	NP_564191	AT1G23260	AtUEV1A

Tomato UBCs

Chromosome loci	Gene name
Solyc08g081270.2.1	SIUBC 6
Solyc05g054550.2.1	SIUBC 7
Solyc12g056100.1.1	SIUBC 8
Solyc08g008220.2.1	SIUBC 9
Solyc05g050230.2.1	SIUBC 10
Solyc03g033410.2.1	SIUBC 11
Solyc07g066080.2.1	SIUBC 12
Solyc07g062570.2.1	SIUBC 13
Solyc10g007260.2.1	SIUBC 13-2
Solyc04g011430.2.1	SIUBC 14
Solyc11g071870.1.1	SIUBC 15
Solyc04g080810.2.1	SIUBC 16
Solyc02g084760.2.1	SIUBC 17
Solyc04g079970.2.1	SIUBC 18
Solyc12g089030.1.1	SIUBC 19
Solyc11g065190.1.1	SIUBC 20
Solyc11g071260.1.1	SIUBC 21
Solyc10g081160.1.1	SIUBC 22
Solyc01g111680.2.1	SIUBC 23
Solyc02g078210.2.1	SIUBC 24
Solyc10g007000.2.1	SIUBC 25
Solyc01g079290.1.1	SIUBC 26
Solyc06g072570.2.1	SIUBC 27
Solyc10g011740.2.1	SIUBC 28
Solyc02g083570.2.1	SIUBC 29
Solyc03g007470.2.1	SIUBC 30
Solyc01g095490.2.1	SIUBC 31
Solyc12g099310.1.1	SIUBC 32
Solyc03g123660.2.1	SIUBC 33
Solyc06g063100.2.1	SIUBC 34
Solyc05g054540.2.1	SIUBC 35
Solyc09g009720.1.1	SIUBC 36
Solyc07g024070.1.1	SIUBC 37
Solyc08g081950.2.1	SIUBC 38

Solyc06g082600.2.1	SIUBC 39
Solyc06g007510.2.1	SIUBC 40
Solyc02g087750.2.1	SIUBC 41
Solyc03g112720.2.1	SIUBC 43
Solyc02g093110.2.1	SIUBC 44
Solyc04g078620.2.1	SIUBC 45
Solyc12g088680.1.1	SIUBC 46
Solyc03g044260.2.1	SIUBC 47
Solyc01g005840.2.1	SICOP 10
Solyc04g007970.2.1	SIUEV1B
Solyc01g007860.2.1	SIUEV1C
Solyc10g083120.1.1	SIUEV1D

***N. benthamina* ubiquitin E2s**

Sequence ID in sol genomics network

(http://solgenomics.net/)	Gene name
Niben101Scf05166g02005.1	<i>NbUBC1</i>
Niben101Scf03194g01006.1	<i>NbUBC2</i>
Niben101Scf02253g03005.1	<i>NbUBC3</i>
Niben101Scf05118g08003.1	<i>NbUBC4</i>
Niben101Scf07327g02015.1	<i>NbUBC5</i>
Niben101Scf04988g01009.1	<i>NbUBC6</i>
Niben101Scf06359g00012.1	<i>NbUBC7</i>
Niben101Scf06668g00002.1	<i>NbUBC8</i>
Niben101Scf12932g00011.1	<i>NbUBC9</i>
Niben101Scf19214g00010.1	<i>NbUBC10</i>
Niben101Scf01664g02017.1	<i>NbUBC11</i>
Niben101Scf05528g01009.1	<i>NbUBC12</i>
Niben101Scf01002g13002.1	<i>NbUBC13</i>
Niben101Scf00398g00015.1	<i>NbUBC13-2</i>
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Niben101Scf03886g02005.1	<i>NbUBC27</i>
Niben101Scf00470g04003.1	<i>NbUBC28</i>
Niben101Scf01664g02017.1	<i>NbUBC29</i>
Niben101Scf08278g00004.1	<i>NbUBC30</i>

Niben101Scf05584g02011.1	<i>NbUBC31</i>
Niben101Scf02111g11001.1	<i>NbUBC32</i>
Niben101Scf00240g02004.1	<i>NbUBC33</i>
Niben101Scf00870g02009.1	<i>NbUBC34</i>
Niben101Scf06359g00012.1	<i>NbUBC35</i>
Niben101Scf09492g00008.1	<i>NbUBC36</i>
Niben101Scf00936g00004.1	<i>NbUBC37</i>
Niben101Scf04673g02005.1	<i>NbUBC38</i>
Niben101Scf00262g04002.1	<i>NbUBC39</i>
Niben101Scf04436g07003.1	<i>NbUBC40</i>
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Human ubiquitin E2s

NCBI Accession number of proteins	Protein name
P49459.2	UBE2A(hHR6A)
CAG28562.1	UBE2B(hHR6B)
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P62837.1	UBE2D2(UbcH5B)
CAG33197.1	UBE2D3(UbcH5C)
Q9Y2X8.1	UBE2D4(HBUCE1)
NP_003332.1	UBE2E1(UbcH6)
CCQ43860.1	UBE2E2
NP_872619.1	UBE2E3(UbcH9)
Q5U203.2	UBE2F(NCE2)
NP_003333.1	UBE2G1(UBE2G)
NP_003334.2	UBE2G2(UBC7)
NP_003335.1	UBE2H(UBCH)
NP_919237.1	UBE2I(Ubc9)
NP_057105.2	UBE2J1(NCUBE1)
NP_919296.1	UBE2J2(NCUBE2)
NP_005330.1	UBE2K(HIP2)
CAG30492.1	UBE2L3(UbcH7)
CAG33407.1	UBE2L6(UbcH8)
NP_003960.1	UBE2M(Ubc12)
NP_003339.1	UBE2N(Ubc13)
NP_071349.3	UBE2O(E2-230K)
Q7Z7E8.1	UBE2Q1(NICE-5)
NP_775740.1	UBE2Q2
P49427.2	UBE2R1(CDC34)
Q6ZWZ2.1	UBE2R2(CDC34B)
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NP_689702.1	UBE2U
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Q15819.4
AAH10900.1
Q9H832.2
Q9NR09.2

UBE2V2(MMS2)
UBE2W
UBE2Z(Use1)
BIRC6(apollon)

Supplemental Table II List of primers used in this study

Name	Sequence(5'-3')	purpose
SI-UBC8-EcoRI-F	ACGGATCCATGGCATCCAAGCGGATTC	SI-UBC8 in the pGEX-4T-1 vector
SI-UBC8-XhoI-R	CGCTCGAGCTATCCCATGGCAAATTTTTC	SI-UBC8 in the pGEX-4T-1 vector
SI-UBC14-GW-F	CACCATGGCTTACACAAGCTAGTC	SI-UBC14 ORF gateway cloning in pDEST17 vector
SI-UBC14-GW-R	TTCTCGAGCTACATTTCTTGACCGTC	SI-UBC14 ORF gateway cloning in pDEST17 vector
SI-UBC31-EcoRI-F	TAGAATTCATGGCTTCGAAACGGATATT	SI-UBC31 in the pGEX-4T-1 vector
SI-UBC31-XhoI-R	TTCTCGAGTTAACCCATTGCATATTTCTGG	SI-UBC31 in the pGEX-4T-1 vector
SI-UBC39-EcoRI-F	AGGAATTCATGGCGTCGAAAGCGCATAT	SI-UBC39 in the pGEX-4T-1 vector
SI-UBC39-XhoI-R	TGCTCGAGTTATCCCATCGCATATTTTGA	SI-UBC39 in the pGEX-4T-1 vector
SI-UBC40-EcoRI-F	TTGAATTCATGGCGTCGAAGAGGATATT	SI-UBC40 in the pGEX-4T-1 vector
SI-UBC40-XhoI-R	AACTCGAGTCATCCCATTCATATTTCTGA	SI-UBC40 in the pGEX-4T-1 vector
SI-UBC36-F-GW	CACCATGGCTTCTTACACAAGCCG	SI-UBC36 ORF gateway cloning in pDEST17 vector
SI-UBC36-R	CGCTCGAGTCACAACATCTCTTGATTTTC	SI-UBC36 ORF gateway cloning in pDEST17 vector
SI-UBC3-EcoRI-F	AGGAATTCATGTTCGACACCGCGGAAG	SI-UBC3 in the pGEX-4T-1 vector
SI-UBC3-XhoI-R	CGCTCGAGTCAGCTGTGTCTGCCAGCTT	SI-UBC3 in the pGEX-4T-1 vector
SI-UBC5-EcoRI-F	TAGGATCCATGTCTTCTCCAAGCAAACG	SI-UBC5 in the pGEX-4T-1 vector
SI-UBC5-XhoI-R	TGCTCGAGTCATGGATCAACAGGGCCT	SI-UBC5 in the pGEX-4T-1 vector
SI-UBC6-EcoRI-F	CAGAATTCATGTCTTCCCCTAGCAAACG	SI-UBC6 in the pGEX-4T-1 vector
SI-UBC6-XhoI-R	TGCTCGAGTTAGGGATCTGCTTTTCCAG	SI-UBC6 in the pGEX-4T-1 vector
SI-UBC12-F-GW	CACCATGGCTTCAAAGAGGATTCAG	SI-UBC12 ORF gateway cloning in pDEST17 vector
SI-UBC12-R	CGCTCGAGTCAACCCATTGCGTATTTCT	SI-UBC12 ORF gateway cloning in pDEST17 vector
SI-UBC16-BamHI-F	GAGGATCCATGACTAGTGTCTTCTGCTTC	SI-UBC16 in the pGEX-4T-1 vector
SI-UBC16-XhoI-R	CACTCGAGTACACTTATCGTCATGGAA	SI-UBC16 in the pGEX-4T-1 vector
SI-UBC17-BamHI-F	AAGGATCCATGTTCGGCTCCTCTGCC	SI-UBC17 in the pGEX-4T-1 vector
SI-UBC17-XhoI-R	TGCTCGAGTCACACCTTATCATCATGGAA	SI-UBC17 in the pGEX-4T-1 vector
SI-UBC20-BamHI-F	GTGGATCCATGGCGACAATGAACAGTGG	SI-UBC20 in the pGEX-4T-1 vector
SI-UBC20-XhoI-R	CGCTCGAGTACACACTAGGCTTGTATAG	SI-UBC20 in the pGEX-4T-1 vector
SI-UBC27-BamHI-F	GAGGATCCATGGTGGACTTGGCTAGGG	SI-UBC27 in the pGEX-4T-1 vector
SI-UBC27-XhoI-R	CGCTCGAGTTAGCTGGACAACAGCTTTTC	SI-UBC27 in the pGEX-4T-1 vector
SI-UBC32-BamHI-F	GTGGATCCATGGCGGAAGACAAGTATAAT	SI-UBC32 in the pGEX-4T-1 vector
SI-UBC32-XhoI-R	CGCTCGAGTTACGATTCATCCATAAAGACA	SI-UBC32 in the pGEX-4T-1 vector
SI-UBC1-EcoRI-F	CGGAATTCATGTTCGACTCCAGCT	SI-UBC1 in the pGEX-4T-1 vector
SI-UBC1-XhoI-R	AACTCGAGTCAGTCAGCAGTCCA	SI-UBC1 in the pGEX-4T-1 vector
SI-UBC2-EcoRI-F	CGGAATTCATGTCAACTCCTTCA	SI-UBC2 in the pGEX-4T-1 vector
SI-UBC2-XhoI-R	AACTCGAGTCAGTCTGCAGTCCA	SI-UBC2 in the pGEX-4T-1 vector
SIUbc7-GW-F	CACCATGGCTTCAACTTCTCCTTC	SI-UBC7 ORF gateway cloning in pDEST15 vector
SIUbc7-GW-R	TTACATCATTTCTTGAGACCG	SI-UBC7 ORF gateway cloning in pDEST15 vector
SI-UBC15-EcoRI-F	CGGAATTCATGTCTTCTCCAAGC	SI-UBC15 in the pGEX-4T-1 vector
SI-UBC15-XhoI-R	AACTCGAGTCAAGATCAGCATG	SI-UBC15 in the pGEX-4T-1 vector
SI-UBC21-EcoRI-F	CGGAATTCATGCAGGCTTCAAGG	SI-UBC21 in the pGEX-4T-1 vector
SI-UBC21-XhoI-R	AACTCGAGTTAGCCCTTCTTGGG	SI-UBC21 in the pGEX-4T-1 vector
SI-UBC22-EcoRI-F	CGGAATTCATGGCAACTAATGAA	SI-UBC22 in the pGEX-4T-1 vector
SI-UBC22-XhoI-R	CGCTCGAGTTATAATCTCTTCAA	SI-UBC22 in the pGEX-4T-1 vector
SI-UBC41-EcoRI-F	CGGAATTCATGTTCGACGCCGGCT	SI-UBC41 in the pGEX-4T-1 vector
SI-UBC41-XhoI-R	AACTCGAGTCAGTCCGCCGTCCA	SI-UBC41 in the pGEX-4T-1 vector
SI-UBC10-ORF-F	CACCATGGCTTCGAAACGAATATTGAA	SI-UBC10 ORF gateway cloning in pDEST15 vector
SI-UBC10-ORF-R	TTAACCATGGCATACTCTG	SI-UBC10 ORF gateway cloning in pDEST15 vector
SI-UBC35-ORF-F	CACCATGGCTTACAGCTTCTCCTTC	SI-UBC35 ORF gateway cloning in pDEST15 vector
SI-UBC35-ORF-R	TTACGTCATTTCTTGGGACCG	SI-UBC35 ORF gateway cloning in pDEST15 vector
SI-UBC26-ORF-F	CACCATGGATGAGGCAAACAAGAAC	SI-UBC26 ORF gateway cloning in pDEST15 vector
SI-UBC26-ORF-R	TTAACTAACCCCTTGGCTGTTTGA	SI-UBC26 ORF gateway cloning in pDEST15 vector
SI-UBC25-ORF-F	CACCATGGAGACTCATAAACAAGTAG	SI-UBC25 ORF gateway cloning in pDEST15 vector
SI-UBC25-ORF-R	GCTACTCAGTCCCATTCTGT	SI-UBC25 ORF gateway cloning in pDEST15 vector
SI-UBC28-ORF-F	CACCATGGCTTCTAAGCGGATATTG	SI-UBC28 ORF gateway cloning in pDEST15 vector
SI-UBC28-ORF-R	TTAACCATAGCATACTCTG	SI-UBC28 ORF gateway cloning in pDEST15 vector
SI-UBC33-ORF-F	CACCATGGCAGAAAAAGCATGTGTAA	SI-UBC33 ORF gateway cloning in pDEST15 vector
SI-UBC33-ORF-R	TCAAAGCTGAAGCAGAGGCA	SI-UBC33 ORF gateway cloning in pDEST15 vector
SI-UBC13-ORF-F	CGGAATTCATGGCTAACAGC	SI-UBC13 in the pGEX-4T-1 vector
SI-UBC13-ORF-R	CCGCTCGAGTCATGCACCACTAG	SI-UBC13 in the pGEX-4T-1 vector
SI-UBC13-2-ORF-F	CGGAATTCATGGCTAACAGC	SI-UBC13-2 in the pGEX-4T-1 vector
SI-UBC13-2-ORF-R	CCGCTCGAGTCATGCACCACTAG	SI-UBC13-2 in the pGEX-4T-1 vector
SI-UBC4-GW-F	CACCATGTCTTCCCCAAGCAAAG	SI-UBC4 ORF gateway cloning in pDEST15 vector

SI-UBC4-GW-R	TTATGGATCAACAGGTCCTG	SI-UBC4 ORF gateway cloning in pDEST15 vector
SI-UBC7-GW-F	CACCATGGCTTCAACTTCTCCTTC	SI-UBC7 ORF gateway cloning in pDEST15 vector
SI-UBC7-GW-R	TTACATCATTTCTTGAGACCG	SI-UBC7 ORF gateway cloning in pDEST15 vector
SI-UBC9-GW-F	CACCATGGCATCCAAGAGGATTCT	SI-UBC9 ORF gateway cloning in pDEST15 vector
SI-UBC9-GW-R	CTAACCCATTGCATACTTTTG	SI-UBC9 ORF gateway cloning in pDEST15 vector
SI-UBC11-GW-F	CACCATGGCATCAAGGAGAATTCAA	SI-UBC11 ORF gateway cloning in pDEST15 vector
SI-UBC11-GW-R	TCAATTCATAGCATATTTTTGGG	SI-UBC11 ORF gateway cloning in pDEST15 vector
SI-UBC23-GW-1F	CACCATGGATGAGTCTGAGTCTAC	SI-UBC23 ORF gateway cloning in pDEST15 vector
SI-UBC23-GW-1R	TCCTGCCAAGCAACATCAAC	SI-UBC23 ORF gateway cloning in pDEST15 vector
SI-UBC23-GW-2F	AGATCCGCAAAGTTGTGGTT	SI-UBC23 ORF gateway cloning in pDEST15 vector
SI-UBC23-GW-2R	GCAGCAGGTAGCCTTGGGA	SI-UBC23 ORF gateway cloning in pDEST15 vector
SI-UBC23-GW-3F	GATTCATGGTCCCAAAGGTC	SI-UBC23 ORF gateway cloning in pDEST15 vector
SI-UBC23-GW23-3R	CTACAATTGGTGGAGATGTTG	SI-UBC23 ORF gateway cloning in pDEST15 vector
SI-UBC24-GW-1F	CACCATGGATACATCTTAAGTGAC	SI-UBC24 ORF gateway cloning in pDEST15 vector
SI-UBC24-GW-1R	GCCAAACAATAATACCGATACA	SI-UBC24 ORF gateway cloning in pDEST15 vector
SI-UBC24-GW-2F	CTGTTTAGGTGATGCGGTTT	SI-UBC24 ORF gateway cloning in pDEST15 vector
SI-UBC24-GW-2R	TTAATCGGACAACACTGACTGC	SI-UBC24 ORF gateway cloning in pDEST15 vector
SI-UBC29-GW-F	CACCATGGCATCCAGGAGAATTCA	SI-UBC29 ORF gateway cloning in pDEST15 vector
SI-UBC29-GW-R	TCAGTTCATGGCATACTTTTG	SI-UBC29 ORF gateway cloning in pDEST15 vector
SI-UBC30-GW-F	CACCATGGCTTCCAAGCGGATCT	SI-UBC30 ORF gateway cloning in pDEST15 vector
SI-UBC30-GW-R	TTAGCCCATGGCATACTTCT	SI-UBC30 ORF gateway cloning in pDEST15 vector
SI-UBC34-GW-F	CACCATGGCAGAAAAGGCATGTGT	SI-UBC34 ORF gateway cloning in pDEST15 vector
SI-UBC34-GW-R	TCAAAGCTGAAGTAGCGGCA	SI-UBC34 ORF gateway cloning in pDEST15 vector
SI-UBC37-GW-1F	CACCATGGCTCAAGAGGCGCGG	SI-UBC37 ORF gateway cloning in pDEST15 vector
SI-UBC37-GW-1R	TGAAATCGGTTGACTGAAGC	SI-UBC37 ORF gateway cloning in pDEST15 vector
SI-UBC37-GW-2F	CAGGCTATAAGCAATTCAGG	SI-UBC37 ORF gateway cloning in pDEST15 vector
SI-UBC37-GW-2R	TCAAGTATTGGATAAAAAGATTG	SI-UBC37 ORF gateway cloning in pDEST15 vector
SI-UBC38 ^{new} -GW-F	CACCATGGCGTCCAAGCGGATTCT	SI-UBC38 ORF gateway cloning in pDEST15 vector
SI-UBC38-GW-R	CTAACCCATGGCGTACTTTT	SI-UBC38 ORF gateway cloning in pDEST15 vector
Nb-UBC9-GW-F	CACCATGGCATCCAAGAGGATTCT	Group III VIGS fragment Nb-UBC9 cloning
Nb-UBC9-VIGS-R	TGTAGCCTGCCAGTGAACAT	Group III VIGS fragment Nb-UBC9 cloning
Nb-UBC28-VIGS-F	GAAGATATGTTTCACTGGCA	Group III VIGS fragment Nb-UBC28 cloning
Nb-UBC28-VIGS-R	TATTTATATTTGGATGAAAACTTTTG	Group III VIGS fragment Nb-UBC28 cloning
Nb-UBC11-VIGS-F	CAAAAGTATTCCATCCAATATA	Group III VIGS fragment Nb-UBC11 cloning
Nb-UBC11-VIGS-R	TCGTCTGGATTTGGATCTGT	Group III VIGS fragment Nb-UBC11 cloning
Nb-UBC39-VIGS-F	ACAGATCCAAACCCAGACGA	Group III VIGS fragment Nb-UBC39 cloning
Nb-UBC39-R	TGCTCGAGTTATCCCATCGCATATTTTGA	Group III VIGS fragment Nb-UBC39 cloning
Nb-UBC12-VIGS-F	ATGGGATAACTCGAGCATCTATCCATTTC	Group III VIGS fragment Nb-UBC12 cloning
Nb-UBC12-VIGS-R	TGCTGTTTATGTTGGGTGG	Group III VIGS fragment Nb-UBC12 cloning
Nb-UBC30-VIGS-F	GGTTTTCCACCCAAACATAAACAGCA	Group III VIGS fragment Nb-UBC30 cloning
Nb-UBC30-VIGS-R	GATTAGGGTCCGTCAGAAGTG	Group III VIGS fragment Nb-UBC30 cloning
Nb-UBC12VIGS-G-F	CACCAGTATTTGTCTGGACATCTTAAAG	VIGS fragment cloning for silencing Nb-UBC12 alone or the four E2 genes Nb-UBC12, 10, 28 and 31.
Nb-UBC12VIGS-G-R	AAGCAAACCTTGGTGAGCAGCACCTTGATACAGT	VIGS fragment cloning for silencing Nb-UBC12 alone or the four E2 genes Nb-UBC12, 10, 28 and 31.
Nb-UBC10VIGS-G-F	CACCAAGGTTTTGCTTTCAATCTGC	VIGS fragment cloning for silencing the three E2 genes Nb-UBC10, 28 and 31 or the four E2 genes Nb-UBC12, 10, 28 and 31.
Nb-UBC10VIGS-G-R	TTATTGTAGCTTGGTGGCAATCTCAGGAACAAGGGG	VIGS fragment cloning for silencing the three E2 genes Nb-UBC10, 28 and 31 or the four E2 genes Nb-UBC12, 10, 28 and 31.
Nb-UBC28VIGS-G-F	CACCAAGCTACAATAATGGGCCC	VIGS fragment cloning for silencing the three E2 genes Nb-UBC10, 28 and 31 or the four E2 genes Nb-UBC12, 10, 28 and 31.
Nb-UBC28VIGS-G-R	AATGGATAGTACTAAAAATAC	VIGS fragment cloning for silencing the three E2 genes Nb-UBC10, 28 and 31 or the four E2 genes Nb-UBC12, 10, 28 and 31.
Nb-UBC11VIGS-F	CACCGACAGCCCTTATGCAGGTGG	VIGS fragment cloning for silencing the three E2 genes Nb-UBC11, 29, 39 and 40.
Nb-UBC11VIGS-R	GAATTCTCTCAGGAGGAAAATGGATGGCC	VIGS fragment cloning for silencing the three E2 genes Nb-UBC11, 29, 39 and 40.
Nb-UBC29VIGS-F	TTTCTCTGAGGAGAATTCTCAAGGAGC	VIGS fragment cloning for silencing the three E2 genes Nb-UBC11, 29, 39 and 40.
Nb-UBC29VIGS-R	CTACTGGACCTGCACTGCATGAAG	VIGS fragment cloning for silencing the three E2 genes Nb-UBC11, 29, 39 and 40.

Nb-UBC39VIGS-F	CACCTTATGCAGGAGGTGTATT	VIGS fragment cloning for silencing the three E2 genes Nb- <i>UBC11</i> , 29, 39 and 40.
Nb-UBC39VIGS-R	TCCAGACAAAATCAGGAGGGAAATGAATTG	VIGS fragment cloning for silencing the three E2 genes Nb- <i>UBC11</i> , 29, 39 and 40.
Nb-UBC40VIGS-F	CTCCTGATTTTGTCTGGATATTCTAAAAGAG	VIGS fragment cloning for silencing the three E2 genes Nb- <i>UBC11</i> , 29, 39 and 40.
Nb-UBC40VIGS-R	AAGGGCTGTCGGTGGATTGGGTCAGTCAATAGGGAG	VIGS fragment cloning for silencing the three E2 genes Nb- <i>UBC11</i> , 29, 39 and 40.
SI-06-ube1-1F	CACCATGCTTCTAGAAAGAGAC	SI- <i>UBA1</i> ORF gateway cloning in pDEST15 vector
SI-06-ube1-1R	GAATGCCAAATGTAGCAGAGG	SI- <i>UBA1</i> ORF gateway cloning in pDEST15 vector
SI-06-ube1-2F	AACGAACCTTGGTCCCTATG	SI- <i>UBA1</i> ORF gateway cloning in pDEST15 vector
SI-06-ube1-2R	CATTCACTTCGGCTGGTGT	SI- <i>UBA1</i> ORF gateway cloning in pDEST15 vector
SI-06-ube1-3F	CCAATGTGCATTTGCATTC	SI- <i>UBA1</i> ORF gateway cloning in pDEST15 vector
SI-06-ube1-3R	GATTACCAAGAAAATTCACGGGA	SI- <i>UBA1</i> ORF gateway cloning in pDEST15 vector
SI-UBC10-XhoI-KpnI-F	GGCTCGAGGTACCATGGCTTCGAAACGAATATTG	SI- <i>UBC10</i> ORF cloning for constructing pA7-nYFP-SI- <i>UBC10</i> and pSPYCE(M)- <i>SIUBC10</i>
SI-UBC10-StuI-PstI-R	GGAGGCCTGCAGACCCATGGCATACTTCTGGG	SI- <i>UBC10</i> ORF cloning for constructing pA7-nYFP-SI- <i>UBC10</i> and pSPYCE(M)- <i>SIUBC10</i>
SI-UBC12-KpnI-XhoI-F	GGTACCCTCGAGATGGCTTCAAAGAGGATTCAG	SI- <i>UBC12</i> ORF cloning for constructing pA7-nYFP-SI- <i>UBC12</i> and pSPYCE(M)- <i>SIUBC12</i>
SI-UBC12-PstI-R	GGGCTGCAGACCCATTGCGTATTCTGGG	SI- <i>UBC12</i> ORF cloning for constructing pA7-nYFP-SI- <i>UBC12</i> and pSPYCE(M)- <i>SIUBC12</i>
SI-UBC25-XhoI-F	CACCTCGAGATGGAGACTCATAAACAAGTAG	SI- <i>UBC25</i> ORF cloning for constructing pA7-nYFP-SI- <i>UBC25</i>
SI-UBC25-SmaI-R	CCCGGGCTCAGTCCCATCTGTGTGTC	SI- <i>UBC25</i> ORF cloning for constructing pA7-nYFP-SI- <i>UBC25</i>
SI-UBC21-XhoI-F	CACCTCGAGATGCAGGCTTCAAGGGCAAGAC	SI- <i>UBC21</i> ORF cloning for constructing pA7-nYFP-SI- <i>UBC21</i>
SI-UBC21-SmaI-R	CCCGGGGCCCTTCTTGGGCATTGC	SI- <i>UBC21</i> ORF cloning for constructing pA7-nYFP-SI- <i>UBC21</i>
SI-UBC16-XhoI-F	CACCTCGAGATGACTAGTGCTTCTGCTTCATC	SI- <i>UBC16</i> ORF cloning for constructing pA7-nYFP-SI- <i>UBC16</i> and pSPYCE(M)- <i>SIUBC16</i>
SI-UBC16-SmaI-R	CCCGGGCACTTTATCGTCATGGAACC	SI- <i>UBC16</i> ORF cloning for constructing pA7-nYFP-SI- <i>UBC16</i> and pSPYCE(M)- <i>SIUBC16</i>
SI-UBA1-XhoI-F	CACCTCGAGATGCTTCTAGAAAGAGACCGG	SI- <i>UBA1</i> ORF cloning for constructing pA7-cYFP-SI- <i>UBA1</i>
SI-UBA1-SmaI-R	CCCGGGACGGAAGTATACAGACACCAG	SI- <i>UBA1</i> ORF cloning for constructing pA7-cYFP-SI- <i>UBA1</i>
AvrptoB-XhoI-KpnI-F	GGCTCGAGGTACCATGGCGGTATCAATAGAGC	<i>AvrPtoB</i> ORF cloning for constructing pA7-cYFP- <i>AvrPtoB</i> and pBTEX-nYFP- <i>AvrPtoB</i>
AvrptoB-StuI-PstI-R	GGAGGCCTGCAGGGGACTATTCTAAAAGCAT	<i>AvrPtoB</i> ORF cloning for constructing pA7-cYFP- <i>AvrPtoB</i> and pBTEX-nYFP- <i>AvrPtoB</i>
Nb-EF1a-F	AGCCTGGTATGGTTGTGACTTTTTG	RT-PCR for reference gene Nb- <i>EF1a</i>
Nb-EF1a-R	CATGGGCTTGGTGGGAATC	RT-PCR for reference gene Nb- <i>EF1a</i>
Nb-UBC8-RT-F	CACCATGGCATCCAACCGATTCTCAAAG	RT-PCR for Nb- <i>UBC8</i>
Nb-UBC8-RT-R	AGCCCCTCCAGAGATGGTCACT	RT-PCR for Nb- <i>UBC8</i>
Nb-UBC9-RT-F	CACCATGGCATCCAAGAGAATTCTGAAAAG	RT-PCR for Nb- <i>UBC9</i>
Nb-UBC9-RT-R	CCAAATATTTGTGTTTCAGCAACTAACCC	RT-PCR for Nb- <i>UBC9</i>
Nb-UBC10-RT-F	CACCATGGCTTCGAAACGAATATTGAA	RT-PCR for Nb- <i>UBC10</i>
Nb-UBC10-RT-R	CCGCCATAGGCAATATTAGCCCA	RT-PCR for Nb- <i>UBC10</i>
Nb-UBC11-RT-F	CACCATGGCATCCAGGAGAATTCA	RT-PCR for Nb- <i>UBC11</i>
Nb-UBC11-RT-R	CAACTCAATTCATAGCAAACCTTTTGG	RT-PCR for Nb- <i>UBC11</i>
Nb-UBC12-RT-F	TCTTACTGTATCCAAGGTGCTGCT	RT-PCR for Nb- <i>UBC12</i>
Nb-UBC12-RT-R	CCAAATGTTTTTCATCCCATGGCATAT	RT-PCR for Nb- <i>UBC12</i>
Nb-UBC28-RT-F	TAGAATTCATGGCTTCGAAACGGATATT	RT-PCR for Nb- <i>UBC28</i>
Nb-UBC28-RT-R	CATGGGTTAAACCGTTACCTATGG	RT-PCR for Nb- <i>UBC28</i>
Nb-UBC29-RT-F	CACCATGGCATCCAGGAGAATTCA	RT-PCR for Nb- <i>UBC29</i>
Nb-UBC29-RT-R	CTTCATGTCTTCAGACTCAGTTCATA	RT-PCR for Nb- <i>UBC29</i>
Nb-UBC30-RT-F	CACCATGGCTTCCAAGCGGATCT	RT-PCR for Nb- <i>UBC30</i>
Nb-UBC30-RT-R	TTAGCCCATGGCATACTTCT	RT-PCR for Nb- <i>UBC30</i>
Nb-UBC31-RT-F	CACCATGGCTTCGAAACGAATATTGAA	RT-PCR for Nb- <i>UBC31</i>
Nb-UBC31-RT-R	GCCGCCATAGGCAATAGTTAGCCC	RT-PCR for Nb- <i>UBC31</i>
Nb-UBC38-RT-F	ACGGATCCATGGCATCCAAGCGGATTC	RT-PCR for Nb- <i>UBC38</i>
Nb-UBC38-RT-R	CTAACCCATGGCGTACTTTT	RT-PCR for Nb- <i>UBC38</i>
Nb-UBC39-RT-F	AGGAATTCATGGCGTCAAGCGCATAT	RT-PCR for Nb- <i>UBC39</i>
Nb-UBC39-RT-R	GGACACTTTCCGCATCATCCCATATA	RT-PCR for Nb- <i>UBC39</i>
Nb-UBC40-RT-F	TTGAATTCATGGCGTCAAGAGGATATT	RT-PCR for Nb- <i>UBC40</i>
Nb-UBC40-RT-R	TCATTACCACAAGCACAATAAGA	RT-PCR for Nb- <i>UBC40</i>
Nb-UBC16-RT-F	GGGCACCAGGAACCTCTGTAT	RT-PCR for Nb- <i>UBC16</i>

Nb-UBC16-RT-R	TGGTATGGGAGTTGGAGTCA	RT-PCR for Nb- <i>UBC16</i>
Nb-UBC17-RT-F	TGGGTGATTGAAGTGATTGGG	RT-PCR for Nb- <i>UBC17</i>
Nb-UBC17-RT-R	TCAAGGGTCAGAAGGACACC	RT-PCR for Nb- <i>UBC17</i>
SI-EF1a-RT-F	TCCAAAGATGGTCAGACCCGTGAA	Real-time PCR for reference gene SI- <i>EF1a</i>
SI-EF1a-RT-R	ATACCTAGCCTTGGAGTACTTGGG	Real-time PCR for reference gene SI- <i>EF1a</i>
SI-UBC8-RT-F	CATTTTGAAGGAACAGTGGAGC	Real-time PCR for SI- <i>UBC8</i>
SI-UBC8-RT-R	CTCCGGGCAGTTGTCTCATA	Real-time PCR for SI- <i>UBC8</i>
SI-UBC9-RT-F	TTCCAAGGTGCTGTGTCAATC	Real-time PCR for SI- <i>UBC9</i>
SI-UBC9-RT-R	ATGGGTCTGAGCAGCAACTAAC	Real-time PCR for SI- <i>UBC9</i>
SI-UBC10-RT-F	TGCTGGGGTGTTTTTTTGGTC	Real-time PCR for SI- <i>UBC10</i>
SI-UBC10-RT-R	AAGGGGTCGTCAGGGTTTGGGT	Real-time PCR for SI- <i>UBC10</i>
SI-UBC11-RT-F	AGGTCCTGTGGCTCAGGATATA	Real-time PCR for SI- <i>UBC11</i>
SI-UBC11-RT-R	CTTAGGGGTTTGAAGGGTAG	Real-time PCR for SI- <i>UBC11</i>
SI-UBC12-RT-F	TCTTACTGTATCCAAGGTGCTGCT	Real-time PCR for SI- <i>UBC12</i>
SI-UBC12-RT-R	GTGTTCAACCCATTGCGTATTCT	Real-time PCR for SI- <i>UBC12</i>
SI-UBC28-RT-F	TACAAGACAGACAGGGCAAATA	Real-time PCR for SI- <i>UBC28</i>
SI-UBC28-RT-R	GGGAAGGTAGAGGACAGAGAGAC	Real-time PCR for SI- <i>UBC28</i>
SI-UBC29-RT-F	CATGCAGTGCAGGTCCAGTAGC	Real-time PCR for SI- <i>UBC29</i>
SI-UBC29-RT-R	CTTGGGAGGTTTGAAGGGTAA	Real-time PCR for SI- <i>UBC29</i>
SI-UBC30-RT-F	AGTCCTTATTCCGGTGGAGTTT	Real-time PCR for SI- <i>UBC30</i>
SI-UBC30-RT-R	CTTCCGTTGCTGTTTATGTTTG	Real-time PCR for SI- <i>UBC30</i>
SI-UBC31-RT-F	AGGTTTTGCTTTCAATTGCTC	Real-time PCR for SI- <i>UBC31</i>
SI-UBC31-RT-R	GGCAGTTGATTTCGTATTGGC	Real-time PCR for SI- <i>UBC31</i>
SI-UBC38-RT-F	TCCTACTTCTGCAGTGTGGT	Real-time PCR for SI- <i>UBC38</i>
SI-UBC38-RT-R	CATTGCTGTTGATGTTCCGGTG	Real-time PCR for SI- <i>UBC38</i>
SI-UBC39-RT-F	TTCCCCCTGATTATCCCTTC	Real-time PCR for SI- <i>UBC39</i>
SI-UBC39-RT-R	GTGGTCTCGTATTGGCCCTGT	Real-time PCR for SI- <i>UBC39</i>
SI-UBC40-RT-F	CCCTTATGCTGGAGGTGTATT	Real-time PCR for SI- <i>UBC40</i>
SI-UBC40-RT-R	ATCATCTGGGTTTGGGTCTGT	Real-time PCR for SI- <i>UBC40</i>
Nb-EF1a-F	TACTGGTGGTTTTGAAGCTG	Real-time PCR for reference gene Nb- <i>EF1a</i>
Nb-EF1a-R	ATACCTAGCCTTGGAGTACTTGGG	Real-time PCR for reference gene Nb- <i>EF1a</i>
Nb-Wrky28-F	GCATTCATGACAAAGAGTGAGGTT	Real-time PCR for Nb- <i>Wrky28</i>
Nb-Wrky28-R	GACATTTTTGACTTGTGCACCTAT	Real-time PCR for Nb- <i>Wrky28</i>
Nb-Pti5-F	CCTCCAAGTTTGAGCTCAGATAGT	Real-time PCR for Nb- <i>Pti5</i>
Nb-Pti5-R	CCAAGAAATTCTCCATGTA	Real-time PCR for Nb- <i>Pti5</i>
Nb-Acre31-F	GAGAAACTGGGATTGCCTGAAGGA	Real-time PCR for Nb- <i>Acre31</i>
Nb-Acre31-R	AACTTGGCCATCGTGATCTTGGTC	Real-time PCR for Nb- <i>Acre31</i>
Nb-Gras2-F	TCATGAGGCGTTACTCGGAGCATT	Real-time PCR for Nb- <i>Gras2</i>
Nb-Gras2-R	TACCTAGCACCAAGCAGATGCAGA	Real-time PCR for Nb- <i>Gras2</i>
AvrPtoB307-smaI-R	CCCGGGTACATGTCTTTCAAGGGCCGTG	<i>AvrPtoB</i> ORF cloning for constructing pSPYNE173- <i>AvrPtoB1-307</i>
SI-UBC8-XhoI-F	CACCCTCGAGATGGCATCCAAGCGGATTCTC	SI- <i>UBC8</i> ORF cloning for constructing pSPYCE(M)- <i>SIUBC8</i>
SI-UBC8-SmaI-R	CCCGGGTCCCATGGCAAATTTTGGAGTC	SI- <i>UBC8</i> ORF cloning for constructing pSPYCE(M)- <i>SIUBC8</i>
SI-UBC9-XhoI-F	CACCCTCGAGATGGCATCCAAGAGGATTCTG	SI- <i>UBC9</i> ORF cloning for constructing pSPYCE(M)- <i>SIUBC9</i>
SI-UBC9-SmaI-R	CCCGGGACCCATTGCATACTTTTGGG	SI- <i>UBC9</i> ORF cloning for constructing pSPYCE(M)- <i>SIUBC9</i>
SI-UBC11-XhoI-F	CACCCTCGAGATGGCATCAAGGAGAATTCAAA	SI- <i>UBC11</i> ORF cloning for constructing pSPYCE(M)- <i>SIUBC11</i>
SI-UBC11-SmaI-R	CCCGGGATTCATAGCATATTTTGGGTC	SI- <i>UBC11</i> ORF cloning for constructing pSPYCE(M)- <i>SIUBC11</i>
SI-UBC28-XhoI-F	CACCCTCGAGATGGCTTCTAAGCGGATATTG	SI- <i>UBC28</i> ORF cloning for constructing pSPYCE(M)- <i>SIUBC28</i>
SI-UBC28-SmaI-R	CCCGGGACCCATAGCATACTTCTGGG	SI- <i>UBC28</i> ORF cloning for constructing pSPYCE(M)- <i>SIUBC28</i>
SI-UBC29-XhoI-F	CACCCTCGAGATGGCATCCAGGAGAATTCAAAAAG	SI- <i>UBC29</i> ORF cloning for constructing pSPYCE(M)- <i>SIUBC29</i>
SI-UBC29-SmaI-R	CCCGGGGTTTCATGGCATACTTTTGGAG	SI- <i>UBC29</i> ORF cloning for constructing pSPYCE(M)- <i>SIUBC29</i>
SI-UBC30-XhoI-F	ggtaccctcgagATGGCTTCCAAGCGGATC	SI- <i>UBC30</i> ORF cloning for constructing pSPYCE(M)- <i>SIUBC30</i>
SI-UBC30-SmaI-R	GGGctcgagGCCCATGGCATACTTCTGGG	SI- <i>UBC30</i> ORF cloning for constructing pSPYCE(M)- <i>SIUBC30</i>
SI-UBC31-XhoI-F	CACCCTCGAGATGGCTTCGAAACGGATATTG	SI- <i>UBC31</i> ORF cloning for constructing pSPYCE(M)- <i>SIUBC31</i>

SI-UBC31-SmaI-R	CCCGGGACCCATTGCATATTTCTGGG	SI-UBC31 ORF cloning for constructing pSPYCE(M)-SIUBC31
SI-UBC38-XhoI-F	CACCCTCGAGATGGCGTCCAAGCGGATTCTG	SI-UBC38 ORF cloning for constructing pSPYCE(M)-SIUBC38
SI-UBC38-SmaI-R	CCCGGGACCCATGGCGTACTTTTGGG	SI-UBC38 ORF cloning for constructing pSPYCE(M)-SIUBC38
SI-UBC39-XhoI-F	CACCCTCGAGATGGCGTCAAGCGCATATTG	SI-UBC39 ORF cloning for constructing pSPYCE(M)-SIUBC39
SI-UBC39-SmaI-R	CCCGGGTCCCATCGCATATTTTGGAG	SI-UBC39 ORF cloning for constructing pSPYCE(M)-SIUBC39
SI-UBC40-XhoI-F	CACCCTCGAGATGGCGTCAAGAGGATATTG	SI-UBC40 ORF cloning for constructing pSPYCE(M)-SIUBC40
SI-UBC40-SmaI-R	CCCGGGTCCCATTGCATATTTCTGAG	SI-UBC40 ORF cloning for constructing pSPYCE(M)-SIUBC40
SIUBC8C85G-F	GGCAGCATTgGCCTTGACATT	SI-UBC8C85G ORF gateway cloning in pDEST15 vector
SIUBC8C85G-R	AATGTCAAGGcAATGCTGCC	SI-UBC10C85G ORF gateway cloning in pDEST15 vector
SIUBC10C85G-F	CAATGGGAGTATAgGCTTGGAC	SI-UBC10C85G ORF gateway cloning in pDEST15 vector
SIUBC10C85G-R	CAGTATGTCCAAGcTATACTC	SI-UBC10C85G ORF gateway cloning in pDEST15 vector