## SUPPLEMENTARY ONLINE MATERIALS

for

Stefan Arenz<sup>1,†</sup>, Maha Abdelshahid<sup>1,†</sup>, Daniel Sohmen<sup>1,†</sup>, Roshani Payoe<sup>2</sup>, Agata L. Starosta<sup>1,§</sup>, Otto Berninghausen<sup>1</sup>, Vasili Hauryliuk<sup>2,3</sup>, Roland Beckmann<sup>1,4</sup>, Daniel N. Wilson<sup>1,4,\*</sup>

<sup>1</sup>Gene Center and Department for Biochemistry, University of Munich, Munich, 81377, Germany

<sup>2</sup> University of Tartu, Institute of Technology, Nooruse 1, 50411 Tartu, Estonia

<sup>3</sup> Laboratory for Molecular Infection Medicine Sweden (MIMS), Umeå University, Building 6K and 6L, University Hospital Area, SE-901 87 Umeå, Sweden

<sup>4</sup> Center for integrated Protein Science Munich (CiPSM), University of Munich, Munich, 81377, Germany

\* To whom correspondence should be addressed. Tel: +49 89 2180 76903; Fax: +49 89 2180 76945; Email: wilson@Imb.uni-muenchen.de.

§ Present Address: Centre for Bacterial Cell Biology, Institute for Cell and Molecular Biosciences, University of Newcastle, Newcastle upon Tyne, United Kingdom.

<sup>†</sup> These authors contributed equally to the paper as first authors.



Figure S1 Activity of *E. coli* RelA to synthesize ppGpp. Cognate deacylated tRNA strongly activates ppGpp synthesis by RelA. Time courses of <sup>3</sup>H-ppGpp synthesis by RelA in the presence (filled circles) or absence (empty circles) of 1.5  $\mu$ M deacylated tRNA<sup>Phe</sup>. In both cases the reaction mixture contained 0.5  $\mu$ M 70S programmed with 1  $\mu$ M MF-mRNA and 1.5  $\mu$ M tRNA<sup>Met</sup>, 100 nM RelA, 100  $\mu$ M ppGpp, 0.3 mM <sup>3</sup>H-GDP and 1 mM ATP. Results are shown as mean values of 3 replicates and error bars indicate standard error of the mean.



**Figure S2:** *In silico* **sorting scheme of the ReIA-SRC cryo-EM dataset.** After removal of nonaligning and edge particles, sorting of the dataset yielded four homogenous sub-datasets. The first (58%; 116,800 particles) contained stoichiometric density for A-, P- and E-tRNAs, the second (18%; 37,629 particles) contained stoichiometric density for A- and P-tRNAs, the third (11%; 21,985 particles) contained stoichiometric density for P-tRNA and the fourth sub-dataset (13%; 24,749 particles) contained stoichiometric density for ReIA, A/R-tRNA and P-tRNA (ReIA-SRC).



**Figure S3: Resolution of the cryo-EM reconstruction of the ReIA-SRC.** (A) Fourier-shell correlation curve of the refined final map, indicating the average resolution of the ReIA-SRC is 3.7 Å. (B,C) (B) Overview and (C) slice through of the ReIA-SRC colored according to the local resolution as calculated using ResMap (1). (D,E) Different views showing the rigid body-fitted homology model of the TGS subdomain of ReIA (green, PDB2EKI) into the cryo-EM density (grey mesh). (F,G) Different views showing the rigid body-fitted homology model of the ACT subdomain of ReIA (orange, PDB2KO1) into the cryo-EM density (grey mesh).



Figure S4: Secondary structure prediction for subdomains of *E. coli* ReIA. The amino acid (AA) sequence of *E. coli* ReIA with the HD (magenta), SYNTH (pink), TGS (green), HELICAL (blue), CC (gold) and ACT (orange) subdomains is shown together with secondary structure predictions for  $\alpha$ -helices and  $\beta$ -strands (2D) based on PSIPRED (2). The secondary structures predictions for TGS and ACT domains were adjusted based on homology models generated from HHPred (3) and Modeller (4).



**Figure S5: Weblogo plots showing conservation for TGS, CC and ACT subdomains of** *E. coli* **ReIA.** The amino acid number for *E. coli* ReIA subdomains is shown on the x-axis and the figure was generated using the Weblogo server (5).

## SUPPLEMENTARY REFERENCES

- 1. Kucukelbir, A., Sigworth, F.J. and Tagare, H.D. (2014) Quantifying the local resolution of cryo-EM density maps. *Nat Methods*, **11**, 63-65.
- Buchan, D., Minneci, F., Nugent, T., Bryson, K. and Jones, D. (2013) Scalable web services for the PSIPRED Protein Analysis Workbench. *Nucleic Acids Research*, **41** W340-W348.
- 3. Soding, J., Biegert, A. and Lupas, A.N. (2005) The HHpred interactive server for protein homology detection and structure prediction. *Nucleic Acids Res*, **33**, W244-248.
- 4. Eswar, N., Eramian, D., Webb, B., Shen, M.Y. and Sali, A. (2008) Protein structure modeling with MODELLER. *Methods Mol Biol*, **426**, 145-159.
- 5. Crooks, G.E., Hon, G., Chandonia, J.M. and Brenner, S.E. (2004) WebLogo: a sequence logo generator. *Genome Res.*, **14**, 1188-1190.