Supplementary Figures

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Supplementary Figure 1. A worm in the microfluidic device. Worms in the device feed on a bacterial suspension that is continuously flown through the chamber. The pillars restrain the worms from escaping while providing enough space for egg laying. Eggs are collected in an egg-collection area.



Supplementary Figure 2. Pumping dynamics depend on food concentrations. (A) Food concentration dependence of average pump duration time (Δt_{PD}). The bar graph shows average pump duration time (Δt_{PD}) with standard errors at respective food concentrations. (B) The number of pumps observed since the beginning of the experiment (time 0) for 3 different worms at each of 3 densities (OD₆₀₀=0.3, 1, and 3), demonstrating the reproducibility of the assay. (C) Comparison of measured pumping counts (dotted curves) with their chord line (i.e. the lines that connect the two end points of the measured curves). The slopes of these chord lines are the average pumping rates.



Supplementary Figure 3. Bursts of fast pumping are longer and more frequent at higher food densities. (A) A representative time series of pharyngeal pumping of a wild-type (N2) worm at OD₆₀₀=1. Each pumping event is indicated by its instantaneous rate, i.e. the inverse of the time interval between that pump and the next. The time series shows long pauses, fast pumping, and pumping at intermediate rates. Blue and red dashed lines indicate the thresholds for fast pumping (250 pumps/min) and long pauses (100 pumps/min), respectively. (B, C) Survival probability of burst duration (B) and the time between bursts (C) at various food concentrations.



Supplementary Figure 4. Differential roles for the serotonergic neurons NSM and HSN in regulating fast pumping. (A) F(r), the fraction of time spent in intervals longer than 1/r, at OD₆₀₀=3. ADF(TeTx): strain in which the tetanus toxin is expressed from an ADF-specific promoter; ADF(*twk-18*): strain in which the activated form of potassium channel *twk-*18 is expressed from a ADF-specific promoter; All others: strains in which *tph-1* has been removed specifically in the indicated neuron(s) using the Cre/Lox approach. (B) The average fraction of time spent in fast pumping by the same worms at OD₆₀₀=1, compared to wild-type (N2) worms and *tph-1* mutants. Error bars as in Fig. 2. (C-F) Fluorescent images of the transcriptional reporter *tph-1p::GFP* from the HSN neurons (C,D) or the NSM neurons (E,F). Panels C and E show intact neurons, while panels D and F are images from the same region of worms in which the respective neurons were ablated by reconstituted caspase-3. (G, H) Comparison of HSN-ablated and NSM-ablated worms, at OD₆₀₀=3 or at 5mM exogenous 5-HT with no food. (G) F(r), the fraction of time spent in intervals longer than 1/r. (H) The average fraction of time spent in fast pumping. Error bars as in Fig. 2. (I) Comparison of the average fraction of time spent in fast pumping. Error bars as in Fig. 2. Statistical significance as in Fig. 4. (J) The average fraction of time spent in fast pumping in L4 larvae. Error bars as in Fig. 2.



Supplementary Figure 5. (A) F(r), the fraction of time spent in intervals longer than 1/r, at OD₆₀₀=3 for two mutants alleles of *ser-1*, a transgenic rescue, and the *ser-7 ser-1(ok345)* double mutant. (B) The average fraction of time spent by these worms in fast pumping at OD₆₀₀=2. Error bars and thresholds as in Fig. 5. Statistical significance of difference from wild-type behavior indicated as in Fig. 4. (C) Histograms of pump duration (Δt_{PD}) in *ser-4(ok512)* at various food concentrations (colored curves), compared to wild-type (N2) at OD₆₀₀=3 (black, dashed). (D) F(r) for several serotonin receptor mutants, at OD₆₀₀=3. (E) The average fraction of time spent in fast pumping in worms carrying combinations of mutant alleles in serotonin receptors, at OD₆₀₀=3. Error bars as in Fig. 2, statistical significance as in Fig. 4. (F) F(r) for two mutants alleles of *ser-4*, along with *ser-7; ser-4(opy024)* double mutant. (G) Ratio between the average pumping rate of *tph-1* worms and that of N2 worms at various food concentrations. Included for comparison is the same ratio for worms feeding on a lawn of bacteria on agar plates, as measured in Cunningham *et al.*³².



Supplementary Figure 6. Stimulated pumping by exogenous serotonin. (A) F(r), the fraction of time spent by wild-type (N2) worms in intervals longer than 1/r in bacteria-free media supplemented with exogenous serotonin. For comparison, black symbols corresponds to worms feeding on bacteria at OD₆₀₀=3 with no serotonin added. The dotted vertical lines indicate the last point of the linear segment of F(r), taken as the threshold rate for fast pumping. (B) Zoom-in on the low-rate end of **Fig. 3A** demonstrates the presence of long pauses at all food concentrations. (C) A representative time series of serotonin-induced pumping, illustrating the suppression of very long pauses. (D) Average pumping rates are plotted against the fraction of fast pumping at different concentrations of serotonin in bacteria-free media. (E, F) *ser-7 ser-1(ok345)* and *ser-7; ser-4(opy024)* double mutants compared to N2 and single mutant allele *ser-1(ok345), ser-4(opy024)* and *ser-7* in a media supplemented with 5mM 5-HT in the absence of food in (E) long pauses and (F) fast pumping. Statistical significance as in Fig. 4. (G) F(r) for wild-type worms and serotonin receptor mutants in bacteriafree media with 5mM serotonin. Zoom-in on the low-rate end demonstrates the suppression of long pauses. (H) Same as in panel G. Dashed lines are the best-fit lines between 100 pumps/min and 250 pumps/min. Dotted vertical lines as in (A).

Supplementary Tables

Supplementary Table 1. Primers

Primer	Sequence (5'-to-3')	
Transgenic rescue of <i>ser-1</i>		
ser-1 F	agtcgacacatttgccaacc	
ser-1 R	cagtgcttggaaggaggaga	
CRISPR: sgRNA oligos		
ser-1 sg1 F	gagaggagaaggctggccgg gttttagagctagaaatagcaag	
ser-1 sg2 F	gcacgggatgagctcacacgg gttttagagctagaaatagcaag	
ser-4 sg1 F	gaaattctaatttaaggtagg gttttagagctagaaatagcaag	
ser-4 sg2 F	gagccgggaattgtaaagagg gttttagagctagaaatagcaag	
CRISPR: PCR primers for homology arms (us: upstream arm; ds: downstream arm)		
ser-1 us F	aacgacggccagtgaattcccgcgg caagttctagtagcaagttgaactag	
ser-1 us R	ttataggccgcctgatgcgccgcgg gaggaaagcgaaaaagtatg	
ser-1 ds F	atagttgcaggaccactgcggccgc aacgacagaggcatcaacc	
ser-1 ds R	atgattacgccaagcttgcggccgc cacactttggcacaaaactg	
ser-4 us F	aacgacggccagtgaattcccgcgg ggttcaggtatactgttttcag	
ser-4 us R	ttataggccgcctgatgcgccgcgg gaatatatgcttattggggg	
ser-4 ds F	atagttgcaggaccactgcggccgc ggcgaacgaaagagtcgaa	
ser-4 ds R	atgattacgccaagcttgcggccgc ctcacgggttactgtagtctttgg	
CRISPR: primer for PCR-verification of insertion at target site		
ser-1 us F	gggttgtaattttgaaattccagg	
ser-1 ds R	gcaccctatgatctacactttttc	
ser-4 us F	cacgccgagattttcagc	
ser-4 ds R	catatcggctactttcggcc	
Pmyo-2 R	gtgagtcgtattacgtacgaggtacc	
unc-54 F	cattcccaatataccaaacataactg	

Supplementary Table 2. Experiment Details

All experiments were done in two independent biological repeats, unless noted otherwise.

Figure	Experiment Details
Fig. 2, Fig. 3	The numbers of animals used for OD ₆₀₀ =0, 0.5, 1, 2, 3, 8 are 62, 61, 113, 64, 33, 84, respectively.
	5mins/animal. The experiments were repeated more than twice for the followings: 4, 5, 3, 4, 5
	repeats for OD ₆₀₀ =0, 1, 2, 3, 8, respectively.
Fig. 4A	The numbers of animals used for OD ₆₀₀ =0, 0(with 5-HT), 0.5, 1, 1(with 5-HT), 2, 3, 8 are 7, 7, 10,
	13, 8, 9, 18, 17, respectively. 5mins/animal. The experiments were repeated more than twice
	for the followings: OD ₆₀₀ =1, 3, 8.
Fig. 4C	The numbers of animals used are (from left to right) are 33, 18, 14, 43, 17, 44, 28, 18, 47, 17,
	113, 15, respectively. 5mins/animal. The experiments were repeated more than twice for the
	followings: N2=4, HSN;NSM=3.
Fig. 5A	The number of animals used are as follows (in the order of increasing concentration): N2 (as in
	Fig. 2); <i>tph-1</i> (as in Fig. 4); <i>ser-1(ok345)</i> = 19, 23, 27, 23, 34, 18; <i>ser-1(opy027)</i> = 28, 44; <i>ser-</i>
	4(ok512) = 13, 19, 27, 21, 61, 16, 23; ser-4(opy024) = 21, 84; ser-5 = 12, 29, 17, 16; ser-7 = 16, 17, 1
	32, 16, 14, 18; <i>mod-1</i> = 14, 17, 16, 20, 19; <i>ser-5;ser-4;mod-1;ser-7 ser-1</i> = 14, 18, 41, 23, 69.
	5mins/animal.
Fig. 5B	The numbers of animals used are (from left to right) are 33, 18, 34, 42, 61, 84, 16, 14, 20, 69.
	5mins/animal. The experiments were repeated more than twice for the followings: N2 =4, ser-
	1(ok345)=3, ser-4(ok512)=4, ser-7=3, ser-5;ser-4;mod-1;ser-7 ser-1=7.
Fig. 6A, B, C	The number of animals used are as follows (in the order of increasing concentration): N2= 62,
	11, 20, 14, 18, 24; <i>ser-1(ok345)</i> = 19, 13, 13, 14; <i>ser-4(ok512)</i> = 13, 16, 15, 13, 13; <i>ser-4(opy024)</i>
Supp. Fig. 6D	= 30, 15; <i>ser-5</i> = 12, 9, 13, 12; <i>ser-7</i> = 16, 7, 9, 15, 11; <i>mod-1</i> = 14, 9, 10, 13; <i>ser-5;ser-4;mod-1;ser-</i>
	<i>7 ser-1</i> = 14, 8, 7, 19, 11. 5mins/animal. The experiments were repeated more than twice for N2
	(0mM)=4 and N2 (5mM)=3.
Fig. 6D, E	The numbers of animals used are (from left to right) are 18, 7, 14, 12, 13, 30, 12, 15, 10, 19.
	5mins/animal. The experiments were repeated more than twice for the followings: N2 =3, <i>ser</i> -
	7=3, ser-5;ser-4;mod-1;ser-7 ser-1=3.
Supp. Fig. 4B	The numbers of animals used are (from left to right) are 113, 13, 13, 47, 17, 19, 10, 14, 24.
	5mins/animal. The experiments were repeated more than twice for the followings: $N2 = 5$,
	ADF(TeTx)=4, HSN;NSM=3.
Supp. Fig. 4G, H	The numbers of animals used are (from left to right) are 28, 20, 47, 30. 5mins/animal.
Supp. Fig. 4I	The numbers of animals used are (from left to right) are 28, 39, 47, 16. 5mins/animal.
Supp. Fig. 4J	The numbers of animals used are 9(N2), 13(NSM). 5mins/animal.
Supp. Fig. 5B	The numbers of animals used are (from left to right) are 64, 9, 23, 18, 21, 17, 16, 16, 23.
	5mins/animal. The experiments were repeated more than twice for the followings: N2 =3, <i>ser</i> -
	1(ok345)=3.
Supp. Fig. 5E	The numbers of animals used are (from left to right) are 33, 14, 34, 25, 103, 84, 80.
	5mins/animal. The experiments were repeated more than twice for the followings: N2=4

Supp. Fig. 6E, F	The numbers of animals used are (from left to right) are 18, 15, 14, 34, 30, 12. 5mins/animal.
	The experiments were repeated more than twice for the followings: N2 =3, <i>ser-7</i> =3.