# Natriuretic Peptide Receptor B modulates the proliferation of the cardiac cells expressing the Stem Cell Antigen-1.

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#### SUPPLEMENTAL METHODS

#### Cell isolation and proliferation

Neonatal mice were sacrificed by decapitation with surgical scissors and the hearts removed. Hearts (1-2 days-old) were enzymatically digested with 0.45 mg/ml collagenase II (Worthington, Biochemical Corporation, USA) and 1 mg/ml pancreatin (Sigma-Aldrich). Cardiomyocytes were separated from the non-myocyte cells (NMCs) by two differential plating. NMCs were first cultured **in the proliferation medium**: a 3:1 mixture of DMEM and Medium 199 (Invitrogen Corp, San Diego, CA, USA) supplemented with 10% horse serum (Oxoid), 5% fetal bovine serum (FBS) (Invitrogen), 10 mM Hepes, 100 U/ml penicillin G, and 100 microg/ml streptomycin.

#### Western blot

Total proteins were extracted from sorted Sca-1<sup>+</sup> cells after lysis in a buffer containing Nonidet P-40 0.5%, NaCl 150 mM, Na-orthovanadate 1 mM, NaF 10 mM, Tris-HCL pH=7.5 10 mM, PMSF 1 mM, EDTA pH=8 1 mM, aprotinin 10  $\mu$ g/ml, leupeptin 10  $\mu$ g/ml, pepstatin 1  $\mu$ g/ml. 30  $\mu$ g of these total proteins were separated by SDS-PAGE, transferred to nitrocellulose membranes (Biorad), and blocked for 1h at room temperature in Odyssey blocking buffer (1/2 in TBS Li-COR Biosciences).

#### Neonatal and infarcted mice treated with BNP

BNP (1 microg/mouse), CNP (1 microg/mouse), or NaCl was injected intraperitoneally (ip) into 3-4 days old C57BL/6 mice (American Peptide Co, Sunnyvale, CA, US) every 2 days during 2 weeks as previously described <sup>1</sup>. Mice were sacrificed 3 days after the

last injection using ip lethal injection of pentobarbital (150 mg/kg). The breathing arrest was controlled and cervical dislocation was performed.

In adult mice, the left anterior descending coronary artery (LAD) was occluded in 10 weeks-old C57 BL/6 mice to induce MI. Mice were anesthetized by ip injection of ketamine/xylazine/acepromazine (65 mg/kg; 15 mg/kg; 2 mg/kg). Mice were moved to a heated plateform. During surgery, the depth of mouse anesthesia was tested by gentle pinch the legs. Directly after the surgery, NaCl or BNP (1 microg/20g in 20 microl) was injected into the myocardium and thereafter BNP (2 microg/mouse) was injected ip every 2 days. After surgery, temgesic (0.1 mg/kg) was injected subcutaneously to reduce the pain, and injections were performed twice per day for up to 4 days. Mice were sacrificed 10 days after MI induction by lethal injection of pentobarbital (150 mg/kg intraperitoneally). The breathing arrest was controlled and cervical dislocation was performed.

#### REFERENCE

 Bielmann, C. *et al.* Brain natriuretic peptide is able to stimulate cardiac progenitor cell proliferation and differentiation in murine hearts after birth. *Basic Res Cardiol* **110**, 455-72 (2015). **Supplemental Table S1.** Flow cytometric characterization of Non-Myocyte Cells (NMCs) isolated directly from murine hearts or after cell culture.

Age of mice	strain	n	days after isolation	NPR-A+	NPR-B+	c-kit+	Sca-1 <sup>+</sup>	NPR-A⁺ /c-kit⁺	NPR-A <sup>+</sup> /Sca-1 <sup>+</sup>	NPR-B <sup>+</sup> /c-kit <sup>+</sup>	NPR-B <sup>+</sup> /Sca-1 <sup>+</sup>
Adult	WT	6	0	3.3±1.0	6. <b>3</b> ±1.1	6.2±1.1	50.7±2.7	2.1±0.7	1. <b>8</b> ±0.2	3.5±0.7	3.7±0.8
Neonatal	WT	6	0	2.9±0.3	5.6±1.1	2.9±0.3	15.3±1.6	2.9±0.3	2.0±0.6	2.7±0.4	4.1±1.4
Neonatal	WT	5	<b>3</b> °	3.8±0.5*	<b>2.6</b> ±1.0*	2.3±0.9	49.0±4.2*	0.6±0.2*	3.3±0.4	0.6±0.2*	2.1±0.9
Neonatal	NPR-A -/-	3	3		1. <b>9</b> ±0.2	0.7±0.2	64.2±9			0.4±0.1	1.9±0.5
Neonatal	NPR-B-/-	6	3	2.9±0.7		1. <b>3</b> ±0.5	52.2±5.6	0.9±0.2	2.3±0.5		

Data represent the percentage of cells positive for the different markers and are expressed as the mean  $\pm$  SEM of the indicated number of experiments (n). Neonatal and adult non-myocyte cells (NMCs) were analysed directly after isolation (0 days after isolation) or ° 3 days after the first passage at confluence, at the onset of BNP treatment. Cells expressing NPR-A and NPR-B (C57BL/6 or WT cells) were compared to cells expressing either only NPR-B (NPR-A KO mice) or only NPR-A (NPR-B deficient mice). \* p<0.05 between WT cells after isolation and after 3 days after the first passage.

Supplemental Table S2. Antibodies used in flow cytometry analysis, cell sorting,

immunohistology and Western blot analysis.

## Flow Cytometry analysis and Cell sorting

rat-anti Sca-1 PE-Cy5 or FITC	1/50	eBiosciences
rat anti-c-kit PE-Cy5 or FITC	1/50	eBiosciences
rat anti-CD31 APC	1/100	eBiosciences
rat anti-PDGFR- $\alpha$ PE	1/80	eBiosciences
rabbit anti-NPR-A	1/50	Abcam
<ul> <li>+ donkey anti-rabbit FITC</li> </ul>	1/1000	Molecular Probes
goat anti-NPR-B	1/20	Santa Cruz Biotechnology
+ chicken anti-goat FITC	1/500	Molecular Probes

## Immunohistology

	Secondary antibodies		
Abcam	horse anti-goat biotin	1/200	Vector
Santa Cruz Biotechnology	horse anti-rabbit biotin	1/200	Vector
Santa Cruz Biotechnology	donkey anti-rabbit Alexa 594	1/500	1
Abcam	donkey anti-goat Alexa 594	1/1000	
Santa Cruz Biotechnology	donkey anti-rat Alexa 488	1/1000	
Abcam	goat anti-rabbit Alexa 594	1/2000	Molecular
Abcam	donkey anti-rabbit Alexa 488	1/1000	Probes
Becton Dickinson	Steptavidine Alexa 594	1/1000	
Dako	Streptavidine Alexa 647	1/1000	
	chicken anti-goat Alexa 488	1/1000	
Abcam			
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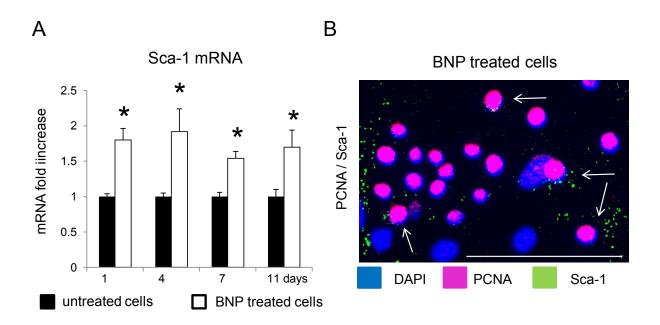
### Western blot analysis

<u>Primary antibodies</u> rabbit anti-phospho- phospholamban	1/1000	Millipore	<u>Secondary antibodies</u> goat anti-rabbit Alexa 680	1/5000	Molecular Probes
mouse anti- phospholamban	1/500	Millipore	Anti-mouse IRDye 800	1/5000 or 1/20000 (for tubulin)	Rockland Immunochemicals
rabbit anti- p38 rabbit anti-phospho p38 Mouse anti-tubulin	1/1000 1/500 1/20 000	Cell Signaling Cell Signaling Sigma		, ,	

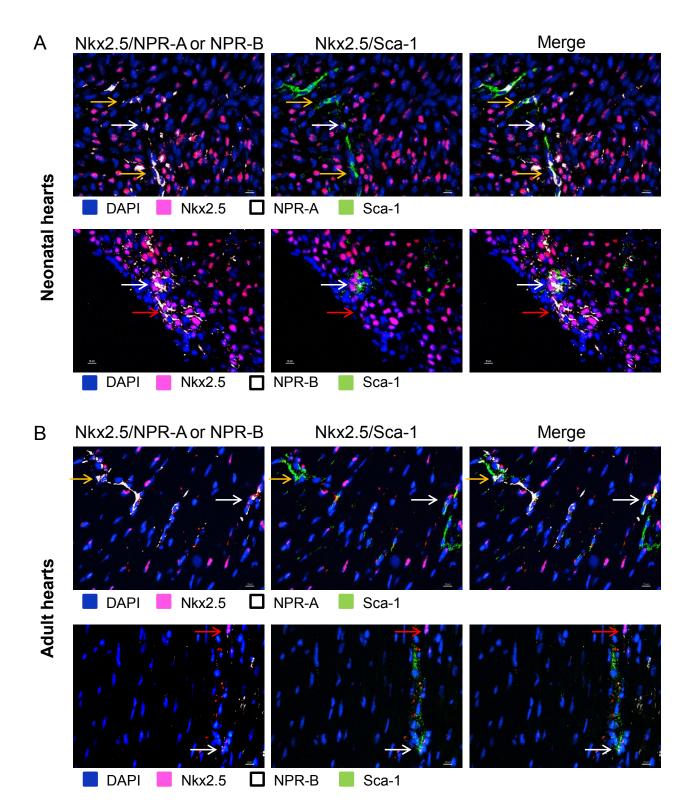
**Supplemental Table S3:** Sequences of primers used in quantitative RT-PCR.

Gene	Sense	Anti-sense	Product size (bp)
Brachyury	CCTCCCTTGTTGCCTTAGAGTAGTT	GCAGATTGTCTTTGGCTACTTTGTC	185
CD90	TGGGTGCAGCAACTGGAGGC	CTCGGGACACCTGCAAGACTGA	181
CD31	GCCTCACCAAGAGAACGGAAGGC	CTGCTTTCGGTGGGGACAGGC	158
c-kit	ATCTGCTCTGCGTCCTGTTG	CTGATTGTGCTGGATGGATG	108
Collagen 1α1	AATGGCACGGCTGTGTGCGA	AACGGGTCCCCTTGGGCCTT	183
Collagen 1α2	GGCCCCCTGGTATGACTGGCT	CGCCACGGGGACCACGAATC	129
DDR2	TTCCCTGCCCAGCGAGTCCA	ACCACTGCACCCTGACTCCTCC	181
eNOS	GGCTGTGGTAGTTAGGGCATC	AGGTTTGGGTTGGGCATCT	165
Gata-4	CTGTCATCTCACTATGGGCA	CCAAGTCCGAGCAGGAATTT	259
Hand2	CCTTCAAGGCGGAGATCAAGA	CCTGTCCGGCCTTTGGTTTT	118
Islet-1	GCCTCAGTCCCAGAGTCATC	AGAGCCTGGTCCTCCTTCTG	308
MIc-2v	GACCCAGATCCAGGAGTTCA	AATTGGACCTGGAGCCTCTT	163
Nanog	CACCCACCCATGCTAGTCTT	ACCCTCAAACTCCTGGTCCT	152
Nkx2.5	CAAGTGCTCTCCTGCTTTCC	GTCCAGCTCCACTGCCTTCT	130
NPR-A	CCAATTATGGCTCCCTGCTA	CGGTACAAGCTCCCACAAAT	198
NPR-B	TCATGACAGCCCATGGGAAA	GGTGACAATGCAGATGTTGG	209
Oct-4	GGATGCTGTGAGCCAAGG	GAACAAAATGATGAGTGACAGACAG	175
PDGFRα	GGGAAGGACTGGAAGCTTGGGGC	AGATGAGGCCCGGCCCTGTGAGG	154
Sca-1	TTTGAGACTTCTTGCCCATC	ACCCAGGATCTCCATACTTTC	159
Sox2	AAGGGTTCTTGCTGGGTTTT	AGACCACGAAAACGGTCTTG	150
Tbx5	GGAAAGATGAGGAATGTTCCAG	GTGTTACAGCTGATGTCCTCCA	223
Tbx20	CCCCGCTGCCAGCCAGGCTCTA	GTGCACCCGTGGCTGGTACTTATGC	167
Tcf21	GGCCAACGACAAGTACGAGA	GCTGTAGTTCCACACAAGCG	129
Troponin T	GCGGAAGAGTGGGAAGAGACA	CCACAGCTCCTTGGCCTTCT	127
Vimentin	GCCGAAAGCACCCTGCAGTCA	GCCTGCAGCTCCTGGATCTCTTCA	146
vWF	GATGCCCCAGTCAGCTCTAC	TCAGCCTCGGACAACATAGA	131
Wnt1	CACGGCACAGGGTATGAGAG	GTTGGGGCCACTCCAGATAC	128

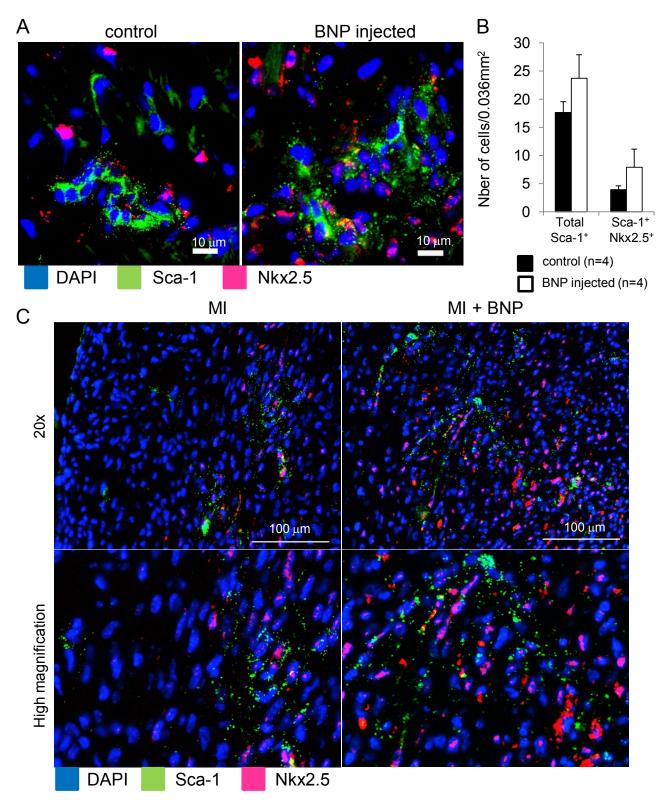
α-ΜΗϹ	AACCAGAGTTTGAGTGACAGAATG	ACTCCGTGCGGATGTCAA	130
β <b>-MHC</b>	ATGAGACGGTGGTGGGTTT	CTTTCTTTGCCTTGCCTTTG	117
18S	ACTTTTGGGGCCTTCGTGTC	GCCCAGAGACTCATTTCTTCTTG	96



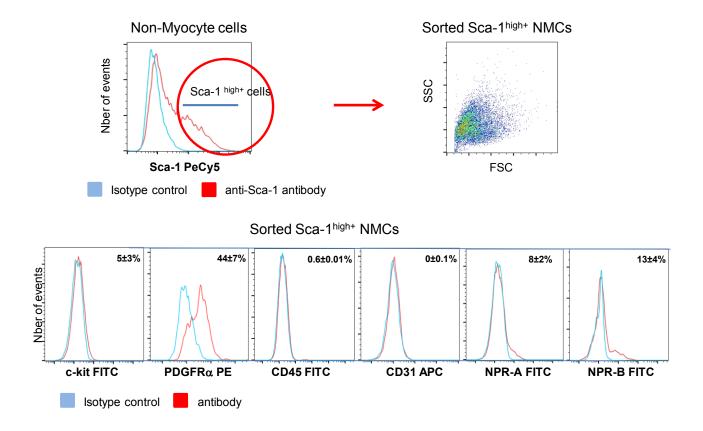
<u>Supplemental Figure 1.</u> BNP induces Sca-1<sup>+</sup> cell proliferation. A. Quantitative relative expression of mRNA coding for Sca-1 in untreated and BNP treated cells. n= at least 7 different experiments per group. B. Representative immunostaining of cells cultured 4 days with BNP and stained with antibodies against the Proliferating Cell Nuclear Antigen (PCNA) (pink), Sca-1 (green) and DAPI (blue). White arrows highlight proliferating Sca-1<sup>+</sup> cells. Scale bar represents 100  $\mu$ m.



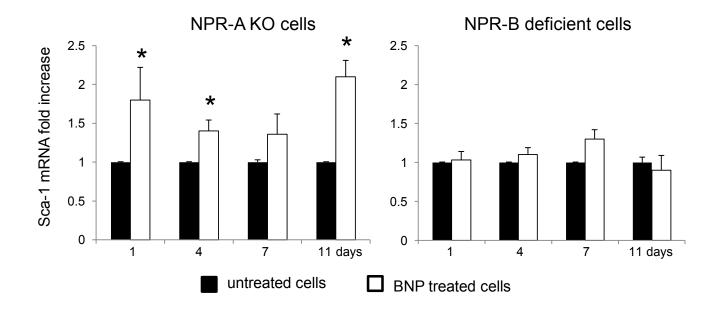
<u>Supplemental Figure 2.</u> NPR-A and NPR-B receptors are expressed on Nkx2.5<sup>+</sup> Sca-1<sup>+</sup> cells in neonatal and adult hearts. Immunostainings of neonatal (A) and adult murine hearts (B). Scale bars represented 10  $\mu$ m. The yellow arrows point to Sca-1<sup>+</sup> Nkx2.5<sup>-</sup> cells expressing NPR-A or NPR-B, the white arrows point to Nkx2.5<sup>+</sup> Sca-1<sup>+</sup> cells and the red arrows point to Sca1<sup>-</sup> cells expressing Nkx2.5 and NPR-B.



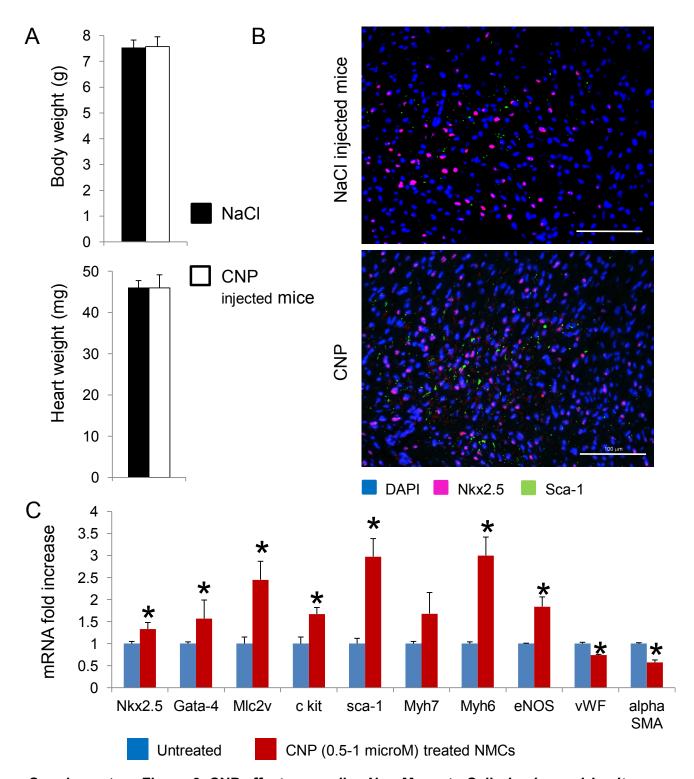
<u>Supplemental Figure 3.</u> BNP treatment in neonatal and infarcted hearts leads to increased number of Sca-1<sup>+</sup> Nkx2.5<sup>+</sup> cells. A. Heart sections from control or BNP injected neonatal mice. B. Quantification of the number of Sca-1<sup>+</sup> cells expressing Nkx2.5 in the hearts of control or BNP injected mice. Data were obtained from at least 6 different pictures per mouse, n = 4 mice per group. Each section covered an area of 0.036 mm<sup>2</sup>. Data are means ± SEM. C. Heart sections, from infarcted mice injected or not with BNP. Representative pictures of x 20 and high magnification of the infarcted area of the infarcted hearts 10 days afer myocardial infarction (MI).



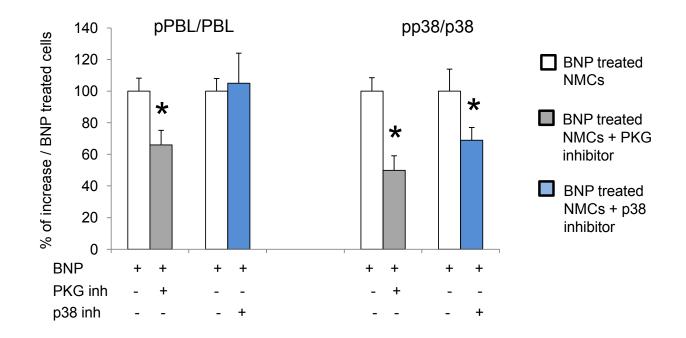
<u>Supplemental Figure 4.</u> Phenotypic characterisation of the sorted Sca-1<sup>high+</sup> cells. Representative histograms and dot plot (right) of non myocyte cell (NMC) sorting for high Sca-1 expression. The histograms allow to characterize the Sca-1<sup>high+</sup> cells for the expression of c-kit, PDGFR $\alpha$ , CD45, CD31 and NPR-A and NPR-B proteins. The numbers represent the percentage of the positive cells compared to the total number of Sca-1<sup>high+</sup>cells and are the means ± SEM of 3-11 different experiments.



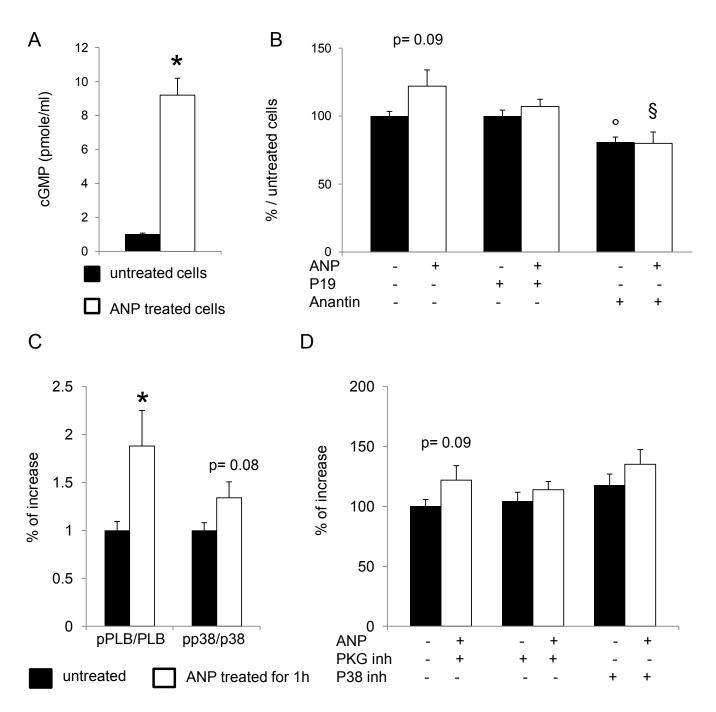
<u>Supplemental Figure 5.</u> BNP induced increased Sca-1 mRNA expression only in presence of NPR-B receptor. Quantitative relative expression of mRNA coding for Sca-1 in untreated and BNP treated NPR-A and NPR-B deficient NMCs (n= at least 6 different experiments at different time point). All the results are means  $\pm$  SEM, \* p<0.05 versus untreated cells.



Supplementary Figure 6. CNP effect on cardiac Non Myocyte Cells *in vivo* and *in vitro*. A-B. Newborn mice (4 days old) were injected with CNP intraperitoneally every 2 days for up to 2 weeks. Mice were sacrificed 2 days after the last injection. CNP treatment had no effect on body or heart weights. However the numbers of Sca-1<sup>+</sup> cells and of Sca-1<sup>+</sup> Nkx2.5<sup>+</sup> cells was increased (B). n=4 mice per group C. Non Myocyte cells were isolated from neonatal hearts and treated with CNP in differentiation medium for up to 3 weeks. In presence of CNP, NMCs upregulated cardiomyocyte specific genes (Nkx2.5, Gata-4, Mlc-2v, Myh6). n= 9 different experiments and \* p< 0.05. vWF: von Willbrand Factor, SMA : smooth muscle actin.



<u>Supplemental Figure 7.</u> BNP induced PKG activation, which is necessary for at least a part of p38 MAPK phosphorylation. NMCs were treated 15-30 min with BNP and with a PKG inhibitor or a p38 inhibitor. n= at least 6 different experiments per group. The data were related to the average of BNP treated NMCs. All the results are means  $\pm$  SEM, \* p<0.05 versus BNP treated cells.



<u>Supplemental Figure 8.</u> ANP has a moderated effect on Sca-1<sup>+</sup> cell proliferation. A. cGMP levels were measured in the non myocyte cell supernatants (n=4-7) 1h after ANP treatment (1µM). B. Sorted Sca-1<sup>high+</sup> cells isolated from wild type hearts were cultured with ANP in presence or not of NPR-B receptor antagonist (P19, 0.5-1 microM) and of an ANF antagonist (Anantin, 0.2 microM). The number of cells was counted after 9-11 days of culture and the results were related to untreated cells. B. Quantification of the data from western blot analysis expressed relatively to the average of untreated cells. Sorted Sca-1<sup>high+</sup> cells were or not treated 1h with ANP. C. Number of sorted Sca-1<sup>high+</sup> cells after 11 days of culture with or without ANP and PKG or p38 inhibitor. The data were related to the average of untreated cells. A-C: n= at least 4 different experiments, All the results are means ± SEM, \* p<0.05 versus untreated cells without inhibitors.