- 1 Supplemental Information
- 2 Mechanistic Study of the Synergistic Antibacterial Activity of Combined Silver3 Nanoparticles and Common Antibiotics
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- 10 Supplemental materials include:
- 11 Figure SI 1. Structures of antibiotics used in current study
- 12 Figure SI 2. Additive antibacterial effect of Ag⁺ and tetracycline against *Salmonella*
- 13 Figure SI 3. UV-vis titration of tetracycline into AgNPs solution
- 14 Table SI 1. Choice of centrifuge speed to separate *Salmonella* cells from AgNPs solutions
- 15 Table SI 2. Effect of antibiotics on Ag⁺ release
- 16 Table SI 3. Effect of *Salmonella* cells on Ag^+ release
- 17 Table SI 4. Estimation of Ag⁺ release from AgNPs
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- 21 Figure SI 1. Structures of ampicillin (AMP), penicillin (PEN), neomycin (NEO),
- 22 kanamycin (KAN), enoxacin (ENO) and tetracycline (TET).
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Figure SI 2. Inhibition of 5 μ M Ag⁺ combined with 0, 0.5, 2, 8, and 16 μ M tetracycline against 1x10⁵ CFU/mL *Salmonella* cells. Average counts were obtained from three repeats.

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29 Additive antibacterial effect of Ag⁺ and tetracycline against Salmonella

As an important control, it needs to be confirmed whether Ag^+ has synergistic effect with antibiotics. Tetracycline was chosen as a representative. 5 μ M Ag^+ was combined with 0, 0.5, 2, 8, and 16 μ M tetracycline to test their inhibition against 1x10⁵ CFU/mL bacterial cells. All experimental conditions remain the same with AgNPs groups. As shown in figure SI 2, all groups inhibit 42.7% - 46.8% of bacterial growth with no significant difference, demonstrating that there is no synergistic effect between Ag^+ and tetracycline.

No evidence shows Ag^+ and tetracycline have interaction to form "complex" like that between AgNPs and tetracycline. Therefore, Ag^+ and tetracycline must carry out antibacterial activity separately even when they are combined together. This means there should be an additive antibacterial effect between these two antibiotics. However, since the *Salmonella* is resistant to tetracycline, the inhibition results observed in this test are actually caused by Ag^+ .





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45 UV-vis titration of tetracycline into AgNPs solution

46 In the experiments of Ag binding to *Salmonella* cells and the Ag⁺ release in the presence 47 of bacterial cells, centrifugation is the key method to separate bacteria cells from AgNPs 48 in solution or separate released Ag⁺ from solutions. If AgNPs would aggregate, this 49 method would not work. To confirm it, 100 µM tetracycline was titrated into 50 µM 50 AgNPs solutions and characterized by UV-vis absorption. The concentration of 51 tetracycline increased from 0 to 33 µM during the titration, but there is no sign of 52 aggregation, as shown in figure SI 2. This proves that in this concentration range of 53 tetracycline used, it does not cause AgNPs to aggregate.

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57 **Table SI 1.** Choice of centrifuge speed to separate *Salmonella* cells from AgNPs 58 solutions. AgNPs (100 μ M) and 10⁶-10⁷ CFU/mL *Salmonella* solutions were used to test 59 how centrifuge spin rate affects the settlement of the cells or the AgNPs. At 200 rpm, 60 nearly 100% cells settle. Then different concentrations of AgNPs solutions were used to 61 test the validity of the separation method of centrifuge at 2,000 rpm for 15 min (below 62 table).

Centrifugation speed, rpm	100 μM AgNPs solution, % remaining in supernatant after centrifugation		10 ⁶ -10 ⁷ CFU/mL <i>Salmonella</i> solution, % remaining in supernatant after centrifugation		
	Average	SD	Average	SD	
0	100.00	0.00	100.00	0.00	
500	99.63	2.40	97.64	2.50	
1000	96.15	2.68	83.58	13.00	
1500	94.09	2.64	30.27	15.34	
2000	88.72	1.28	5.76	5.46	
2500	80.64	1.71	3.63	4.18	
3000	68.64	2.45	0.44	0.83	
3500	60.78	2.26	\	\	

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% remaining in supernatant after centrifugation at 2,000 rpm for 15 min

	Average	SD
AgNPs 100µM	88.72	1.28
AgNPs 50µM	91.58	1.85
AgNPs 20µM	92.47	1.3
AgNPs 10µM	94.19	0.58
AgNPs 5µM	96.31	1.27
AgNPs 1µM *	99.10 *	3.95 *

AgNPs concentrations	1 µM		50 µM		
	Ag^+ , nM	Ag release, %	Ag^+ , nM	Ag release, %	
AgNPs	96.9 ± 20.1	9.7 ± 2.0	1046.1 ± 258.3	2.1 ± 0.5	
AgNPs+10 µM AMP	103.2 ± 27.4	10.3 ± 2.7	1068.1 ± 344.1	2.1 ± 0.7	
AgNPs+10 µM PEN	97.4 ± 43.6	9.7 ± 4.4	1145.3 ± 319.0	2.3 ± 0.6	
AgNPs+10 µM ENO	134.1 ± 34.9	13.4 ± 3.5	1395.6 ± 307.1	2.8 ± 0.6	
AgNPs+10 µM KAN	134.9 ± 36.3	13.5 ± 3.6	1485.3 ± 372.8	3.0 ± 0.7	
AgNPs+10 µM NEO	154.2 ± 27.6	15.4 ± 2.8	1579.3 ± 341.6	3.2 ± 0.7	
AgNPs+10 µM TET	144.8 ± 20.7	14.5 ± 2.1	1446.6 ± 281.5	2.9 ± 0.6	

65 **Table SI 2.** Ag^+ release concentrations and percentages from 1 μ M and 50 μ M AgNPs 66 solutions in presence of various antibiotics (10 μ M).

68 Effect of antibiotics on Ag⁺ release

As shown in table SI 2, ampicillin and penicillin have no significant effect on the Ag⁺ release from 1 μ M or 50 μ M AgNPs solutions. Enoxacin, kanamycin, neomycin and tetracycline enhance Ag⁺ release by about 3-5% and 0.9-1.1% in cases of 1 μ M and 50 μ M AgNPs, respectively. Results both AgNPs concentrations confirm that the interaction between antibiotics and AgNPs (formation of complexes) facilitate the Ag⁺ release from AgNPs.

When the concentration of AgNPs increases from 1 μ M to 50 μ M (50 times increase), the released Ag⁺ concentrations don't increase by 50 times, but by 10 times. And the Ag⁺ release percentage even decreased by about 75%, from 8% decreased to 2% in AgNPs only group. This demonstrates no linear relationship between Ag⁺ release and AgNPs concentration. In other words, AgNPs concentration influences Ag⁺ release but the whole equilibrium system of solution determines the final concentration of Ag⁺.

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Concentrations	1 µM		50 µM		
	Ag^+ , nM	Ag release, %	Ag^+ , nM	Ag release, %	
AgNPs	96.9 ± 20.1	9.7 ± 2.0	1046.1 ± 258.3	2.1 ± 0.5	
AgNPs + Cell	120.8 ± 34.1	12.1 ± 3.4	1181.3 ± 201.3	2.4 ± 0.4	
AgNPs +10 µM PEN	97.4 ± 43.6	9.7 ± 4.4	1145.3 ± 319.0	2.3 ± 0.6	
AgNPs + 10 µM PEN + Cell	122.8 ± 25.7	12.3 ± 2.6	1274.4 ± 305.4	2.5 ± 0.6	
AgNPs + 10µM TET	144.8 ± 20.7	14.5 ± 2.1	1446.6 ± 281.5	2.9 ± 0.6	
AgNPs + 10 µM TET + Cell	182.4 ± 25.8	18.2 ± 2.6	1627.7 ± 203.3	3.3 ± 0.4	

Table SI 3. Ag^+ release concentrations and percentages from 1 μ M and 50 μ M AgNPs solutions in presence of *Salmonella* cells.

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87 Effect of *Salmonella* cells on Ag⁺ release

The presence of Salmonella further enhances Ag⁺ release in all experimental groups 88 regardless of the AgNPs concentrations, as shown in table SI 3. The Ag⁺ ion release from 89 AgNPs depends on the coating agents, size and concentration of AgNPs as well as other 90 91 environmental factors, including temperature, pH values, dissolved dioxygen and protons ¹⁻³. In current experimental conditions, antibiotic molecules that have interaction with 92 93 AgNPs enhance the Ag⁺ ion release from AgNPs. Since AgNPs surface absorbs and is covered with Ag⁺ ions, the interaction between antibiotics and AgNPs facilitates the 94 95 release of Ag⁺ ion. Especially, the competition between antibiotics and citrate causes the leaving of citrate ions, which helps the leaving of Ag⁺ ions from AgNPs by forming 96 citrate complex with Ag^{+ 4}. On the other hand, the presence of antibiotics will bring an 97 98 environment of more dissolved dioxygen and protons, promoting conversion from Ag(0)to Ag(I) ^{5, 6}. For the same reason, the complex biological system of bacterial cells will 99 promote conversion of AgNPs to Ag(I) and trigger enhanced Ag^+ release. 100

Layer of a single AgNP (outside to inside)	Radius r, nm	Surface area, S= $4\pi r^2$, nm ²	Estimated Number, N = $S/S_{Ag}, \times$ 10^4	Total Ag ⁺ upon complete release, \times 10^4	Total Ag^+ release of 30 AgNPs, $\times 10^5$	Total Ag^+ release of 37 AgNPs, $\times 10^5$	Total Ag^+ release of 73 AgNPs, $\times 10^5$
1	14.9	2789.9	3.0	3.0	9.0	11.1	21.9
2	14.6	2662.5	2.9	5.9	17.6	21.7	42.8
3	14.2	2538.2	2.7	8.6	25.8	31.8	62.8
4	13.9	2416.8	2.6	11.2	33.6	41.4	81.7
5	13.5	2298.4	2.5	13.7	41.0	50.6	99.8
6	13.2	2182.9	2.3	16.0	48.1	59.3	116.9
7	12.8	2070.5	2.2	18.2	54.7	67.5	133.2
8	12.5	1961.0	2.1	20.4	61.1	75.3	148.6
9	12.1	1854.5	2.0	22.4	67.1	82.7	163.2
10	11.8	1750.9	1.9	24.2	72.7	89.7	176.9
11	11.5	1650.4	1.8	26.0	78.0	96.2	189.9
12	11.1	1552.8	1.7	27.7	83.0	102.4	202.1

103 **Table SI 4.** Estimated number of released Ag⁺ from every surface layer of AgNPs.

104 Notes: Average diameter of 29.8 nm is used for spherical AgNPs, giving an original 105 radius of 14.9 nm. Van der Waals radius of Ag atom is known to be 172 pm, yielding an area of AgNPs cross-section of $S_{Ag} = \pi r^2 = 0.09294 \text{ nm}^2$. 106

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108 Estimation of Ag⁺ release from each AgNP

109 The rough number of Ag atoms on the surface of every single AgNP is estimated by the 110 surface area of AgNPs dividing by the cross section area of a single Ag atom, as listed in table SI 4. Assuming Ag⁺ ions are released from the surface of AgNPs layer by layer, we 111 112 can estimate the Ag^+ release capability of different layer of every single AgNP.

In the case of 1 µM AgNPs interacting with 1x107 CFU/mL Salmonella, 30 and 37 113 114 AgNPs out of 73 AgNPs have interaction or bind to every Salmonella cell in absence and presence of tetracycline, respectively. And released Ag⁺ concentrations are 121 and 182 115 nM, indicating Ag⁺ ions to *Salmonella* cell ratios are 7.28×10^6 : 1 and 10.96×10^6 : 1 in 116 absence and presence of tetracycline, respectively. There is an increase of $3.68 \times 10^6 \text{ Ag}^+$ 117 release triggered by a single Salmonella cell. Compared with total Ag⁺ release of 37 118 119 AgNPs listed in table SI 4, combination of tetracycline and Salmonella cells facilitate the 120 release of about 3 layers of Ag on the surface of AgNPs.

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