Biophysical Journal, Volume 112

Supplemental Information

Cytoplasmic RNA-Protein Particles Exhibit Non-Gaussian Subdiffusive

Behavior

Thomas J. Lampo, Stella Stylianidou, Mikael P. Backlund, Paul A. Wiggins, and Andrew J. Spakowitz

Cytoplasmic RNA-Protein Particles Exhibit Non-Gaussian Subdiffusive Behavior: Supplemental Information

Thomas J. Lampo¹, Stella Stylianidou², Mikael P. Backlund³, W.

E. Moerner³, Paul A. Wiggins^{2,4,5}, and Andrew J. Spakowitz^{1,6,7,8}

1. Department of Chemical Engineering, Stanford University, Stanford CA 94305

2. Department of Physics, University of Washington, Seattle WA 98195

3. Department of Chemistry, Stanford University, Stanford CA 94305

4. Department of Bioengineering, Washington University, Seattle WA 98195

5. Department of Microbiology, Washington University, Seattle WA 98195

6. Department of Applied Physics, Stanford University, Stanford CA 94305
7. Department of Materials Science, Stanford University, Stanford CA 94305

8. Biophysics Program, Stanford University, Stanford CA 94305

- [1] S. Stylianidou, N. J. Kuwada, and P. A. Wiggins, Biophysical journal 107, 2684 (2014).
- [2] M. A. Thompson, J. M. Casolari, M. Badieirostami, P. O. Brown, and W. Moerner, Proceedings of the National Academy of Sciences 107, 17864 (2010).
- [3] S. C. Weber, M. A. Thompson, W. E. Moerner, A. J. Spakowitz, and J. A. Theriot, Biophysical Journal 102, 2443 (2012).



FIG. 1: Representative trajectories of RNA-protein particles. (ab) Sample trajectories of RNA-protein particles diffusing in the *E. coli* cytoplasm for (a) 1 s and (b) 1 min intervals between position measurements. (c) Sample trajectories placed arbitrarily in a circle with a diameter of 3 μm (typical size of an *S. cerevisiae* cell).



FIG. 2: **RNA-protein particles exhibit viscoelastic behavior.** (a-d) Velocity autocorrelation function of RNA-protein particles in *E. coli* for particle position measurements taken at one second intervals (a,b) and one minute intervals (c,d) [1] and corrected for drift due to cell growth. (e,f) Velocity autocorrelation function of RNA-protein particles in *S. cerevisiae* [2]. The velocity autocorrelation function is defined as $C_v^{(\delta)}(\Delta t) = \langle v_{\delta}(\Delta t) \cdot v_{\delta}(0) \rangle$ where the velocity $v_{\delta}(t) = [x(t+\delta) - x(t)]/\delta$ for the discrete time interval δ . (b), (d), and (f) are the same as (a), (c), and (e) respectively but with a differently scaled x-axis to show that the negative correlation peak occurs at $\Delta t/\delta = 1$. The black lines in (b), (d), and (f) are theoretical predictions for fractional Brownian motion with the measured values of α from the eMSD inserted into the equation $[(\eta+1)^{\alpha}+|\eta-1|^{\alpha}-2\eta^{\alpha}]/2$, where $\eta = \Delta t/\delta$ [3].



FIG. 3: **The effect of track length on the diffusivity distribution.** The distribution of diffusivities normalized by their mean for 10,000 fractional Brownian motion simulations of varying track lengths. Diffusivities are calculated using a power law fit to the first 15 points of each trajectory's time-averaged MSD. Distributions are plotted using a kernel density estimation with a Gaussian kernel and width determined by Silverman's rule of thumb.



FIG. 4: The relation between probve diffusivity and fluorescence intensity. Particle diffusivities as a function of their mean intensity for the *e. coli* 1 s (a), *E. coli* 1 min (b), and the *S. cerevisiae* data (c). The black dashed line represents a best fit of the model y = A/x with fit parameter A. We show R^2 and Pearson correlation coefficient (ρ) for each model fit to the data on a loglog scale.



FIG. 5: The effect of finite track lengths on the diffusivity autocorrelation function. Diffusivity autocorrelation function for fractional Brownian motion simulations that have the same data sparsity and 10x the size of the *E. coli* 1 s data set (a) and the *S. cerevisiae* (b).