

# Supporting Information

## Discovery and optimization of a selective ligand for the Switch/Sucrose Non-Fermenting-related bromodomains of Polybromo protein-1 by the use of virtual screening and hydration analysis.

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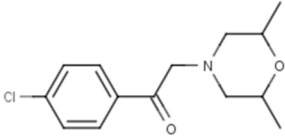
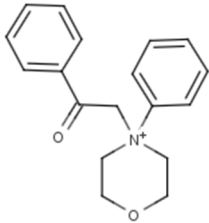
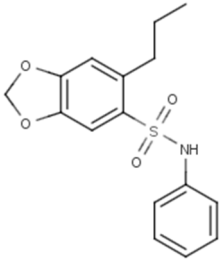
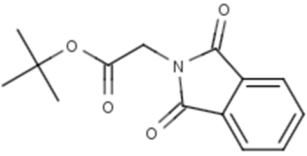
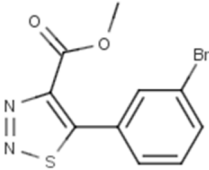
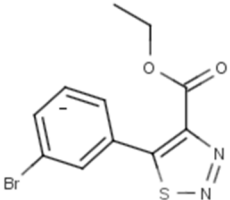
Table S6. ITC data for the studied compounds.

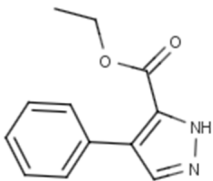
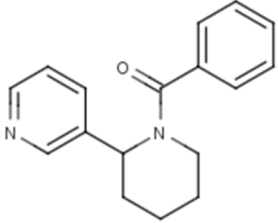
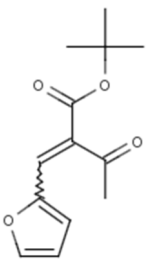
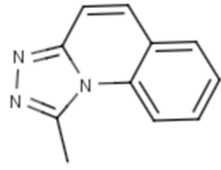
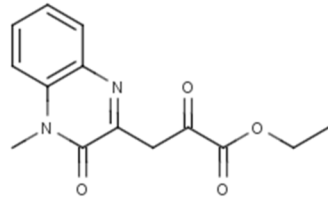
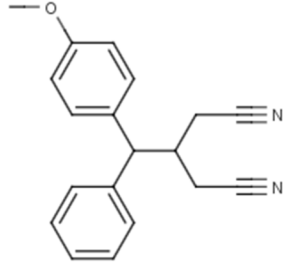
Figure S1. The network of hydrogen bonds anchoring the buried water molecule in the **9**-PB1(5) complex.

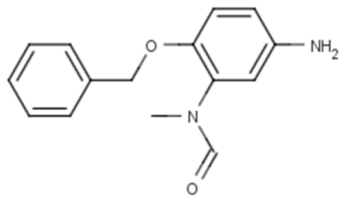
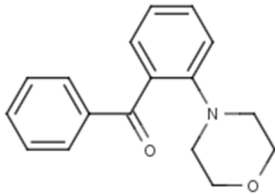
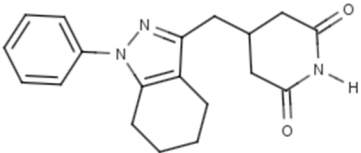
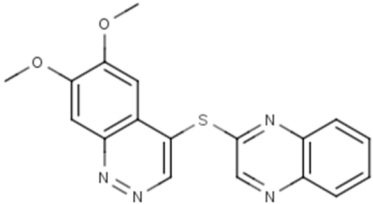
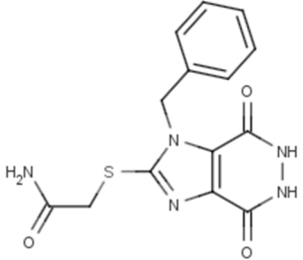
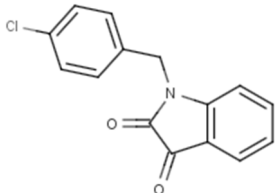
Figure S2. 1BR-TERT cells viability assay of **12**.

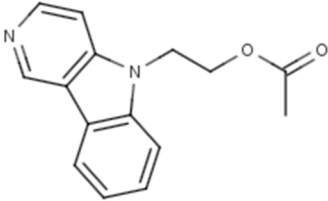
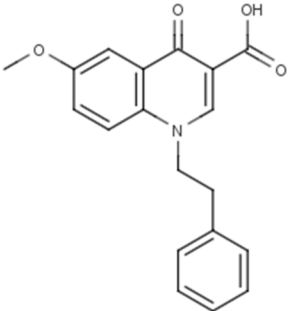
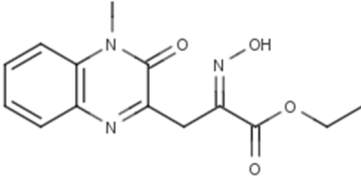
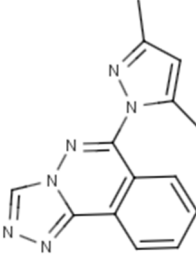
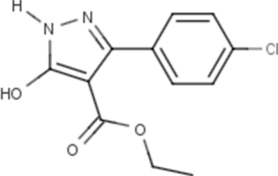
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MOLPRINT2D	Daylight	no scaling
Radial	Carhart	square root feature
Radial	Carhart	square root unity
Radial	Daylight	no scaling
Radial	Daylight	square root feature
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Radial	Hybrid	square root feature
Radial	Hybrid	square root feature
Radial	Mol2	no scaling
Radial	Mol2	square root feature

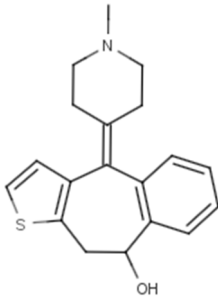
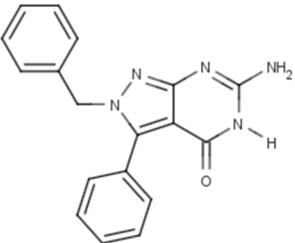
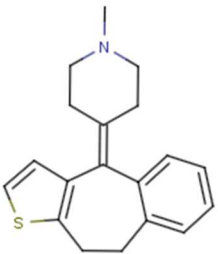
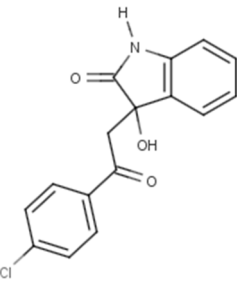
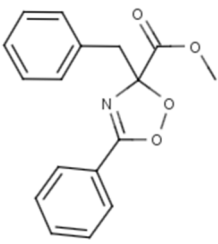
**Table S1.** The 11 most efficient FP combinations of Canvas used subsequently for screening. These FP screens afforded high enrichment quantified as the enrichment factor at 20% of screened library ( $EF_{20\%}$ ). The 11 FP combinations affording high recovery of known actives showed an  $EF_{20\%}$  greater than 2.25 while the remaining combinations resulted to lower  $EF_{20\%}$  ranging from 0 to 1.25. The  $EF_{20\%}$  ranges from 0 (no enrichment) to 5 (maximum enrichment).

Compound	Structure	$T_m$ (°C)	
		PB1(5)	BRD4(2)
NSC32894		-1.9	-2.0
NSC35427		-2.3	-1.8
NSC38116		0.0	0.0
NSC45837 (3)		0.0	0.8
NSC48742 (4)		-0.2	<b>2.0</b>
NSC48745		-2.4	-0.5

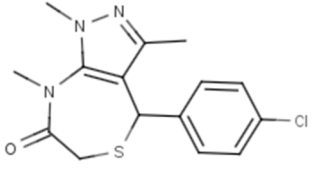
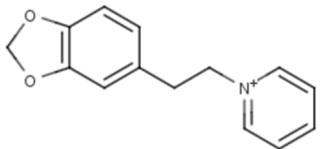
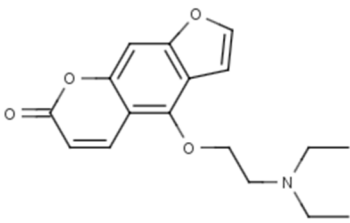
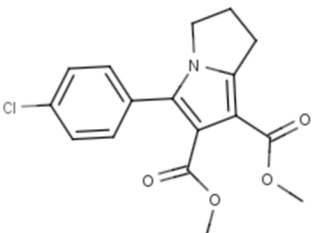
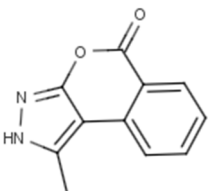
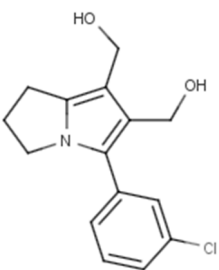
NSC55495 (5)		-0.2	<b>1.6</b>
NSC55917 (6)		0.6	<b>1.8</b>
NSC75086		-0.4	-0.6
NSC76484 (8)		0.1	<b>3.2</b>
NSC86927		-1.5	0.4
NSC97717 (7)		-0.4	<b>1.5</b>

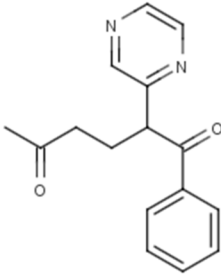
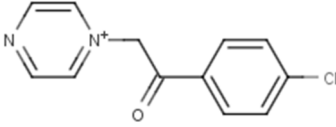
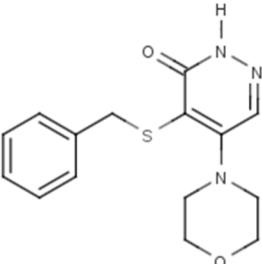
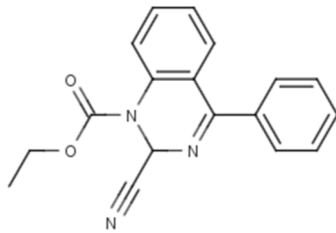
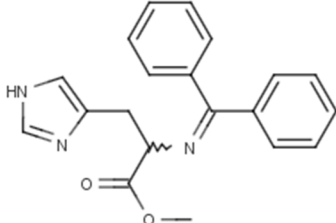
NSC99768		-2.8	0.3
NSC111167		-0.4	1.1
NSC112143		-0.1	0.4
NSC114077		0.0	0.0
NSC114088		-0.3	0.1
NSC127077		-1.4	-4.4

NSC164955		-0.9	0.3
NSC211010		-0.3	0.9
NSC251944		-0.8	-0.2
NSC270404		1.0	0.5
NSC270420		0.0	-0.4

NSC282480		0.2	0.4
NSC282740		-0.3	0.3
NSC291564		-1.2	-1.4
NSC297830		-0.3	-0.9
NSC303574		0.2	-1.0



NSC313672		-1.1	-0.3
NSC340031		0.0	-0.2
NSC341960		-1.1	0.4
NSC356242		0.2	0.2
NSC356476 (9)		<b>4.1</b>	<b>1.9</b>
NSC358365		0.5	-0.5

NSC400371		0.1	0.2
NSC400609		-1.4	-1.5
NSC409712		-5.1	-3.7
NSC607601		0.0	-0.2
NSC612976		-1.1	-0.6
NSC669640		0.1	-0.5

**Table S2.** The 40 compounds from NCI/DTP that were evaluated in the DSF screen and their  $T_m$  values towards PB1(5) and BRD4(2). The compounds were screened at 200 $\mu$ M while the protein was at 20 $\mu$ M. Shown are the compounds, their NSC code numbers and the respective  $T_m$  values. Stabilization values exceeding 1.5 °C are marked in bold. Compound **9** showed significant affinity for PB1(5) and was thus subjected to co-crystallization and X-ray crystallography to determine its binding mode to PB1(5) and suggest possible routes for optimization.

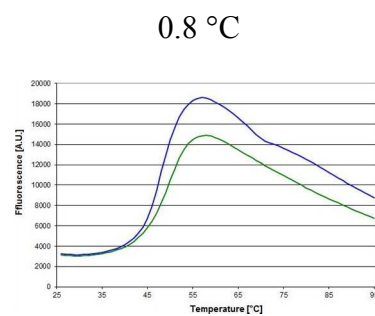
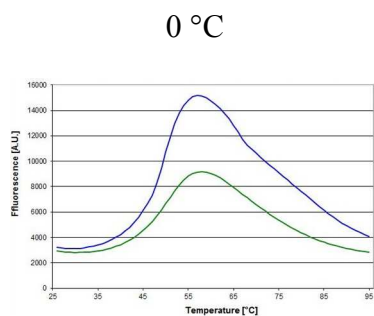
Compound

$T_m$  PB1(5)

$T_m$  BRD4(2)

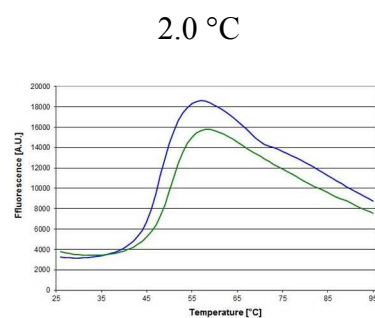
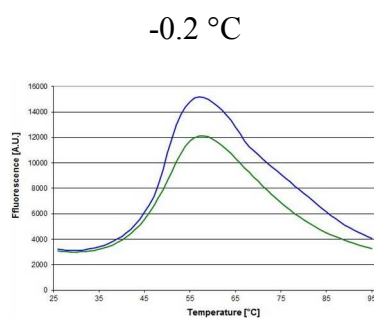
3 (NSC45837)

C, R



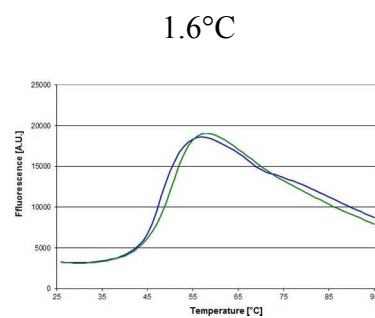
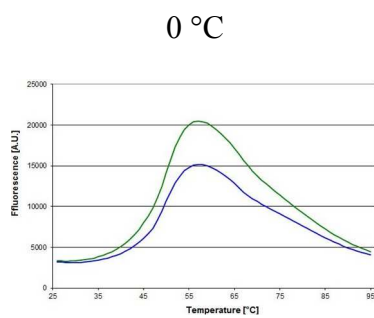
4 (NSC48742)

C, R



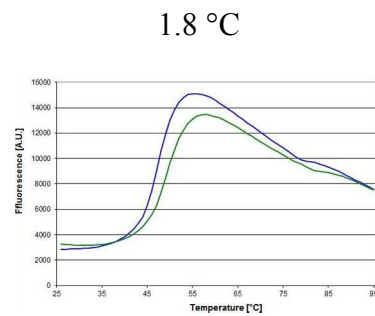
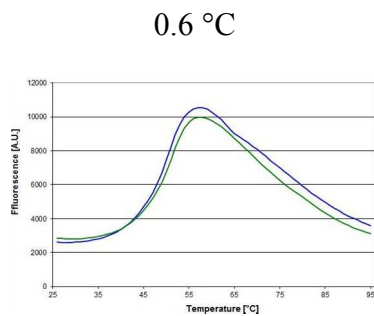
5 (NSC55495)

G



6 (NSC55917)

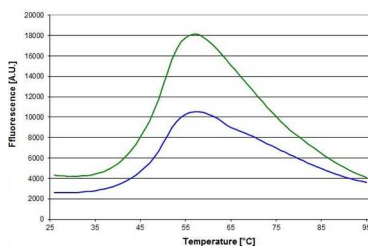
G



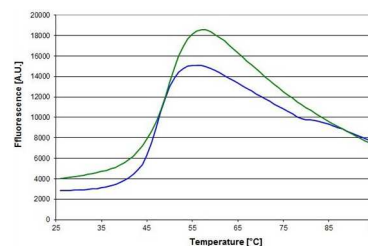
7 (NSC97717)

G

-0.4 °C



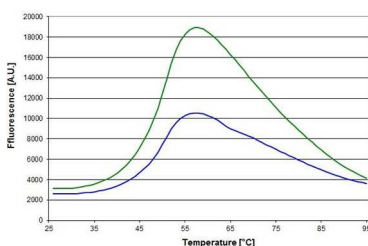
1.5 °C



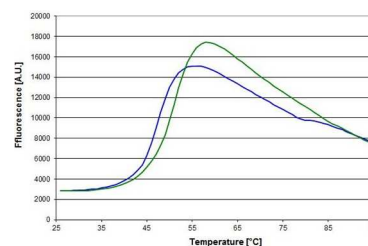
8 (NSC76484)

C, R

0.1 °C



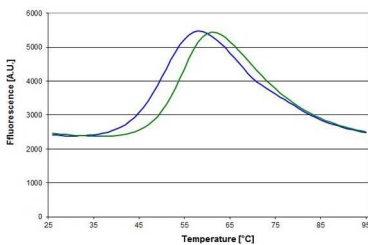
3.2 °C



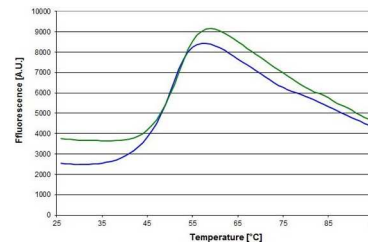
9 (NSC356476)

G

4.1 °C



1.9 °C



**Table S3.** The  $T_m$  curves of the seven compounds showing measurable binding affinity in the DSF screen. The compounds were screened at 200 $\mu$ M while the protein was at 20 $\mu$ M. Shown are the NSC number of the compounds and the  $T_m$  in  $^{\circ}$ C observed for PB1(5) and BRD4(2). In the panel BRD2(2), BRD3(2), BRD4(1) and BRDT were included in addition to BRD4(2) and PB1(5) but no binding was observed for those proteins. The screening methodology affording each compound is shown below the NSC code; C: Canvas; G: Glide; R: ROCS.

PDB ID	5III	5HRV	5HRW
Protein	PB1(5)	PB1(5)	PB1(5)
Compound	<b>9</b>	<b>10</b>	<b>11</b>
Space group	P2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>	C 2 2 2 <sub>1</sub>	P 2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>
Cell dimensions:			
a, b, c (Å)	41.11 57.97 106.01	57.82 140.3 41.55	41.11 59.10 105.50
α, β, γ (deg)	90.00 90.00 90.00	90.00 90.00 90.00	90.00 90.00 90.00
Resolution* (Å)	24.10 (2.02)	28.91 (1.70)	29.55 (1.80)
Unique observations*	17373 (2488)	19011 (883)	23971 (1312)
Completeness* (%)	99.8 (100.0)	99.3 (89.7)	97.4 (93.2)
Redundancy*	4.3 (4.3)	7.8 (4.2)	5.0 (4.8)
Rmerge*	0.070 (0.671)	0.043 (0.360)	0.053 (0.735)
I/σI*	12.5 (2.0)	28.2 (3.9)	18.5 (2.2)
<b>Refinement</b>			
Resolution (Å)	2.02	1.70	1.80
R <sub>work</sub> / R <sub>free</sub> (%)	18.8/26.5	17.78 / 20.76	20.21 / 23.57
Number of atoms			
(protein/other/water)	1826/30/113	970 / 20 / 99	1796 / 59 / 81
B-factors (Å <sup>2</sup> )			
(protein/other/water)	36.24/25.55/39.24	26.39/22.55/34.40	29.69/33.56/33.37
r.m.s.d bonds (Å)	0.015	0.017	0.019
r.m.s.d angles (°)	1.381	1.554	1.931
Ramachandran			
Favoured (%)	98.62	100.00	99.53
Allowed (%)	1.38	0.00	0.00
Disallowed (%)	0.00	0.00	0.47

\* Values in parentheses correspond to the highest resolution shell.

PDB ID	5HRX	5II2	5IID
Protein	PB1(5)	PB1(5)	PB1(5)
Compound	<b>12</b>	<b>15</b>	<b>14</b>
Space group	P 2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>	P2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>	P2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>
Cell dimensions:			
a, b, c (Å)	41.18 58.42 105.80	41.52 58.51 138.23	41.50 56.40 139.48
$\alpha$ , $\beta$ , $\gamma$ (deg)	90.00 90.00 90.00	90.00 90.00 90.00	90.00 90.00 90.00
Resolution* (Å)	29.21 (1.73)	25.00 (2.21-2.10)	19.39 (2.53-2.40)
Unique observations*	26823 (1398)	127884 (16300)	62035 (6533)
Completeness* (%)	97.8 (96.3)	99.0 (95.3)	99.3 (97.0)
Redundancy*	5.6 (5.0)	6.3 (5.9)	4.6 (3.5)
Rmerge*	0.051 (0.893)	0.128 (0.368)	0.104 (0.784)
I/ $\sigma$ I*	17.3 (1.8)	9.1 (4.0)	10.4 (1.5)
<b>Refinement</b>			
Resolution (Å)	1.73	2.10	2.40
R <sub>work</sub> / R <sub>free</sub> (%)	21.38 / 25.37	17.6/22.7	22.1/28.1
Number of atoms			
(protein/other/water)	1810/44/75	1878/57/82	1894/40/49
B-factors (Å <sup>2</sup> )			
(protein/other/water)	32.71/29.78/36.95	28.13/22.88/33.88	45.22/45.64/41.39
r.m.s.d bonds (Å)	0.019	0.016	0.014
r.m.s.d angles (°)	1.888	1.623	1.471
Ramachandran			
Favoured (%)	99.53	98.65	99.12
Allowed (%)	0.00	1.35	0.88
Disallowed (%)	0.47	0.00	0.00

\* Values in parentheses correspond to the highest resolution shell.

**Table S4.** Data collection and refinement statistics.

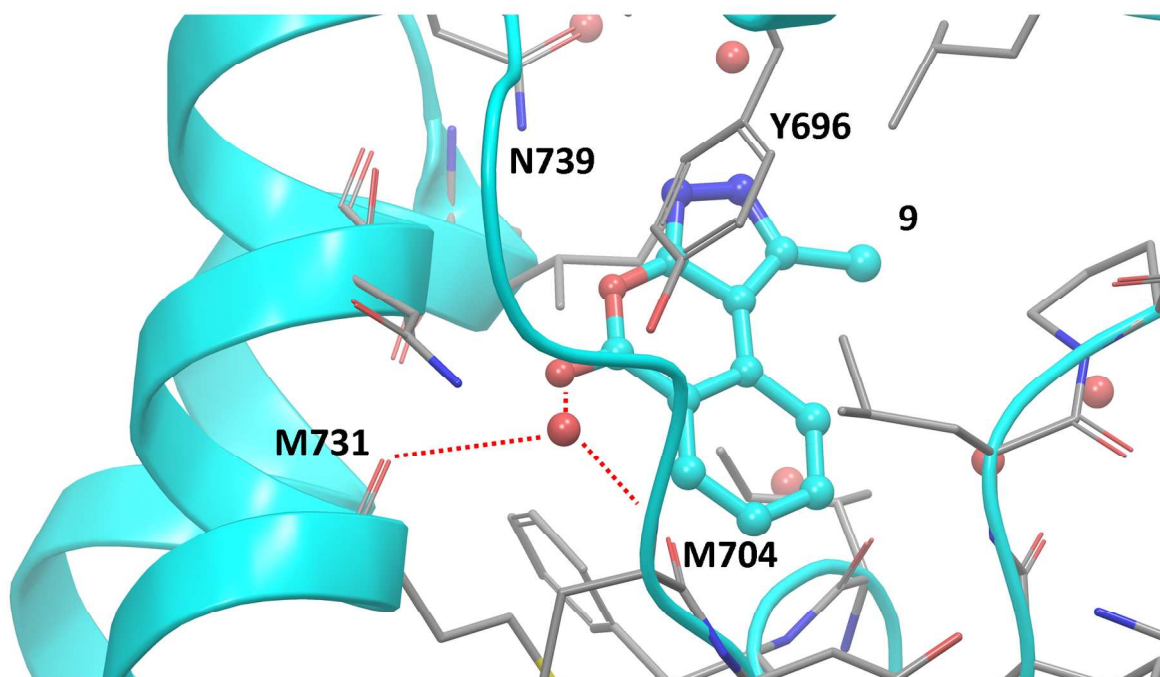
Protein	<b>9</b>	<b>10</b>	<b>11</b>	<b>12</b>	<b>2</b>	<b>1</b>
PB1(1)	0.3 ± 0.8	0.7 ± 0.4	0.4 ± 0.4	0.8 ± 0.9	-1.4 ± 0.5	0.6 ± 0.6
PB1(2)	-0.4 ± 0.4	0.2 ± 0.1	0.1 ± 0.2	0.3 ± 0.1	0.9 ± 0.1	-0.7 ± 0.1
PB1(3)	0 ± 0.2	0.6 ± 0.2	0.4 ± 0.3	1.1 ± 0.2	1.3 ± 0	1.4 ± 0.6
PB1(4)	0.1 ± 0.3	0.3 ± 0.2	0.1 ± 0.3	0.2 ± 0.1	0.1 ± 0	-0.6 ± 0.5
PB1(5)	0.7 ± 0.3	2.8 ± 0.2	1.1 ± 0.2	2.2 ± 0.3	9.1 ± 0.1	-0.5 ± 0.8
PB1(6)	-0.1 ± 0.2	0 ± 0.2	-0.2 ± 0.4	-0.1 ± 0.2	-0.1 ± 0	-1.6 ± 0.3
SMARCA 2A	0.4 ± 0.1	0.5 ± 0.3	0.1 ± 0.2	0.7 ± 0.2	6.3 ± 0.1	0.3 ± 0.2
SMARCA 2B	-0.1 ± 0.3	0.3 ± 0.2	0.1 ± 0.2	0.5 ± 0.3	4.4 ± 0.2	0.6 ± 0.2
SMARCA 4A	0.5 ± 0.4	0.3 ± 0.2	-0.2 ± 0.3	0.5 ± 0.3	5.6 ± 0.1	-0.6 ± 0.2
TAF1(1)	-0.6	0.3	0.1	0	0 ± 0	-0.15 ± 0.1
TAF1(2)	0.03	0.3	-0.3	0.1	-0.1 ± 0.3	0.25 ± 0.1
TAF1(1:2)	0.2	-0.7	-0.6	-1.5	-0.8 ± 0.1	-1.3 ± 0
TAF1L(1)	-0.7	0.3	0.3	-0.6	0 ± 0.1	-0.2 ± 0.4
TAF1L(2)	0.4	0.5	-0.1	-1	-0.3 ± 0.5	-0.1 ± 0.1
TAF1L(1:2)	0.4	-0.8	-0.8	-0.8	-1.2 ± 0.4	-2.4 ± 0.3
BRD1	0.1	-0.3	-0.9	-0.6	0.1 ± 0.1	-0.1 ± 0.1
BRD2(1)	0	-0.8	-1	-0.9	-0.7 ± 0.2	7.1 ± 0.6
BRD4(1)	0	-0.5	-0.7	-0.4	-0.6 ± 0	8.8 ± 0
BRD4(2)	-0.1	-0.3	-0.5	0.1	0.2 ± 0	10.1 ± 0.1

**Table S5.** DSF assay summary.  $\Delta T_m$  (°C) values are shown. The compounds were screened at 100 $\mu$ M while the protein was at 2.5 $\mu$ M. Compounds **9-12** were measured in triplicate on PB1 and SMARCA. **1** and **2** were measured in duplicate.

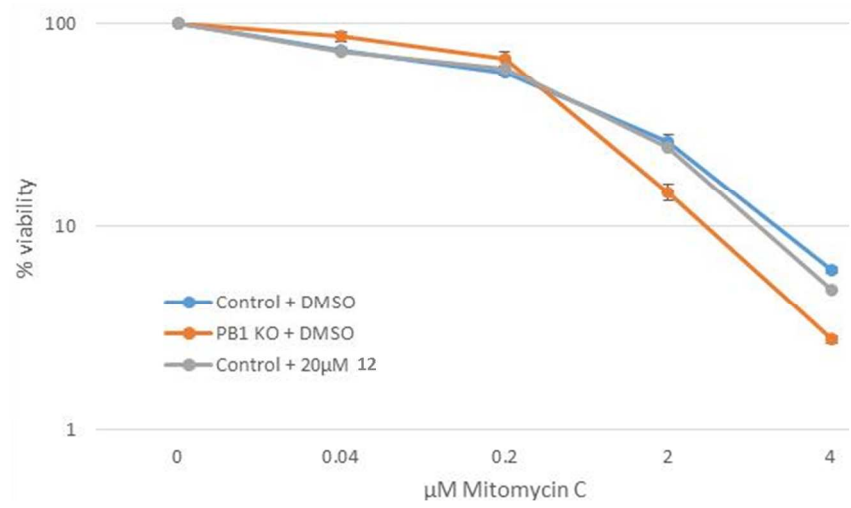
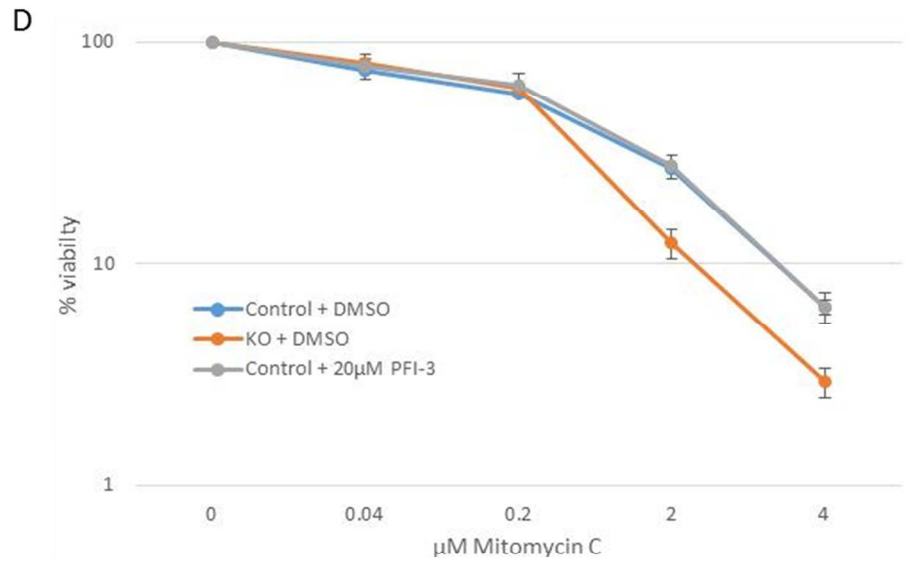
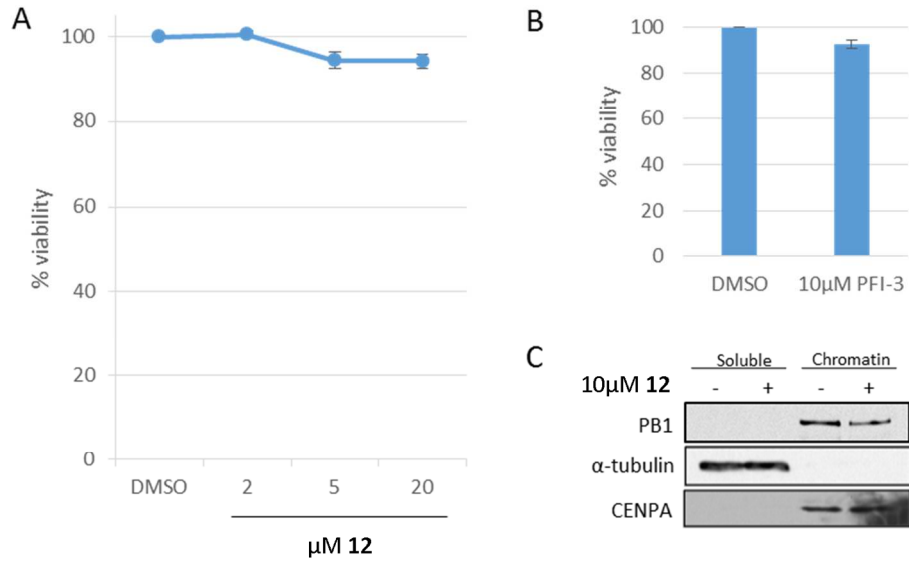


Protein – Compound	$K_d$ ( $\mu\text{M}$ )	n	$\Delta G$ (kcal/mol)	$\Delta H$ (kcal/mol)	$-T\Delta S$ (kcal/mol)
PB1(5) – 2	0.042	1.2	-9.7	-7.9	-1.8
PB1(5) – 9	11.5	1.0	-6.5	-5.6	-0.9
PB1(5) – 10	3.4	1.2	-7.2	-6.7	-0.6
PB1(5) – 11	3.3	0.8	-7.2	-2.9	-4.3
PB1(5) – 12	5.1	1.1	-7.0	-6.4	-0.6
PB1(3) – 12	No binding				
SMARCA2(2A) – 12	No binding				
BRD4(1) – 12	No binding				

**Table S6.** Isothermal titration calorimetry summary. Protein (200 $\mu\text{M}$ ) was titrated into compound solution (15 $\mu\text{M}$ ) at 15°C.



**Figure S1.** The buried water molecule that is not displaced upon binding of **9** in PB1(5) is stabilized by an extended network of H-bonds through surrounding residues such as the sidechain of Y696, the backbone carbonyls of M704 and M731 as well as the carbonyl of **9**.



**Figure S2.** Compound **12** slightly reduces viability of 1BR-hTERT human fibroblast cells. (A) Viability curve for 1BR-hTERT cells treated with DMSO or **12**. (B) Viability graph for cells as in (A) treated with DMSO or **2**. (C) Cells treated with DMSO or **12** were processed for chromatin fractionation as outlined in the experimental procedures. Samples were blotted for PB1,  $\alpha$ -tubulin or CENPA as indicated. (D) Viability curve for U2OS PB1 knock-out cells treated with **2** (upper graph) or **12** (lower graph) in presence of mitomycin C (MMC).