

The intergenic region of the maize defensin-like protein genes *Def1* and *Def2* functions as a embryo-specific asymmetric bidirectional promoter

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Supplementary Files

The following supplementary material is available at *JXB* online.

Fig. S1 Comparative analysis of the tissue specificity of the polar and bidirectional promoters in transgenic maize plants. Root, stem, and leaf tissues of trefoil-stage transgenic maize plants were used for GUS histochemical staining. Red arrow indicates trace GUS expression.

Fig. S2 Comparative analysis of the tissue specificity of the polar and bidirectional promoters in transgenic maize plants. Root, stem, leaf, and sheath tissues of large, trumpet-stage transgenic maize plants and tassels, silk, and pollen of transgenic maize plants were used for GUS histochemical staining. Red arrow indicates trace GUS expression.

Fig. S3 Comparative analysis of the expression patterns of the polar and bidirectional promoters during early developmental stages of transgenic maize plants. (A) Analysis of promoter tissue specificity using GUS histochemical staining in transgenic maize. (B) Analysis of the tissue specificity of *GFP::P_{zmDef1}::GUS* in transgenic maize. Red arrow indicates GUS staining sites in the aleurone layer and in embryos; Control tissues corresponding to the various developmental stages are shown on the right side of each photograph. DAP, days after pollination; bars = 1 mm.

Fig. S4 Analysis of bidirectional promoter tissue specificity in transgenic maize.

Fig. S5 Characterization of the tissue specificity and expression strength of the *P_{Zmdef2}* in transgenic maize plants. (A) Qualitative analysis of bidirectional expression activities of *P_{Zmdef2}* in transgenic maize seeds; Bright-I and GFP-I, intact seeds imaged in bright field mode and using a GFP filter set, respectively; Bright-L, GFP-L, and GUS-L, longitudinal sections imaged in bright field mode, using a GFP filter set, and in bright field mode after histochemical staining, respectively. Control tissues

corresponding to the various developmental stages are shown on the right side of each photograph; bars = 1 mm. (B) Quantification of GUS activities level during various developmental stages of seeds from transgenic maize plants. En, endosperm; Em, embryo.

Fig. S6 The activity of different *GFP::mP_{ZmBD1}::GUS* constructs assayed using maize immature embryo transient transformation. (A) to (W) represent the 23 constructs pbd1GG3 to pbd23GG3, respectively. (X), negative control. Bars = 0.5 mm.

Table S1. Promoter sequences and cloning primers used in this work.

Figure S1

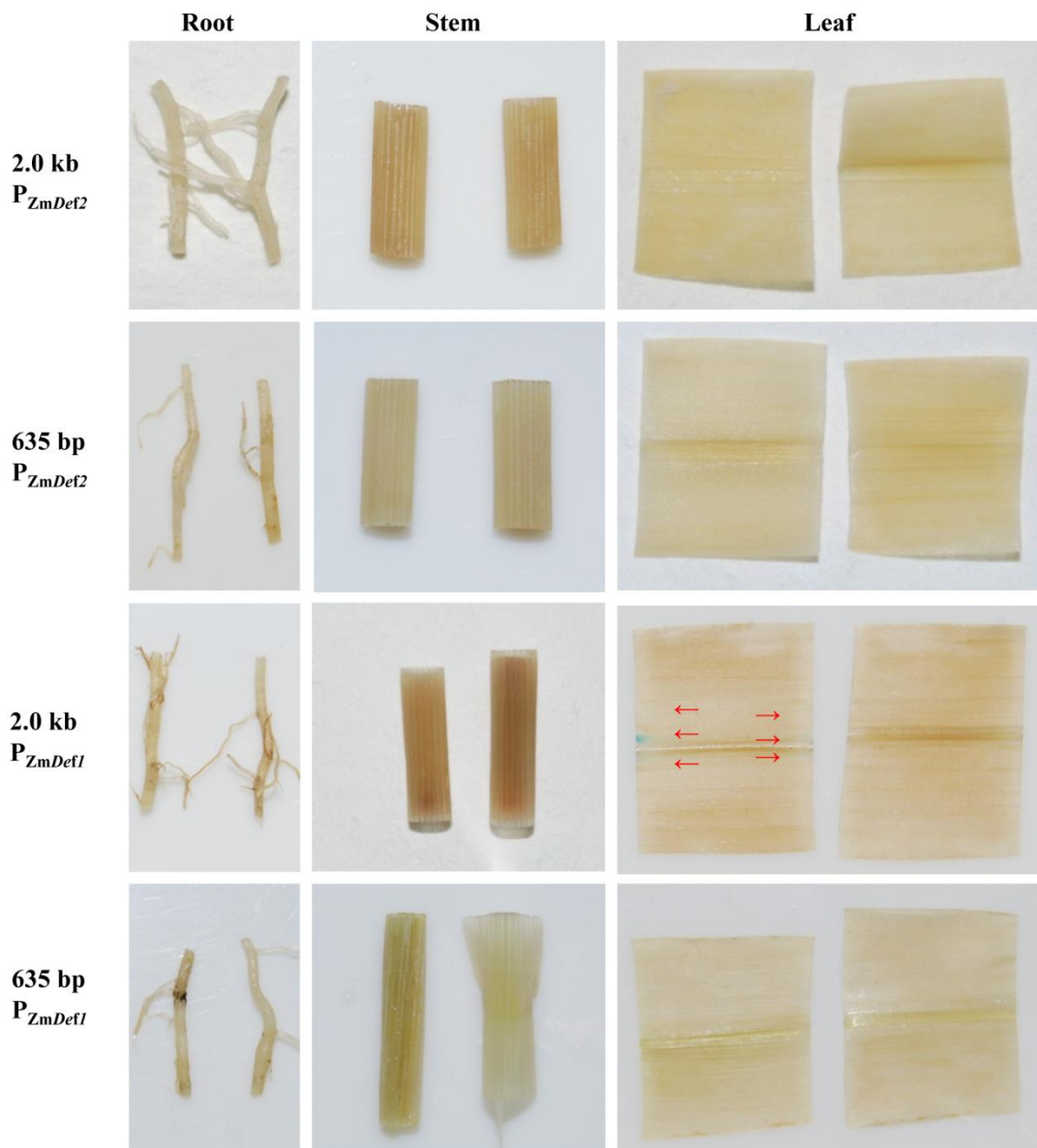


Figure S2

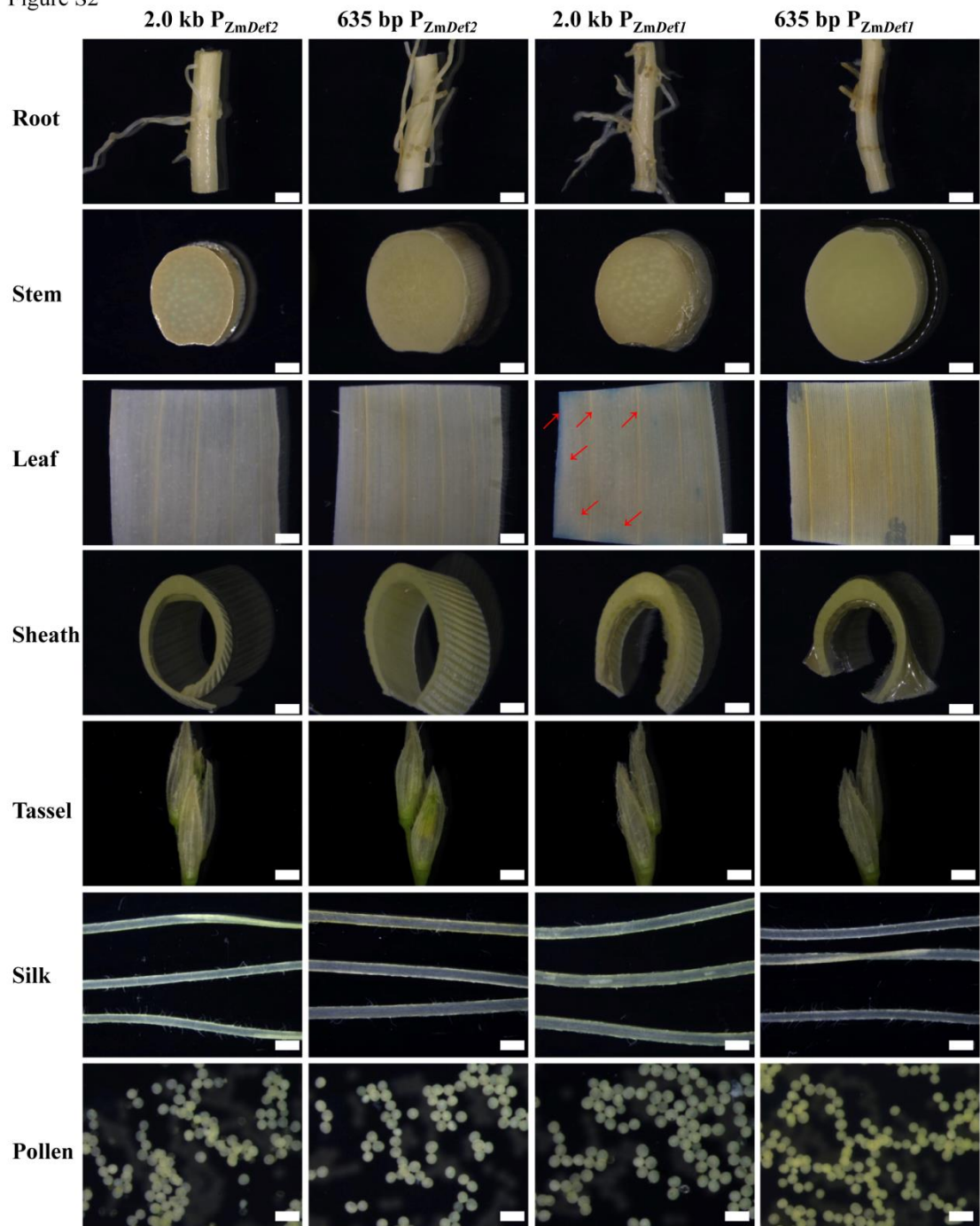


Figure S3

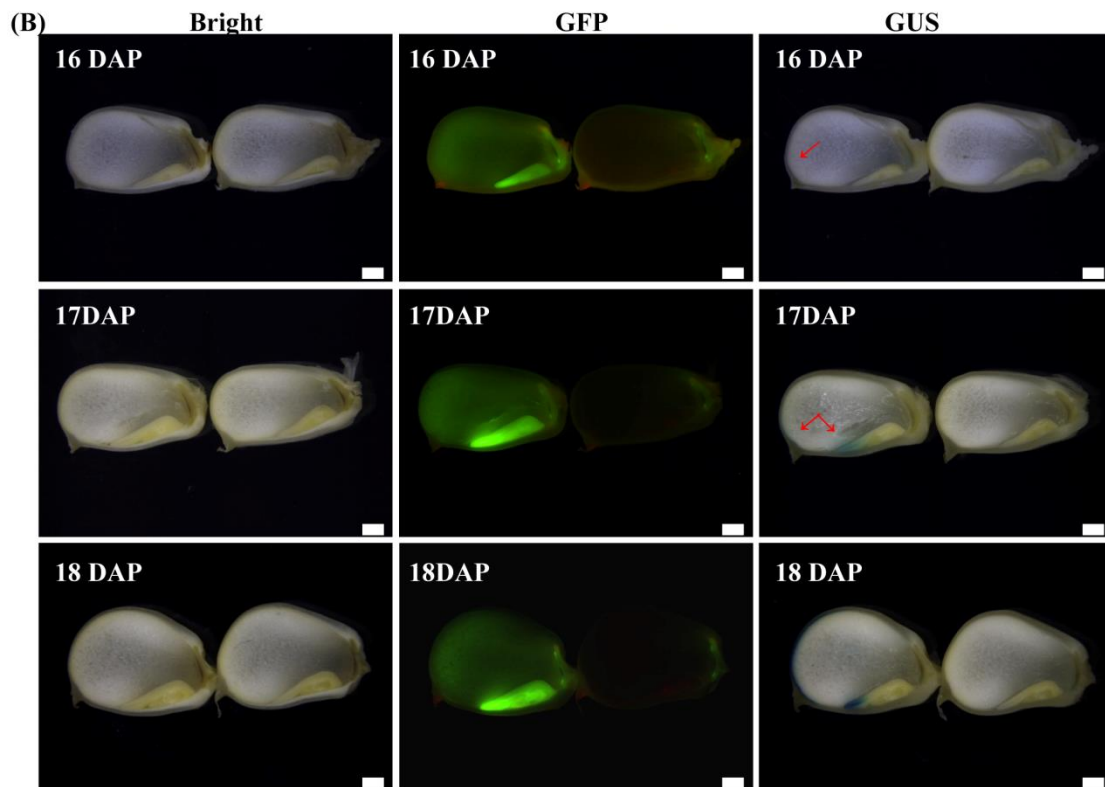
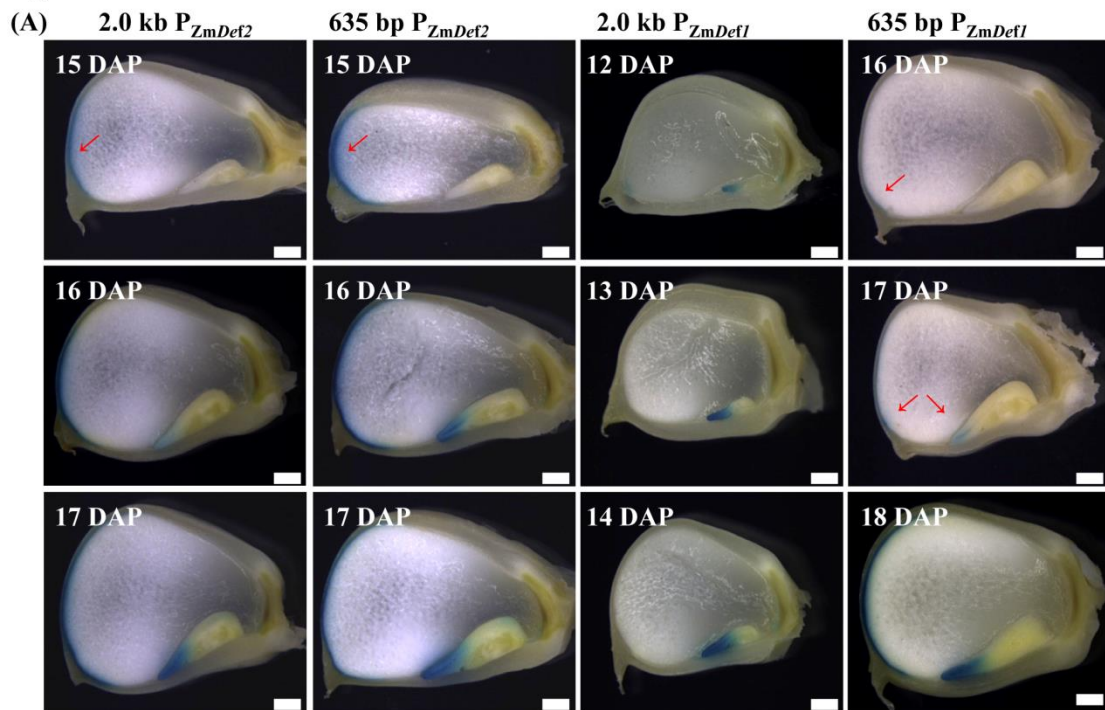


Figure S4

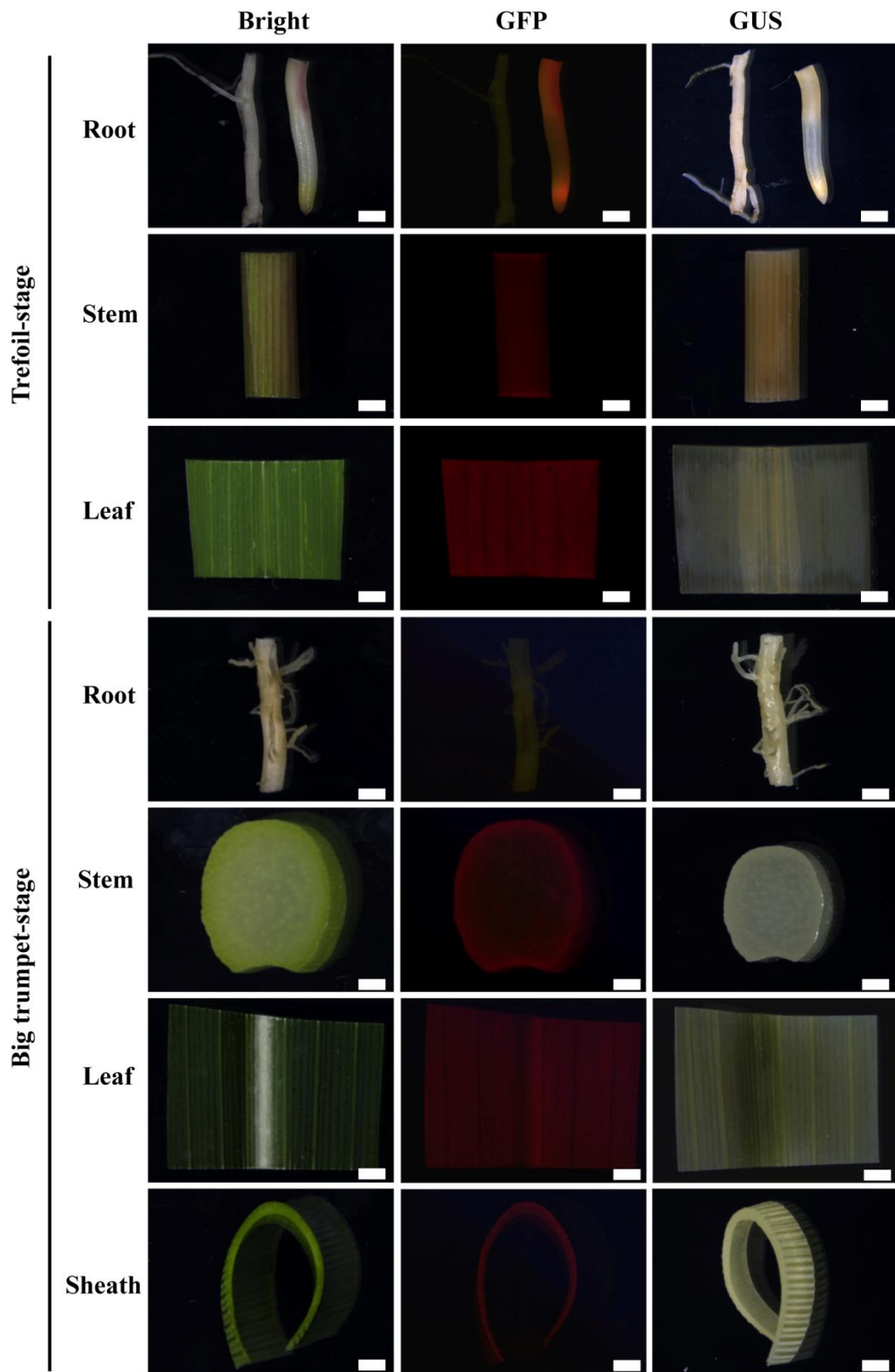
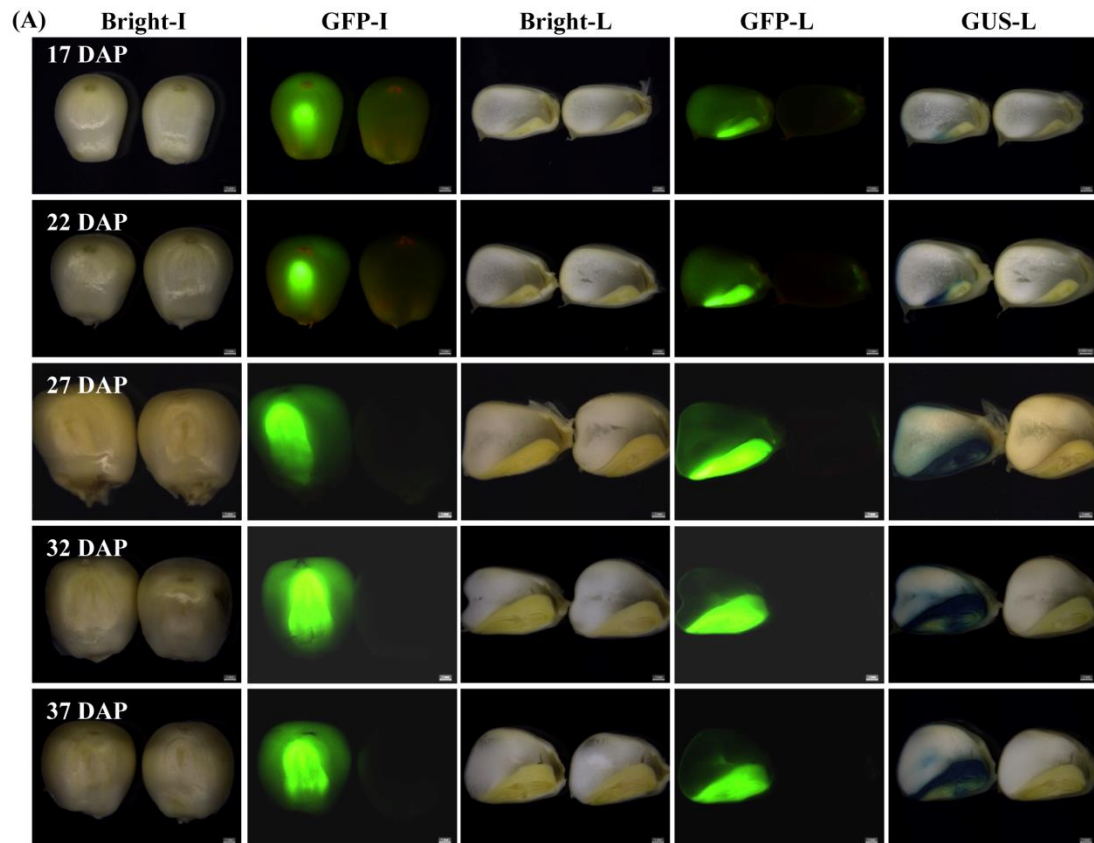


Figure S5



(B)

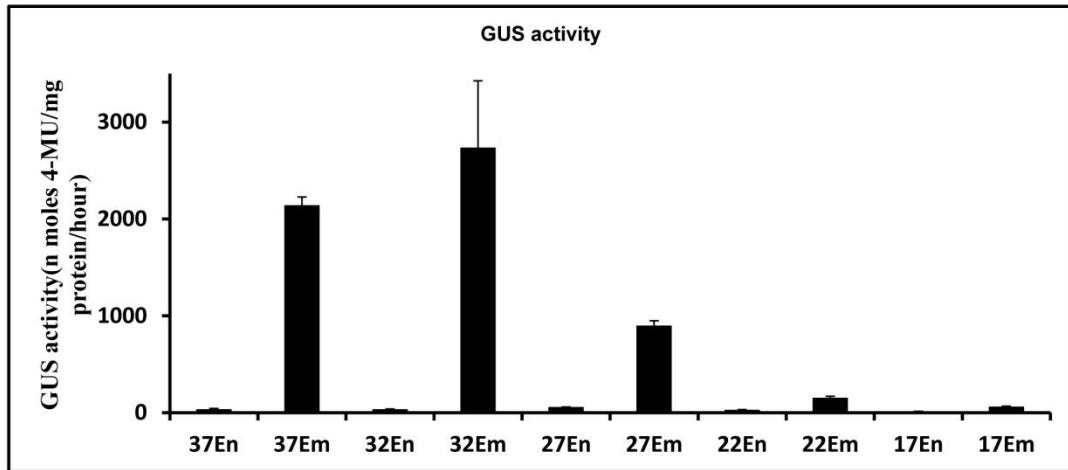


Figure S6

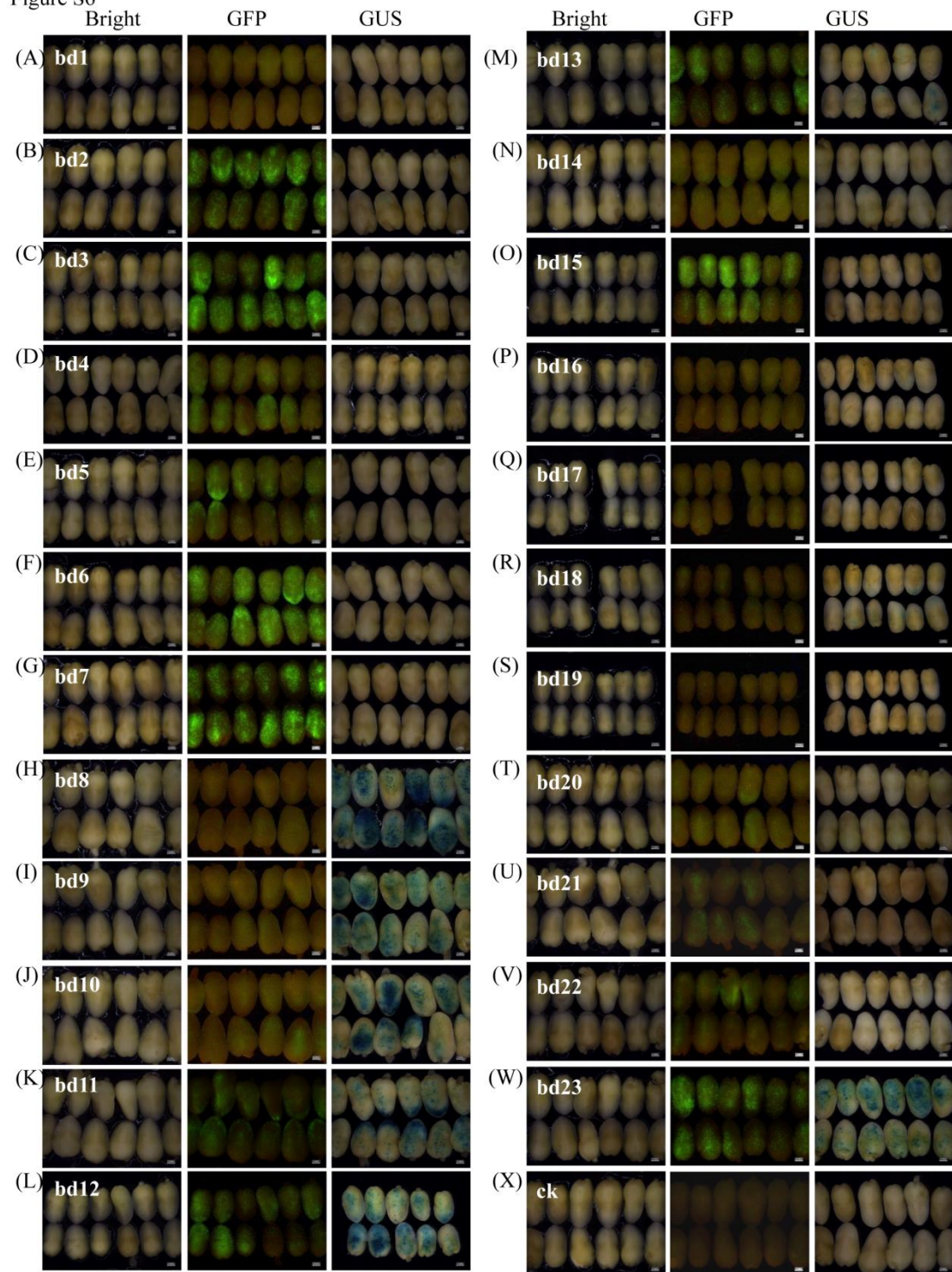


Table S1. Promoter sequences and cloning primers used in this work.

primers used for RT-PCR		
<i>Def1</i>	DEF1F	5-TGCTGCTCCTCATCGTCGTTGC-3
	DEF1R	5-TTGCCGCCGCCGTAGCCTTC-3
<i>Def2</i>	DEF2F	5-AGTCCAGGGCGACCGTGTG-3
	DEF2R	5-CGAGTGGTGCTGGCTCTTGC-3
<i>Actin</i>	AC200F	5-ATGTTTCCTGGGATTGCCGAT-3
	AC200R	5-CCAGTTTCGTCATACTCTCCCTTG-3
primers used for promoters clone		
2.0 kb P _{ZmDef1}	DEF1f	5-AAGCTTCGTCAGCAGTAGTTCTATTTTCAG-3
	DEF1r	5-ATTCCGGCCGGCTCTGCTGCTTGTGC-3
2.0 kb P _{ZmDef2}	DEF2f	5-AAAACTTTGGAGCCTAAACCTCTAAATTAAG-3
	DEF2r	5-CTGCAGCGCCGGCCGGCGTGGATCGGAT-3
635 bp P _{ZmBD1}	DEF1f1	5-aagcttATTCCGGCCGGCTCTGCTGCTTG-3
	DEF2r	5-CTGCAGCGCCGGCCGGCGTGGATCGGAT-3

Promoter sequences:

>635 bp P_{ZmBD1}

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